

# **Final Remedial Investigation Report**

**New York State Department of  
Environmental Conservation  
Investigation and Design  
Engineering Services  
Standby Contract No. D004437  
Work Assignment No. D004437-35**

Former Paul Miller Dry  
Cleaners Site  
(Site No: 243018)  
Port Richmond, New York

September 2012



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September 26, 2012

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PROJECT: NYSDEC Standby Contract No. D004437  
Work Assignment No.: D004437-35  
Former Paul Miller Dry Cleaners Site

SUBJECT: Final Remedial Investigation Report  
Former Paul Miller Dry Cleaners Site  
Port Richmond, New York  
Site ID No. 243018

Dear Mr. Sarnowicz:

Camp Dresser McKee & Smith (CDM Smith) is pleased to submit our Final Remedial Investigation Report for the above referenced work assignment. This report was prepared in accordance with DER-10 and Task 4 of this work assignment.

If you have any questions, or need additional information, please call me at 732-590-4674.

Very truly yours,

Seth Kellogg, PG  
Senior Project Manager  
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cc: Project File  
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# Section 1

## Introduction

This Remedial Investigation (RI) report for the former Paul Miller Dry Cleaners site, located in Port Richmond, Richmond County, New York (NY), herein referred to as “the Site,” was prepared by CDM for the New York State Department of Environmental Conservation (NYSDEC) under the Engineering Services for Investigation and Design, Standby Contract No. D004437.

This report details the results of the field investigation conducted in accordance with the *Remedial Investigation and Feasibility Study Work Plan* (RI/FS Work Plan) dated September 2009. This RI report was developed in accordance with the “State Superfund Standby Contract Work Assignment D004437-35, RI/FS, former Paul Miller Dry Cleaners, Site No. 243018.” The Work Plan and this RI follow the guidelines set forth in the *“Division of Environmental Remediation (DER)-10 Technical Guidance for Site Investigation and Remediation”* (NYSDEC, 2010).

### 1.1 Purpose of the Report

The purpose of this RI report is to present the results of the data gathered to help determine the most effective remedial alternatives to address groundwater contamination at the Site. The goal of the investigation was to define the nature and extent of site-related contamination in soil and groundwater in order to assess risks to ecological and human receptors.

The RI report was prepared in accordance with the following documents:

- New York State Department of Environmental Conservation (NYSDEC) *DER-10 Technical Guidance for Site Investigation and Remediation* (NYSDEC, 2010)
- Remedial Investigation and Feasibility Study Work Plan, Volume I, September 2009
- CDM Generic Quality Assurance Project Plan (QAPP), February 2008 and all associated standards operating procedures
- Health and Safety Plan, March 2011

### 1.2 Site Description and Background

#### 1.1.1 Site Description

Located in Port Richmond, NY, the Site occupies a 0.39 acre parcel in a commercial area, as depicted in Figure 1-1. The former dry cleaner building is currently being used as a fast food restaurant. The front (southern) portion of the building is slab on grade. The rear (northern) portion of the building has a basement, which is currently used for storage of extraneous equipment. The basement contains 4 rooms, which appear to have been boiler room, two storage rooms and a fur vault.

The Site is relatively flat with its entire surface area covered with concrete and/or asphalt. Review of the USGS *Arthur Kill, New York 7.5-Minute topographic map* (1981) indicates that ground surface is approximately 30 feet above mean sea level (amsl). The survey conducted as part of this RI confirmed

that the Site grades topographically from approximately 28 feet amsl in the south to approximately 25 feet amsl in the north-northwest. A tax map depicting the Site is available through the gis.nyc.gov website (Figure 1-2).

### 1.1.2 Operational History

Historical documents, including aerial photographs and city directories indicate that the site was undeveloped until approximately 1960. Paul Miller Dry Cleaners operated at the site from approximately 1960 through 1995. As of 2000, the site has been a fast food restaurant. Section 2.3 of the 2009 RI Report provided a detailed review of the site development and history and operational history(CDM 2009).

## 1.3 Previous Investigations

In 1994, the owner of the shopping center in which the Site is located conducted an environmental investigation at the Site. Subsequently, in May 2000, NYSDEC retained Lawler, Matusky & Skelly Engineers LLP (LMS) to conduct an Immediate Investigation Work Assignment (IIWA) of the Site. The objectives of the IIWA were to determine groundwater flow in the vicinity of the site, identify the nature and extent of groundwater contamination as related to historic Site activities, and to assess whether the site poses a threat to public health or the environment.

### 1.3.1 Soil Quality

Piezometers were installed and boring logs constructed by LMS. According to their logs, the Site is underlain by heterogeneous soils characteristic of the glacial till that covers much of Staten Island. The soils were identified in LMS borings logs as being reddish to brown in color and comprised of sands and silts to clays with some gravel.

No soil samples were collected for laboratory analysis. However, the presence of volatile organics (as detected with a photoionization detector or via olfactory or visual observations) was not noted in any of the three boring logs available from the LMS IIWA report (boring logs P-1 through P-3).

### 1.3.2 Groundwater Quality

During the course of the LMS 2000 IIWA, seven piezometers were installed and sampled. These seven wells supplemented previously existing monitoring wells installed by others. No additional information is available for these wells. During the CDM site visit, five piezometers and two monitoring wells were located on the Site property. Two additional monitoring wells were noted to the north of the building in the shopping center.

Groundwater results from the LMS IIWA identified the highest concentrations of chlorinated VOCs in groundwater immediately adjacent to the east side of the building. The contamination was determined to be migrating vertically downward as evidenced by higher PCE concentrations in the deeper piezometers.

Based on the results of their investigation, LMS recommended that a soil gas survey be conducted in the area to identify the potential impact of PCE contamination to indoor air at Buildings A, B, and C. Additionally, LMS recommended that a deeper well be installed to vertically delineate groundwater contamination in the vicinity of P-3D on the east side of the building. Continuous soil cores were proposed for collection to better characterize the subsurface stratigraphy in the vicinity of the Site.

CDM performed a RI at the site in 2009. The field investigation included the following:

- Geophysical survey of proposed monitoring well locations for utility clearance; and survey of east side of Site for buried structures, such as sumps and underground storage tanks (USTs);
- Collection of 13 subsurface soil samples ;
- Installation of 13 groundwater monitoring wells and subsequent collection of groundwater samples;
- Topographic and site survey of Site topography, building corners and features, and investigation locations;
- Installation of temporary sub-slab soil vapor sampling ports and subsequent collection of sub-slab soil vapor samples, indoor air samples, and two ambient air samples;
- Stage, characterized and dispose of Investigation Derived Wastes.

Subsurface soil samples were collected for volatile organic compound (VOC) and semi-volatile organic compound (SVOC) analysis; three of these samples were run for full list Target Compound List (TCL)/Target Analyte List (TAL) analysis. No residual source areas in unsaturated soils that may be still contributing to groundwater contamination were detected. Tetrachloroethene (PCE) was only detected at concentrations slightly above the unrestricted use soil cleanup objectives at one location (MW-11S) from a depth of 15 to 15.5 feet below ground surface (bgs). PCE and its associated breakdown products were not detected in vadose zone soil samples collected during the RI. The RI determined that soil was not the primary media of concern.

Monitoring wells MW-8S, MW-9S/9D, MW-10S/10D, MW-11S/11D, MW-12S, MW-13S/13D, MW-14S, MW-15D, MW-16S were installed as part of this RI. Prior to groundwater sampling in monitoring wells, a synoptic round of groundwater levels was measured. Groundwater samples were analyzed for VOCs and SVOCs; three of these samples were also run for full list TCL/TAL analysis. Results showed groundwater contamination at the Site consists primarily of PCE and its associated breakdown products: trichloroethene (TCE), 1,1-dichloroethene (1,1-DCE), cis-1,2-dichloroethene (cis-1,2-DCE), trans-1,2-dichloroethene (trans-1,2-DCE), and vinyl chloride (VC). Chlorobenzene (CB) was also detected. Two well results and oil-water interface probe readings confirmed the likely presence of dense non-aqueous phase liquid (DNAPL) at MW-12S and MW-14S. Since MW-15D functions as a deep well paired with MW-14S and concentrations in MW-15D were not indicative of the presence of DNAPL, it was concluded that DNAPL has perched atop the low permeability layers encountered at MW-14S. DNAPL is also suspected atop low permeability layers in the vicinity of MW-12S.

A soil vapor investigation was conducted in order to determine if vapor phase contaminants are present at concentrations that could impact indoor air quality. The investigation included collecting four sub-slab soil vapor samples, five indoor air samples, one duplicate sample of each, and two ambient air samples for VOC analysis. TCE and PCE were detected at concentrations exceeding guidance values in an indoor air sample in Building A (the building onsite). This sample also showed the presence of vinyl chloride, cis-1,2-DCE, chloroform, and toluene at concentrations above background values.

The RI concluded that it is likely any PCE released at the Site has since migrated vertically through the vadose and saturated zones to ultimately accumulate atop low permeability layers. PCE NAPL has continued to migrate along these layers, functioning as a source for dissolved phase PCE groundwater contamination. The RI found evidence of reductive dechlorination taking place in the aquifer at the Site as there are detections of degradation products (trans- and cis-1,2-DCE and VC), and oxidation-reduction potentials in groundwater indicate that conditions are slightly reducing.

## 1.4 Report Organization

This RI report is organized in the following manner. Tables, figures, and appendices are presented after Section 7.

- Section 1      INTRODUCTION presents the Site background and history, location, operational and remedial history, potential sources, and the project objectives.
- Section 2      STUDY AREA INVESTIGATIONS provides the investigation procedures and any variations which may have influenced sampling procedures or analytical results.
- Section 3      PHYSICAL CHARACTERISTICS OF THE STUDY AREA describes the physical conditions of the Site and surroundings, including a general description of soils, geology, hydrogeology, topography, and groundwater levels and flow direction.
- Section 4      NATURE AND EXTENT OF CONTAMINATION presents and evaluates the analytical results of the environmental samples collected at the Site, as compared to the recommended New York State standards.
- Section 5      CONTAMINANT FATE AND TRANSPORT evaluates the physical and chemical characteristics of the site contaminants. It presents a conceptual model of the site sources, contaminant migration pathways, and receptors.
- Section 6      CONCLUSIONS presents the conclusions based upon the results of the remedial investigation and presents recommendations for potential future work.
- Section 7      REFERENCES

## Section 2

# Study Area Investigations

The RI was designed to characterize the nature and extent of contamination at the Site and to provide data to support the Feasibility Study. The following subsections describe the field investigation that was conducted from May 2011 through March 2012 in support of the RI. All field work, except where noted, was conducted in accordance with the CDM Quality Assurance Project Plan (QAPP) which has been provided to NYSDEC for Contract Number D-004437 and referenced within the Former Paul Miller Remedial Investigation and Feasibility Study Work Plan.

The RI consisted of the following field activities:

- Electrical Conductivity (EC) / Membrane interface probe (MIP) Investigation
- MIP confirmatory soil samples
- MIP confirmatory groundwater samples
- Sub-slab soil sampling
- Sub-slab groundwater sampling
- Monitoring well rehabilitation and piezometer abandonment
- Monitoring well sampling
- Slug tests; and
- Two rounds of synoptic water levels

## 2.1 Electrical Conductivity and Membrane Interface Probe Investigation

Electrical Conductivity (EC) and Membrane Interface Probe (MIP) technology was used to evaluate stratigraphy and screen soil and groundwater for VOCs at 35 locations. Figures 2-1 shows the EC/MIP locations. The objective of the EC/MIP survey was to provide screening-level characterization of VOC contamination in the subsurface and to provide high-resolution estimates of soil stratigraphy. The locations of the screening points were selected based on historical data and aerial photographs.

The probe was advanced using a Geoprobe direct push technology (DPT) rig. The 1.5-inch diameter probe was pushed into the subsurface at a penetration rate of approximately 1-foot per minute. The tip of the probe contains a thermister, which provides a heat source to volatilize VOCs. The gases, that are produced, pass into the probe through a permeable membrane and enter a sampling loop. The gases are then transported to the surface and pass through the photoionization detector (PID), flame ionization detector (FID), and electron capture detector (ECD). The detectors analyze the gas and provide an immediate qualitative readout of volatile organic content. The probe also records electrical conductivity of the subsurface. The data were displayed on a monitor and were downloaded to a laptop computer.

MIP results were analyzed on a daily basis and were used to direct sampling on an ongoing basis. In addition, the MIP response data was used to provide screening-level data on the vertical distribution of VOC contamination in groundwater. The MIP data were also used to direct the confirmatory soil and groundwater sampling, which are discussed in Section 2.1.2 and 2.1.3. A complete set of EC/MIP logs is included in Appendix A.

### 2.1.1 MIP Confirmatory Soil Sampling

Following review of the MIP results, intervals with significant detections were selected for confirmatory soil sampling and analysis. Soil borings were advanced using the DPT method and a soil sample was collected from the specified interval. Sample locations are shown on Figure 2-1 and sample details are provided in Table 2-1. Soil samples were collected using a decontaminated stainless steel trowel and bowl per procedures the generic QAPP. Soil samples were tested at Mitkem Laboratories in Rhode Island and were tested for VOCs.

### 2.1.2 MIP Confirmatory Groundwater Screening

Following review of the MIP results, intervals with significant detections were selected for confirmatory groundwater sampling and analysis. Borings were advanced using the DPT method and a groundwater screening sample was collected from the specified interval.

To collect groundwater screening samples, a retractable well screen was driven to the target depth of a groundwater screening borehole using the DPT method. The well screen was subsequently opened to create a four-foot sampling interval. New, sample-dedicated high density polyethylene (HDPE) tubing with a check valve attached at the bottom was inserted through the drill rods into the well screen and the tubing was connected to a peristaltic pump at the surface. Water was purged from the well screen and temporarily collected in five-gallon buckets. During purging, a calibrated water quality meter was used to measure pH, specific conductivity, dissolved oxygen (DO), temperature, and oxidation-reduction potential (ORP) at five minute intervals. Turbidity was measured at five-minute intervals using a calibrated LaMotte turbidity meter. Purging continued until the water quality parameters had been stable for 3 consecutive readings or parameters stabilized as much as subsurface conditions would allow within a 1-hour time period.

Sample locations are shown on Figure 2-1 and sample details are provided in Table 2-2. Groundwater samples were collected directly into the 40 ml pre-preserved VOA vials per procedures the generic QAPP. Groundwater samples were tested at Mitkem Laboratories in Rhode Island and were tested for VOCs.

## 2.2 Sub-slab Soil Screening

Sub-slab soil sampling was conducted to evaluate the potential for DNAPL to be located under the building foundation. Thirteen samples were collected from eight boreholes (Table 2-3). Six boreholes were installed through the basement, B-1a, B-1b, B-2, B-3, B-4, B-5 and B-6. Borings B-1a, B-1b and B-2 were located in the boiler room, borings B-3 and B-4 were located in the former fur vault and borings B-5 and B-6 were located in the storage room. An angled boring (B-7) was also installed under the building in the center of the eastern wall of the building (Figure 2-2)

Soil borings were advanced using DPT methods and sampled continuously. Soil cores were retrieved and the lithology was logged in the field at each of the sample locations. Three soil samples were planned for each boring. A shallow sample was to be taken immediately below the slab 0-5ft, a deep sample was to be taken at the boreholes terminal depth, and the final sample was to be at a selected

depth interval based on the presence of visual contamination or when elevated levels of organic vapors (VOC) were detected on a photoionization detector (PID). The soil was screened with a PID with an 11.7 lamp, but no readings above background were detected.

In B-1a, B-1b, B-2, B-5 and B-6 two soil samples were collected; a shallow sample labeled with an "A" and a deep sample labeled with a "B". In B-3 and B-4 only the shallow "A" samples were retrieved. In B-7 only the deep "B" sample was retrieved. B-3 and B-4 hit refusal after 1.5 feet of penetration. It is suspected that there is an additional slab underneath the elevated slab in the fur vault. This slab could not be penetrated by the small DPT rig used inside the building. Some samples were recovered from these locations (B-3 and B-4) however the sample mostly contained gravel. At B-7-A and B-7-B the drill rig was conducting an angle boring through the sidewalk to obtain soil samples below the estimated location of the former dry cleaning machine.. Upon attempting to collect shallow sample B-7-A aiming for 5 ft below the slab under the former dry cleaning machine, the drill rig hit refusal on the foundation of the Former Paul Miller Dry Cleaners. The sample at B-7-A was not collected due to refusal at 3.5ft bgs.

Soil samples were collected using a decontaminated stainless steel trowel and bowl per procedures the generic QAPP. Soil samples were tested at Mitkem Laboratories in Rhode Island and were tested for VOCs.

## 2.3 Sub-slab Groundwater Screening

Sub-slab groundwater screening sampling was conducted to evaluate the potential for DNAPL to be located under the building foundation. Six samples were collected from eight boreholes (Table 2-4). Six boreholes were installed through the basement, B-1a, B-1b, B-2, B-5 and B-6. An angled boring (B-7) was also installed under the building in the center of the eastern wall of the building (Figure 2-2). Temporary wells were installed in B-1a, B-1b, B-2, B-5, B-6 and B-7-B. These wells were inserted directly into the soil boring location upon completion of the soil boring. The PVC screens were inserted to 5 ft bgs at all of the sample locations for groundwater except for B-7-B where the well was installed with 5 ft of casing and 10 ft of screen to a depth of 12.5 ft bgs.

At each of these temporary wells groundwater samples were collected. Most samples were collected via the "Low Stress (Low Flow) Purging and Sampling" method as described in the QAPP. The temporary wells at locations B-1a and B-7 had a low rate of recharge making it necessary to recover groundwater samples without stabilizing the water quality parameters. Groundwater samples were analyzed at Mitkem Laboratories in Rhode Island for VOCs.

## 2.4 Monitoring Well Rehabilitation and Piezometer Abandonment

During a site visit in November 2011, it was noted that several of the surface completions for the monitoring wells and piezometers were in poor repair and not functioning properly. Thirteen locations were inspected as a part of the rehabilitation and decommission process and it was determined that P4-D, MW-1, MW-12S, MW-15D, MW-16S, MW-14S, and MW-8S should be rehabilitated and P-1, P-2, P3-S, P3-D, P-5 and P-4S should be abandoned. Locations P-4S, P4-D and P-1 could not be located and are believed to be under a concrete pad, constructed as part of the drive through on the adjacent property.

The monitoring wells were rehabilitated by removing the concrete pad and flush-mount cap and replacing them with a new concrete pad and new flush-mount cap. The concrete pads were broken up using the direct push rig fitted with a chisel bit. The debris and flush-mount cap were removed. The new flush-mount cap was installed. Then the concrete was poured around the cap and leveled. Cones were placed around the drying concrete pads to alert pedestrians of a potential hazard.

The piezometers were decommissioned in accordance with CP-43 by removing the concrete pad and flush-mount casing, and pulling the PVC casing and well screen. The well casing and screen were easily removed and did not appear to be properly grouted into place. The holes were filled with neat cement to depth of 6 inches below the ground surface (bgs). The decommissioned locations had cones placed over them and were given over night to allow the cement to harden. The following day the holes were filled with eco-friendly asphalt and tamped flush with the ground surface.

## 2.5 Monitoring Well Sampling

During March 2012, monitoring well sampling was conducted at 17 existing monitoring wells. Monitoring well samples were collected via the "Low Stress (Low Flow) Purging and Sampling" method as described in the QAPP. During purging, the water quality parameters temperature, turbidity, specific conductance, dissolved oxygen, pH and oxidation-reduction potential were recorded (Table 2-5). Samples were analyzed onsite, for ferrous iron, using the Hach colorimetric method. Well purge water was contained and transferred to 55-gallon drums. Section 2.11 discusses control and disposal of investigation derived waste.

Monitoring well samples were submitted to Spectrum Laboratories for VOC, nitrate/nitrite, sulfate, chloride, alkalinity, total organic carbon (TOC), and methane, ethane, ethene (MEE) analysis. VOCs were analyzed by per the NYSDEC-approved Work Plan. .

## 2.6 Slug Testing

Slug tests were performed at the Site to estimate the hydraulic conductivity (K) of water-bearing strata. Six "shallow" wells, screened approximately 25 to 35 feet below ground surface (bgs), and two deep wells, screened approximately 60 to 70 feet bgs were tested. The shallow strata exhibit the highest contaminant concentrations on site and site specific estimations of hydraulic conductivity will allow a more effective review of remedial options during the feasibility study stage. The two deeper wells were tested to understanding the hydraulic conductivity beneath the most contaminated zone to help evaluate plume behavior for different remedial alternatives.

Slug tests were performed using a weighted cylinder to displace the water and a pressure transducer to measure and record water level changes over time. At each well tested, the pressure transducer was deployed and programmed to record depth to water on a logarithmic time basis (water levels were recorded at a rate of four times per second at the start of the test, and the time interval between measurements increased logarithmically throughout the test). Manual water levels were measured periodically to confirm the accuracy of the pressure transducer.

The slug was tied to a rope and lowered to a depth just above the static water level. A few seconds after activating the data logger, the slug was rapidly lowered into the water, displacing the water level upward to initiate the falling head test. After the water level recovered to the static level (i.e. minimum 90 percent recovery), the data logger was turned off and reprogrammed for a new test. A few seconds

after re-activating the data logger the slug was rapidly removed from the well, lowering the water level to initiate the rising head test. When the water level returned to the static level, the data logger was turned off and the equipment was removed from the well, decontaminated, and moved to the next well.

The slug test analysis and results are discussed in Section 3.2

## 2.7 Synoptic Water Levels

Two rounds of synoptic water levels were collected in November 2011 and March 2012. Water level measurements were converted to elevation and used to evaluate the vertical and horizontal groundwater flow at the site. Prior to measuring water levels, the water-tight caps were removed and the water levels were allowed to equilibrate with atmospheric pressure. The depth to water at each well was then measured to an accuracy of 0.01 feet with an electric water level indicator. The water level indicator was decontaminated with distilled water and non-phosphate detergent between wells. Table 2-6 contain synoptic water level data for March 2012.

## 2.8 Indoor Air Sampling

Air quality sampling badges for PCE were placed in two locations in and around the bank building to the west of the former Paul Miller Dry Cleaners in March, 2011. Table 2-7 summarizes the indoor air samples collected. A Passive air sampling badge and a duplicate were placed in the basement of the bank building, and in the vicinity of the bank teller drive through, approximately four feet off the ground. The badges were kept in place for 24 hours, and then collected and shipped to the laboratory for PCE analysis.

Attempts were made to conduct passive air sampling at the Kentucky Fried Chicken restaurant to the east of the Paul Miller building, but no site access for the sampling was granted by the restaurant management.

## 2.9 Topographic Survey

Sample locations and monitoring wells were surveyed by Bryant Associates, a New York-licensed surveyor, based in Syracuse, New York, surveyed the four existing monitoring wells, and 35 MIP locations. The horizontal datum, used by Bryant, was New York State Plane, Long Island NAD 83/96, and the vertical datum used was NAVD88.

## 2.10 Control of Investigation-Derived Waste

Investigation derived waste from each sampling location was containerized in 55-gallon drums and disposed of off-site. A total of four drums containing soil cuttings and two drums containing purge water were generated during the investigation. IDW generated during this investigation was disposed of in one shipment. Composite and representative soil and water samples were collected. The samples were analyzed for the full TCLP list and RCRA characteristics. The drums were stored in a designated area at the Site until they could be removed by a waste hauler. The drums were removed from the Site by Innovative Recycling Technologies, Inc. The laboratory data for the waste characterization samples is provided in Appendix B.

## 2.11 Laboratory Analysis and Validation

All samples were analyzed by Spectrum Analytical. The soil samples were analyzed for VOCs by EPA method 8260B, SVOCs by EPA method 8270C, pesticides by EPA method 8081, PCBs by EPA method 8082, metals by EPA methods 6010/7471, and mercury by Method 7471. Groundwater samples were analyzed for VOCs by EPA method 8260B, SVOCs by EPA method 8270C, pesticides by EPA method 8081, PCBs by EPA method 8082, metals by EPA methods 6010/7040, mercury by EPA Method 7470 and methane, ethane, and ethene by Method RSK 175. Groundwater samples were also analyzed for sulfide by Method 4500-S,F, alkalinity by Method 2320, chloride, nitrate, nitrite, and sulfate by EPA Method 300.0, hardness by Method 2340B, TDS by Method 2540C, TSS by Method 2540D, ammonia by Method 4500, Kjeldahl nitrogram by Method 4500-NORG, and total organic carbon by Method 5310B.

NYSDEC ASP Category B data deliverables were obtained for all analyses except waste characterization analyses. The analytical data packages are provided in Appendix C.

All samples collected, with the exception of the waste characterization samples, were validated in accordance with NYSDEC Data Usability Summary Report (DUSR) guidance by a party that is independent of the laboratory that performed the analyses and CDM. A usability analysis was conducted by Nancy Potak, a qualified data validator. The DUSRs are provided in Appendix D.

## 2.12 Field Documentation

Field notebooks were utilized during all on-site work. A dedicated field notebook was maintained by the field technician overseeing the Site activities. In addition to a copy of the field notes, copies of all original sampling forms and purge forms used during the field activities are provided in Appendix E.

## Section 3

# Physical Characteristics of the Study Area

### 3.1 Topography and Drainage

The Former Paul Miller Site lies at approximately 25 feet amsl and has been leveled and paved for development. Local topography slopes gently from the southwest to the northeast in the vicinity of the Site (USGS Arthur Kill, New York 7.5-Minute topographic map, 2011). Palmer's Run formerly flowed west to east and bisected the paved area just north of the site. The Site's former, natural topography, likely sloped towards Palmer's Run,. Despite having been filled in, the channel, and possibly its branches,still have an influence on groundwater flow at the Site.

### 3.2 Regional Geology and Hydrogeology

The Site is located within the Atlantic Coastal Plain Physiographic Province. A history of coastal submergence and emergence spanning the Cretaceous Period, significant differential erosion during the Cenozoic, and glaciation during the Quaternary Period is reflected in the present day geology of Staten Island.

#### 3.2.1 Bedrock

In the middle of Staten Island is a lens-shaped body of serpentine that unconformably overlies the Manhattan Schist, the local basement rock. The serpentine ophiolite, a piece of ancient ocean crust that was obducted onto the North American plate during the Taconic Orogeny in the Paleozoic Era. Over time, the highly mafic minerals of the former mafic to ultramafic oceanic rocks altered chemically into the serpentine seen today. To the west of the Serpentine lens lie the sedimentary rocks of the Triassic Newark Supergroup, which unconformably overlie the basement rocks and are intruded by a northeast-southwest trending Jurassic sill of diabasic composition.

#### 3.2.2 Unconsolidated Sedimentary Deposits

To the east of the serpentine ophiolite, the geology of Staten Island is characterized by a large southeastward-thickening wedge of glacial till and outwash. The till and outwash unconformably overlie a series of unconsolidated marine coastal plain sands, silts, and clays from the Cretaceous period, which, in turn, unconformably overlie the gently eastward-dipping Manhattan Schist, the local basement rock.

As identified in *The Glacial Geology of New York City and Vicinity* by (Sanders and Merguerian, 1994), to the west of the ophiolite the Newark Supergroup is unconformably overlain by the Harbor Hill Formation, a widespread Quaternary ground moraine deposit comprised of reddish-brown glacial till and outwash. This unconsolidated sequence is representative of the subsurface materials that immediately underlie the Site.

The Harbor Hill Formation is approximately 100 to 150 feet thick in the area of the Site. Groundwater in these deposits occurs under water-table or semi-confined conditions depending on the nature of the subsurface at any given location. The general flow of groundwater in the unconsolidated glacial till is to the north towards Kill van Kull (USGS, 1987).

The consolidated rock units of the Newark Supergroup and the overlying unconsolidated deposits are hydraulically connected, and groundwater flows both vertically and horizontally within them. However, the majority of the groundwater flow occurs within the glacial unconsolidated deposits due to its greater hydraulic conductivities.

### 3.3 Site Geology and Hydrogeology

The Site is underlain by the unconsolidated glacial till of the Harbor Hill Formation, which has likely been reworked by Palmer's Run and its tributaries. The Harbor Hill Formation is estimated to be 100-150 feet thick at the site. Borings and wells did not extend below a depth of 100 feet bgs, so the total thickness was not confirmed.

Site stratigraphy was evaluated from lithologic descriptions collected during soil borings and the electrical conductivity (EC) investigation (Appendix A). Lithologic logs indicate that the geologic deposits at the Site are predominantly fine to medium sand, with silty sand lenses, and local deposits of coarse sand and gravel (e.g. MW-11D). In some cases blow counts were low (e.g. a maximum of 15 blows per 6 inches at MW-15D), indicating fairly loose soils; in some cases the soils exhibited higher blow counts indicating more compact soils (e.g. blow counts generally at least 30 per six inches at MW-14S).

Generally, the upper 35 feet are a heterogenous mix of silt and sand. The silt and sand appear to be locally stratified, but the units are not continuous across boreholes. Below 35 feet bgs, the lithologic descriptions and the EC logs show a transition to well graded sand to the terminal depth of the borings at approximately 80 feet bgs. Figure 3-1 is a representative cross section of the site's general geology.

Slug tests were conducted on eight wells. Table 3-1 summarizes the results of the slug tests and the data evaluation is detailed in Appendix F. The calculated hydraulic conductivities range from 0.05 ft/day (MW-14S) to 69 feet per day (MW-15D). The relative hydraulic conductivities generally compare favorably with the strata screened at each well, as summarized below:

- MW-15D exhibited the highest hydraulic conductivity at 59-69 ft/day; this is consistent with its strata – medium sand.
- MW-11D, MW-12S and MW-13S exhibited hydraulic conductivity values ranging from 3 to 6 ft/day; MW-11D and MW-13S both primarily screened in siltier sand than MW-15D.
- MW-8S, MW-9S and MW-16S exhibited K values of less than 1 ft/day. MW-9S is screened in silty medium sand and MW-16S is screened in fine to silty sand; strata at these wells exhibited low blow counts and the lower K values are consistent with the finer formation material. MW-8S is screened in fine sand and sandy gravel with clay layers exhibiting somewhat higher blow counts, both consistent with the lower K values exhibited.
- MW-14S exhibited the lowest K value, 0.05 ft/day. This well is screened in silty sand and sandy silt, and its blow counts were among the highest of the wells tested, which are both consistent with the low K.

The heterogeneity of the glacial overburden has created a very complex groundwater flow system. Figures 3-2, 3-3 and 3-4 show the potentiometric surfaces at the water table, 30 feet bgs and 70 feet bgs, respectively. These surfaces were calculated using the water levels measured in March 2012

(Table 2-6). The three potentiometric surfaces were plotted separately to illustrate how contaminant migration differs with depth at the site.

The water table potentiometric surface (Figure 3-1) was developed using the four wells at the site that are screened across the water table. This potentiometric surface shows a groundwater high at MW-3, which is likely related to the local geology or to a stormwater drainage feature. The groundwater flow at the water table is to both the north and south from MW-3.

The shallow potentiometric surface (Figure 3-2) represents groundwater flow at 30 feet bgs. This potentiometric surface was constructed using data from the shallow (S) wells. Groundwater flow from the Site in this interval is to the northwest overall, but it should also be noted that groundwater from the northeast of the site is flowing south-southwest and converging just north of the site. This flow pattern is likely influenced by the geology, given the likelihood of preferential flow paths in the till. Preferential flows paths are indicated by the hydraulic conductivity measurements, which vary by two orders of magnitude in the shallow wells. The shallow potentiometric surface is also consistent with the Site's former, natural topography, where groundwater in the immediate vicinity of the Site likely flowed towards Palmer's Run. Despite having been filled in, it is likely that the presence of the channel, and possibly its branches, still influence groundwater flow at the Site.

The deep potentiometric surface (Figure 3-3) represents groundwater flow at 70 feet bgs. This potentiometric surface was constructed using data from the deep (D) wells. Groundwater flow in this interval is to the north-northeast and is consistent with regional groundwater flow towards Kill Van Kull.

## Section 4

### Nature and Extent of Contamination

This section presents the environmental sampling results and analytical methods used to evaluate contamination at the Site. Section 4.1 describes the approach for selecting site-specific screening criteria, selection of representative contaminants, and data presentation. Section 4.2 presents soil, groundwater, and MIP investigation results. Section 4.3 presents an interpretation of the data and an overall evaluation of contamination at the Site.

#### 4.1 Approach to the Evaluation of Contamination

The characterization and evaluation of the nature and extent of contamination is focused on those constituents identified as representative contaminants, which were generally determined by evaluating historical information on contaminant sources, exceedances of screening criteria, the frequency and magnitude of the exceedances, and background concentrations. However, all other detected contaminants were also subject to the media-specific screening process and are discussed briefly in the text.

##### 4.1.1 Selection of Screening Criteria

The soil analytical results were compared to the Unrestricted Use Soil Cleanup Objectives (6 NYCRR Part 375-6.8 (a), December 14, 2006). The groundwater analytical results were compared to the New York State Standards and Guidance Values for Class GA Groundwater (NYSDEC TOGS 1.1.1).

The 2006 NYSDOH Vapor Intrusion guidance indicates that the State of New York does not have any standards, criteria or guidance values for subsurface soil vapor. However, Table 3-1 of the guidance document provides guidance values for indoor and outdoor air against which methylene chloride, PCE, and TCE may be compared. Additionally, background concentrations derived from background studies are available in Appendix C of the guidance document. In the case of the Site, the 90th percentile values presented in Table C2, "EPA 2001: Building assessment and survey evaluation (BASE) database," are appropriate for comparison to the Site's indoor and outdoor air sample results. This value is 100 µg/m<sup>3</sup> for PCE

##### 4.1.2 Selection of Representative Contaminants

Representative contaminants were selected to focus the evaluation of contamination on site related contaminants, which will be drivers of remediation. To select the site related contaminants, CDM reviewed the historical investigations and considered the chemicals and processes used at the Site.

The Site is located in an urbanized area which has been developed for approximately 50 years and operated as a dry cleaners from approximately 1960 – 1995. Based on this analysis, it was determined that PCE and its degradation products, TCE, cis-1,2-DCE, and VC were most representative of site related contamination . These compounds generally exhibit the greatest spatial distribution and highest detected levels. Other contaminants detected at the site will not be evaluated in detail.

### 4.1.3 Data Presentation

The analytical results from the RI were entered into the site database for evaluation purposes. The data were exported to a geographic information system (GIS) and geological evaluation and visualization software for analysis and graphical presentation. The graphical data presented includes VOC data for aqueous samples ( $\mu\text{g}/\text{L}$ ) and soil samples ( $\text{mg}/\text{kg}$ ).

All analytical data were reviewed to ensure that they meet the project requirements for representativeness, completeness, precision, and accuracy.

Groundwater and soil analytical data were compared to the appropriate medium-specific screening criteria. The discussion below focuses primarily on the representative contaminants. Concentrations of site related compounds are displayed on the figures in Section 4.

## 4.2 Investigation Results

### 4.2.1 Membrane Interface Probe Investigation

A subsurface investigation using a membrane interface probe (MIP) was conducted in May 2011 to aid in delineating the horizontal and vertical extent of contamination. The membrane interface probe (MIP) houses 4 detectors including the electron captor detector (ECD), flame ionization detector (FID), and the photoionization detector (PID). The FID measures volatile organic hydrocarbons, and the PID measures aromatic compounds. However, the compounds of interest at our site are CVOCs. The ECD measures the relative concentration of CVOCs in the soil and aquifer adjacent to a sampling window in the probe body. The ECD can detect the presence of gross VOCs but does not produce concentration data or specific compound speciation; the data is used as a screening tool. The MIP logs are included in Appendix A. Investigation locations can be seen on Figure 2-1, and MIP results are on Figure 4-1.

The highest ECD responses for the largest depth interval were detected in MIP-2, MIP-2C, MIP-2CR, MIP-3, MIP-4, MIP-5, MIP-5B, and MIP-6. With the exception of MIP-5B, these boreholes are all located around the site building. MIP-2C and MIP-2CR are located near the northwest corner of the building. MIP-2 is located further northwest of these boreholes. MIP-5 and MIP-6 are located near the front end of the building. MIP-5B is located directly west of the north end of the site building. ECD readings at MIP-2CR, MIP-5, and MIP-6 show a sustained high response for a large depth interval. In this case, it is possible that contamination flooded the detector at a certain depth due to the presence of NAPL. Since the detector is flooded with NAPL, the probe will continue to read high concentrations until the NAPL has worked its way out of the detector. The time it takes for the NAPL to leave the detector is unknown; therefore, the full depth of the contaminated interval is unknown. However, in general, excluding MIP-3 and MIP-4, contamination appears to be in the 10 ft amsl to -15 ft amsl interval (between 10 and 35 feet below ground surface). MIP-3 and MIP-4 are located northeast of the Site; contamination in these boreholes is in a deeper interval, below approximately 0 ft amsl.

Moving downgradient from these boreholes, the ECD responses decrease in occurrence, magnitude, and thickness of the depth interval. Responses are seen at MIP-1, MIP-1B, and MIP-2B. The general interval of contamination at MIP-1 and MIP-1B, which are northeast of the site, is between 10 and 20 ft bgs (15 and 5 ft amsl). The interval of contamination at MIP-2B, which is northwest of the site, is deeper, between 35 and 40 ft bgs (-10 and -15 ft amsl).

## 4.2.2 Soil Investigation

### 4.2.2.1 MIP Confirmatory Soil Sampling

During the MIP investigation in May 2011, 14 soil samples were collected from below the water table, including two duplicates. Eleven of these samples were co-located with the MIP in order to confirm the ECD data. Figure 2-1 presents the sample locations. Analytical data is presented in Table 4-1. The total depth of the 14 soil screening points ranged from 10 feet bgs at location MIP6 to 45 feet bgs at MIP3. Soil samples were collected at two different depths at MIP5 and MIP6 locations.

As shown on Figure 4-1, PCE was detected in 8 of soil samples (MIP1, MIP2, MIP3, MIP5, MIP5B, and MIP6) used to confirm the MIP results. Shallow and deep samples collected at MIP5 and MIP6 both detected PCE. TCE was detected in one sample at MIP3. *Cis*-1,2-DCE was detected in three samples (MIP1B, MIP2, and MIP3). VC was not detected in any sample. No samples exceeded screening criteria. ECD peaks matched depths of the soil screening samples that detected contamination. Groundwater screening samples were not taken in these boreholes. However, given the low soil organic carbon-

water partitioning coefficients ( $K_{oc}$ ) of VOCs, it is likely that there is groundwater contamination at these depths which would explain the ECD readings. The ECD responds to VOC concentrations in both the soil and the groundwater.

Other VOCs detected include acetone, methylene chloride, and naphthalene but samples did not exceed soil screening criteria. Acetone and methylene chloride were detected below lab reporting limits and may be lab contaminants. No SVOCs were detected in soil samples at concentrations exceeding the unrestricted use soil cleanup objectives. No pesticides or PCBs were detected in the soil samples. Nickel was detected at a concentration of 31 mg/kg in sample MIP1B and at 77.9 mg/kg in sample MIP5B, which exceed the unrestricted use soil cleanup criteria of 30 mg/kg. Chromium was detected at a concentration of 34 mg/kg in sample MIP5B which exceeds the soil screening criteria of 30 mg/kg. No other metals were detected at concentrations exceeding the unrestricted use soil cleanup objectives in soil samples.

### 4.2.2.2 Sub-Slab Soil Screening

CDM conducted a sub-slab investigation in November 2011 to characterize VOC contamination in the soil beneath the building at the site. During this investigation, 13 soil samples were collected. Seven samples were collected in the vadose zone at six inches below the slab; six soil samples were collected beneath the water table at multiple depths, ranging from 5 ft to 12.5 ft below the slab. Figure 2-2 depicts the sample locations; analytical data is presented in Table 4-2.

As shown on Figure 4-2 PCE was detected in six soil samples collected from the vadose zone ranging from 2.7 J  $\mu$ g/kg to 110 J  $\mu$ g/kg, and in three soil samples collected from beneath the water table ranging from 4.5 J  $\mu$ g/kg to 55  $\mu$ g/kg . TCE was detected in two soil samples collected above the water table ranging from 6.6 J  $\mu$ g/kg to 16 J  $\mu$ g/kg, and in two saturated soil samples ranging from 1.2 J  $\mu$ g/kg to 3.2 J  $\mu$ g/kg. *Cis*-1,2-DCE was detected in three dry soil samples ranging from 3.5 J  $\mu$ g/kg to 12  $\mu$ g/kg. The maximum concentrations detected were all at PM-SB-5-A. VC was not detected in any samples. No samples exceeded soil screening criteria. The reporting limit for these compounds ranged between 5 and 6.8  $\mu$ g/kg.

Other VOCs detected include 1,2,4-trimethylbenzene, acetone, methylene chloride, naphthalene, and toluene but samples did not exceed soil screening criteria. SVOCs, pesticides, PCBs, or metals were not analyzed in these soil samples.

## 4.2.3 Groundwater Investigation

### 4.2.3.1 MIP Confirmatory Groundwater Screening

During the MIP investigation, CDM collected four groundwater screening samples from two locations, MIP-5 and MIP-6, and one duplicate. Samples from MIP-5 were collected at depths of 15 ft bgs and 28 ft bgs; samples from MIP-6 were collected at depths of 15 ft bgs and 33.5 ft bgs. Analytical data is presented in Table 4-4 and Figure 4-3.

PCE concentrations exceeded the screening criteria in all samples; concentrations ranged from 150 µg/L (deep sample at MIP-5) to 32,000 D µg/L (shallow sample at MIP-5). PCE concentrations in the shallow samples taken at locations MIP-5 and MIP-6 were indicative of a possible presence of DNAPL. DNAPL is suspected to be present when the groundwater concentration is greater than one percent of its pure-phase solubility (150,000 µg/L for PCE). TCE exceeded the groundwater screening criteria in samples at MIP-6 at shallow and deep concentrations of 9 µg/L and 8.7 µg/L, respectively. It was detected in the deep sample at MIP-5 but did not exceed groundwater screening criteria. *Cis*-1,2-DCE concentrations exceeded groundwater screening criteria in three samples. The shallow sample at MIP5 showed the highest concentration at 780 µg/L. VC was not detected in any samples. These results confirm the information gathered from the ECD measurements taken in these boreholes. The ECD readings suggest that the probe had been fouled by NAPL in MIP-5 at approximately the same depth the shallow sample was taken which detected concentrations indicative of NAPL. Concentrations detected in MIP6 confirm the high ECD readings and suggest possible DNAPL which could lead to possible probe fouling as well. VOC concentrations in soil confirmatory samples taken in these boreholes did not exceed soil screening criteria. However, since VOCs have a low  $K_{oc}$  meaning these compounds are more hydrophilic, absorption would not be expected which leads to low soil concentrations.

Methyl tert-butyl ether was the only other VOC detected. It was detected at 1.7 J µg/L, which is below both the reporting limit and the groundwater screening criterion. SVOCs detected include bis(2-ethylhexyl)phthalate and diethyl phthalate. No pesticides or PCBs were detected. Iron, manganese, and nickel exceeded their groundwater screening criteria in four, three, and one sample, respectively. The maximum concentration of iron was 2,660 µg/L found in the deep sample at MIP6. The maximum concentration of manganese was 2,320 µg/L found in the shallow sample at MIP5. Aluminum and magnesium were detected but their concentrations did not exceed groundwater screening criteria.

### 4.2.3.2 Sub-Slab Groundwater Screening

In November 2011, CDM conducted a sub-slab investigation, as discussed in Section 2. Six groundwater screening samples were collected to characterize the aqueous VOC contamination beneath the building on the site. Five samples were taken at approximately 8-9 ft bgs. One sample was taken at an angle at about 12.5 ft below the slab. The analytical results from the investigation are in Table 4-5. As illustrated on Figure 4-3, concentrations of PCE and *cis*-1,2-DCE exceeded groundwater screening criteria in all six samples. TCE concentrations exceeded groundwater screening criteria in three samples, and VC concentrations exceeded groundwater screening criteria in one sample.

PCE concentrations ranged from 11 J µg/L (PM-GWS-6) to 150 J µg/L (PM-GWS-2). TCE concentrations ranged from 5.2 µg/L (PM-GWS-1A) to 16 µg/L (PM-GWS-2). *Cis*-1,2-DCE ranged from 11 µg/L (PM-GWS-1A) to 90 J µg/L (PM-GWS-2). VC was detected in one well at a concentration of 2.1 J µg/L. The maximum detections for all four compounds were in sample PM-GWS-2 which is located near the east side of the building, closest to MW-14S. These concentrations are not indicative of NAPL.

There isn't a correlation between the groundwater screening samples and the soil screening samples taken at this location which is expected since the samples were taken at different depths.

Other VOCs detected include 2-butanone, acetone, and carbon disulfide. No Samples were not analyzed for SVOCs, pesticides, PCBs, or metals.

#### **4.2.3.3 Monitoring Well Sampling**

In March 2012, CDM collected groundwater samples from the study area monitoring wells, as discussed in Section 2. The analytical results from the sampling event are in Table 4-5. Results for Monitored Natural Attenuation (MNA) parameters are presented and discussed in Section 5. As illustrated on Figure 4-4, PCE and *cis*-1,2-DCE were most frequently detected at concentrations above screening criteria in these monitoring wells. *Cis*-1,2-DCE was detected in more wells than PCE suggesting that degradation is occurring. TCE detection concentrations were also mostly above screening criteria. VC was the least frequently detected representative compound; however, most of the detections were above screening criteria.

PCE concentrations ranged from 5 µg/L to 100,000 µg/L. The highest concentrations were detected in wells screened between 24 and 35 ft bgs, which is the interval where ECD responses were observed during the MIP investigation. These are the "S" series wells indicating shallow screens. Concentrations greater than 1,500 µg/L (one percent of the solubility of PCE, e.g., indicative of the potential presence of NAPL) were detected in MW-14S (100,000 µg/L), MW-12S (71,000 µg/L), MW-11S (9,000 µg/L), MW-13S (2,500 µg/L), and MW-16S (2,200 µg/L). MW-14S is located near the east side of the site building. MW-11S, MW-12S, and MW-13S are located near the northeast corner of the building. MW-16S is located near the south end (the front) of the building. Concentrations in MW-14S and MW-16S have decreased since 2008. Concentrations in MW-11S, MW-13S, and MW-12S have increased. These increased concentrations may reflect migration of contamination, such as the movement of DNAPL under the force of gravity.

TCE concentrations ranged from 2.6 J µg/L to 2400 µg/L. The highest concentrations detected were located in MW-11S, MW-14S, and MW-13S. TCE concentrations have increased in these wells since 2008 which suggests degradation of PCE is occurring at the site.

*Cis*-1,2-DCE concentrations ranged from 1.3 J µg/L to 11,000 µg/L. The highest concentrations detected were located in MW-11S, MW-13S, and MW-01 which is located near the front of the site building and screened at the water table (shallow than the S series wells). Concentrations of *cis*-1,2-DCE in MW-11S and MW-13S have increased since 2008 which also suggests degradation of PCE is occurring at the site. MW-01 was not sampled in 2008.

VC concentrations ranged from 0.58 J µg/L to 1,200 JD µg/L. The highest VC concentration was detected in MW-01 which is a shallow well located near the southeast corner of the building.

No VOCs were detected in wells screened below 35 feet bgs (the deep D series wells) except for MW-15D which displayed a detection of *cis*-1,2-DCE and an exceedance of PCE with a concentration of 5 µg/L. This result is equal to the screening criteria, and much lower than concentrations seen in the shallow wells. In 2008, high PCE concentrations were detected in MW-13D and MW-15D. The lack of contamination in these wells in 2012 could be explained by the nature of NAPL. NAPL may have migrated from the shallow zone downward under the influence of gravity, and was potentially in the vicinity of the deep wells in 2008. Since then, either the NAPL has continued to sink and the most of

the residual contamination has since been transported by advection downgradient, the NAPL dissolved, or it has moved to an unsampled location.

Other VOCs detected above screening criteria include 1,1,2-trichloroethane, 1,1-DCE, and trans-1,2-DCE. The latter two compounds are degradation byproducts of PCE and are site related. VOCs detected below screening criteria include 1,2-dichlorobenzene, 1,4-dichlorobenzene, chlorobenzene, chloroform, chloromethane, ethane, ethene, methane, methyl tert-butyl ether, and toluene. It is not known if these are site related detections. Samples were not analyzed for SVOCs, metals, PCBs, or pesticides.

### 4.2.3 Air Investigation

Passive air sampling badges were placed in the bank building adjacent to the former Paul Miller site in March of 2011. Analytical results for the target compound, PCE, are presented on Table 4-7 and the laboratory data package is included as Appendix G. No PCE was detected above the detection limit of 1.0 µg/m<sup>3</sup>.

Since access was denied to the other nearby building, the restuaruant east of the former Paul Miller building, PCE in air concentrations are unknown in this building.

## 4.3 Discussion of Contamination

To gain further insight into the horizontal and vertical distribution of the PCE, TCE, cis-1,2-DCE, and VC contamination in groundwater, a series of cross sections was generated. A cross-section transect location map is provided in Figure 4-5. It should be noted that the points exhibited in the cross sections were projected onto the cross section baselines. As mentioned in Section 2, the groundwater

screening and MIP points encountered refusal at some locations. The shallow portion of the aquifer (above 35 ft bgs) consists of unconsolidated glacial till comprised of gravel, silty sands, silt, and clay. The deep aquifer consists of mostly silty sand and well-graded sand.

### 4.3.1 DNAPL

The first cross section, A to A' shown in Figure 4-6, runs north to south and cuts through the eastern portion of the site building. It is oriented in the direction of shallow groundwater flow. PCE concentrations indicative of the presence of DNAPL were detected near the south end and the northeast corner of the building. These two locations are suspected source areas. The highest PCE concentrations were detected in MW-14S near the east side of the building. Concentrations decreased exponentially directly downgradient from this well but concentrations indicating the presence of DNAPL were observed in the two wells closest to the northeast corner of the building and MW-14S. MW-01, MW-16S, and MIP6 are upgradient from this source location. However, the contaminant concentrations in these wells/boreholes are indicative of the potential for a second source location near the front of the building.

A second cross section, B to B' shown in Figure 4-7, runs parallel to the A to A' cross section along the west side of the building. The shallow groundwater screening sample at MIP5 displayed NAPL-like concentrations. This is also indicative of the potential for an additional source.

### 4.3.2 Dissolved Phase Contamination

Concentrations indicative of dissolved phase PCE contamination were observed downgradient of the suspected source locations in MW-08S (downgradient of the second suspected source location) on

cross section B-B' and MW-09S in cross section C-C', Figure 4-8, and MW-10S in cross section A-A'. MW-03, displayed in cross section D-D' and on Figure 4-9, located near the northwest corner of the building did not contain high concentrations of contaminants; however, this well was sampled at approximately 15 ft amsl which is above the general depth interval where contamination in this area is observed. The ECD readings and a soil sample taken from a deeper depth in this area give indication of contamination at depths below the sampled depth. Dissolved phase contamination was also observed in the subslab groundwater screening samples. However, there is much uncertainty about the contamination under the building since some boreholes hit refusal at very shallow depths and due to the potential for an additional slab. The highest concentrations of TCE and cis-1,2-DCE occur at MW-11S located near the northeast corner of the building; the highest VC concentration occurs at MW-01 which is located near the south end of the building. The presence of DNAPL at the site will continue to cause widespread dissolved phase PCE contamination as it migrates, potentially leading to higher concentrations of daughter products as it degrades.

#### 4.3.3 Extent of Contamination

The horizontal distribution of contamination in the shallow zone above 35 feet bgs is illustrated on Figure 4-10. The MIP and monitoring well investigation results indicate that contamination extends from MIP5 and MIP6 in the south area of the site to MIP5B to the west, MW13S to the east, MIP1 to the northwest, MIP2 to the north, and MW10S to the northeast. The extent of contamination has been delineated with either groundwater sample results below SCGs or MIP results indicating a lack of contamination in the four directions around the site building. It should be noted that MIP readings were used to discern the presence or absence of contamination in areas where there are not any wells. However, since the MIP tool is qualitative and does not provide quantitative measurements of contaminant concentrations like groundwater samples do, the delineation provided by these locations is an estimate.

The vertical distribution of contamination in the south and north areas differs. In general, the greatest levels of contamination are observed in the shallow aquifer, above 35 ft bgs (-10 ft amsl). In the south area of the site, contamination was observed at approximately 15 ft amsl and -2 ft amsl with the higher concentrations being detected at the shallower depth. In the north area of the site, near the back of the building, contamination is observed in the 5 ft amsl to -10 ft amsl interval. Contamination in the deep wells was not detected or detected at low concentrations.

The EC probe readings and boring log information indicate a greater percentage of sand below approximately -12 ft amsl. The previous RI (2009) reported high concentrations of contaminants in this zone. The more conductive deposits in this zone could explain the lack of identified contamination. Comparing the ECD readings, groundwater screening results, and groundwater results to the soil information, it is suggested that any contamination below this elevation dilutes and disperses more quickly due to the higher hydraulic conductivity.

### 4.4 Qualitative Human Health Risk Assessment

Utilizing the analytical results collected during the current investigation, the following exposure pathways were assessed:

## Exhibit 4-A. Qualitative Human Health Risk Assessment

Environmental Media & Exposure Route	Human Exposure Assessment
Direct contact with surface soils (and incidental ingestion)	People are not coming into contact with surface soils because contaminated soils are covered by pavement and concrete.
Direct contact with subsurface soils (and incidental ingestion)	Contaminated soils are present from the surface to the water table. Proper health and safety practices should be followed to limit potential exposure during ground-intrusive work at the site.
Direct contact with groundwater ( including ingestion of groundwater)	Contaminated groundwater is not being used for domestic uses including drinking water, as the area is served by public water supply. Contaminated groundwater is present at approximately 3 to 9 feet bgs, therefore human exposure is possible during intrusive activities.
Inhalation of air (exposures related to soil vapor intrusion)	The 2009 RI identified indoor air concentrations of PCE and PCE degradation products at concentrations exceeding indoor air quality criteria (CDM, 2009). Analysis has concluded that soil and groundwater contamination from beneath the building are not a significant contributing factor to indoor air quality in the on-site structure. Indoor air quality appears to be impacted by contaminated building structure materials, rather than vapor intrusion from the environment. No mitigation measures have been installed to date.

Based on analytical results, an exposure risk exists for soil, groundwater, and air. Proper health and safety practices should be followed to limit potential exposure.

## Section 5

# Contaminant Fate and Transport

This section examines the primary chemical and physical processes that affect the fate and transport of the contaminants in groundwater and soils at the Site. The focus is on the four indicator contaminants at the site: PCE, TCE, *cis*-1,2-DCE, and VC. An understanding of the fate and transport of the indicator contaminants aids the evaluation of potential current and future exposure risks and focuses the evaluation of remedial technologies in the feasibility study. This section provides the following:

- A summary of the indicator contaminants and corresponding chemical and physical properties
- A discussion of processes that affect the fate of the indicator contaminants in the environment
- A discussion of processes that affect transport potential of the indicator contaminants
- A discussion of transport mechanisms and pathways
- An evaluation of natural attenuation
- Summary of conceptual site model

## 5.1 Indicator Contaminants

Indicator contaminants were selected based on a review of the analytical data collected during the RI, analysis of the spatial distribution of contamination, and the historical data for the Site. Hence, in this section the fate and transport of the following indicator contaminants are examined:

- PCE
- TCE
- *cis*-1,2-DCE
- VC

## 5.2 Chemical and Physical Properties of Indicator Contaminants

To predict the fate, or persistence and potential transport of indicator contaminants that are present in soil, and groundwater, it is necessary to identify which contaminants are likely to migrate or degrade. These processes depend on a given chemical's physical and chemical properties and the properties of the media through which it migrates. **Table 5-1** presents the chemical and physical properties of the indicator contaminants. The properties are defined in the following paragraphs and further discussed in the next section.

The solubility of a chemical is defined as the upper limit of its dissolved concentration in water at a specified temperature. Concentrations in excess of solubility may indicate a co-solvent effect or the

presence of a non-aqueous phase liquid. As shown on **Table 5-1**, all VOCs have relatively high water solubilities: 150 mg/L for PCE, 1,366 mg/L for TCE, 3,500 mg/L for cis-1,2-DCE, and 2,760 mg/L for VC.

Henry's Law constant provides a measure of the extent of chemical partitioning between air (vapor phase) and water (dissolved phase) at equilibrium. The higher the Henry's Law constant, the more likely a chemical is to volatilize. All of the indicator contaminants have Henry's Law constants greater than  $10^{-3}$  atmosphere-m<sup>3</sup>/mole (atm-m<sup>3</sup>/mol), which indicates they will volatilize from water.

Vapor pressure is the pressure exerted by a chemical vapor, at any given temperature, in equilibrium with its solid or liquid form. It is used to calculate the rate of volatilization of a pure substance from a surface or to estimate a Henry's Law constant for chemicals with low water solubility. The higher the vapor pressure, the more likely a chemical is to exist in a gaseous state. All indicator contaminants have relatively high vapor pressure, ranging from 18.47 (PCE) to 2,600 millimeters (mm) of mercury (Hg) (VC), which indicates that these VOCs will evaporate rapidly from the near-surface soil.

The organic carbon partition coefficient ( $K_{oc}$ ) provides a measure of the extent of chemical partitioning between organic carbon and water at equilibrium. The higher the  $K_{oc}$ , the more likely a chemical is to bind to soil or sediment rather than to remain dissolved in water. The range of  $K_{oc}$  values for indicator contaminants is from 15.4 L/kg (VC) to 265 L/kg (PCE), which indicates that these indicator contaminants have a relatively low potential to bind to soil or sediment and are mobile in water.

The soil-water partition coefficient ( $K_d$ ) provides a soil- or sediment-specific measure of the extent of chemical partitioning between soil or sediment and water, adjusted for dependence upon organic carbon.  $K_d$  is adjusted using the fraction of organic carbon ( $f_{oc}$ ) of the soil/sediment as shown in the formula  $K_d = K_{oc} \times f_{oc}$ . A higher  $K_d$ , indicates that a chemical is more likely to bind to soil or sediment rather than to remain in the dissolved phase, thereby reducing its transport capability. As no data were collected at the Site for fraction of organic carbon, one can either use literature  $K_d$  values or assume an  $f_{oc}$  of 0.1 percent (0.1%), which is typical for coastal plain environments. The literature  $K_d$  values are presented in **Table 5-1** and range from 0.071 cm<sup>3</sup>/g (cis-1,2-DCE) to 0.33 cm<sup>3</sup>/g (TCE), which indicate low adsorption for these indicator contaminants. The calculated  $K_d$  values based on a 0.1%  $f_{oc}$  were even lower than the literature  $K_d$  values, also suggesting the low adsorption for these indicator contaminants.

The octanol-water partition coefficient ( $K_{ow}$ ) provides a measure of the extent of chemical partitioning between water and octanol at equilibrium. The greater the  $K_{ow}$ , the more likely a chemical is to partition to octanol rather than to remain in water. Octanol is used as a surrogate for lipids, and  $K_{ow}$  is used to predict bioconcentration in living organisms. All indicator contaminants have relatively low  $K_{ow}$  indicating that they have low potential to bioconcentrate in living organisms.

## 5.3 Environmental Fate

Contaminant fate describes the length of time that a contaminant will remain in its original chemical state in the environment. Chemicals that persist in a given medium are those that form insoluble precipitates, or resist biodegradation, hydrolysis, and volatilization. The fate of metals depends on the partitioning between soluble and in-soluble particulate solid phases. Partitioning is affected by adsorption, precipitation, co-precipitation, and complexation. These processes are governed by pH,

Eh, ionic strength of the water, concentration of the complexing ions, and the concentration and type of metals.

### 5.3.1 Processes that Affect Fate and Transport

The major processes that affect the fate, or persistence, of the indicator contaminants are volatilization, degradation, hydrolysis, dissolution, and precipitation. The most persistent chemicals are those that form insoluble compounds, precipitate, or do not hydrolyze or biodegrade.

**Dissolution** - Dissolution is the process of dissolving, changing, or separating a substance into component parts or changing it from a solid to a fluid state. Mechanisms that cause or enhance dissolution include solution by heat, moisture liquefaction, melting, or decomposition.

**Precipitation** - In chemistry, precipitation is the condensation of a solid from a solution. This occurs when the solution is saturated, whereupon the solid forms, and usually sinks to the bottom of the solution. Contaminants are converted to an insoluble form (particle) by the chemical reaction with precipitating reagents, the particles formed by this reaction are then removed from solution by settling and/or filtration.

**Hydrolysis** - Hydrolysis is a chemical decomposition process that uses water to split chemical bonds of substances. There are two types of hydrolysis, acidic and enzymatic. Hydrolysis occurs in certain inorganic salts in solution, in nearly all non-metallic chlorides, in esters, and in other organic substances.

**Biodegradation** - Biodegradation is the breakdown of organic contaminants by microbial organisms into smaller compounds. The microbial organisms transform the contaminants through metabolic or enzymatic processes. Biodegradation processes vary greatly, but frequently the final product of the degradation is carbon dioxide or methane. Biodegradation can occur under aerobic conditions, where oxygen is present in sufficient concentration, or under anaerobic conditions, where oxygen is lacking.

**Volatilization** – Volatilization is the conversion of a chemical compounds from a solid state or liquid phase to vapor phase, by application of either heat and/or reducing pressure to overcome the binding force within the original state.

**Matrix Diffusion** – Matrix diffusion is the general term used to describe a set of physical process that cause the movement of contaminants from groundwater or non-aqueous phase liquid (NAPL) into the pore space of low-permeability matrices such as clay, tight silts, or bedrock. A concentration gradient between contaminants dissolved in groundwater and pore water in the matrix can drive contaminants into the matrix. The movement of a NAPL into the matrix could occur by advection or by matric potential. Diffusion can also occur in the solid phase, though much more slowly, for example by diffusion below the surface in a carbon film on a sand grain. This process can contribute to retardation of plume migration. Back diffusion of contaminants from the matrix can be an ongoing source of groundwater contamination.

### 5.3.2 Fate of Indicator Contaminants

The fate of VOCs is dictated by their volatility and degradation. Their presence in surface soils is usually short-lived provided that there is no continuous source. The fate of the indicator contaminants is discussed below.

**Tetrachloroethene** - The dominant fate of PCE that may be present in soils is volatilization. Volatilization is also an important fate process of PCE in groundwater based on its Henry's Law constant of  $1.73 \times 10^{-2}$  atm-m<sup>3</sup>/mol.

Based on its K<sub>oc</sub> value of 265 milliliters per gram (mL/g), PCE is moderately mobile in soils. Consequently, PCE has the potential to migrate through the soil into groundwater. PCE has a specific gravity greater than water (1.62) indicating that pure liquid phase PCE will sink when dissolved in groundwater. The solubility of PCE in water is 150 milligrams per Liter (mg/L).

Chlorinated hydrocarbon compounds, such as PCE and TCE, can be progressively dechlorinated via reductive dechlorination, and ultimately degrade to innocuous end products such as carbon dioxide, ethane, ethene, and water under appropriate subsurface conditions. This process will be discussed in detail in Section 5.5.

The widespread PCE contamination in groundwater indicates that it is a primary contaminant at the Site.

**Trichloroethene** – The dominant fate of TCE that may be present in vadose zone soils is volatilization. Volatilization can be an important fate process of TCE in groundwater based on its Henry's Law constant of 0.011 atm-m<sup>3</sup>/mol, especially TCE near the top of the water table.

Due to its high mobility in soils (i.e., high solubility - 1,366 mg/L, low K<sub>oc</sub> value etc.), TCE is less likely to bind to soil, and thus has the potential to migrate through the soil into groundwater. TCE has a specific gravity greater than water (1.465), indicating that pure liquid phase TCE will sink when dissolved in groundwater.

Under anaerobic conditions, as might be seen in soil microsites, flooded soils, or within aquifers, TCE is slowly biodegraded via reductive dechlorination (refer to the PCE degradation pathway); however the extent and rate of degradation are dependent upon the strength of the reducing environment, as often observed with the light non-aqueous phase liquid (LNAPL) presence. Biodegradation in soil and groundwater may occur at a relatively slow rate with half-lives on order of months to a year (Lucius *et al.* 1990).

The widespread TCE contamination in groundwater indicates that it is a primary contaminant at the Site. Its presence is likely due to the degradation of PCE.

**cis-1,2-Dichloroethene** - The dominant fate process of *cis*-1,2-DCE in soil and groundwater is volatilization, because of its high vapor pressure (180 mm Hg) and Henry's Law constant ( $4.1 \times 10^{-3}$  atm-m<sup>3</sup>/mol).

Based on a K<sub>oc</sub> value of 43.8 L/kg, little adsorption to soil is expected for *cis*-1,2-DCE (ATSDR 1996). Without significant adsorption to soil, *cis*-1,2-DCE can leach into groundwater where biodegradation should occur (Hazardous Substance Database [HSDB] 2005). The presence of *cis*-1,2-DCE in groundwater, especially under sandy soil conditions (Barber *et al.* 1988), substantiates its leachability. The relatively low K<sub>oc</sub> and high vapor pressure of *cis*-1,2-DCE indicate that this compound should also readily volatilize from moist soil surfaces (ATSDR 1996). *Cis*-1,2-DCE undergoes slow reductive dechlorination under anaerobic conditions (Fogel *et al.* 1986, ATSDR 1996).

In the reductive dechlorination process, *cis*-1,2-DCE is a breakdown product of the PCE spilled at the site. Under the proper conditions, *cis*-1,2-DCE degrades to VC. The accumulation of *cis*-1,2-DCE in groundwater at the Site indicates that more *cis*-1,2-DCE is being produced than is being degraded.

**Vinyl Chloride** – VC in soil, and in shallow groundwater, can volatilize because of its high vapor pressure (2,600 mm Hg) and Henry's Law constant ( $2.78 \times 10^{-2}$  atm-m<sup>3</sup>/mol). Vinyl chloride is an anaerobic biodegradation product of higher chlorinated VOCs such as PCE.

Vinyl chloride is soluble in water and thus can leach through the soil and enter groundwater before evaporation can occur (ATSDR 2006). A K<sub>oc</sub> value of 15.38 suggests a very low sorption tendency, meaning that this compound would be highly mobile in water. The literature K<sub>d</sub> value of 0.037 cm<sup>3</sup>/g indicates that VC would be highly to moderately mobile in soil. Thus, VC has the potential to leach into groundwater as shown by its historical and frequent detection in the Site groundwater. Vinyl chloride is degraded by both anaerobic and aerobic degradation processes.

## 5.4 Contaminant Transport Mechanisms

Contaminant transport pathways and exposure mechanisms provide the ways for chemicals in contaminated media at the Site to travel from the area of deposition and to potentially leave the property.

**DNAPL Migration** - The Site is underlain by unconsolidated glacial deposits consisting of unsorted sands, silts, clays, gravel and boulders. Site-related contaminants released to the subsurface will migrate downward through the aquifer. If the quantity of solvent reaching the water table is sufficient, some of it can remain in an undissolved state and continue to move downward under the influence of gravity until a physical barrier to flow is encountered, such as a boulder or clay lens.

DNAPL can seep into the pore spaces of silts and clay, where it can reside for long durations. DNAPL accumulated in sands would be exposed to great volumes of water flowing by, which would help dilute it. However, the conductivity of clay, boulders, and silt layers are much lower than that of sands. DNAPL trapped in the pore spaces of these lower permeability layers would not attenuate as rapidly. Instead, it would diffuse, adsorb, and continue to migrate through the less permeable materials and along the surfaces of the layers.

**Leaching to Groundwater** - The entire Site is developed with buildings, asphalt, or concrete. As such, rainwater is collected in the stormwater system, and only a minimal amount infiltrates the subsurface (through cracks in the pavement etc.). Therefore, leaching of contamination into percolating stormwater is expected to be minimal.

**Groundwater Transport** - All of the indicator contaminants exhibit relatively high water solubility and/or low K<sub>oc</sub> values and, therefore, high leachability. Contaminants in groundwater can be transported in either a dissolved or particle-sorbed state. The lithology along with the general topography, is the primary influence on the physical behavior of groundwater. Potential migration/exposure mechanisms for groundwater contaminants include groundwater use. Groundwater transport processes are detailed in Section 5.4.2.

**Volatilization** - Volatilization is a minor transport process for VOCs and at the Site would have occurred mainly from surface soil contaminants. Subsurface contaminants at or above the water table may also volatilize. Since contaminants were not detected in subsurface soils at or above the water table at concentrations above their respective screening criteria, volatilization is not expected to be a

significant transport mechanism from onsite soils. However, accumulation of vapors can occur underneath these structures and pose risk to human health via vapor intrusion (i.e., discharge into on-site buildings), due to the presence of asphalt parking area and building foundations in portion of the Site which essentially act as a cap. The majority of indoor air contaminant levels in the on site building have been attributed to contaminated building structure materials, and not to soil or groundwater contamination.

**Advection** - Advection describes the process of solute migration, which due to the average bulk movement of groundwater, is typically the most important factor governing the transport of contaminants in groundwater. Advection defines the direction and velocity of a plume's center of mass. The advective transport term is computed using velocities determined by solving the groundwater flow equation, which is a function of hydraulic conductivity, hydraulic gradient, and flow cross-sectional area. Average linear groundwater velocity ( $v$ ) is a function of hydraulic conductivity, hydraulic gradient, and effective porosity ( $\eta$ ). Effective porosity values typically fall within the range of values of specific yield and total porosity. Specific yield (i.e., the amount of water released from storage per unit drop in piezometric head) represents the lower limit of reasonable effective porosity values.

**Dispersion and Dilution** - Dilution consists of a series of phenomena, including groundwater mixing (i.e., flushing with uncontaminated or less contaminated groundwater) and hydrodynamic dispersion, which play an important role in naturally attenuating groundwater contamination in a non-destructive manner, as opposed to the natural attenuation specifically via biodegradation that will be discussed later in this section.

Specifically, hydrodynamic dispersion describes the spread of contaminants around an average groundwater flow path, beyond the region they would normally occupy due to advection alone. Hydrodynamic dispersion is the sum of two processes, mechanical dispersion and molecular diffusion. Mechanical dispersion results from mixing that occurs as a consequence of local variations in groundwater velocity and the aquifer's matrix. Molecular diffusion results from variations in solute concentrations within the groundwater system. In general, except when groundwater velocities are very low (e.g., a few centimeters a year), this diffusion effect is generally secondary to, and often negligible, compared to the mechanical dispersion effect. However, at the Site, the diffusion effect is an important factor in the migration of contaminants into tight silts and clays at the site.

**Retardation** - Dissolved-phase contaminant transport velocity is generally retarded in comparison to groundwater by adsorption processes. The degrees of adsorption and retardation are typically controlled by soil/water partitioning relations, which are a function of the chemical-specific partition coefficient and the soil organic carbon content. A typical method of generally describing this phenomenon in solute transport evaluations is by using a retardation factor.

When contaminated soil and groundwater are in contact, dissolved chemicals partition between them according to soil/water partitioning relations. Some of the chemical mass is irreversibly adsorbed. However, much of the mass is able to adsorb or desorb over time to maintain equilibrium mass balance. In general, adsorption occurs faster than desorption. For this reason, adsorption is commonly associated with retarding chemical transport (e.g., chemical transport velocity is slower than groundwater velocity), whereas desorption is commonly associated with chemical persistence (e.g., contaminated soils leachability to groundwater, attenuating groundwater concentration trends exhibit a "tailing" effect).

Due to the lack of site-specific data such as soil organic carbon content and effective porosity, retardation was not calculated for the indicator contaminants. However, for plumes characteristic of the VOC contaminants encountered at the Site, the contaminant's mass moves at essentially the same rate as the average linear groundwater velocity.

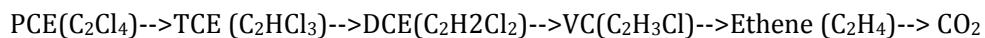
## 5.5 Natural Attenuation of VOCs in Groundwater

A preliminary assessment to evaluate the natural attenuation potential of the Site was performed. The natural attenuation evaluation consists of evaluating the available historically detected concentrations of the contaminants which are associated with the degradation of PCE, including TCE, cis-1,2-DCE, and VC in conjunction with the natural attenuation indicator parameter data, for evidence indicating potential occurrence of biodegradation processes.

### 5.5.1 Background on Natural Attenuation Processes

Natural attenuation refers to all of the naturally occurring processes (biodegradation, and other abiotic processes such as dispersion, sorption, volatilization etc.) that affect the fate and transport of contaminants in soil and groundwater, and achieving a reduction in the total mass, toxicity, mobility, volume, or concentration of a contaminant in soil and groundwater. Often misconstrued as "do-nothing", under proper conditions, these processes can be effective in containing and remediating such contamination in a time frame comparable to other alternatives.

Chlorinated ethene compounds, such as PCE and TCE, can be progressively dechlorinated via reductive dechlorination. If sufficient electron donors and appropriate microorganisms are present, complete reductive dechlorination of the highly chlorinated compounds may occur. During this process, the chlorine atoms are replaced by hydrogen, resulting in compounds with reduced carbon and less chlorine. Therefore, the chlorinated solvent is acting as an electron acceptor and needs other carbon substrates to act as electron donors. As such, PCE degrades to TCE which subsequently degrades to DCE, with the cis isomer (cis-1, 2-DCE) predominant over the trans isomer, then VC and, ultimately, ethene and carbon dioxide as shown below (Chapelle 1993; Wiedemeier *et al.* 1998). The complete degradation of PCE and TCE is favored by sequential anoxic/oxic conditions (Chapelle 1996).



Biodegradation of chlorinated hydrocarbon compounds may occur through three different pathways, which are described as below:

**Electron Acceptor Reactions** – this process involves a reductive dechlorination reaction, during which the chlorinated hydrocarbon compound serves as electron acceptor, and a chlorine atom is replaced by a hydrogen atom. Concurrent with the degradation of the chlorinated hydrocarbon compound, the accumulation of less chlorinated daughter products and an increase of chloride ions will occur. Generally, the more chlorinated compounds are more susceptible to reductive dechlorination. For instance, TCE is the more susceptible to reductive dechlorination, followed by cis 1,2 DCE and then vinyl chloride (USGS, 2002). Reductive dechlorination can occur under a range of reducing conditions, however, it occurs more rapidly under sulfate reducing and methanogenic conditions, as opposed to nitrate reducing and iron reducing conditions (Bouwer 1994). Sufficient source of electron donors must be available to sustain microbial activity. This could be anthropogenic carbon such as petroleum hydrocarbons (i.e., BTEX etc.), landfill leachate rich in organic content, or natural organic matter.

**Electron Donor Reactions** – This process involves the transfer of electrons either from less chlorinated hydrocarbon such as vinyl chloride under aerobic and some anaerobic conditions. The reactions provide energy for microorganism growth and reproduction. It is generally believed that PCE and TCE do not participate in such reactions as microorganisms cannot gain enough energy to sustain growth. As daughter products of PCE and TCE degradation (cis 1,2 DCE and vinyl chloride) occurring at source area (often associated with more reducing conditions) are transported downgradient to less reducing conditions or even aerobic conditions by groundwater flow, they can be degraded through this pathway.

**Cometabolism** - This process involves the fortuitous degradation of a chlorinated ethenes catalyzed by an enzyme or cofactor that is produced by microorganisms for other purposes.

### 5.5.2 Natural Attenuation Evaluation

The US Department of the Navy document, *Technical Guidelines for Evaluating Monitored Natural Attenuation of Petroleum Hydrocarbons and Chlorinated Solvents in Ground Water at Naval and Marine Corps Facilities* (Navy 1998), provides guidelines for assessing the potential for natural attenuation at Sites. The USEPA document (1998) *Technical Protocol for Evaluating Natural Attenuation of Chlorinated Solvents in Ground Water* is also useful. The following information helps to evaluate the occurrence of natural attenuation.

- Change in contaminant mass over time
- Presence/absence of degradation byproducts
- Suitable geochemical (ReDox) conditions
- Available carbon (e.g., electron donor)

During the March 2012 sampling event, natural attenuation indicator parameters were collected and analyzed in order to evaluate whether the subsurface conditions are conducive to in-situ natural degradation of the chlorinated hydrocarbon compounds over time. The natural attenuation indicator parameters that were collected from all the monitoring wells include pH, specific conductivity, DO, temperature, oxidation reduction potential (Eh/ORP), and ferrous ion. The natural attenuation indicator parameters that were collected and sent off-site for laboratory analysis include alkalinity as calcium carbonate, nitrate/nitrite, chloride, sulfate, TOC, MEE.

Table 5-2 provides a summary of the natural attenuation indicator parameters collected from monitoring wells for analysis and an explanation of each parameter's significance. Concentrations of the indicator compounds are also listed to show the presence/absence of daughter products. The concentrations are presented by moles instead of by weight; this allows a direct evaluation of the amount of degradation that has occurred.

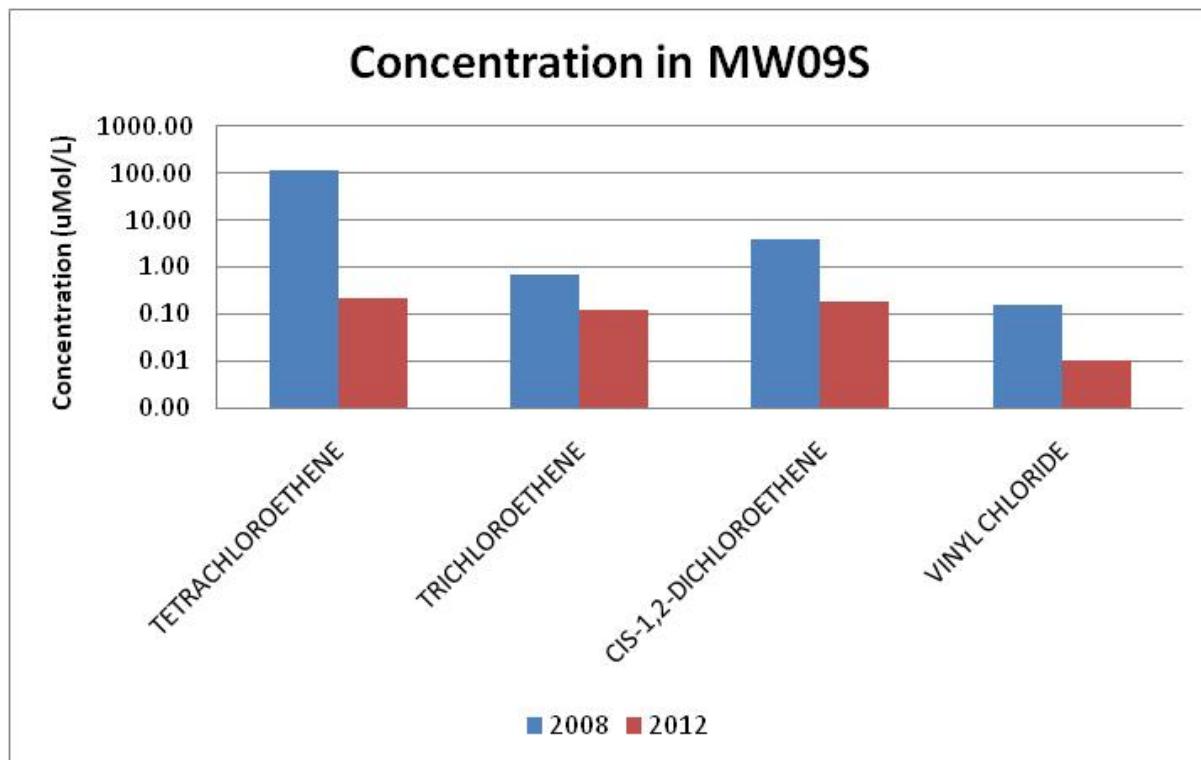
Table 5-3 presents the results of the natural attenuation analysis in different zones of the site. Degradation products of PCE were detected in all the wells where contamination was detected, indicating that at least some degree of reductive dechlorination of PCE has occurred in the past. ReDox conditions varied across the site and according to depth. In general, the most favorable conditions for the reductive dechlorination of PCE and TCE (methanogenic conditions) were identified

in the very shallow wells, MW01, 02, 03, and 04. The remaining wells showed either aerobic or nitrate reducing conditions, redox conditions that are considered unfavorable for natural attenuation of PCE and TCE, but where cis-1,2-DCE and VC are known to be slowly degradable.

Reductive dechlorination requires an organic carbon substrate to be effective. At the site, there is a general relationship evident between the amount of total organic carbon measured and methanogenic conditions. The very shallow wells show high organic carbon and methanogenic conditions, whereas no organic carbon was detected in the deep wells, and more oxidizing conditions are dominant. This indicates that the presence of measured total organic carbon is a proxy for suitable conditions for reductive dechlorination. There is generally a low amount of organic carbon in groundwater at the site. Of particular concern is the lack of measurable carbon in the downgradient well MW09S. The lack of available carbon could limit the future microbial reductive dechlorination of contaminants at the site.

Another line of evidence for natural attenuation is to verify if the plume has been expanding over time. Exhibit 5-A, below, shows the molar concentrations of the indicator contaminants as measured in 2008 and in 2012 (log scale) in the downgradient well MW09S. All four of the indicator compounds showed a decrease in concentration of between one and three orders of magnitude. It should be noted that just two monitoring rounds is too few to fully evaluate trends in concentration. Furthermore, volatilization, dilution, and dispersion can also cause effective decreases in concentration over time, as well as abiotic degradation of VOCs. It is unknown what contribution these other attenuation mechanisms are making to mass reduction.

Exhibit 5-A. Change in Concentrations in MW09S between 2008 and 2012



### 5.5.3 Summary of the Evaluation of Natural Attenuation

Across the site, the presence of cis-1,2-DCE and VC indicates that dechlorination of PCE has at least occurred at some point in the past. Based on the 2012 round of data, it appears that the more favorable conditions for natural attenuation of PCE and TCE are present in the shallowest parts of the aquifer. Moving deeper into the aquifer and also downgradient, conditions become more oxidizing. This means that conditions are not very suitable for microbial degradation of the parent products, but potentially the daughter products cis-1,2-DCE and VC can be degraded in these areas to non-toxic byproducts.

The relative lack of organic carbon and the less reducing conditions in the more contaminated depths of the aquifer indicate that further microbial reductive dechlorination of PCE and TCE could be limited. Given the finite amount of carbon in the pre-release subsurface, one would expect declining rates of attenuation over time as the available carbon was utilized. Dilution and dispersion also lead to decreasing concentrations. Volatilization and abiotic degradation are other attenuation mechanisms that may be causing the observed concentration reductions.

The monitoring data from 2008 and from 2012 indicate that dissolved phase concentrations in subsurface areas lateral, downgradient, and vertical to the source area are decreasing. More data would be needed to determine if these decreasing concentrations are a temporal trend.

## 5.6 Conceptual Site Model

### 5.6.1 Physical Setting

Groundwater at the site is found at an elevation of approximately 19 feet amsl, which corresponds to a relatively thin vadose zone of between three and nine feet thick. The geology at the site is characterized by a heterogeneous mix of soil types above 35 feet bgs, and a relatively homogenous sand or silty sand below 35 feet bgs. The top heterogeneous unit consists of glacial deposits of various permutations of clay, silt, and sand. The unit has relatively low hydraulic conductivity (less than 0.5 feet per day). Due to its more sandy composition, the unit below 35 feet bgs has much higher hydraulic conductivity (greater than 30 feet per day). In the shallow stratum, groundwater flows in a north/northwestern direction from the former Paul Miller building; in the deeper stratum, groundwater flows to the northeast, eventually discharging into Kill Van Kull approximately one mile distant. Figures 3-1 through 3-4 display the general geology of the site in cross section and the potentiometric surfaces.

### 5.6.2 Contaminant Sources, Migration Pathways, and Fate

High concentrations of VOCs indicative of DNAPL (e.g., aqueous concentrations greater than 1% of the solubility of PCE) were found at multiple locations around the former Paul Miller Dry Cleaners building. This data indicates that waste PCE from the dry cleaning facility may have been disposed of onto the ground in multiple locations under and around the building.

Once in the subsurface, much of the PCE moved by gravity as DNAPL downward through the vadose and saturated zones. Further downward NAPL travel was impeded by locally stratified and less permeable clays and tight silts in the glacial deposits in the upper 35 feet.

NAPL accumulated on top of and between these deposits and is likely still present (as confirmed by the S series monitoring wells and MIP results). NAPL also spilled over the edges of the deposits and continued to travel deeper into the aquifer. Over time, the following processes have likely occurred:

- a portion of the contaminant mass diffused into the clay and silt matrices in the top 35 feet;
- a portion volatilized and rose into the vadose zone and to the surface;
- a portion was diluted by groundwater in the top 35 feet; and
- a portion continued to travel downward into the more homogenous sand below 35 feet, and potentially down to bedrock.

The sum of these actions has resulted in a decrease in the volume of DNAPL in the subsurface below the points of disposal over time, and distribution of dissolved phase mass across different geological matrices.

Since the geology is heterogeneous in the top 35 feet, dissolved phase contamination migrates along preferential pathways with higher hydraulic conductivity. Dilution of the residual DNAPL and back-diffusion of dissolved contaminants from the silt and clay matrices are serving as continual sources of dissolved phase groundwater contamination. The observed heterogeneity means that the mass flux at different elevations in the upper 35 feet of the subsurface and rates of diffusion into and out of the less permeable soils will vary considerably.

DNAPL that continued downward has likely been diluted by groundwater in the sandy stratum. Groundwater travels northeast in this stratum. Dissolved contamination will be carried downgradient by advection. Minimal retardation is expected given the low expected fraction organic carbon in this sandy unit. The sample results from the well immediately downgradient of the Paul Miller building (MW-13D) reinforce this assertion: 6,300 JD ug/L of PCE in 2008 and non-detect in 2012, indicating that mass is not being retained in the unit. It is also possible that DNAPL has sunk further until another low-permeability layer such as bedrock was encountered. Bedrock is expected in this area at a depth of over 100 feet bgs. No wells were screened on top of bedrock, and thus the occurrence of DNAPL here is unknown.

Biodegradation has occurred to varying degrees across the site. The presence of degradation byproducts of PCE indicate that microbial degradation has occurred at some point in the past. The measured geochemistry of the site indicates that the most favorable conditions for natural attenuation of PCE and TCE are present in the shallowest parts of the aquifer (methanogenic redox conditions and available carbon). Moving deeper in the glacial deposits and into the deeper sand stratum, conditions become more oxidizing and less conducive to microbial degradation. Further biodegradation is potentially limited at the site by the lack of available carbon for microbial growth. Overall, the sample results in 2012 appeared to show a decrease in concentrations across most of the wells compared to 2008 data. This observed reduction is likely due to a combination of biodegradation and dilution/dispersion.

A neighboring site, the former Charlton Cleaners, also has a known PCE contamination problem in groundwater. The former Charlton Cleaners building is approximately 300 feet northeast of the former Paul Miller drycleaners building. Based on the Charlton Cleaners RI (LBG 2006), the southwestern extent of Charlton Cleaners contamination does not appear to be fully delineated, especially in the heterogeneous soil above 35ft bgs. The 30 foot bgs potentiometric surface for the

Paul Miller site (Figure 3-3) indicates that groundwater from the Charlton Cleaners may be flowing in part towards Paul Miller. The two PCE plumes may potentially comingle in the immediate vicinity northeast of the Paul Miller site near well MW-10S. The potential plume commingling area is shown on Figure 4-10.

### 5.6.3 Receptors

No users of groundwater potentially impacted by releases from the former Paul Miller Dry Cleaners have been identified. The 2009 RI (CDM, 2009) identified indoor air and sub-slab vapor concentrations that warrant mitigation. Workers and customers of the current site tenant are therefore potential receptors.

## Section 6

# Conclusions and Recommendations

This section presents the conclusions and recommendations, which are based upon the analytical results of the soil and groundwater sampling and the MIP investigation.

## 6.1 Conclusions

### 6.1.1 Soil

Based upon the results of this investigation, contamination in soil is not the primary concern at the site. No residual source materials were identified in the vadose zone that could be still contributing to the groundwater contamination at the site. PCE and its associated breakdown products were not detected above the Unrestricted Use SCGs in any soil samples collected from above the water table.

### 6.1.2 Groundwater

Groundwater concentrations exceeding screening criteria at the site consist primarily of PCE and its associated daughter products: TCE, cis-1,2-DCE, and VC. Concentrations detected in samples from monitoring wells and groundwater screening samples east, south, and west of the building indicate the potential presence of NAPL.

The horizontal distribution of dissolved phase contamination in the shallow zone above 35 feet bgs is illustrated on Figure 4-10. The MIP and monitoring well investigation results indicate that contamination extends from MIP5 and MIP6 in the south area of the site to MIP5B to the west, MW13S to the east, MIP1 to the northwest, MIP2 to the north, and MW10S to the northeast. The extent of contamination has been delineated with either groundwater sample results below SCGs or MIP results indicating a lack of contamination in the four directions around the site building. It should be noted that MIP readings were used to discern the presence or absence of contamination in areas where there are not any wells. However, since the MIP tool is qualitative and does not provide quantitative measurements of contaminant concentrations like groundwater samples do, the delineation provided by these locations is an estimate. While the quality of data and degree of estimation is sufficient to conclude that the extent of contamination on and off site has been adequately defined for the purposes of developing remedial alternatives, additional monitoring well installation may be considered during the remedial design if additional quantitative data is deemed necessary.

There was only one detection of PCE at 5 µg/L in the deep wells screened in the sandy stratum below 35 feet bgs. It is likely that due to the high transmissivity of this unit, dissolved phase contamination entering the sandy stratum is diluted and dispersed quickly. A MIP result (MIP-4C) northeast of the building indicated potential contamination in this stratum. However, the Charlton Cleaners site is in this area; the Paul Miller plume and the Charlton Cleaners plume may potentially be commingling in this area (Figure 4-10). Based on the current and historic distribution of contamination at the Site and the transmissivity of the sandy unit below 35 feet, there is no evidence that DNAPL saturated the overburden aquifer and collected above the bedrock. However, no confirmatory samples were collected above the bedrock.

### 6.1.3 Soil Vapor and Indoor Air

Passive air sampling badges placed in the bank building to the west of the former Paul Miller dry cleaners site for 24 hours did not detect PCE contamination. Given the extent of the plume and the groundwater flow directions determined in this RI, the bank building is not expected to be impacted by the Paul Miller PCE plume. No sampling was conducted at the other adjacent building, the Kentucky Fried Chicken restaurant to the east, because the property owner refused access.

## 6.2 Recommendations

Based on the findings of the RI, no additional data or activities will be required prior to FS development. However, the following additional activities may be needed to complete a remedial design for the site:

- Continued monitoring of the indoor air for the two adjacent buildings
- Additional delineation is recommended to characterize the potential commingling of the Charlton Cleaners plume and the former Paul Miller Dry Cleaners plume. A recommended approach to this activity would involve developing a potentiometric surface that encompasses both sites, compound-specific isotope analysis to differentiate the contributions of each source to mass in the commingling zone, and additional groundwater screening and/or monitoring well installation and sampling.

## Section 7

### References

- ATSDR. 1996. *Toxicological Profile for 1,2-Dichloroethene*. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. August.
- ATSDR. 1997. *Toxicological Profile for Tetrachloroethene*. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. September.
- ATSDR. 1997. *Toxicological Profile for Trichloroethene*. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. September.
- ATSDR. 2006. *Toxicological Profile for Vinyl Chloride*. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. July.
- Bouwer, E.J., 1994. Bioremediation of chlorinated solvents using alternate electron acceptors, In Handbook of Bioremediation: (Norris, R.D., Hinchee, R.E., Brown, R., McCarty, P.L, Semprini, L., Wilson, J.T., Campbell, D.H., Reinhard, M., Bouwer, E.J., Borden, R.C., Vogel, T.M., Thomas, J.M., and Ward, C.H., Eds.), Lewis Publishers, Boca Raton, FL, p. 149 – 175.
- Camp Dresser & McKee Inc. (CDM). 2009. *Draft Remedial Investigation Report* (Site No.: 2-43-018), Port Richmond, New York. September.
- Chapelle, F.H. 1996. Identifying redox conditions that favor the natural attenuation of chlorinated ethenes in contaminated ground-water systems. Symposium on Natural Attenuation of Chlorinated Organics in Ground Water. EPA/540/R-96/509. p. 17-20.
- Chapelle, F.H., 1993. *Ground-Water Microbiology and Geochemistry*: John Wiley & Sons. Inc. New York. 424 p.
- Fogel, M. M., A.R. Taddeo and S. Fogel. 1986. Biodegradation of chlorinated ethenes by a methane-utilizing mixed culture. *Appl Environ Microbial* 51:720-724.
- Freeze, R.A. and J.A. Cherry. 1979. *Groundwater*, Prentice Hall, Inc., Englewood Cliffs, New Jersey.
- Leggette, Brashears & Graham, Inc (LBG), 2006. *Remedial Investigation Report for the Former Charlton Cleaners Facility*. June.
- Lucius, J.E. et al. 1990. Properties and Hazards of 108 Selected Substances. *USGS Open File Report 90-4089*, pp 559.
- New York State Department of Environmental Conservation (NYSDEC). 2010. *DER-10 Technical Guidance for Site Investigation and Remediation*. June
- New York State Department of Health (NYSDOH). 2006. *Final Guidance for Evaluating Soil Vapor Intrusion in the State of New York*. October.

- Reilly, T.E., Frank, H.T. Buxton, and G.D. Bennett. 1987. *A Conceptual Framework for Groundwater Solute-Transport Studies with Emphasis on Physical Mechanisms of Solute Movement*, USGS Water Resources Investigations Report 87-419.
- United States Department of Navy. 1998. *Technical Guidelines for Evaluating Monitored Natural Attenuation of Petroleum Hydrocarbons and Chlorinated Solvents in Ground Water at Naval and Marine Corps Facilities*.
- United States Environmental Protection Agency, 1998. *Technical Protocol for Evaluating Natural Attenuation of Chlorinated Solvents in Ground Water*.
- United States Geological Survey (USGS). 2002. *Natural Attenuation of Chlorinated Volatile Organic Compounds in Ground Water at Operable Unit 1, Naval Undersea Warfare Center, Division Keyport, Washington*.
- Wiedemeier, T.H., Rifai, H.S., Newell, C.J., and Wilson, J.T. 1998. *Natural Attenuation of Fuels and Solvents in the Subsurface*: John Wiley and Sons. New York.

## Tables

**Table 2-1**  
**Membrane Interface Probe Confirmatory Soil Sample Summary**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID	Sample Location	Sample Depth (ft bgs)	Date	Time	Notes
80230-MIP-5C-36.5	MIP-5C	36.5	6/24/2011	9:19	
80230-MIP-6B-42.5	MIP-6B	42.5	6/24/2011	10:57	
80230-MW-12S-35	MW-12S	35	6/24/2011	14:37	
80230-DUP3-SO	MIP-6B	50	6/24/2011	21:00	Duplicate of 80230-MIP-6B-42.5
80230-MIP-4C-41	MIP-4C	41	6/17/2011	10:20	
80230-MIP-4B-45	MIP-4B	45	6/17/2011	12:25	MS/MSD
80230-MIP-2C-35	MIP-2C	35	6/16/2011	15:25	
80230-MIP-5E-16	MIP-5E	16	6/16/2011	16:18	
80230-MIP-3-30	MIP-3	30	5/25/2011	9:45	
80230-MIP-1B-14	MIP-1B	14	5/25/2011	12:40	
80230-MIP-5B-22	MIP-5B	22	5/25/2011	15:00	MS/MSD
80230-DUP1-SO	MIP-3	30	5/25/2011	24:00	Duplicate of 80230-MIP-3-30
80230-MIP-3B-45	MIP-3B	45	5/26/2011	9:20	MS/MSD
80230-MIP-2B-40	MIP-2B	40	5/26/2011	10:30	
80230-MIP-1D-37.5	MIP-1D	37.5	5/26/2011	11:40	
80230-MIP-1C-26	MIP-1C	26	5/26/2011	12:25	
80230-MIP-5-11	MIP-5	11	5/24/2011	12:50	
80230-MIP-5-30	MIP-5	30	5/24/2011	13:07	
80230-MIP-1-16.5	MIP-1	16.5	5/24/2011	15:35	
80230-MIP-2-30	MIP-2	30	5/24/2011	16:45	
80230-MIP-6-30	MIP-6	30	5/23/2011	14:00	
80230-MIP-7-38	MIP-7	38	5/23/2011	12:05	
80230-DUP1-SO	MIP-7	38	5/23/2011	19:00	Duplicate of 80230-MIP-7-28
80230-MIP-6-10	MIP-6	10	5/23/2011	13:35	

Notes:

ID - identification

ft bgs - feet below ground surface

MIP = Membrane Interface Probe

MS/MSD - matrix spike/matrix spike duplicate

**Table 2-2**  
**Membrane Interface Probe Confirmatory Groundwater Sample Summary**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID	Sample Location	Sample Depth (ft bgs)	Date	Time	Notes
80230-GW-MIP-5-15	MIP-5	15	5/24/2011	12:10	
80230-DUP1-GW	MIP-5	15	5/24/2011	18:00	Duplicate of 80230-GW-MIP-5-15
80230-GW-MIP-5-28	MIP-5	28	5/24/2011	10:55	MS/MSD
80230-GW-MIP-6-33.5	MIP-6	33.5	5/23/2011	16:53	
80230-GW-MIP-6-15	MIP-6	15	5/23/2011	17:30	

Notes:

ID - identification

ft bgs - feet below ground surface

MS/MSD - matrix spike/matrix spike duplicate

MIP = Membrane Interface Probe

**Table 2-3**  
**Sub-slab Soil Sample Summary**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID	Sample Location	Sample Depth (ft bgs)	Date	Time	Notes
PM-SB-7-B-1112011	SB-7	12.5	11/2/2011	13:55	
PM-SB-1B-B-1112011	SB-1B	6	11/2/2011	11:17	
PM-SB-1B-A-1112011	SB-1B	0.5	11/2/2011	11:10	
PM-SB-99-1112011	SB-1B	0.5	11/2/2011	11:10	Duplicate of PM-SB-1b-A-1112011
PM-SB-6-A-1112011	SB-6	0.5	11/1/2011	13:30	
PM-SB-6-B-1112011	SB-6	5	11/1/2011	13:34	
PM-SB-1A-A-1112011	SB-1A	0.5	11/1/2012	11:17	
PM-SB-1A-B-1112011	SB-1A	5	11/1/2012	11:22	
PM-SB-2-A-1112011	SB-2	0.5	11/1/2012	10:55	
PM-SB-2-B-1112011	SB-2	5	11/1/2012	11:03	
PM-SB-3-A-1112011	SB-3	0.5	11/1/2012	12:17	
PM-SB-4-A-1112011	SB-4	0.5	11/1/2012	12:15	
PM-SB-5-A-1112011	SB-5	0.5	11/1/2012	12:33	MS/MSD
PM-SB-5-B-1112011	SB-5	6	11/1/2012	12:35	

Notes:

ID - identification

ft bgs - feet below ground surface

MS/MSD - matrix spike/matrix spike duplicate

SB = soil boring

**Table 2-4**  
**Sub-slab Groundwater Sample Summary**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID	Sample Location	Sample Depth (ft bgs)	Date	Time	Notes
PM-GWS-5-110311	SB-5	8-9	11/3/2011	11:20	
PM-GWS-6-110311	SB-6	8-9	11/3/2011	10:45	
PM-GWS-99-110311	SB-1A	8-9	11/3/2011	9:50	duplicate of PM-GWS-1A-110311
PM-GWS-1A-110311	SB-1A	8-9	11/3/2011	9:50	
PM-GWS-7-110211	SB-7	12.5	11/2/2011	15:10	
PM-GWS-1B-110211	SB-1B	8-9	11/2/2011	14:15	
PM-GWS-2-110211	SB-2	8-9	11/2/2011	12:30	

Notes:

ID - identification

ft bgs - feet below ground surface

MS/MSD - matrix spike/matrix spike duplicate

GWS = Groundwater screening

**Table 2-5**  
**Monitoring Well Sample Summary**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID	Sample Location	Sample Depth (ft bgs)	Date	Time	Notes
80230-MW09D-030512	MW-09D	65	3/5/2012	10:50	MS/MSD
80230-MW09S-030512	MW-09S	30	3/5/2012	10:55	
80230-MW10D-030512	MW-10D	65	3/5/2012	13:45	
80230-MW10S-030512	MW-10S	27	3/5/2012	14:00	
80230-MW16S-030612	MW-16S	30	3/6/2012	9:15	
80230-MW01-030612	MW-01	13	3/6/2012	9:10	
80230-MW04-030612	MW-04	9	3/6/2012	12:10	
80230-MW02-030612	MW-02	9	3/6/2012	12:00	
80230-MW15D-030712	MW-15D	65	3/7/2012	8:55	
80230-MW13S-030712	MW-13S	30	3/7/2012	9:20	
80230-MW13D-030712	MW-13D	65	3/7/2012	9:10	
80230-MW08S-030712	MW-08S	30	3/7/2012	11:40	
80230-MW03-030712	MW-03	10	3/7/2012	11:50	
80230-MW11D-030812	MW-11D	65	3/8/2012	9:00	
80230-MW14S-030812	MW-14S	29	3/8/2012	9:00	
80230-MW11S-030812	MW-11S	30	3/8/2012	9:25	
80230-MW12S-030812	MW-12S	30	3/8/2012	11:15	
80230-MW112S-030812	MW-12S	30	3/8/2012	11:15	Duplicate of 80230-MW-12S-030812

Notes:

Sample depth is the middle of the screen

ID - identification

ft bgs - feet below ground surface

MS/MSD - matrix spike/matrix spike duplicate

MW = Monitoring Well

**Table 2-6**  
**Groundwater Elevations March 2012**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Well Name	Northing	Easting	Screen top (ft bgs)	Screen bottom (ft bgs)	Measuring Point Elevations (feet amsl)	Depth to Water (feet bgs)	GW Elevation (Feet amsl)
MW-1	166800.81	946094.63	10	20	28.46	8.94	19.52
MW-2	166999.07	946080.65	5	15	23.96	3.41	20.55
MW-3	166953.80	946095.78	6	16	25.36	3.60	21.76
MW-4	166998.25	946118.12	4	14	24.65	3.56	21.09
MW-8S	166857.43	946018.44	25	35	26.24	7.00	19.24
MW-9S	167042.86	946053.45	25	35	23.3	4.30	19.00
MW-9D	167039.68	946056.62	60	70	23.37	4.32	19.05
MW-10S	167032.46	946144.29	22	32	23.89	4.55	19.34
MW-10D	167035.28	946141.10	60	70	24.36	5.40	18.96
MW-11S	166952.51	946150.01	25	35	25.1	6.00	19.10
MW-11D	166951.23	946144.21	60	70	25.02	5.95	19.07
MW-12S	166927.43	946128.12	25	35	25.24	6.13	19.11
MW-13S	166941.34	946167.86	25	35	27.47	8.40	19.07
MW-13D	166942.04	946170.55	60	70	27.35	8.28	19.07
MW-14S	166880.06	946133.46	24	34	26.26	6.70	19.56
MW-15D	166886.17	946144.74	60	70	25.85	6.68	19.17
MW-16S	166776.64	946099.45	25	35	28.93	9.34	19.59

Notes:

ID - identification

ft amsl - feet above mean sea level

ft bgs - feet below ground surface

GW - groundwater

MW = monitoring well

**Table 2-7**  
**Air Sample Summary**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

<b>Sample Name</b>	<b>Sample Location</b>	<b>Date</b>	<b>Exposure Duration</b>	<b>Notes</b>
PMBANK1	Basement	3/10/2011	24 hr	
PMBANBKDUP	Basement	3/10/2011	24 hr	Duplicate of PMBANK
PMAMBAIR	Drive Through	3/10/2011	24 hr	

**Table 3-1**  
**Slug Test Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Location	Test	Stickup	Total Well Depth	Well Depth	Depth to Water	Aquifer Base	Aquifer Thickness	Top of Screen	Depth to top of Screen from water level (ft)	Screen Length	Effective Screen Length	K	Method	Remarks	Strata Notes (from well installation logs)
		(ft)	(ft bgs)	(ft bgs)	(ft TIC)	(ft bgs)	(ft)	(ft bgs)	(ft)	(ft)	(ft)	(ft/day)			
MW-8S	FH	0	35	35	7.49	70	62.51	25	17.51	10	10	0.62	Bouwer & Rice		Fine sand, loose sand gravel; clayey layers; blow counts 20s-30s; max 50/2"
MW-8S	FH - manual fit	0	35	35	7.49	70	62.51	25	17.51	10	10	0.70	Bouwer & Rice	Preferred fit	
MW-8S	RH	0	35	35	7.49	70	62.51	25	17.51	10	10	0.54	Bouwer & Rice	Preferred fit	
MW-9S	FH	0	35	35	4.76	70	65.24	25	20.24	10	10	0.43	Bouwer & Rice	Preferred fit	
MW-9S	RH	0	35	35	4.76	70	65.24	25	20.24	10	10	0.63	Bouwer & Rice		Silty medium sand, low blow counts - max 20/6"
MW-9S	RH - manual fit	0	35	35	4.76	70	65.24	25	20.24	10	10	0.46	Bouwer & Rice		
MW-11D	FH	0	70	70	6.42	70	63.58	60	53.58	10	10	3.1	Bouwer & Rice	Preferred fit	
MW-11D	RH	0	70	70	6.42	70	63.58	60	53.58	10	10	4.7	Bouwer & Rice		Med-coarse sand, gravel; blow counts 23-53 per 6"
MW-11D	RH - manual fit	0	70	70	6.42	70	63.58	60	53.58	10	10	6.66	Bouwer & Rice	Preferred fit	
MW-12S	FH	0	35	35	6.63	70	63.37	25	18.37	10	10	3.6	Bouwer & Rice	Preferred fit	
MW-12S	RH	0	35	35	6.63	70	63.37	25	18.37	10	10	2.3	Bouwer & Rice		Dense sand, silt, blow counts generally 22-60 per 6", max 50/2"
MW-12S	RH - manual fit	0	35	35	6.63	70	63.37	25	18.37	10	10	3.8	Bouwer & Rice	Preferred fit	
MW-13S	FH	0	35	35	8.89	70	61.11	25	16.11	10	10	4.9	Bouwer & Rice	Preferred fit	
MW-13S	RH	0	35	35	8.89	70	61.11	25	16.11	10	10	2.6	Bouwer & Rice		Loose fine sand; silty, gravelly; Low blow counts - max 23 per 6"
MW-13S	RH - manual fit	0	35	35	8.89	70	61.11	25	16.11	10	10	4.1	Bouwer & Rice	Preferred fit	
MW-14S	FH	0	34	34	7.75	70	62.25	24	16.25	10	10	0.07	Bouwer & Rice		
MW-14S	FH - manual	0	34	34	7.75	70	62.25	24	16.25	10	10	0.05	Bouwer & Rice	Preferred fit	
MW-14S	RH	0	34	34	7.75	70	62.25	24	16.25	10	10	0.07	Bouwer & Rice		Silty sand, sandy silt; blow counts primarily 30s-40s per 6", up to 50/3"
MW-14S	RH - manual fit	0	34	34	7.75	70	62.25	24	16.25	10	10	0.05	Bouwer & Rice	Preferred fit	
MW-15D	FH	0	70	70	7.15	70	62.85	60	52.85	10	10	69.0	Springer-Gelhar	Preferred Test	
MW-15D	RH	0	70	70	7.15	70	62.85	60	52.85	10	10	59.5	Springer-Gelhar	Preferred Test	Medium sand, low blow counts; maximum 15 per 6"
MW-15D	FH2	0	70	70	7.15	70	62.85	60	52.85	10	10	38.7	Springer-Gelhar	Slow insertion	
MW-15D	RH2	0	70	70	7.15	70	62.85	60	52.85	10	10	37.1	Springer-Gelhar	Slow insertion	
MW-16S	FH	0	35	35	9.82	70	60.18	25	15.18	10	10	0.22	Bouwer & Rice	Preferred fit	
MW-16S	RH	0	35	35	9.82	70	60.18	25	15.18	10	10	0.17	Bouwer & Rice	Preferred fit	Fine sand, silty sand; wet layers, dense layers; low blow counts - max 17 per 6"

Notes:

ft - feet

K - hydraulic conductivity

TIC - measurement from top of inner casing

bgs - below ground surface

RH - rising head

FH - falling head

**Table 4-1**  
**MIP Soil Screening Results - Detections**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID	80230-MIP1-16.5	80230-MIP1B-14	80230-MIP1C-26	80230-MIP1D-37.5	80230-MIP2-30	80230-MIP2B-40	80230-MIP2C-35	80230-MIP3-30	80230-DUP2-SO	80230-MIP3B-45
	Sample Location	Depth	MIP-1 16.5	MIP-1B 14	MIP-1C 26	MIP-1D 37.5	MIP-2 30	MIP-2B 40	MIP-2C 35	MIP-3 30	MIP-3 30	MIP-3B 45	
		Sampling Date	05/24/2011	05/25/2011	05/26/2011	05/26/2011	05/24/2011	05/26/2011	06/16/2011	05/25/2011	05/25/2011	05/26/2011	
<b>Chemical</b>	<b>CAS#</b>	<b>NYSDEC Unrestricted Use Soil Cleanup Objectives (1)</b>		<b>Result</b>	<b>Result</b>	<b>Result</b>	<b>Result</b>	<b>Result</b>	<b>Result</b>	<b>Result</b>	<b>Result</b>	<b>Result</b>	
<b>Volatiles (µg/kg)</b>													
1,2-Dichloropropane	78-87-5	NL								3.9 J			
Acetone	67-64-1	50			R						R	R	
Cis-1,2-Dichloroethene	156-59-2	250		1.3 J				1.5 J		55	5.4	2.3 J	
Methylene Chloride	75-09-2	50	2.4 J					2.6 J				1.7 J	
Naphthalene	91-20-3	12000											
Tetrachloroethene	127-18-4	1300		8.1				53		1900 J	6.4	2.2 J	
Trichloroethene	79-01-6	470								7	1.9 J		
<b>Semi-Volatiles (µg/kg)</b>													
Benzo(a)Anthracene	56-55-3	1000											
Benzo(a)Pyrene	50-32-8	1000											
Benzo(b)Fluoranthene	205-99-2	1000											
Benzo(g,h,i)Perylene	191-24-2	100000											
Benzo(k)Fluoranthene	207-08-9	800											
Bis(2-ethylhexyl)Phthalate	117-81-7	NL	360 J					160 J		48 J			
Chrysene	218-01-9	1000											
Dibenz(a,h)Anthracene	53-70-3	330											
Fluoranthene	206-44-0	100000				50 J	55 J						
Indeno(1,2,3-cd)Pyrene	193-39-5	500											
Phenanthrene	85-01-8	100000											
Pyrene	129-00-0	100000				37 J	45 J						
<b>Inorganics (mg/kg)</b>													
Aluminum	7429-90-5	NL		10800		5510		3260		4530		3910	
Antimony	7440-36-0	NL				0.35 BNJ						0.52 BNJ	
Arsenic	7440-38-2	13		2.4 J		2.4 J		1.7 J		1.5		3.6 J	
Barium	7440-39-3	350		81.2		69.3 J		59.1 J		34.9		60.1 J	
Beryllium	7440-41-7	7.2		0.86		0.59		0.25 B		0.44		0.29	
Cadmium	7440-43-9	2.5		0.18 B		0.2		0.071 B				0.11 B	
Calcium	7440-70-2	NL		2900		947		905		2000		926	
Chromium	7440-47-3	30		26.3 J		16.3		11.3		11.1		11	
Cobalt	7440-48-4	NL		11.2 J		6.4 J		5.6 J		5.1		4.9 J	
Copper	7440-50-8	50		30.3 J		16.4 J		10.9 J		188		10.9 J	
Iron	7439-89-6	NL		15600 J		16200 J		11700 J		11400		12200 J	
Lead	7439-92-1	63		24.4		6.2		2.8		4.8		2.7	
Magnesium	7439-95-4	NL		5820 J		3230		1680		1900		1860	
Manganese	7439-96-5	1600		194		336 J		267 J		205		375 J	
Mercury	7439-97-6	0.18		0.005 B									
Nickel	7440-02-0	30		31		14.7		7.6		8.5		7.7	
Potassium	7440-09-7	NL		1950 J		1170		399		621		450	
Selenium	7782-49-2	3.9		0.51 B		0.57 B							
Sodium	7440-23-5	NL		720 J		90.3		61.5		72.9		60.2	
Thallium	7440-28-0	NL		2.8		1.3		1.2				1.1	
Vanadium	7440-62-2	NL		39.3 J		23.8 J		22.8 J		18.2		19.9 J	
Zinc	7440-66-6	109		61.3 J		30.2		13.8		19.7		17.5	
<b>Miscellaneous</b>													
Moisture (%)	MOIST	NL	14	11	10	8 J	7.4 J	15	20	9.5 J	9.9 J	14	

Notes:

ID - identification

NL - not listed

µg/kg - milligrams per kilogram

mg/kg - milligrams per kilogram

Indicates exceedance

1. New York State Register and Official Compilation of Codes, Rules and Regulations of the State of New York - Chapter IV - Quality Services - Subpart 375-6: Remedial Program Soil Cleanup Objectives (6 NYCRR Part 375-6.8 (a)), December 14, 2006

Laboratory Data Qualifiers

U - The compound was not detected at the indicated concentration.

J - Data indicates the presence of a compound that meets the identification criteria.

The result is less than the quantitation limit but greater. The concentration given is an approximate value.

B - The analyte was found in the laboratory blank as well as the sample. This indicates possible laboratory contamination of the environmental sample.

E (Inorganics) - The reported value is estimated because of the presence of interference.

N - Spiked sample recovery not within control limits

**Table 4-1**  
**MIP Soil Screening Results - Detections**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID	80230-MIP4B-45	80230-MIP4C-41	80230-MIP5-11	80230-MIP5-30	80230-MIP5B-22	80230-MIP5E-16	80230-MIP6-10	80230-MIP6-30	80230-MIP6B-42.5	80230-DUP3-SO	80230-MIP7-38	80230-DUP1-SO
			Sample Location	MIP-4B	MIP-4C	MIP-5	MIP-5	MIP-5B	MIP-5E	MIP-6	MIP-6	MIP-6B	MIP-6B	MIP-7	MIP-7
			Depth	45	41	11	30	22	16	10	30	42.5	38	38	38
Sampling Date	6/17/2011		6/17/2011		05/24/2011	05/24/2011	05/24/2011	05/25/2011	06/16/2011	05/23/2011	05/23/2011	06/24/2011	06/24/2011	05/23/2011	05/23/2011
Chemical	CAS#	NYSDEC Unrestricted Use Soil Cleanup Objectives (1)	Result	Result	Result	Result	Result	Result	Result	Result	Result	Result	Result	Result	Result
<b>Volatiles (µg/kg)</b>															
1,2-Dichloropropane	78-87-5	NL	2.8J	2.9J					2.5J						
Acetone	67-64-1	50							R						
Cis-1,2-Dichloroethene	156-59-2	250													
Methylene Chloride	75-09-2	50	1.9J		2.6J	1.9J				2J	1.9J			1.7J	2.1J
Naphthalene	91-20-3	12000			47JB	6	18	16	4.7J	3.1J	4J		1.3J		
Tetrachloroethene	127-18-4	1300											1.6J		
Trichloroethene	79-01-6	470													
<b>Semi-Volatiles (µg/kg)</b>															
Benzo(a)Anthracene	56-55-3	1000											48J	150J	
Benzo(a)Pyrene	50-32-8	1000											59J	180J	
Benzo(b)Fluoranthene	205-99-2	1000											100J	280J	
Benzo(g,h,i)Perylene	191-24-2	100000											61J	160J	
Benzo(k)Fluoranthene	207-08-9	800												120J	
Bis(2-ethylhexyl)Phthalate	117-81-7	NL				160J				68J	50J	54J		74J	69J
Chrysene	218-01-9	1000											100J	260J	
Dibeno(a,h)Anthracene	53-70-3	330												38J	
Fluoranthene	206-44-0	100000											130J	350J	
Indeno(1,2,3-cd)Pyrene	193-39-5	500											46J	120J	
Phenanthrene	85-01-8	100000											37J	83J	
Pyrene	129-00-0	100000											95J	250J	
<b>Inorganics (mg/kg)</b>															
Aluminum	7429-90-5	NL					5060		7990						
Antimony	7440-36-0	NL					0.34BNJ								
Arsenic	7440-38-2	13					2.8J		3.7						
Barium	7440-39-3	350					45.5		62.6						
Beryllium	7440-41-7	7.2					0.47		0.9						
Cadmium	7440-43-9	2.5					0.17								
Calcium	7440-70-2	NL					1130		2970						
Chromium	7440-47-3	30					34J		18.3						
Cobalt	7440-48-4	NL					11.8J		8.4						
Copper	7440-50-8	50					10.8J		10.8						
Iron	7439-89-6	NL					15200J		18900						
Lead	7439-92-1	63					8		12.4						
Magnesium	7439-95-4	NL					2890J		4980						
Manganese	7439-96-5	1600					348		652						
Mercury	7439-97-6	0.18													
Nickel	7440-02-0	30					77.9		19.4						
Potassium	7440-09-7	NL					953J		1850						
Selenium	7782-49-2	3.9					85.6J		158						
Sodium	7440-23-5	NL					1.1								
Thallium	7440-28-0	NL					17.8J		24.7						
Vanadium	7440-62-2	NL					27.2J		53						
Zinc	7440-66-6	109													
<b>Miscellaneous</b>															
Moisture (%)	MOIST	NL	15	14	14	7.3J	14	14	12	14	7.3J	7.7J	8.5J	11	

Notes:

ID - identification

NL - not listed

µg/kg - milligrams per kilogram

mg/kg - milligrams per kilogram

**Indicates exceedance**

1. New York State Register and Official Compilation of Codes, Rules and Regulations of the State of New York - Chapter IV - Quality Services - Subpart 375-6: Remedial Program Soil Cleanup Objectives (6 NYCRR Part 375-6.8 (a)), December 14, 2006

**Laboratory Data Qualifiers**

U - The compound was not detected at the indicated concentration.

J - Data indicates the presence of a compound that meets the identification criteria.

The result is less than the quantitation limit but greater. The concentration given is an approximate value.

B - The analyte was found in the laboratory blank as well as the sample. This indicates possible laboratory contamination of the environmental sample.

E (Inorganics) - The reported value is estimated because of the presence of interference.

N - Spiked sample recovery not within control limits

**Table 4-2**  
**Sub-slab Soil Screening Results - Detections**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Depth Sampling Date	PM-SB-1A-A-1112011 PM-SB-1A-A 0.5 11/01/2011	PM-SB-1A-B-1112011 PM-SB-1A-B 5 11/01/2011	PM-SB-1B-A-1122011 PM-SB-1B-A 0.5 11/02/2011	PM-SB-99-110211 PM-SB-1B-A-DUP 0.5 11/02/2011	PM-SB-1B-B-1122011 PM-SB-1B-B 6 11/02/2011	PM-SB-2-A-1112011 PM-SB-2-A 0.5 11/01/2011	PM-SB-2-B-1112011 PM-SB-2-B 5 11/01/2011
Chemical	CAS#	NYSDEC Unrestricted Use Soil Cleanup Objectives (1)	Result	Result	Result	Result	Result	Result	Result	Result
<b>Volatiles (µg/kg)</b>										
1,2,4-Trimethylbenzene	95-63-6	3600								
Acetone	67-64-1	50	17 J	5.3 J						
Cis-1,2-Dichloroethene	156-59-2	250		3.2 J					3.5 J	14
Methylene Chloride	75-09-2	50			3 J			2.9 J		2 J
Naphthalene	91-20-3	12000								
Tetrachloroethene	127-18-4	1300		55 J	2.7 J	2.3 J		5.9 J		9.2 J
Toluene	108-88-3	700								
Trichloroethene	79-01-6	470		3.2 J						1.2 J
<b>Miscellaneous</b>										
Moisture (%)	MOIST	NL	16	11	17	19	13	16	18	

Notes:

ID - identification

NL - not listed

µg/kg - milligrams per kilogram

mg/kg - milligrams per kilogram

**Indicates exceedance**

1. New York State Register and Official Compilation of Codes, Rules and Regulations of the State of New York - Chapter IV - Quality Services - Subpart 375-6: Remedial Program Soil Cleanup Objectives (6 NYCRR Part 375-6.8 (a)), December 14, 2006

**Laboratory Data Qualifiers**

J - Data indicates the presence of a compound that meets the identification criteria.

The result is less than the quantitation limit but greater. The concentration given is an approximate value.

**Table 4-2**  
**Sub-slab Soil Screening Results - Detections**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID Sample Location Depth Sampling Date			PM-SB-3-A-1112011 PM-SB-3-A 0.5 11/01/2011	PM-SB-4-A-1112011 PM-SB-4-A 0.5 11/01/2011	PM-SB-5-A-1112011 PM-SB-5-A 0.5 11/01/2011	PM-SB-5-B-1112011 PM-SB-5-B 6 11/01/2011	PM-SB-6-A-1112011 PM-SB-6-A 0.5 11/01/2011	PM-SB-6-B-1112011 PM-SB-6-B 5 11/01/2011	PM-SB-7-B-1122011 PM-SB-7-B 12.5 11/02/2011
Chemical	CAS#	NYSDEC Unrestricted Use Soil Cleanup Objectives (1)	Result	Result	Result	Result	Result	Result	Result
<b>Volatiles (µg/kg)</b>									
1,2,4-Trimethylbenzene	95-63-6	3600		1.8 J					
Acetone	67-64-1	50		14 J					
Cis-1,2-Dichloroethene	156-59-2	250			12		12		
Methylene Chloride	75-09-2	50		5 J		3.2 J	3.8 J	3 J	2.9 J
Naphthalene	91-20-3	12000		5.8 J					
Tetrachloroethene	127-18-4	1300	22 J	16 J	110 J	4.5 J	26 J		
Toluene	108-88-3	700		3.8 J	2.3 J	1.9 J			
Trichloroethene	79-01-6	470			16 J		6.6 J		
<b>Miscellaneous</b>									
Moisture (%)	MOIST	NL	18	17	30	13	28	15	13

Notes:

ID - identification

NL - not listed

µg/kg - milligrams per kilogram

mg/kg - milligrams per kilogram

Indicates exceedance

1. New York State Register and Official Compilation of Codes, Rules and Regulations  
of the State of New York - Chapter IV - Quality Services - Subpart 375-6: Remedial  
Program Soil Cleanup Objectives (6 NYCRR Part 375-6.8 (a)), December 14, 2006

Laboratory Data Qualifiers

J - Data indicates the presence of a compound that meets the identification criteria.

The result is less than the quantitation limit but greater. The concentration given  
is an approximate value.

**Table 4-3**  
**Monitoring Well Soil Results - Detections**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

		Sample ID	80230-MW12S-35
		Sample Location	MW-12S
		Depth	35
		Sampling Date	6/24/2011
Chemical	CAS#	NYSDEC Unrestricted Use Soil Cleanup Objectives (1)	Result
<b>Volatiles (µg/kg)</b>			
1,2,4-Trimethylbenzene	95-63-6	3600	1.3 J
Acetone	67-64-1	50	
Cis-1,2-Dichloroethene	156-59-2	250	2.3 J
Methylene Chloride	75-09-2	50	
Naphthalene	91-20-3	12000	2.1 J
Tetrachloroethene	127-18-4	1300	1000
Toluene	108-88-3	700	
Trichloroethene	79-01-6	470	
<b>Miscellaneous</b>			
Moisture (%)	MOIST	NL	12

Notes:

ID - identification

NL - not listed

µg/kg - milligrams per kilogram

mg/kg - milligrams per kilogram

**Indicates exceedance**

1. New York State Register and Official Compilation of Codes, Rules and Regulations of the State of New York - Chapter IV - Quality Services - Subpart 375-6: Remedial Program Soil Cleanup Objectives (6 NYCRR Part 375-6.8 (a)), December 14, 2006

**Laboratory Data Qualifiers**

J - Data indicates the presence of a compound that meets the identification criteria. The result is less than the quantitation limit but greater. The concentration given is an approximate value.

**Table 4-4**  
**MIP Groundwater Screening Data - Detections**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical	Sample ID Sample Location Depth Sampling Date	80230-GW-MIP5-15 MIP5 15 05/24/2011	80230-GW-MIP5-15-DUP MIP5 28 05/24/2011	80230-GW-MIP5-28 MIP5 28 05/24/2011	80230-GW-MIP6-15 MIP6 15 05/23/2011	80230-GW-MIP6-33.5 MIP6 33.5 05/23/2011
Chemical	NEW YORK STATE CLASS GA	Result	Result	Result	Result	Result
<b>Volatiles (µg/L)</b>						
1,1,2-Trichloroethane	1					
1,1-Dichloroethene	5					
1,2-Dichlorobenzene	3					
1,4-Dichlorobenzene	3					
2-Butanone (Mek)	NL					
Acetone	NL					
Carbon Disulfide	60					
Chlorobenzene	5					
Chloroform	7					
Chloromethane	5					
Cis-1,2-Dichloroethene	5	780		1.7 J	21	11
Methyl Tert-Butyl Ether (MTBE)	NL				1.7 J	
Tetrachloroethylene	5	32000 D	33000	150	2100 D	380 D
Toluene	5					
Trans-1,2-Dichloroethene	5					
Trichloroethylene	5			2.3 J	9	8.7
Vinyl Chloride	2					
<b>Semi-Volatiles (µg/L)</b>						
Bis(2-Ethylhexyl)Phthalate	5			1.7 J		
Diethylphthalate	NL		1.1 J			
<b>Inorganics (µg/L)</b>						
Aluminum	NL	516	489	492	249	714
Barium	1000	81 B	79.2 B	92.6 BEJ	153 BEJ	114 BEJ
Calcium	NL	101000	98000	120000	111000	168000
Chromium	50	2.4 B	2.2 B	4 B	1.9 B	11 B
Cobalt	NL	7.2 B	7.4 B	1.7 B	4.8 B	4.8 B
Copper	200	4.1 B	4 B			4.5 B
Iron	300	1700	1600	1410	747	2660
Magnesium	NL	38100	37200	32900	64700	59100
Manganese	300	2320	2250	202	1100	479
Nickel	100	38.6 B	37.5 B	16.9 B	31.2 B	118
Potassium	NL	6800	6790	3840	5540	4570
Sodium	NL	138000	138000	62400	277000	120000
Vanadium	NL	1.9 B	2.2 B	1.9 B	1.2 B	2.3 B
Zinc	NL	39.1 B	38.5 B	23.6 B	32 B	29.2 B

Notes:

JD - identification

µg/kg - milligrams per kilogram

mg/kg - milligrams per kilogram

NL - not listed

**Indicates exceedance**

(1) NYSDEC, June 1998, TOGS 1.1.1, Ambient Water Quality Standards and Guidance Values and Groundwater Effluent Limitations

Laboratory Data Qualifiers

J - Data indicates the presence of a compound that meets the identification criteria. The result is less than the quantitation limit but greater. The concentration given is an approximate value.

B - The analyte was found in the laboratory blank as well as the sample. This indicates possible laboratory contamination of the environmental sample.

P - For dual column analysis, the percent difference between the quantitated concentrations on the two columns is greater than 40%.

\* - For dual column analysis, the lowest quantitated concentration is being reported due to coeluting interference.

E (Organics) - Indicates the analyte's concentration exceeds the calibrated range of the instrument for that specific analysis.

D - The reported value is from a secondary analysis with a dilution factor. The original analysis exceeded the calibration range.

R - The reported value was rejected.

**Table 4-5**  
**Sub-slab Groundwater Screening Data - Detections**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID Sample Location Approximate Depth Sampling Date			PM-GWS-1A-110311 PM-GWS-1A 8-9 11/03/2011	PM-GWS-1A-110311-DUP PM-GWS-1A-DUP 8-9 11/03/2011	PM-GWS-1B-1122011 PM-GWS-1B 8-9 11/02/2011	PM-GWS-2-1122011 PM-GWS-2 8-9 11/02/2011	PM-GWS-5-110311 PM-GWS-5 8-9 11/03/2011	PM-GWS-6-110311 PM-GWS-6 8-9 11/03/2011	PM-GWS-7-1122011 PM-GWS-7 12.5 11/02/2011
Chemical	CAS#	NEW YORK STATE CLASS GA	Result	Result	Result	Result	Result	Result	Result
<b>Volatiles (µg/L)</b>									
1,1,2-Trichloroethane	79-00-5	1							
1,1-Dichloroethene	75-35-4	5							
1,2-Dichlorobenzene	95-50-1	3							
1,4-Dichlorobenzene	106-46-7	3							
2-Butanone (Mek)	78-93-3	NL							140 J
Acetone	67-64-1	NL		4.5 J				8.3 J	89 J
Carbon Disulfide	75-15-0	60							2.1 J
Chlorobenzene	108-90-7	5							
Chloroform	67-66-3	7							
Chloromethane	74-87-3	5							
Cis-1,2-Dichloroethene	156-59-2	5	11	12	12 J	90 J	15 J	12 J	27 J
Ethane	74-84-0	NL							
Ethene	74-85-1	NL							
Methane	74-82-8	NL							
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	NL							
Tetrachloroethene	127-18-4	5	53	57	17 J	150 J	51	11 J	18 J
Toluene	108-88-3	5	2.2 J	2.5 J					
Trans-1,2-Dichloroethene	156-60-5	5							
Trichloroethene	79-01-6	5	5.2	5.5	2.8 J	16	14	4.3 J	3.1 J
Vinyl Chloride	75-01-4	2				2.1 J			

Notes:

ID - identification

µg/kg - milligrams per kilogram

mg/kg - milligrams per kilogram

NL - not listed

**Indicates exceedance**

(1) NYSDEC, June 1998, TOGS 1.1.1, Ambient Water Quality Standards and Guidance Values and Groundwater Effluent Limitations.

Laboratory Data Qualifiers

J - Data indicates the presence of a compound that meets the identification criteria. The result is less than the quantitation limit but greater. The concentration given is an approximate value.

B - The analyte was found in the laboratory blank as well as the sample. This indicates possible laboratory contamination of the environmental sample.

P - For dual column analysis, the percent difference between the quantitated concentrations on the two columns is greater than 40%.

\* - For dual column analysis, the lowest quantitated concentration is being reported due to coeluting interference.

E (Organics) - Indicates the analyte's concentration exceeds the calibrated range of the instrument for that specific analysis.

D - The reported value is from a secondary analysis with a dilution factor. The original analysis exceeded the calibration range.

R - The reported value was rejected.

**Table 4-6**  
**Monitoring Well Data - Detections**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID			80230-MW01-030612	80230-MW02-030612	80230-MW03-030712	80230-MW04-030612	80230-MW08S-030712	80230-MW09D-030512	80230-MW09S-030512	80230-MW10D-030512	80230-MW10S-030512
Sample Location	MW01	MW02	MW03	MW04	MW08S	MW09D	MW09S	MW10D	MW10S		
Depth	13	9	10	9	30	65	30	65	27		
Sampling Date	03/06/2012	03/06/2012	03/07/2012	03/06/2012	03/07/2012	03/05/2012	03/05/2012	03/05/2012	03/05/2012		
Chemical	CAS#	New York State Standards and Guidance Values for Class GA Groundwater (1)	Result	Result	Result	Result	Result	Result	Result	Result	
<b>Volatiles (µg/L)</b>											
1,1,2-Trichloroethane	79-00-5	1	0.6 J				1.1 J				
1,1-Dichloroethene	75-35-4	5	2.8 J					0.65 J			
1,2-Dichlorobenzene	95-50-1	3									
1,4-Dichlorobenzene	106-46-7	3									
2-Butanone (MEK)	78-93-3	NL									
Acetone	67-64-1	NL									
Carbon Disulfide	75-15-0	60									
Chlorobenzene	108-90-7	5	1.1 J								
Chloroform	67-66-3	7				0.81 J		0.66 J			
Chloromethane	74-87-3	5		0.58 J				0.92 J			
Cis-1,2-Dichloroethene	156-59-2	5	4300 D	22	38		26		17	140	
Ethane	74-84-0	NL	310							3.2	
Ethene	74-85-1	NL	700								
Methane	74-82-8	NL	1400	960	130	1500			2.1	45	
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	NL								0.76 J	
Tetrachloroethene	127-18-4	5	750 D				1300 D		34	6.1	
Toluene	108-88-3	5	2 J								
Trans-1,2-Dichloroethene	156-60-5	5	98				10		16	2.6 J	
Trichloroethene	79-01-6	5	130								
Vinyl Chloride	75-01-4	2	1200 JD	11 J		2.8 J			0.58 J	1.3 J	
<b>Miscellaneous (mg/L)</b>											
Alkalinity, Total (as CaCO <sub>3</sub> )	ALK	NL	260	300	600	420	170	210	230	130	1000
Ammonia as N	7664-41-7	2	R	R							
Chloride	16887-00-6	250	860	230	130	430	71	90	72	88	150
Hardness As CaCO <sub>3</sub>	CACOA-H	NL	120	300	440	230	230	320	320	330	620
Nitrate-NO <sub>3</sub>	14797-55-8	10	0.82	0.65			5.1	5.8	3.3	4.2	
Nitrite-NO <sub>2</sub>	14797-65-0	1									
Nitrogen, Kjeldahl	KN	NL	R	R	R	R	R	R	R		
Sulfate	14808-79-8	NL	45	57	2.1 J	15	35	45	44	48	61
Sulfide	18496-25-8	NL	0.4		0.056		0.048		0.083		0.046
Total Dissolved Solids	TDS	NL	1800	800	910	1000	360	370	390	420	690
Total Organic Carbon	TOC	NL	12	12	45	11					5.7 J
Total Suspended Solids	TSS	NL	140	40	80	21	95	68	540	87	230

Notes:

ID - identification

µg/kg - milligrams per kilogram

mg/kg - milligrams per kilogram

NL - not listed

Indicates exceedance

(1) NYSDEC, June 1998, TOGS 1.1.1, Ambient Water Quality Standards and Guidance Values and Groundwater Effluent Limitations.

#### Laboratory Data Qualifiers

J - Data indicates the presence of a compound that meets the identification criteria. The result is less than the quantitation limit but greater. The concentration given is an approximate value.

B - The analyte was found in the laboratory blank as well as the sample. This indicates possible laboratory contamination of the environmental sample.

P - For dual column analysis, the percent difference between the quantitated concentrations on the two columns is greater than 40%.

\* - For dual column analysis, the lowest quantitated concentration is being reported due to coeluting interference.

E (Organics) - Indicates the analyte's concentration exceeds the calibrated range of the instrument for that specific analysis.

D - The reported value is from a secondary analysis with a dilution factor. The original analysis exceeded the calibration range.

R - The reported value was rejected.

**Table 4-6**  
**Monitoring Well Data - Detections**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID Sample Location Depth Sampling Date			80230-MW11D-030812 MW11D 65 03/08/2011	80230-MW11S-030812 MW11S 30 03/08/2011	80230-MW12S-030812 MW12S 30 03/08/2012	80230-MW12S-030812-DUP MW12S -- 03/08/2012	80230-MW13D-030712 MW13D 65 03/07/2012	80230-MW13S-030712 MW13S 30 03/07/2012	80230-MW14S-030812 MW14S 29 03/08/2012	80230-MW15D-030712 MW15D 65 03/07/2012	80230-MW16S-030612 MW16S 30 03/06/2012
Chemical	CAS#	New York State Standards and Guidance Values for Class GA Groundwater (1)	Result	Result	Result	Result	Result	Result	Result	Result	Result
<b>Volatiles (µg/L)</b>											
1,1,2-Trichloroethane	79-00-5	1									
1,1-Dichloroethene	75-35-4	5			15				9.4		
1,2-Dichlorobenzene	95-50-1	3			2.2 J				1.1 J		
1,4-Dichlorobenzene	106-46-7	3							0.92 J		
2-Butanone (MEK)	78-93-3	NL									
Acetone	67-64-1	NL									
Carbon Disulfide	75-15-0	60									
Chlorobenzene	108-90-7	5			2.6 J				3.1 J		
Chloroform	67-66-3	7									
Chloromethane	74-87-3	5									
Cis-1,2-Dichloroethene	156-59-2	5			11000 D	350 J	380 J		7000 D	850	1.3 J
Ethane	74-84-0	NL			10	3.7	4.3		9.3	90	
Ethene	74-85-1	NL							41	33	
Methane	74-82-8	NL			170	31	35		95	39	2
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	NL			2.1 J				3.1 J		
Tetrachloroethene	127-18-4	5			9000 D	71000 D	72000 D		2500 D	100000 D	5
Toluene	108-88-3	5									2200 D
Trans-1,2-Dichloroethene	156-60-5	5			8.9				160		
Trichloroethene	79-01-6	5			2400 D				950 D	1300	64
Vinyl Chloride	75-01-4	2			37 J				210 JD		
<b>Miscellaneous (mg/L)</b>											
Alkalinity, Total (as CaCO <sub>3</sub> )	ALK	NL	300	360	200	210	330	270	350	310	400
Ammonia as N	7664-41-7	2									R
Chloride	16887-00-6	250	88	280	98	97	91	250	290	99	480
Hardness As CaCO <sub>3</sub>	CACO-H	NL	390	550	290	300	340	560	760	360	600
Nitrate-NO <sub>3</sub>	14797-55-8	10	4.5	0.32	5.4	5.1	4.8			5	1.9
Nitrite-NO <sub>2</sub>	14797-65-0	1			0.12 J	0.13 J					
Nitrogen, Kjeldahl	KN	NL									R
Sulfate	14808-79-8	NL	47	94	33	33	46	120	100	47	52
Sulfide	18496-25-8	NL	0.096	0.095	0.034	0.033			0.1	0.11	0.038
Total Dissolved Solids	TDS	NL	590	1000	570	590	520	730	1200	400	1200
Total Organic Carbon	TOC	NL		2.9 J	2 J			6.1 J	3.3 J		
Total Suspended Solids	TSS	NL	420	190	120	100	190	24	180	260	60

Notes:

ID - identification

µg/kg - milligrams per kilogram

mg/kg - milligrams per kilogram

NL - not listed

Indicates exceedance

(1) NYSDEC, June 1998, TOGS 1.1.1, Ambient Water Quality Standards and Guidance Values and Groundwater Effluent Limitations.

Laboratory Data Qualifiers

J - Data indicates the presence of a compound that meets the identification criteria. The result is less than the quantitation limit but greater. The concentration given is an approximate value.

B - The analyte was found in the laboratory blank as well as the sample. This indicates possible laboratory contamination of the environmental sample.

P - For dual column analysis, the percent difference between the quantitated concentrations on the two columns is greater than 40%.

\* - For dual column analysis, the lowest quantitated concentration is being reported due to coeluting interference.

E (Organics) - Indicates the analyte's concentration exceeds the calibrated range of the instrument for that specific analysis.

D - The reported value is from a secondary analysis with a dilution factor. The original analysis exceeded the calibration range.

R - The reported value was rejected.

**Table 4-7**  
**Air Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID Sample Location Sampling Date		PMBANK Basement 3/11/2011	PMB ANKDUP Basement 3/11/2011	PMAMBAIR Drive Through 3/11/2011	
Chemical	NYSDOH Vapor Intrusion Guidance	Result	Result	Result	
<b>Volatiles (<math>\mu\text{g}/\text{m}^3</math>)</b>					
Tetrachloroethene	100	<1	<1	<1	<1

Notes:

ID - identification

$\mu\text{g}/\text{m}^3$  - micrograms per cubic meter

Indicates exceedance

**Table 5-1**  
**Fate and Transport Properties of Site-Related Contaminants**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Compound	CAS Number	Molecular Weight (g/mol)	Water Solubility @25 deg. C (mg/L)	Vapor Pressure @25 deg. C *@20 deg.C (mm Hg)	Henry's Law Constant @25 deg. C (atm-m^3/mol)	Literature Koc (L/kg)	log Kow	Kd Literature based cm^3/g	Retardation Factor (Rf) Calc from Literature Kd	Adsorption based on Literature Kds	Volatilization from Water							
cis-1,2-Dichloroethene (cis-1,2-DCE)	156-59-2	96.95	a1	3.5E+03	a1	1.8E+02	a1	4.08E-03	a1	38.28	b	1.9	b	7.1E-02	b	NC	Low	High
Trichloroethene (TCE)	79-01-6	131.4	a2	1.366E+03	a2	7.4E+01	a2	1.1E-02	a2	94.3	b	2.4	b	3.3E-01	b	NC	Low	High
Tetrachloroethene (PCE)	127-18-4	165.83	a3	1.5E+02	a3	1.847E+01	a3	1.8E-02	a3	265	b	3.4	b	3.1E-01	b	NC	Low	High
Vinyl chloride (VC)	75-01-4	62.5	a4	2.76E+03	a4	2.6E+03	a4	2.8E-02	a4	15.38	b	1.4	b	3.7E-02	b	NC	Low	High

**Site Variables:**

Fraction Organic Carbon, foc = Data unavailable (assume 0.1% for coastal plain environments)

Soil Bulk Density, pb = Data unavailable

Effective Porosity, η = Data unavailable

Adsorption is	"Low" if Kd <	0.5
	"High" if Kd >	2
	"Moderate" if Kd is in-between	
Volatilization from Water is	"Low" if H <	1.E-07
	"High" if H >	1.E-03
	"Moderate" if H is in-between	
Mobility is	"High" if Rf <	10
	"Low" if Rf >	1000
	"Moderate" if Rf is in-between	

**Equations:**

Koc = Soil Organic Carbon/Water Partition Coefficient, L/kg

Kow = n-Octanol/Water Partition Coefficient, dimensionless

Kd = Soil/Water Partition Coefficient [= Koc X foc for organics], cm<sup>3</sup>/g

**Abbreviations:**

atm-m<sup>3</sup>/mol = atmospheres-meters cubed per mole

C = Celcius

calc = calculated

cm<sup>3</sup>/g = cubic centimeters per gram

deg.

= degrees

gm/cm<sup>3</sup> = grams per centimeter cubed

g/mL = gram per milliliter

g/mol = grams per mole

H = Henry's Constant

Kow = n-Octanol/Water Partition Coefficient, dimensionless

L/kg = liters per kilogram

mg/L = milligrams per liter

mm Hg = millimeters of mercury

NC = not calculated

**References:**

- a. Agency for Toxic Substances and Disease Registry (ATSDR). Toxicological Profiles. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. (<http://www.atsdr.cdc.gov/toxpro2.html>)
  - 1: ATSDR. 1996. Toxicological Profile for 1,2-Dichloroethene. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. August.
  - 2: ATSDR. 1997. Toxicological Profile for Trichloroethene. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. September.
  - 3: ATSDR. 1997. Toxicological Profile for Tetrachlorethene. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. September.
  - 4: ATSDR. 2006. Toxicological Profile for Vinyl chloride. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. July.
  - 5: ATSDR. 2006. Toxicological Profile for 1,1,1-Trichloroethane. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. July.
  - 6: ATSDR. 1990. Toxicological Profile for 1,1-Dichloroethane. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. December.
  - 7: ATSDR. 1994. Toxicological Profile for 1,1-Dichloroethene. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. May.
  - 8: ATSDR. 1998. Toxicological Profile for Chloroethane. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service. December.
- b. EPA. 2005. Human Heath Risk Assessment Protocol for Hazardous Waste Combustion Facilities, Support Materials. Chemical-Specific Data. EPA 530-R-05-006. September. HHRAP Companion Database. (<http://www.epa.gov/epaoswer/hazwaste/combust/riskvol.htm#volume2>)

**Table 5-2**  
**Natural Attenuation Parameters**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

	Wells screened at water table						Screened at water table						Deep Wells					
	Downgradient of source zones			Cross-gradient	Down-gradient	Closer to Charlton Cleaners	Under suspected primary discharge				Near suspected additional discharge							
	Units	MW02	MW03	MW04	MW08S	MW09S	MW10S	MW11S	MW12S	MW13S	MW14S	MW16S	MW01	MW09D	MW10D	MW11D	MW13D	MW15D
Tetrachloroethene	uMol/L	ND	ND	ND	7.84 D	0.21	0.04	54.27 D	428.15 D	15.08 D	603.03 D	13.27 D	4.52 D	ND	ND	ND	ND	0.03
Trichloroethene	uMol/L	ND	ND	ND	0.08	0.12	0.02 J	18.26 D	ND	7.23 D	9.89	0.49	0.99	ND	ND	ND	ND	ND
cis -1,2-DCE	uMol/L	0.23	0.39	ND	0.27	0.18	1.44	113.46 D	3.61 J	72.2 D	8.77	0.35	44.35 D	ND	ND	ND	ND	0.01 J
Vinyl Chloride	uMol/L	0.18 J	ND	0.04 J	ND	0.01 J	0.02 J	0.59 J	ND	3.36 JD	ND	ND	19.2 JD	ND	ND	ND	ND	ND
Alkalinity, Total (as CaCO <sub>3</sub> )	mg/L	300	600	420	170	230	1000	360	200	270	350	400	260	210	130	300	330	310
Oxidation-Reduction Potential	millivolts	-63.5	-125.2	-60.7	157.3	-29.4	-31.8	83.5	85.4	-37.5	19.8	129.9	-96.8	126.7	46	114.8	80.4	155.2
pH	--	6.86	7.06	7	7.01	6.82	6.64	6.19	6.78	6.42	6.62	6.85	6.52	7.04	7.26	7.3	7.25	7.14
Specific Conductance	MS/cm	1.667	1.408	1.902	0.564	0.804	1.409	1.456	0.692	1.43	1.687	1.859	4.496	0.809	0.773	0.796	0.79	0.722
Temperature	deg C	13.4	13.46	14.89	17.99	16.61	17.89	17.13	17.39	16.72	16.3	15.37	15.9	15.42	15.33	15.75	15.94	15.32
Total Organic Carbon	mg/L	12	45	11	ND	ND	5.7	2.9	2	6.1	3.3	ND	12	ND	ND	ND	ND	ND
Chloride	mg/L	230	130	430	71	72	150	280	98	250	290	480	860	90	88	88	91	99
Dissolved Oxygen	mg/L	0.79	0.55	1.17	4.77	0.42	1.5	0.94	1.81	2.17	2.71	1.63	1.73	2.39	0.67	1.53	1.81	2.06
Nitrate-NO <sub>3</sub>	mg/L	0.65	ND	ND	5.1	3.3	ND	0.32	5.4	ND	ND	1.9	0.82	5.8	4.2	4.5	4.8	5
Nitrite-NO <sub>2</sub>	mg/L	ND	ND	ND	ND	ND	ND	ND	0.12	ND	ND	ND	ND	ND	ND	ND	ND	ND
Ferrous Iron	mg/L	9.2	25.6	5.1	0.05	ND	1.63	0.06	0.22	0.53	0.65	0.07	5.4	ND	ND	0.67	0.01	0.13
Sulfate	mg/L	57	2.1	15	35	44	61	94	33	120	100	52	45	45	48	47	46	47
Sulfide	mg/L	ND	0.056	ND	0.048	0.083	0.046	0.095	0.034	ND	0.1	0.038	0.4	ND	ND	0.096	ND	0.11
Ethane	ug/L	ND	ND	ND	ND	ND	3.2	10	3.7	9.3	90	ND	310	ND	ND	ND	ND	ND
Ethene	ug/L	ND	ND	ND	ND	ND	ND	ND	ND	41	33	ND	700	ND	ND	ND	ND	ND
Methane	ug/L	960	130	1500	ND	2.1	45	170	31	95	39	ND	1400	ND	ND	ND	ND	2

Concentrations Measured in March, 2012

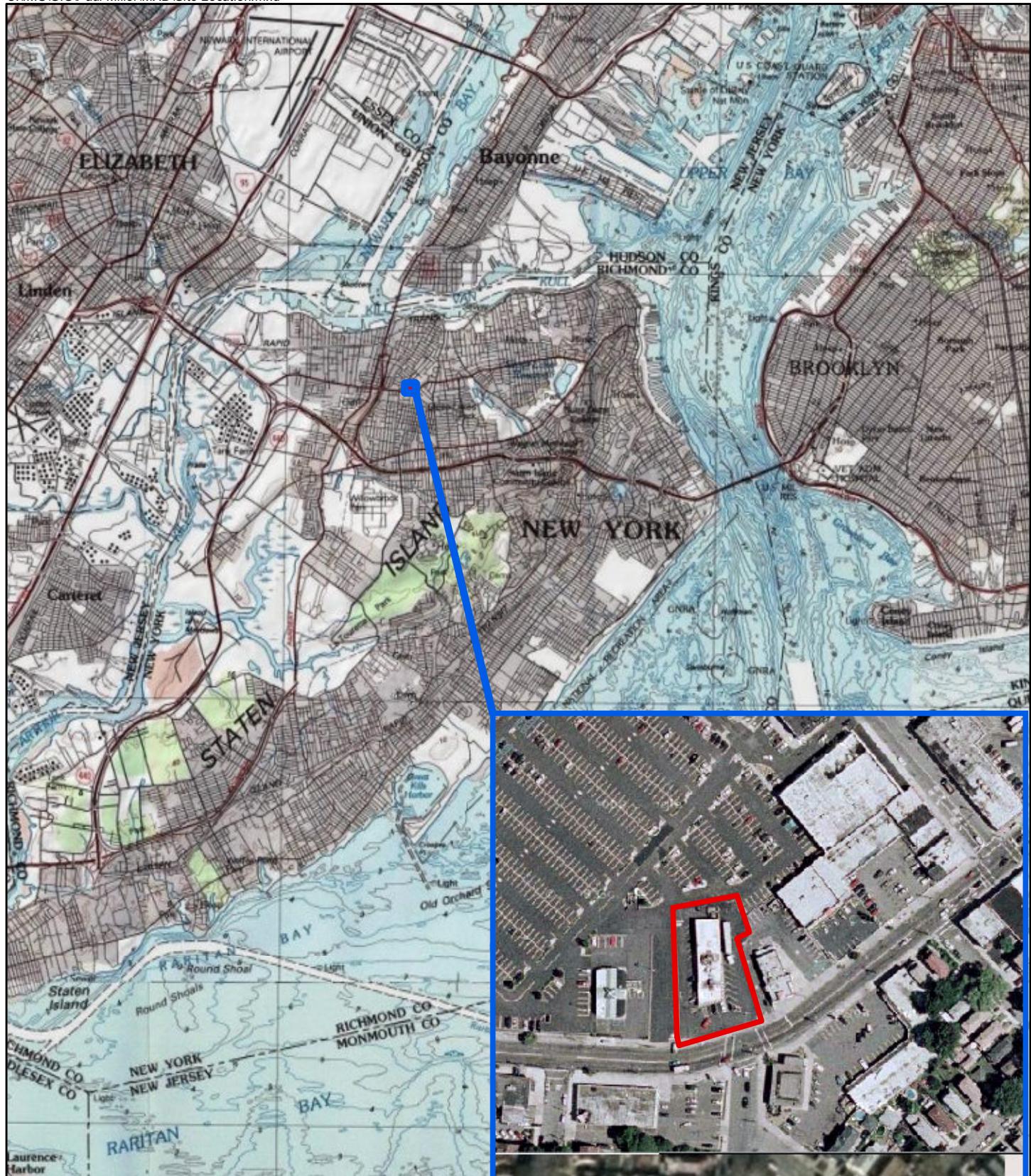
**EXPLANATION OF SIGNIFICANCE:**

<b>pH</b>	Reductive pathway for chlorinated ethenes is most favorable at a relatively neutral pH (5 to 9 pH units).
<b>Dissolved Oxygen</b>	Oxygen is the preferred electron acceptor for microorganisms that facilitate the biodegradation of chlorinated hydrocarbon compounds, followed by nitrate, iron (III), and sulfate. With the presence of high DO concentrations, the reductive pathway of chlorinated ethenes will be suppressed (DO > 0.5 mg/L) or even inhibited (DO > 5 mg/L); on the other hand, VC is oxidized under aerobic conditions.
<b>Temperature</b>	Active biological growth can generally occur under temperatures ranging from 10 to 35 degrees Celsius (°C), and biochemical process can be accelerated when the temperature is above 20 °C.
<b>Oxidation Reduction Potential</b>	ORP is an indicator of the ability of a solution to accept or transfer electrons. The most common electron acceptors in subsurface include DO, nitrate, manganese (IV), iron (III), sulfate, and carbonate, which require increasingly reducing conditions. The reductive pathway of chlorinated hydrocarbon compounds occurs more rapidly under sulfate-reducing and methanogenic conditions.
<b>Nitrate/Nitrite</b>	Once the available DO has been depleted, nitrate is the next preferred electron acceptor for anaerobic biodegradation of hydrocarbon compounds. When at high concentrations (greater than 1 mg/L), nitrate can compete with or even inhibit the reductive pathway of chlorinated ethenes (EPA 1998).
<b>Ferrous Iron</b>	Once the available DO and nitrate sources have been depleted, biologic available iron (III) in the subsurface solid matrix can be utilized as alternate electron acceptor. Iron (III) is reduced into more soluble form, ferrous iron, and thus enters the groundwater. An increase of ferrous iron concentration compared to background levels often indicates an anaerobic biodegradation using iron (III) as an electron acceptor is occurring or has occurred at a Site.
<b>Sulfate</b>	Once the available DO has been depleted and only if insufficient nitrate and iron (III) sources are present, sulfate may be utilized as an alternate electron acceptor. Sulfate is reduced during anaerobic biodegradation to sulfide. At high concentrations (greater than 20 mg/L), sulfate may compete with reductive pathway of chlorinated ethenes.
<b>Chloride</b>	Chloride is an end-product of biodegradation of chlorinated ethenes. Therefore, elevated chloride concentrations above Site background levels (i.e., greater than two times) indicate biodegradation.
<b>Methane</b>	Methane is a by-product of anaerobic biodegradation. Therefore, elevated methane concentrations (above Site background levels) are indicative of biodegradation in an anaerobic environment. As discussed earlier, reductive dechlorination occurs more rapidly under methanogenic conditions. Furthermore, ultimate reductive daughter products will accumulate when methane concentration is greater than 0.5 mg/L in groundwater.
<b>Alkalinity</b>	Increased biological activities often results in elevated carbon dioxide concentrations in groundwater, and hence increased alkalinity due to the reaction between carbon dioxide and aquifer minerals. Therefore, elevated alkalinity (i.e., greater than two times of Site background levels) is indicative of biological activity and potential biodegradation.
<b>Ethene/Ethane</b>	Ethene is daughter product of reduction of vinyl chloride, while ethane can be daughter product of reduction of ethene or chloroethane. Therefore, elevated ethane and/or ethane concentrations (above Site background levels and greater than 10 µg/L) are indicative of biodegradation in an anaerobic environment.
<b>Total Organic Carbon</b>	By acting as electron donor, organic carbon is necessary for sustaining bacteria growth and reductive dechlorination. TOC is an indicator of the carbon content in the subsurface.

**Table 5-3**  
**Natural Attenuation Analysis**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

	<b>Wells screened at the water table (MW-02, MW-03, MW-04)</b>	<b>Under suspected primary discharge (MW-11S, MW-12S, MW-13S, MW-14S)</b>	<b>Under suspected additional discharge (MW-01 and MW-16S)</b>	<b>Downgradient (MW-09S)</b>	<b>Cross-gradient (MW-08S)</b>	<b>Deep Wells (MW-09D, MW-10D, MW-11D, MW-13D, MW-15D)</b>
<b>Degradation Byproducts</b>	Yes	Yes	Yes – more so in MW-01 than MW-16S	Yes	<i>Cis</i> -1,2-DCE, but no VC. Relatively little compared to parent products	N/A except for MW15D, which has <i>cis</i> -1,2-DCE
<b>ReDox conditions</b>	Methanogenic	Aerobic to nitrate reducing	MW-01 is methanogenic; MW-16S is aerobic	Nitrate reducing	Aerobic	Aerobic to nitrate reducing
<b>Available carbon</b>	Carbon limited, except for MW-03	Carbon limited	Carbon limited	Carbon limited	Carbon limited	Carbon limited

## Figures



Site Boundary



Map source: USGS

0 1 2 4 Miles

**Figure 1-1**  
**Site Location**  
**Former Paul Miller Dry Cleaners**  
**Port Richmond, NY**

**CDM  
Smith**

**FINANCE  
NEW YORK**  
MARTHA E. STARK  
COMMISSIONER

- Boroughs
- Tax Blocks
- Tax Lots
- Boundary Lines
- Streets
- Edge of Pavement
- Building Footprints
- Community Districts



Figure 1-2  
Tax Map  
Former Paul Miller Dry Cleaners  
Port Richmond, NY

**Legend**

● MIP Location



## MIP Investigation Locations

0 25 50 75 100  
Feet

**CDM  
Smith**

**Figure 2-1**

Former Paul Miller Dry Cleaners  
Port Richmond, NY



### Legend

- Groundwater Screening Location
- Soil Boring Location
- Groundwater Monitoring Well Location



**Groundwater Screening, Monitoring Well, and Soil Boring Investigation Locations**

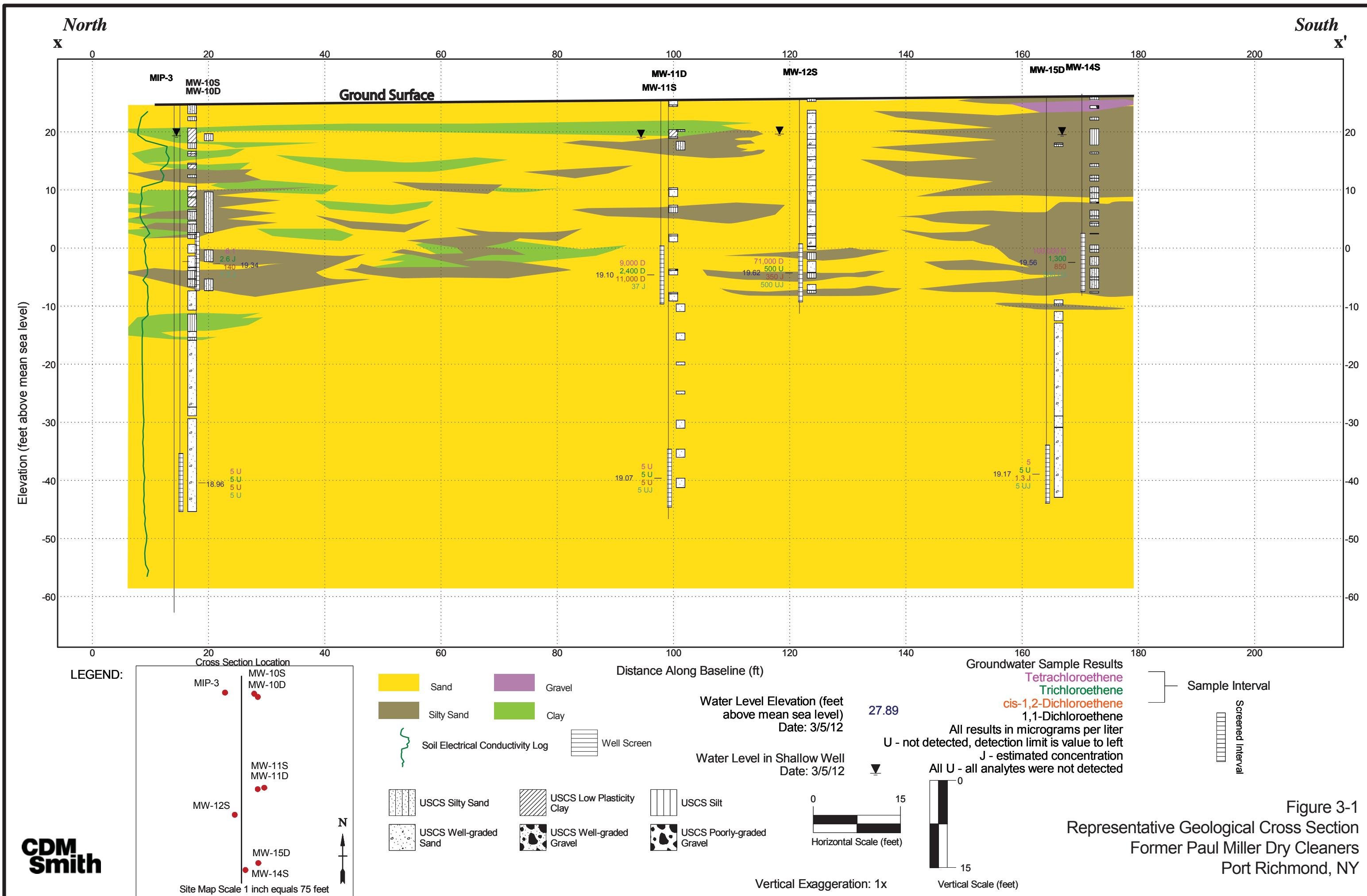
**Figure 2-2**

Former Paul Miller Dry Cleaners  
Port Richmond, NY

Note: Two data frames are shown in order to clearly show all locations.

0 10 20 30 40  
Feet

**CDM  
Smith**



**Legend**

- March 2012 Water Table (ft amsl)
- Groundwater Monitoring Well Location (water level elevation in ft amsl)

**Potentiometric Surface at the Water Table**

Note: This potentiometric surface affects the wells shown.  
Elevations measured on 3/5/12.

0 10 20 30 40  
Feet

**Figure 3-2**

Former Paul Miller Dry Cleaners  
Port Richmond, NY

**CDM  
Smith**

**Legend**

● Groundwater Monitoring Well Location (water level elevation ft amsl)

— March 2012 Shallow Potentiometric Surface (ft amsl)



### Potentiometric Surface at 30 ft bgs

Note: This potentiometric surface

affects the wells shown.

Elevations measured on 3/5/12.

0 10 20 30 40  
Feet

**Figure 3-3**

Former Paul Miller Dry Cleaners  
Port Richmond, NY

**CDM  
Smith**

**Legend**

- Groundwater Monitoring Well Location (groundwater elevation ft amsl)
- March 2012 Deep Potentiometric Surface (ft amsl)

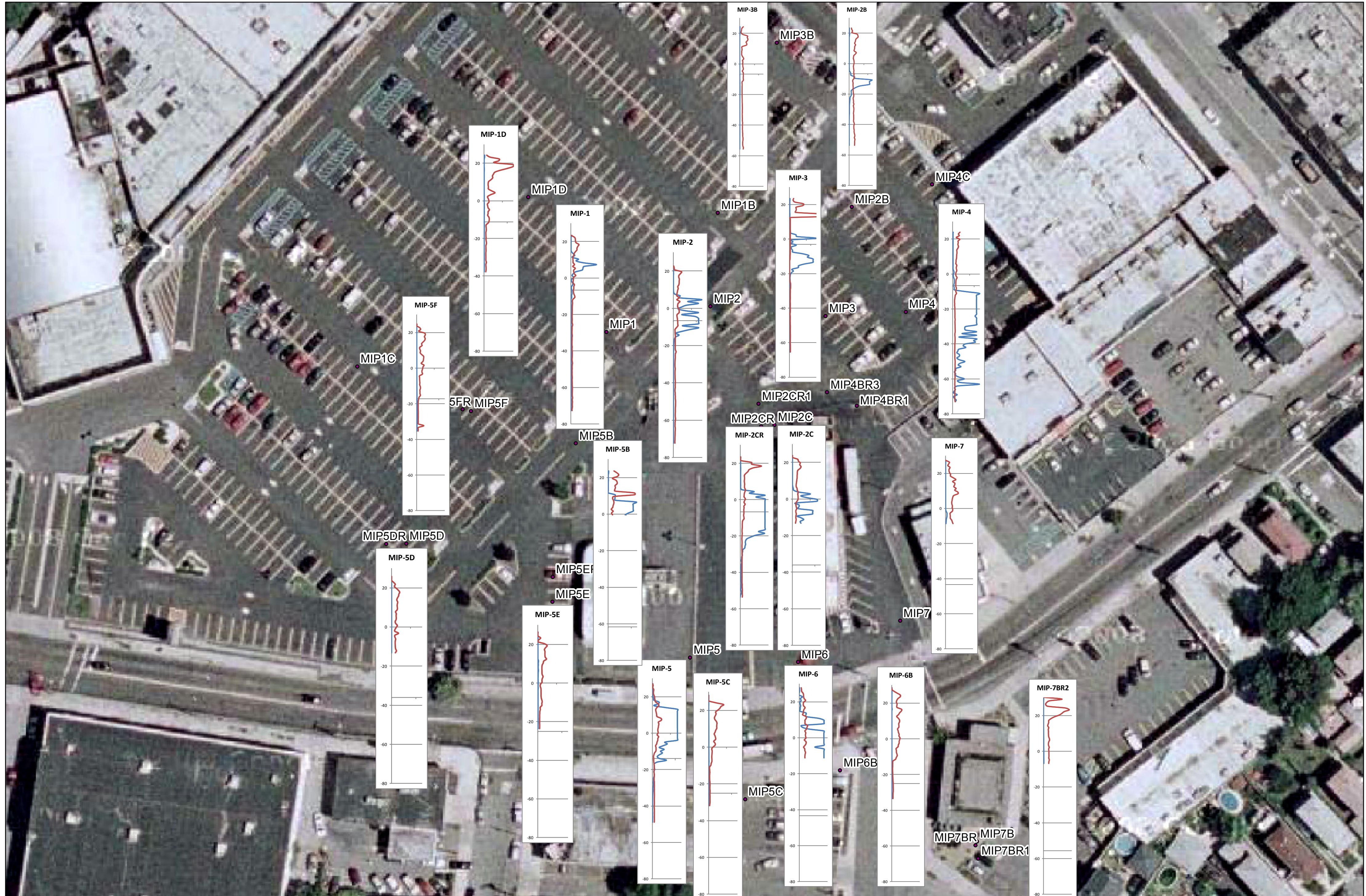
**Potentiometric Surface at 70 ft bgs**

Note: This potentiometric surface  
affects the wells shown.  
Elevations measured 3/5/12.

0 10 20 30 40  
Feet

**Figure 3-4**  
Former Paul Miller Dry Cleaners  
Port Richmond, NY

**CDM  
Smith**



### Legend



• MIP Locations

### NOTES:

- 1) Y axis units are feet above mean sea level.
- 2) Data collected between May and June, 2011.
- 3) Red line presents relative Soil Conductivity. HIGHER results indicate MORE fine grain sizes (such as silt and clay). LOWER results indicate MORE sand.
- 4) Blue line presents Electron Capture Detector (ECD) results. HIGHER results indicate HIGHER detections of VOCs in the soil or aquifer.

### MIP Results

0 25 50 100 150 200 Feet

**Figure 4-1**

Former Paul Miller Drycleaners  
Port Richmond, NY

**CDM  
Smith**

MW-1

ENCLOSURE II

SB-6A	
Depth	0.5
Date	11/1/2011
cis-1,2-DCE	12
TCE	6.6 J
PCE	26 J
Vinyl Chloride	6.7 U

SB-6B	
Depth	5
Date	11/1/2011
cis-1,2-DCE	5.7 U
TCE	5.7 U
PCE	5.7 U
Vinyl Chloride	5.7 U

SB-4A	
Depth	0.5
Date	11/1/2011
cis-1,2-DCE	5.9 U
TCE	5.9 U
PCE	16 J
Vinyl Chloride	5.9 U

PM-SB-6-A PM-SB-6-B

PM-SB-4-A

SB-3A	
Depth	0.5
Date	11/1/2011
cis-1,2-DCE	5.9 U
TCE	5.9 U
PCE	22 J
Vinyl Chloride	5.9 U

PM-SB-5-A PM-SB-5-B

PM-SB-3-A

SB-5A	
Depth	0.5
Date	11/1/2011
cis-1,2-DCE	12
TCE	16 J
PCE	110 J
Vinyl Chloride	6.8 U

SB-5B	
Depth	6
Date	11/1/2011
cis-1,2-DCE	5.4 U
TCE	5.4 U
PCE	4.5 J
Vinyl Chloride	5.4 U

☒ MW-12S

SB-1A-A	
Depth	0.5
Date	11/1/2011
cis-1,2-DCE	5.8 U
TCE	5.8 U
PCE	5.8 U
Vinyl Chloride	5.8 U

SB-1B-A	
Depth	0.5
Date	11/2/2011
cis-1,2-DCE	5.9 U
TCE	5.9 U
PCE	2.7 J
Vinyl Chloride	5.9 U

SB-2A	
Depth	0.5
Date	11/1/2011
cis-1,2-DCE	3.5 J
TCE	5.9 U
PCE	5.9 J
Vinyl Chloride	5.9 U

SB-1A-B	
Depth	5
Date	11/1/2011
cis-1,2-DCE	2.3 J
TCE	2.3 J
PCE	55 J
Vinyl Chloride	5 U

SB-1B-B	
Depth	6
Date	11/2/2011
cis-1,2-DCE	5.5 U
TCE	5.5 U
PCE	5.5 U
Vinyl Chloride	5.5 U

SB-2B	
Depth	5
Date	11/1/2011
cis-1,2-DCE	14
TCE	1.2 J
PCE	9.2 J
Vinyl Chloride	6 U

☒ MW-14S

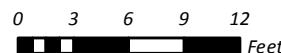
☒ MW-15D

SB-7B	
Depth	12.5
Date	11/2/2011
cis-1,2-DCE	5.2 U
TCE	5.2 U
PCE	5.2 U
Vinyl Chloride	5.2 U

PM-SB-7-B

## Subslab Soil Sample Results

Depth units are feet below slab.  
Analyte units are ug/kg.

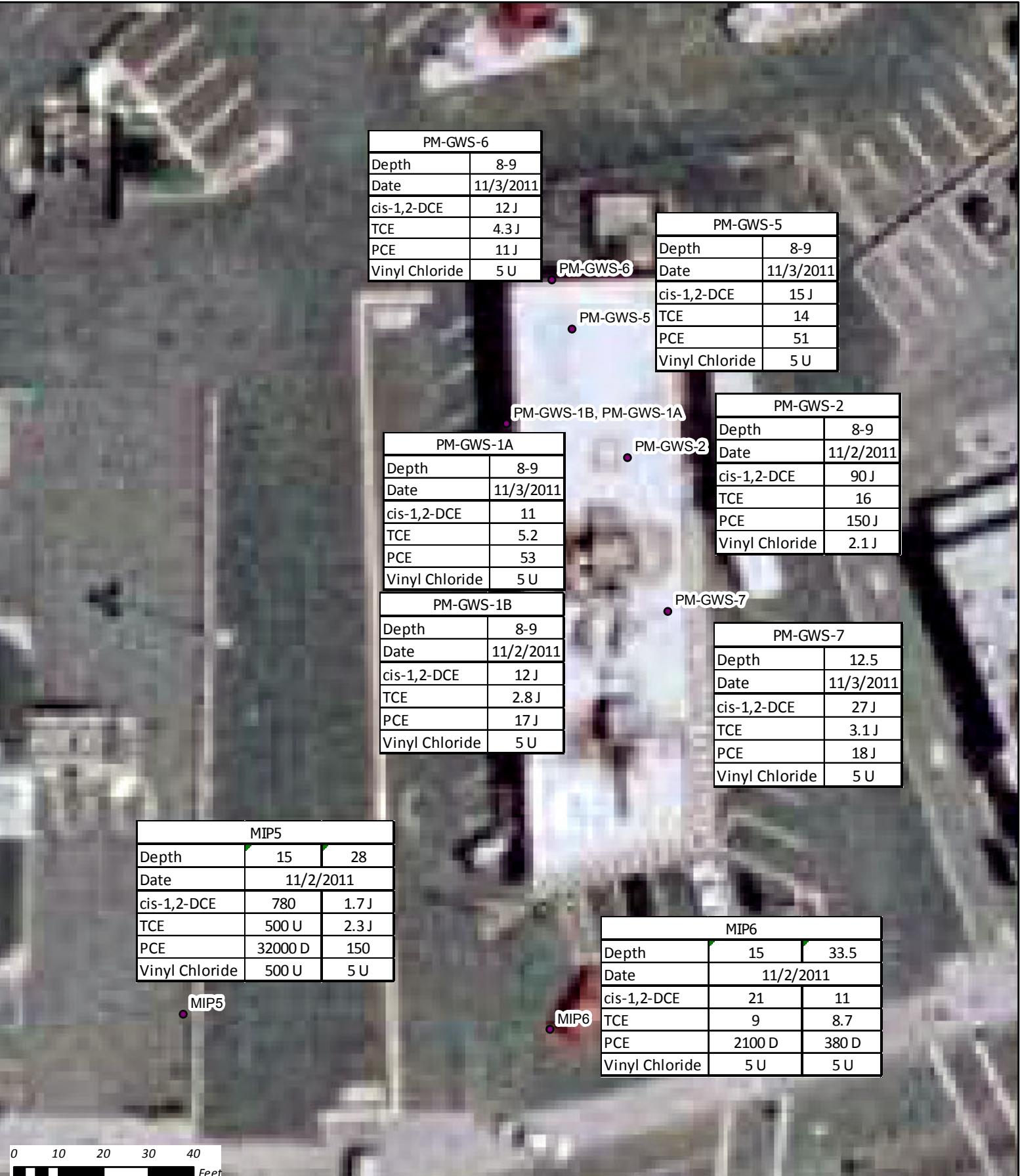


Former Paul Miller Dry Cleaners  
Port Richmond, NY

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Figure 4-2



#### Legend

- Sample\_Locations

Depth units are feet bgs.  
Analyte units are ug/L.

#### Groundwater Screening Results

**Figure 4-3**

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Concentrations are in ug/L  
See Data Usability Summary Report for explanation of qualifiers

MW09S		
Date	10/13/2008	3/5/2012
Vinyl Chloride		0.58 J
cis-1,2-DCE		17
TCE		16
PCE	2400 JD	34

MW09D		
Date	10/13/2008	3/5/2012
Vinyl Chloride		5 UJ
cis-1,2-DCE		5 UJ
TCE		5 UJ
PCE		5 UJ

MW09S  
MW09D

MW10S		
Date	10/13/2008	3/5/2012
Vinyl Chloride		2.60
cis-1,2-DCE		110 D
TCE		8.4 J
PCE		6.1

MW-10D		
Date	10/13/2008	3/5/2012
Vinyl Chloride		5 U
cis-1,2-DCE		5 U
TCE		5 U
PCE		5 U

MW10D MW10S

MW02		
Date	10/14/2008	3/6/2012
Vinyl Chloride		11 J
cis-1,2-DCE		22
TCE		5 U
PCE		5 U

MW02

MW04		
Date	10/14/2008	3/6/2012
Vinyl Chloride		3
cis-1,2-DCE		11
TCE		5 U
PCE		5 U

MW04

MW03		
Date	10/14/2008	3/7/2012
Vinyl Chloride		5 UJ
cis-1,2-DCE		38
TCE		5 U
PCE		5 U

MW03

MW08S		
Date	10/13/2008	3/7/2012
Vinyl Chloride	9.70	5 UJ
cis-1,2-DCE	371 D	26
TCE	92 J	10
PCE	19220 JD	1300 D

MW08S

MW14S		
Date	10/15/2008	3/8/2012
Vinyl Chloride		4.5
cis-1,2-DCE		183 D
TCE		882.2
PCE		160000 D

MW14S

MW-1S		
Date	10/14/2008	3/6/2012
Vinyl Chloride		1200 JD
cis-1,2-DCE		4300 D
TCE		130
PCE		750 D

MW01

MW-16S		
Date	10/14/2008	3/6/2012
Vinyl Chloride		5 UJ
cis-1,2-DCE		100 J
TCE		700 JD
PCE		6500 JD

MW16S

MW-13S		
Date	10/14/2008	3/7/2012
Vinyl Chloride		6.8 J
cis-1,2-DCE		1700 D
TCE		457 JD
PCE		641 D

MW-13D		
Date	10/14/2008	3/7/2012
Vinyl Chloride		5 UJ
cis-1,2-DCE		5 U
TCE		5 U
PCE		6300 JD

#### Legend

- Monitoring Well

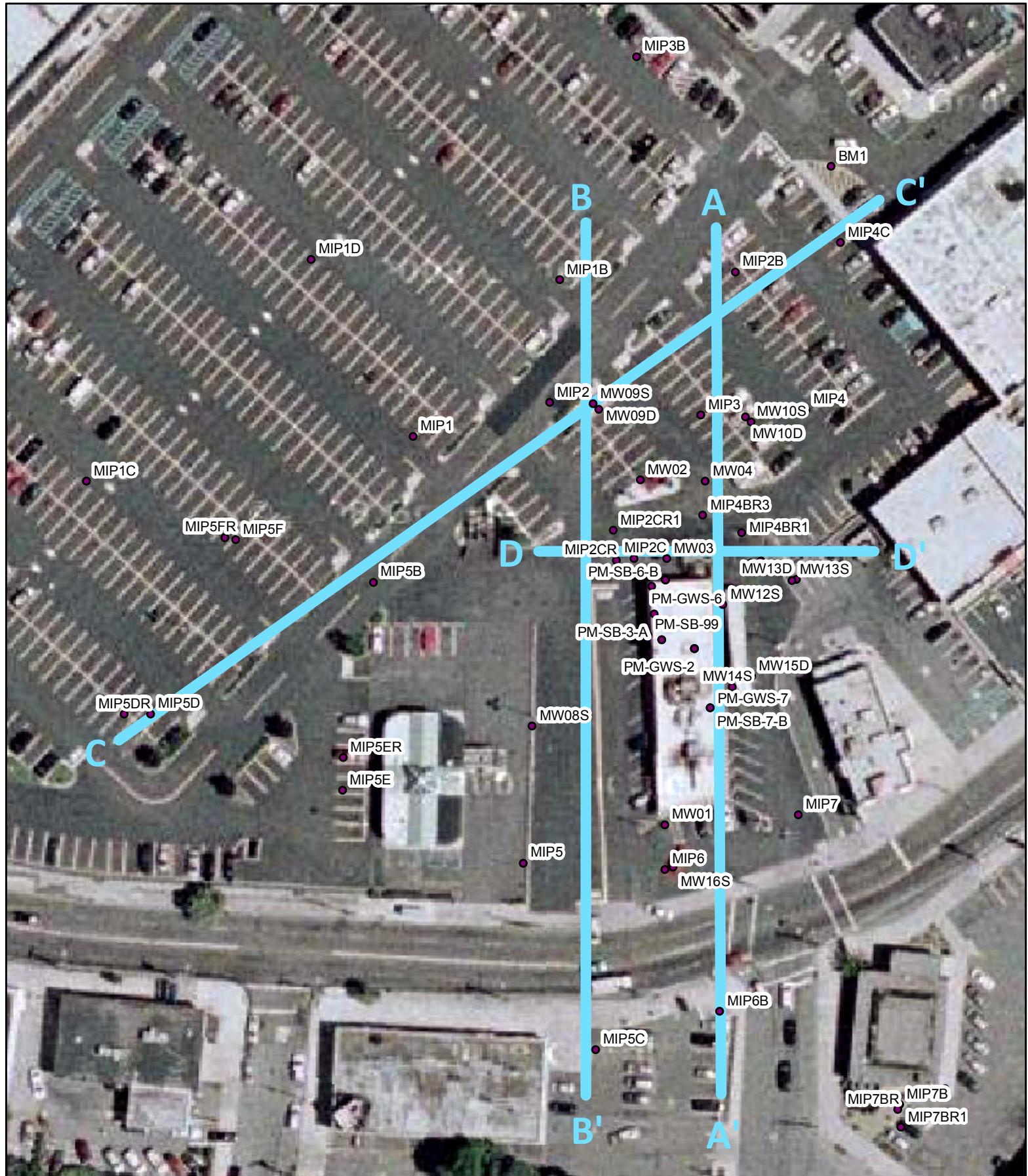


0 25 50 100 Feet

### Groundwater Exceedances

Figure 4-4  
Former Paul Miller Dry Cleaners  
Port Richmond, NY

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## Cross Section Locations



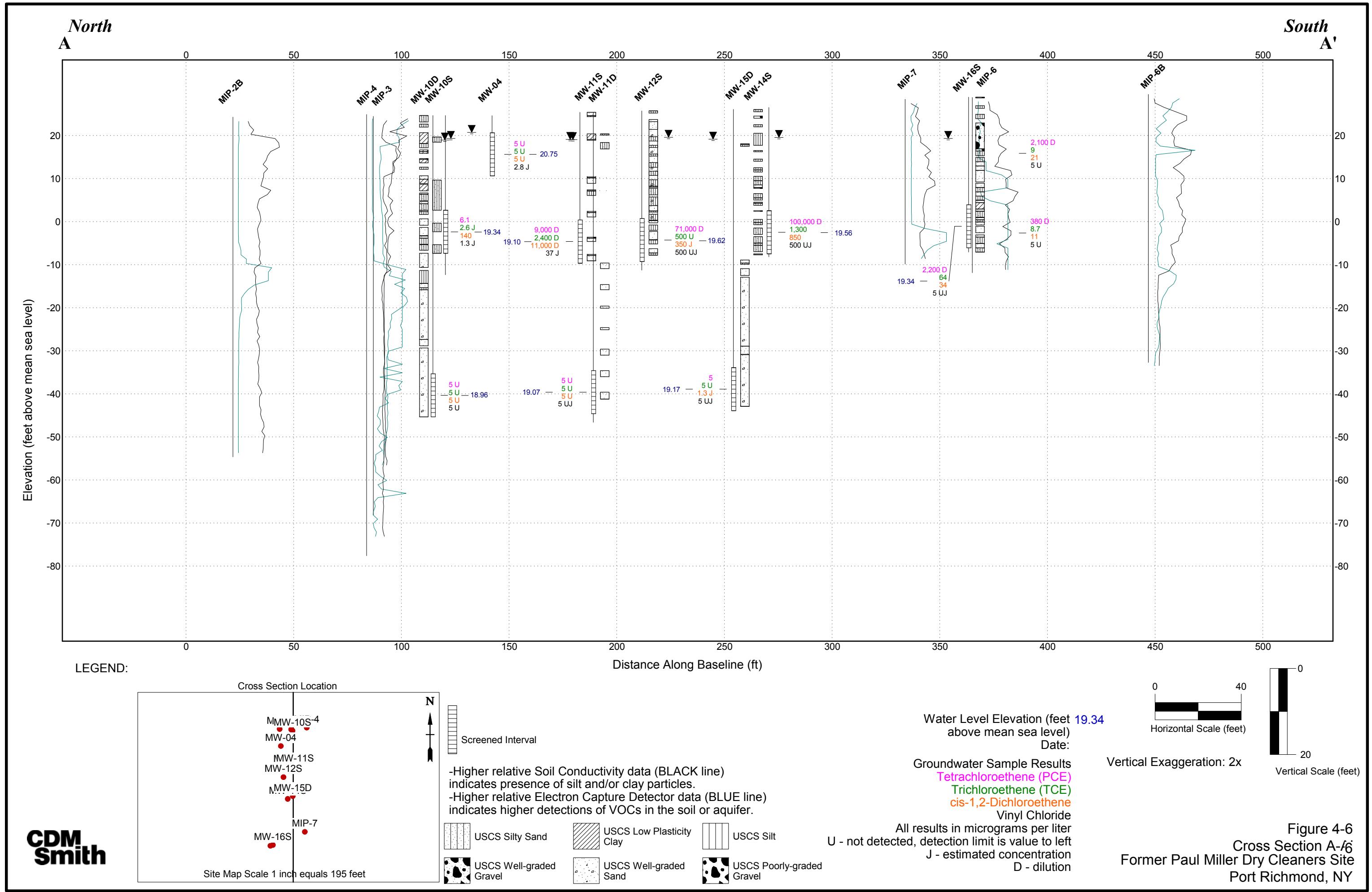
### Legend

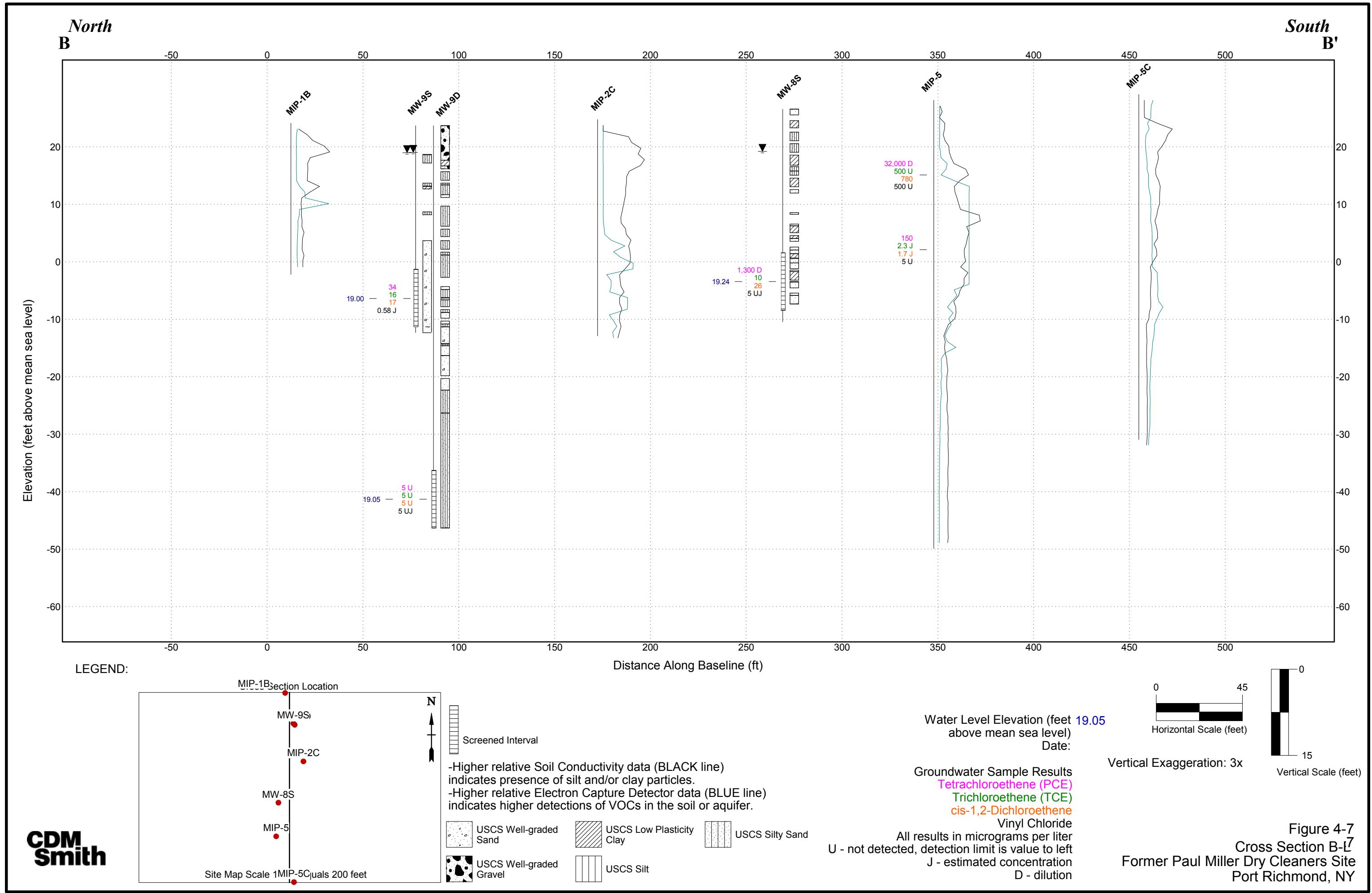
- Sample Locations
- Cross Section Locations

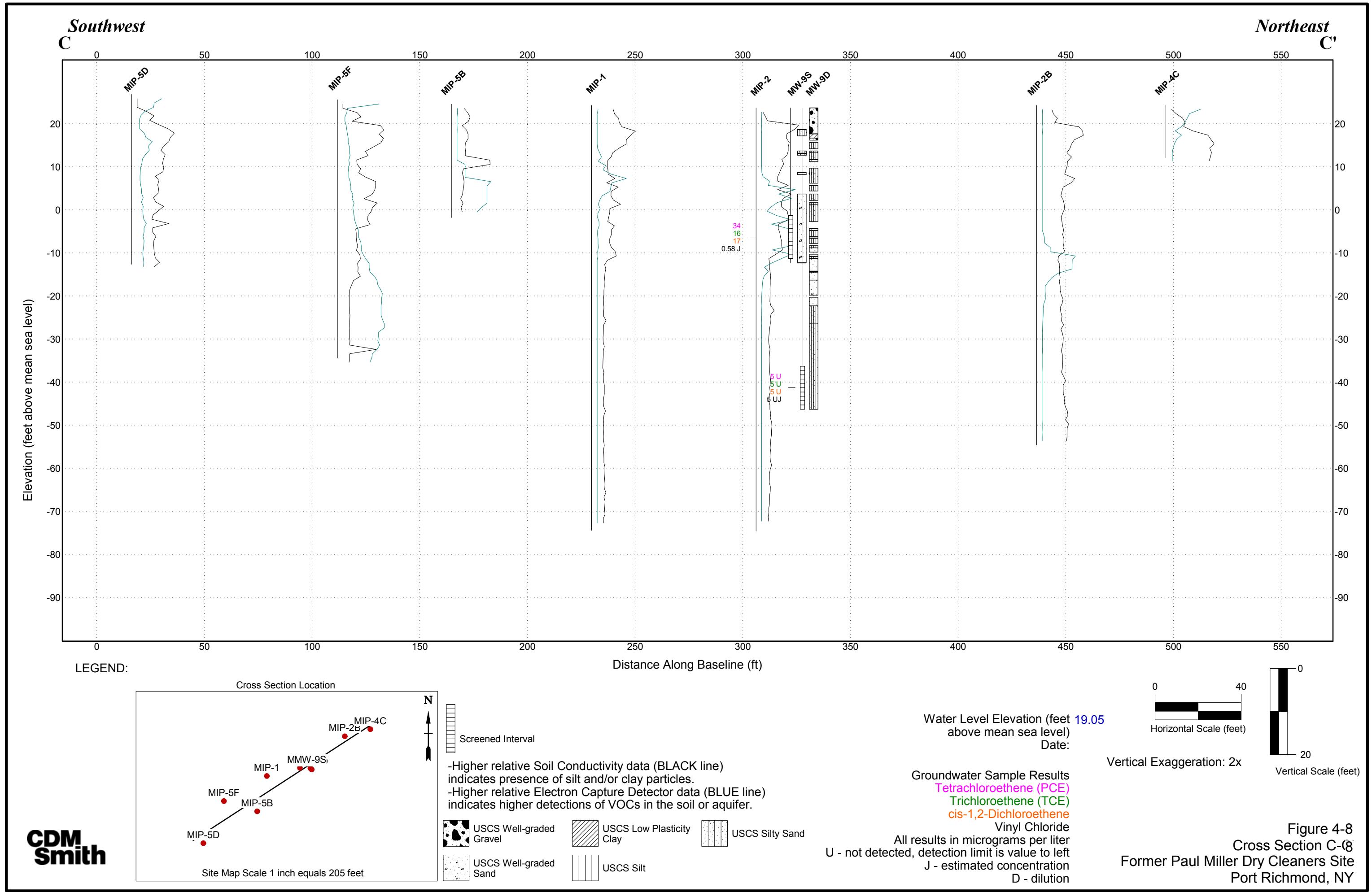
0 25 50 75 100  
Feet

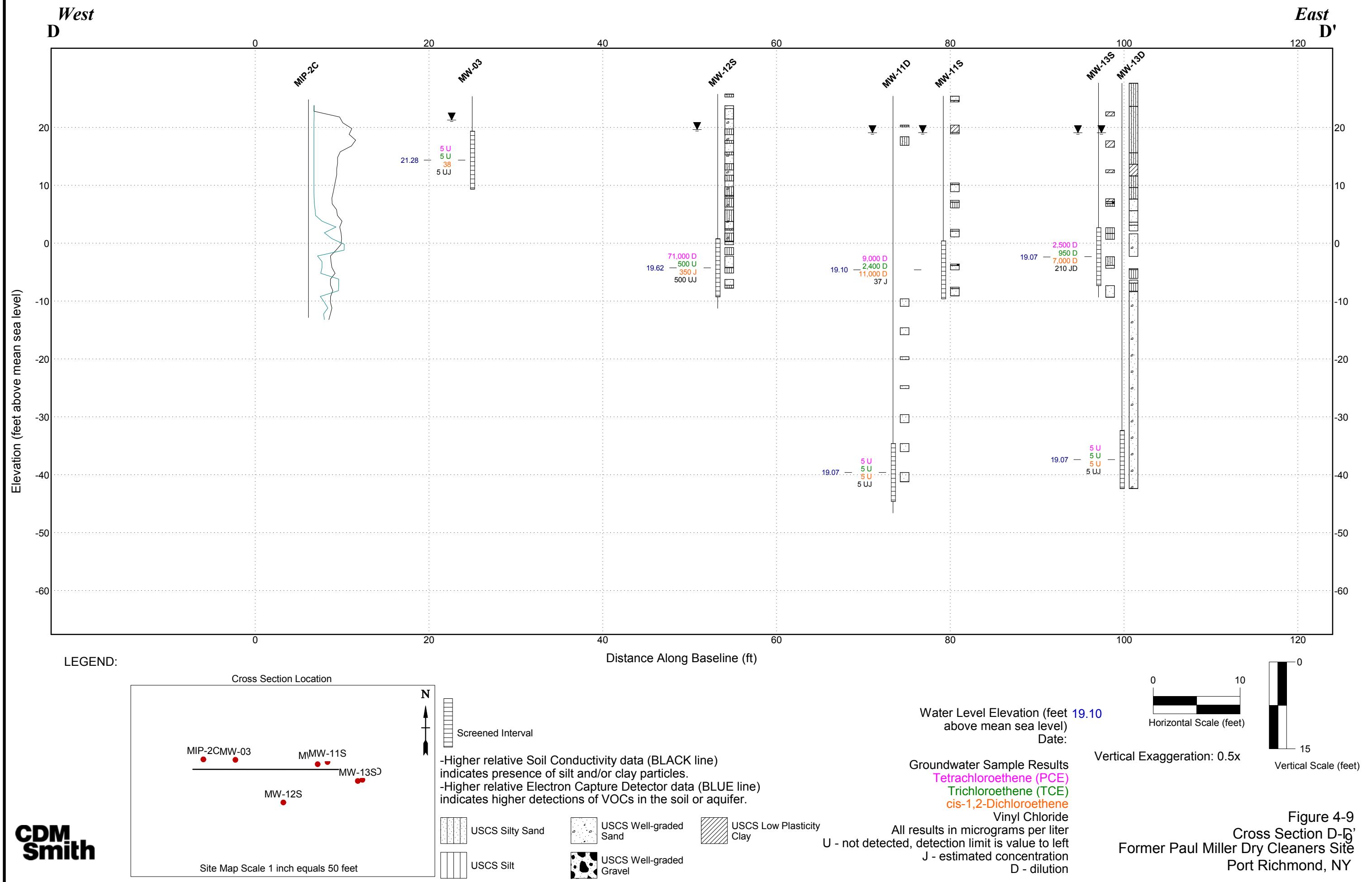
**Figure 4-5**  
Former Paul Miller Dry Cleaners  
Port Richmond, NY

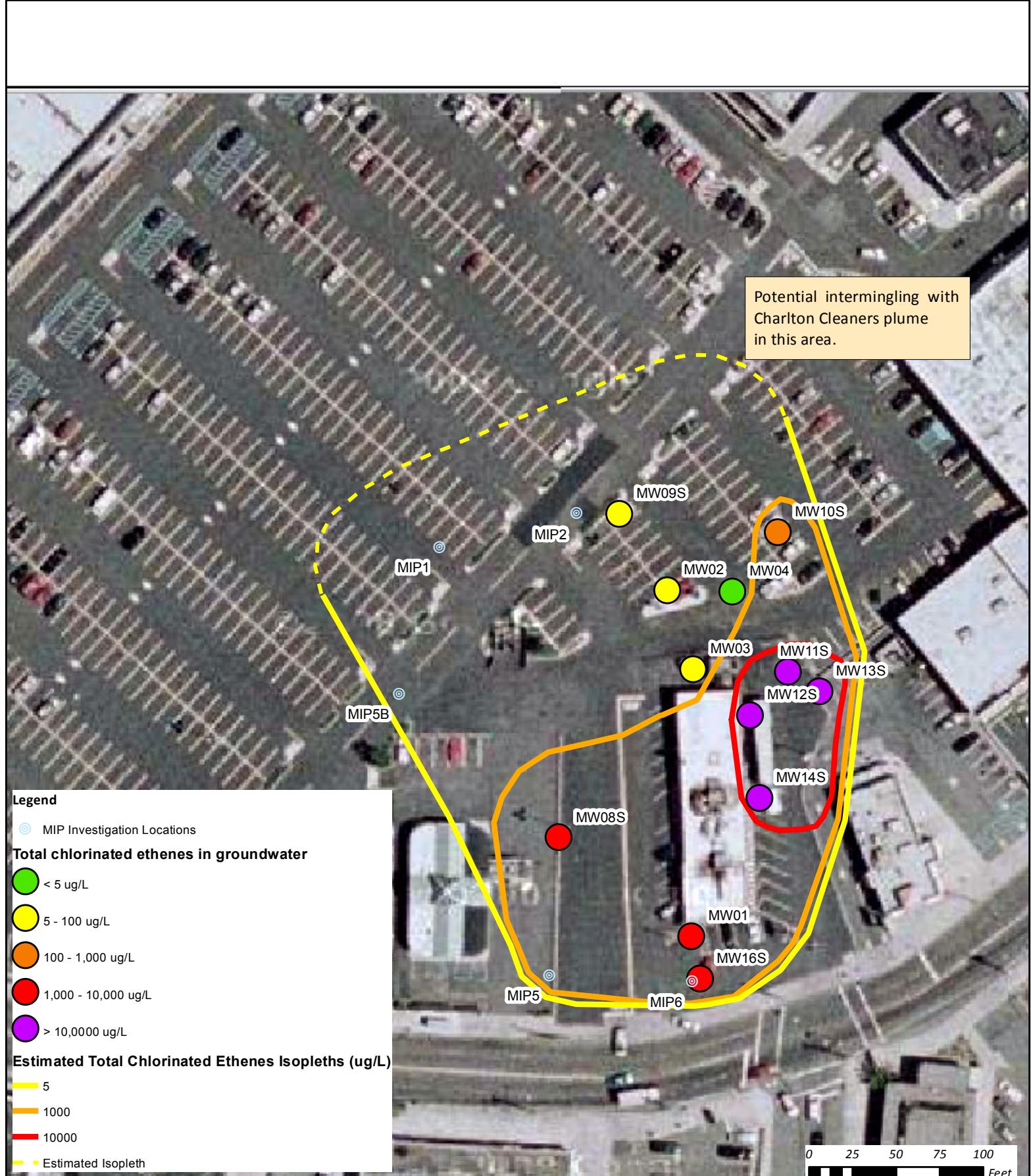
**CDM  
Smith**











### Shallow Groundwater - Total Chlorinated Ethenes Isopleths

Figure 4-10

Former Paul Miller Dry Cleaners  
Port Richmond, NY

Note:

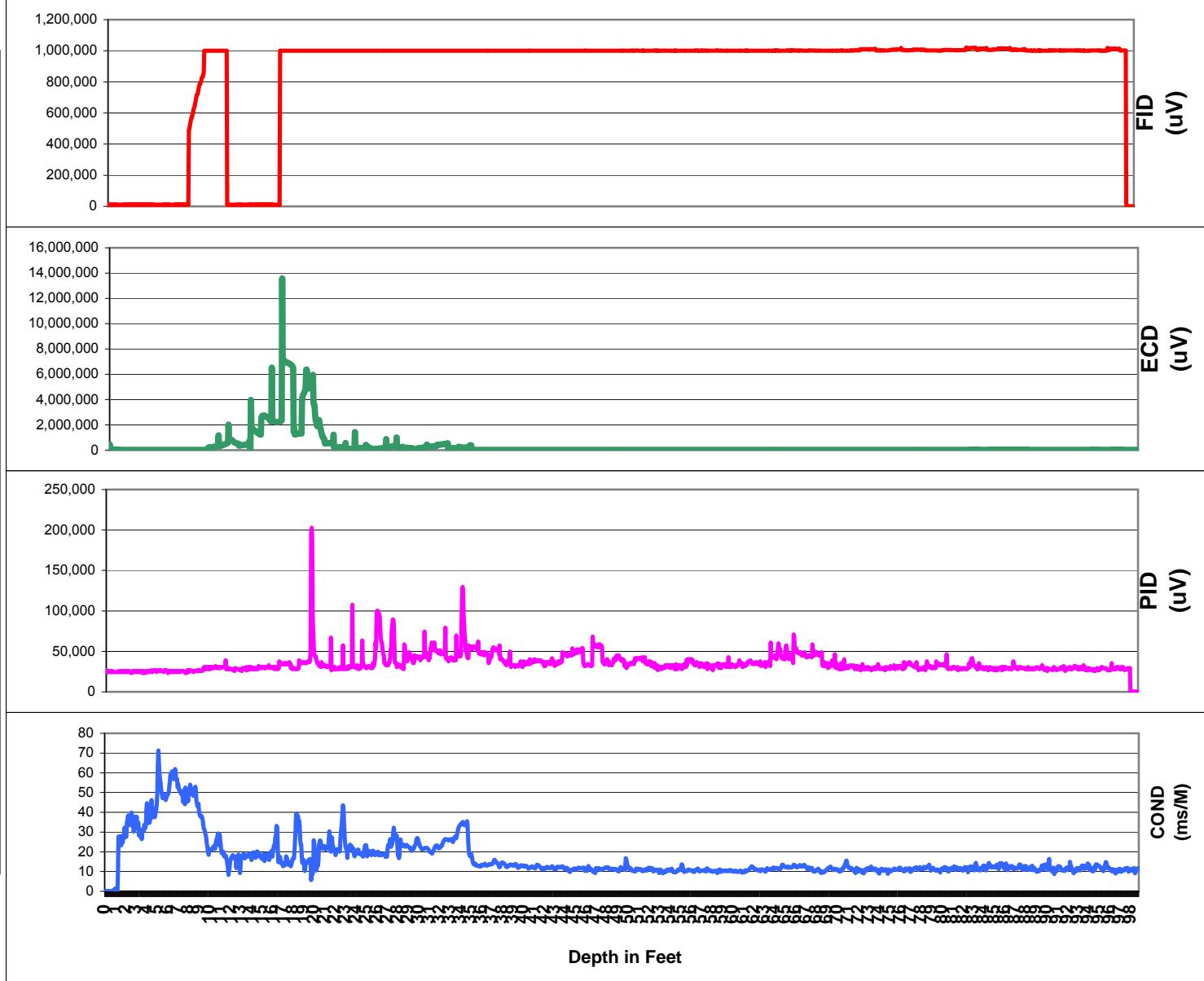
- 1) Shallow groundwater gradient is north/northwest
- 2) Shallow groundwater is less than 35 feet bgs

**CDM  
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## Appendix A

### Membrane Interface Probe Results

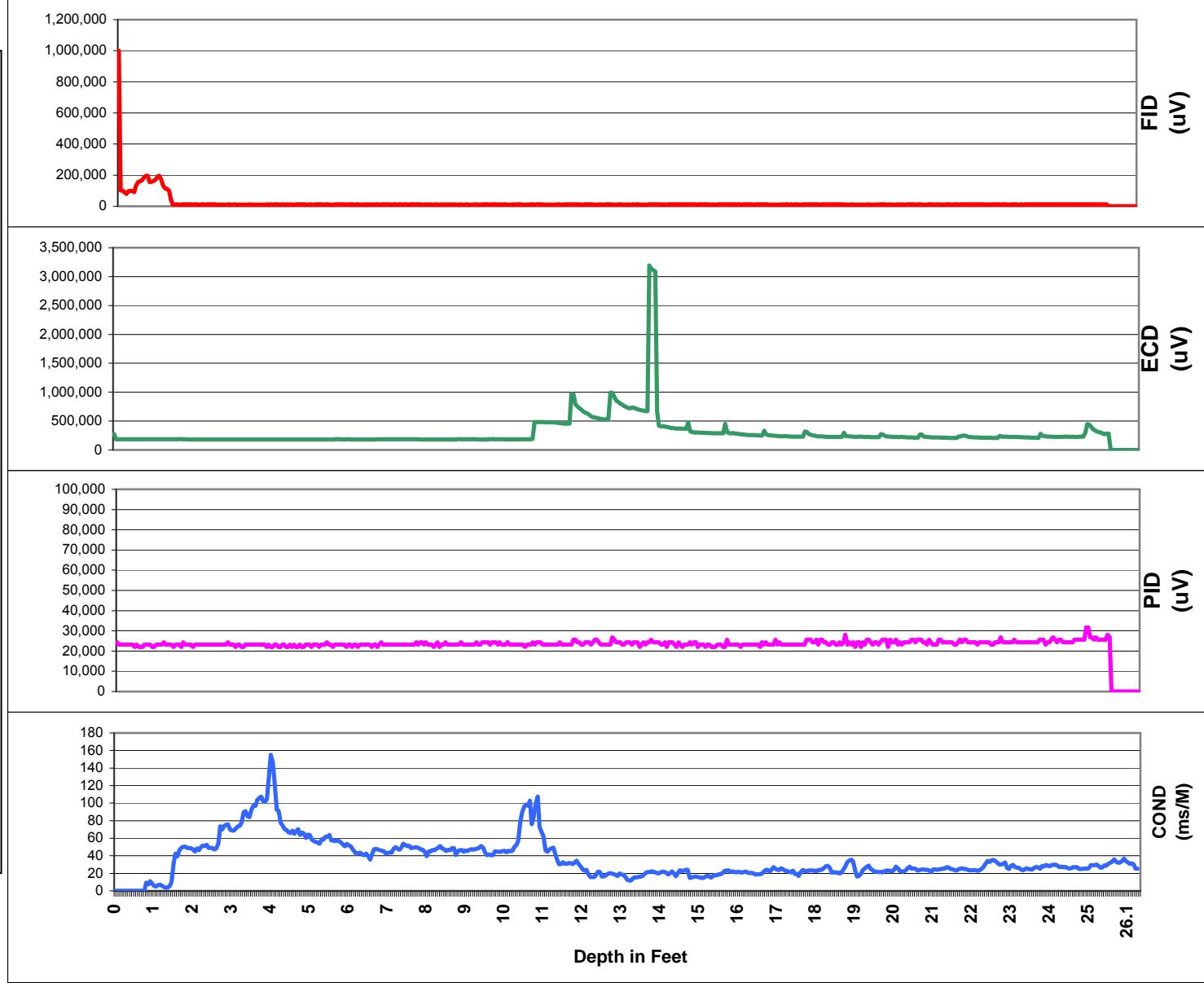
**ZEBRA EC/MIP Summary Log, Point CDMMIP1**  
**Staten Island, NY**



for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

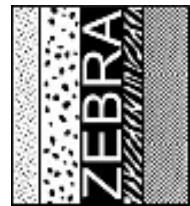
Date: 5/10/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 1 of 0

**ZEBRA EC/MIP Summary Log, Point CDMMIP1B**  
**Staten Island, NY**

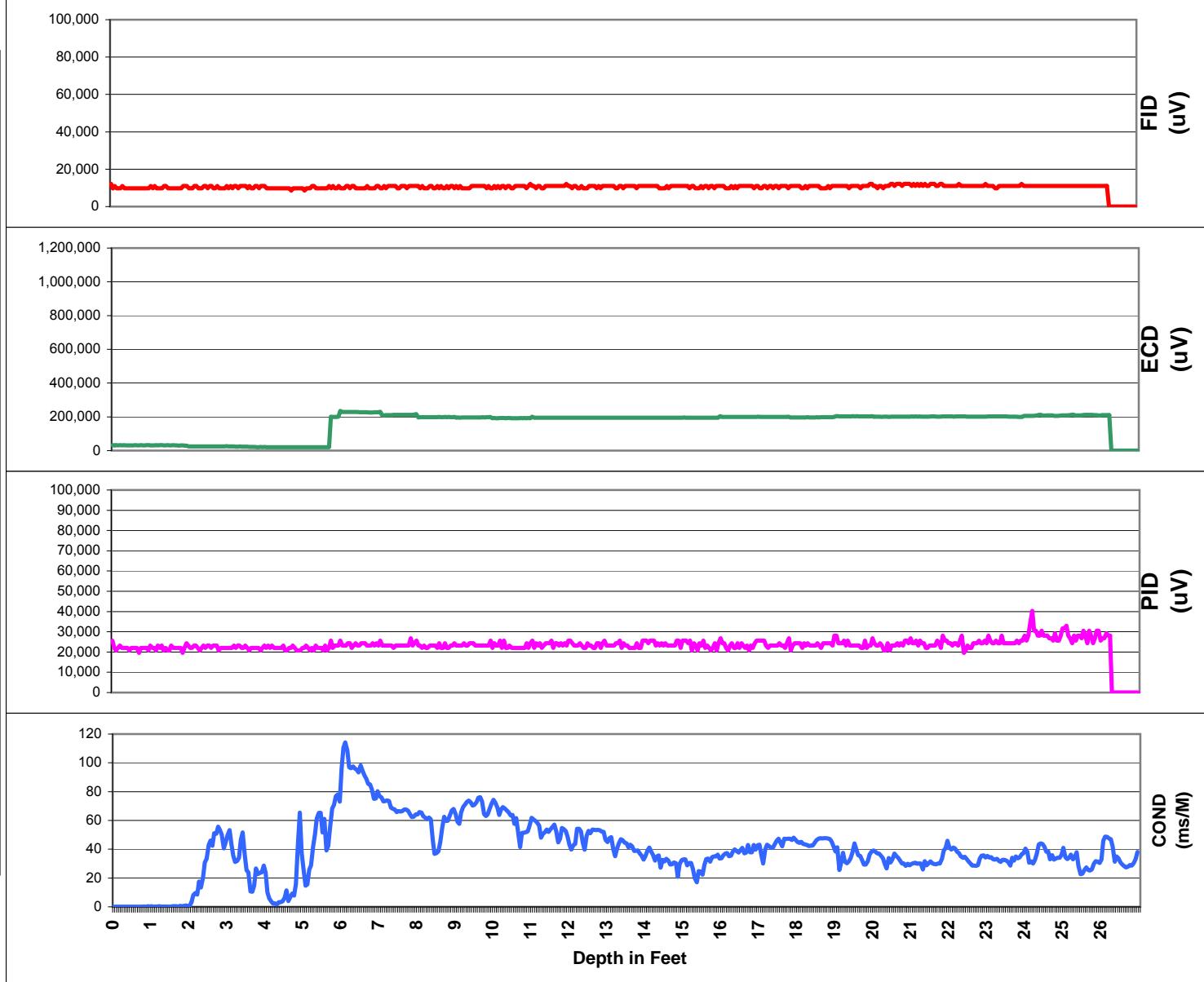


Date: 5/10/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 8 of 0

for: CDM  
by: Zebra Environmental  
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Lynbrook, NY 11563  
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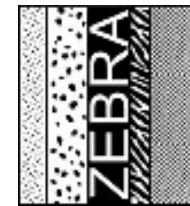
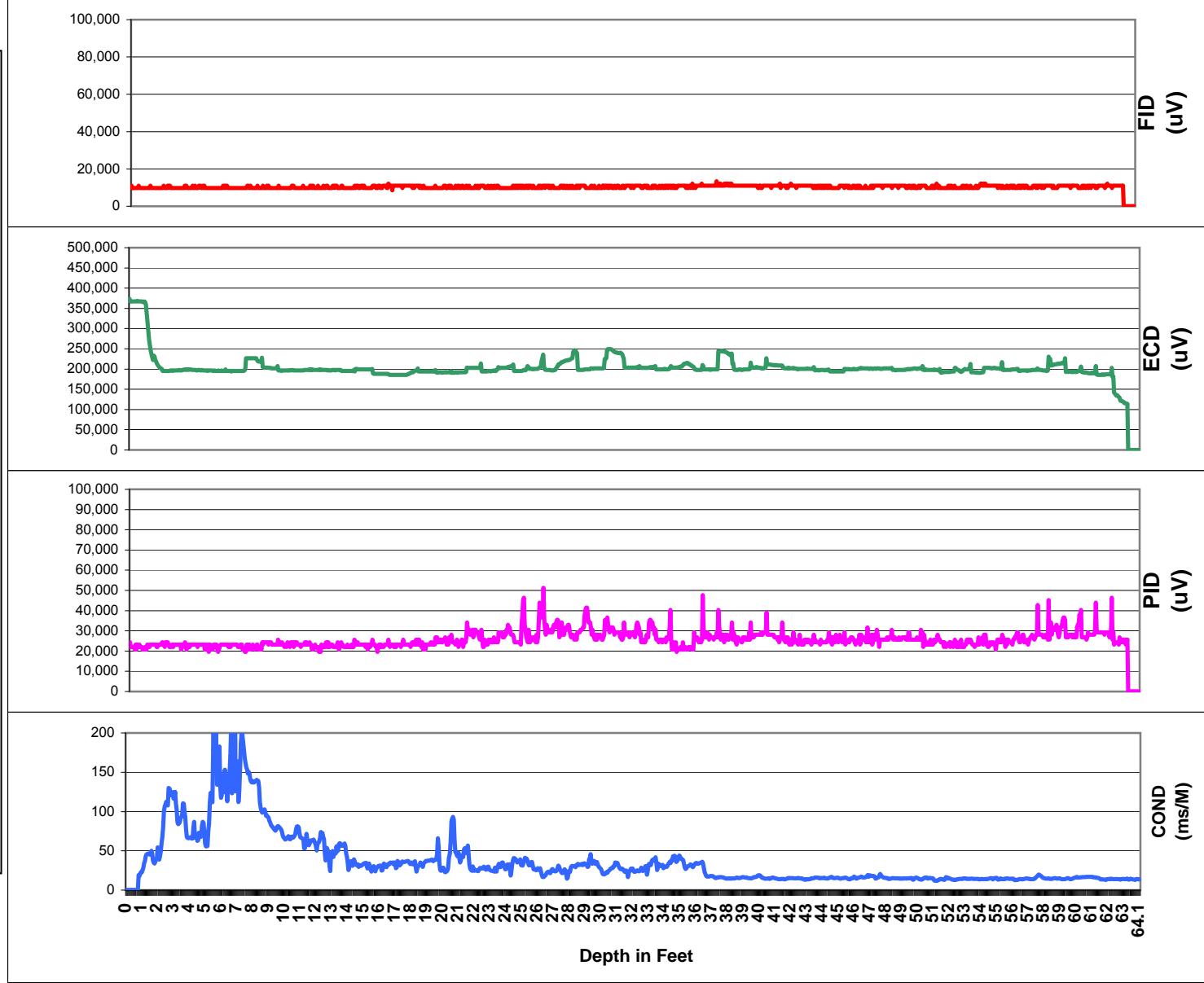
**ZEBRA EC/MIP Summary Log, Point CDMMP1C**  
**Staten Island, NY**



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by: Zebra Environmental  
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Lynbrook, NY 11563  
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Date: 5/21/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 12 of 0

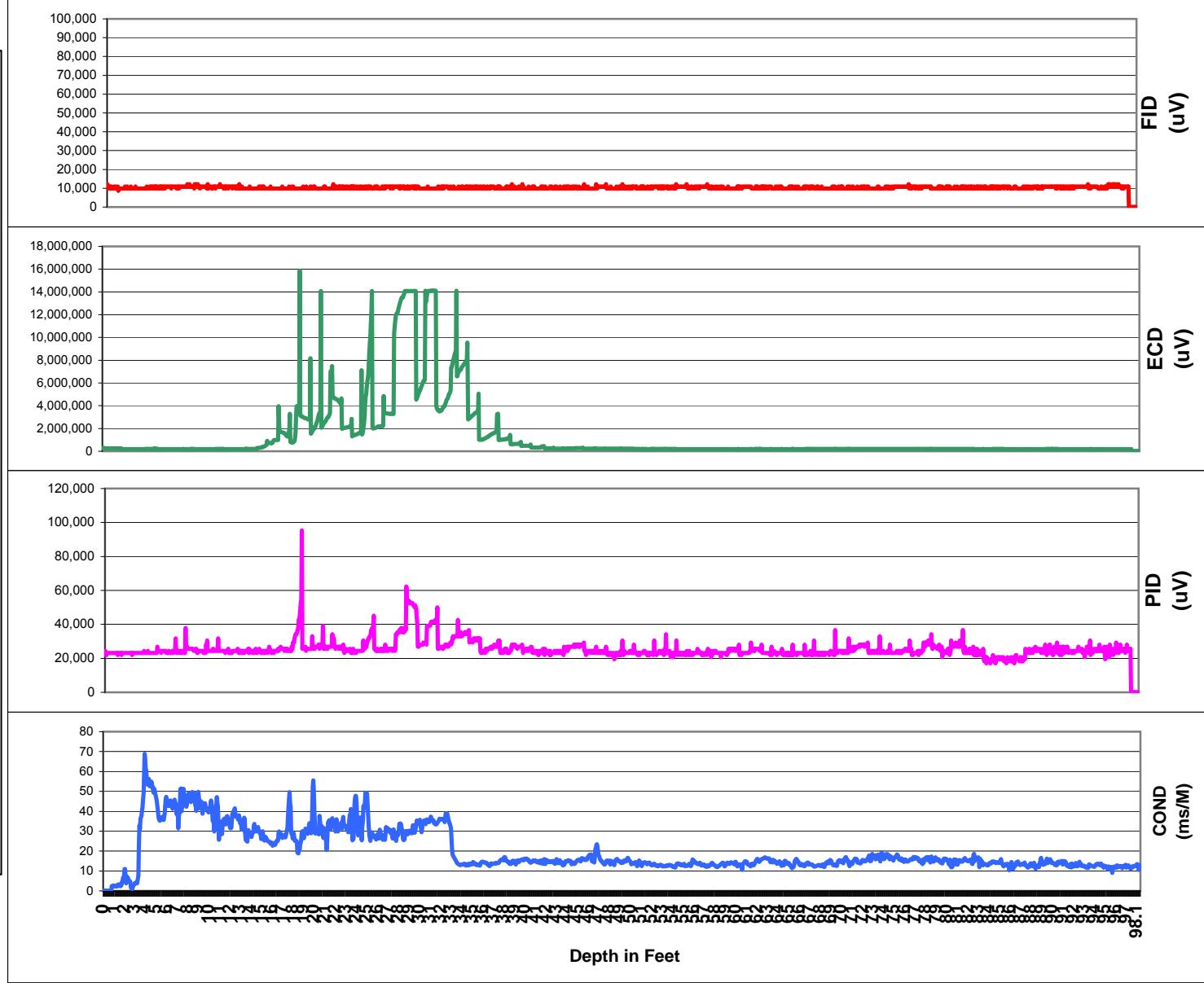
**ZEBRA EC/MIP Summary Log, Point CDMMIPI  
Staten Island, NY**



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by: Zebra Environmental  
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Lynbrook, NY 11563  
(516) 596-6300

Date: 5/21/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 11 of 0

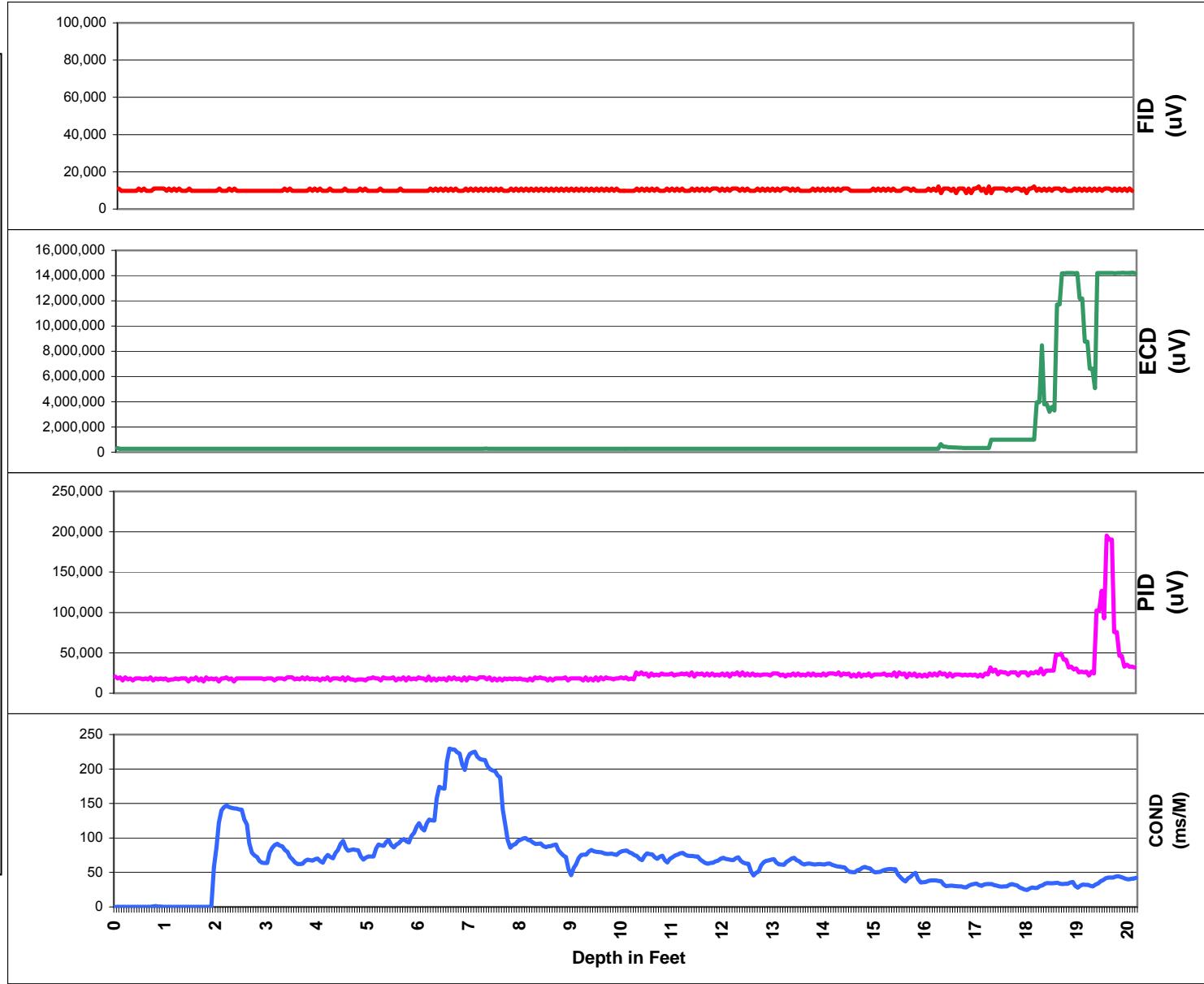
**ZEBRA EC/MIP Summary Log, Point CDMMIP2**  
**Staten Island, NY**



for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

Date: 5/11/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 2 of 0

**ZEBRA EC/MIP Summary Log, Point CDMMIP2C2**  
**Staten Island, NY**

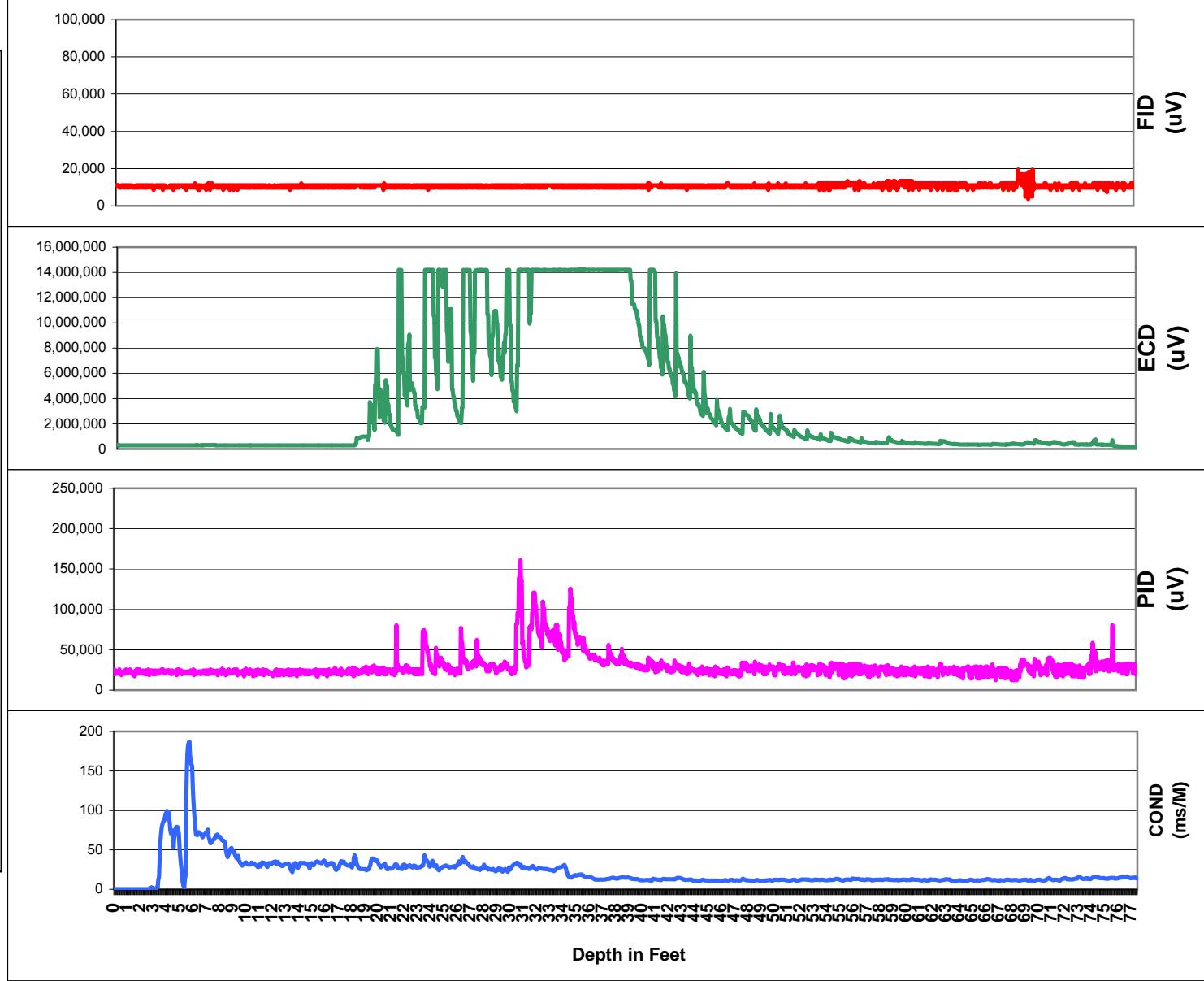


Date: 6/16/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 33 of 0

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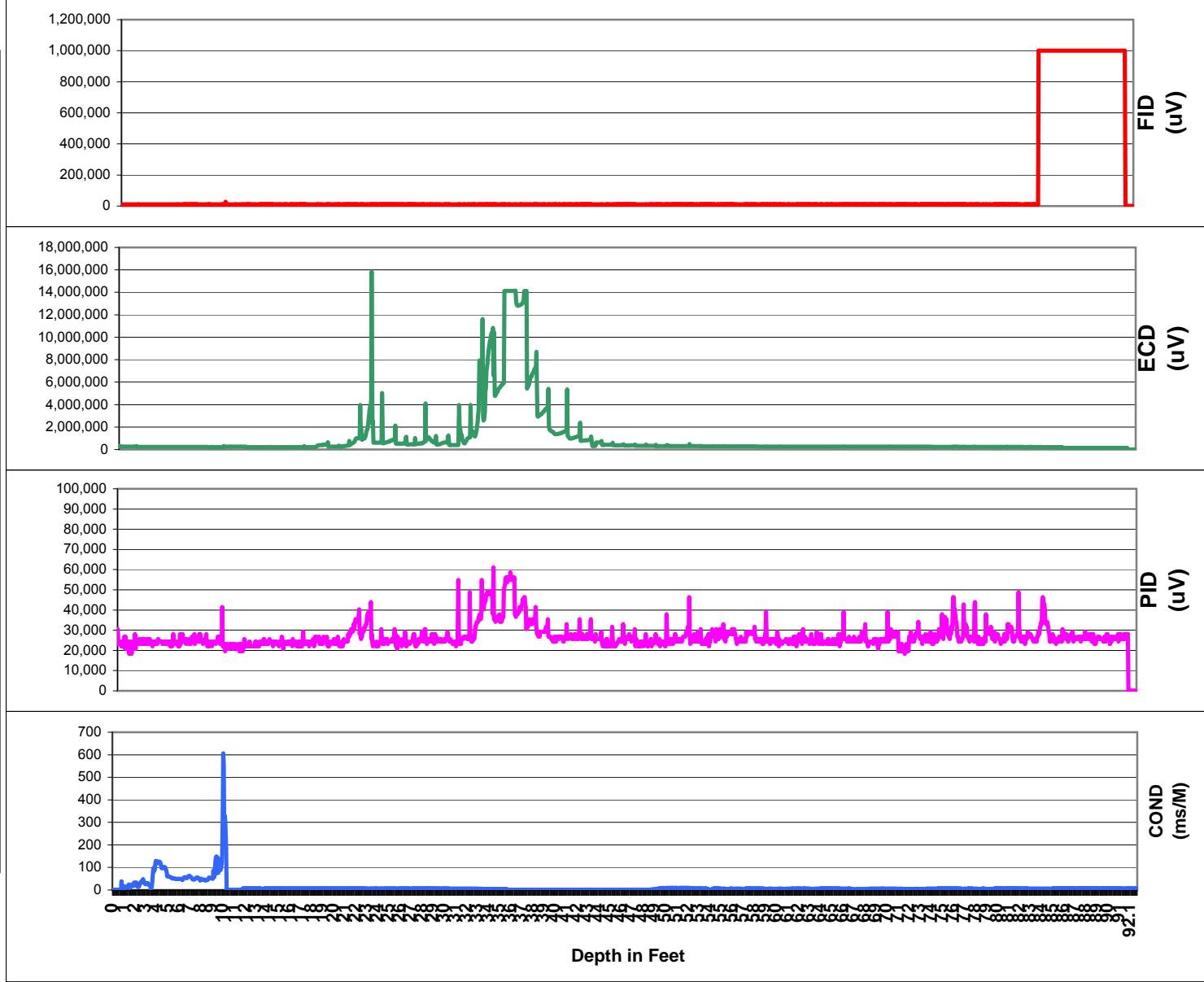
**ZEBRA EC/MIP Summary Log, Point CDMMIP2C3**  
**Staten Island, NY**



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Date: 6/16/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 34 of 0

**ZEBRA EC/MIP Summary Log, Point CDMMIP3**  
**Staten Island, NY**

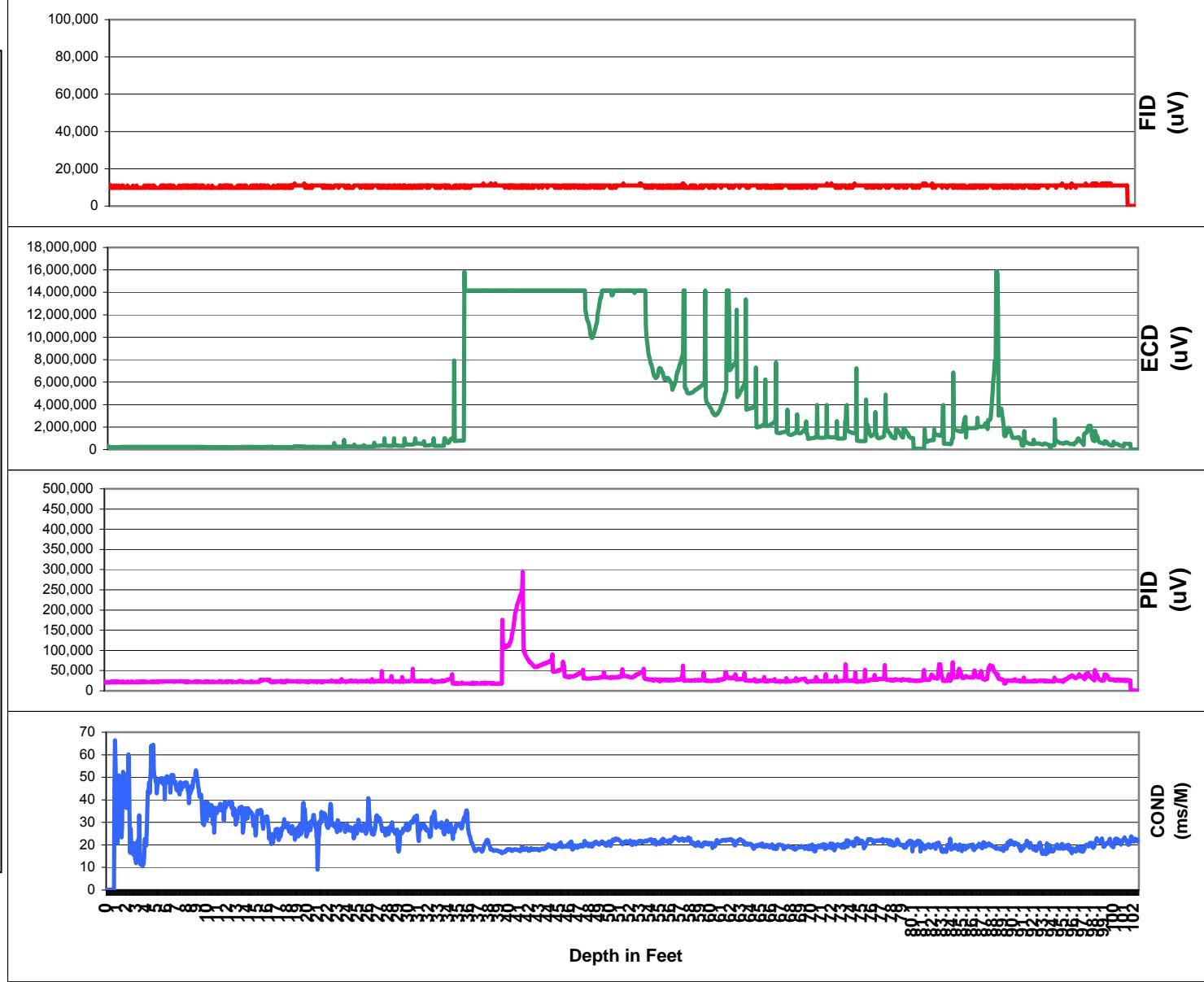


Date: 5/12/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 3 of 0

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by: Zebra Environmental  
30 No. Prospect Avenue  
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**ZEBRA EC/MIP Summary Log, Point CDMMIP4**  
**Staten Island, NY**

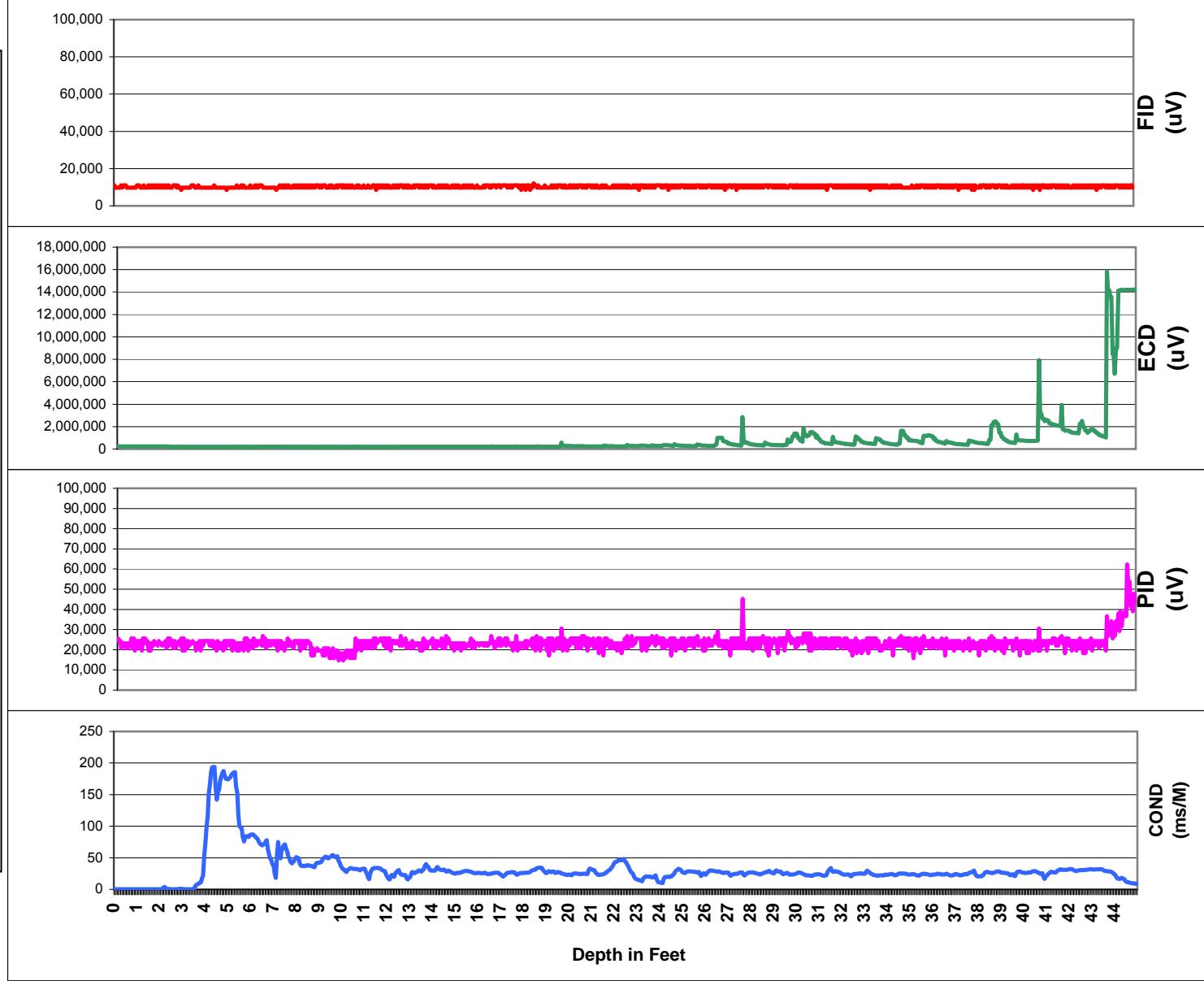


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by: Zebra Environmental  
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Lynbrook, NY 11563  
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**ZEBRA**

Date: 5/13/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 4 of 0

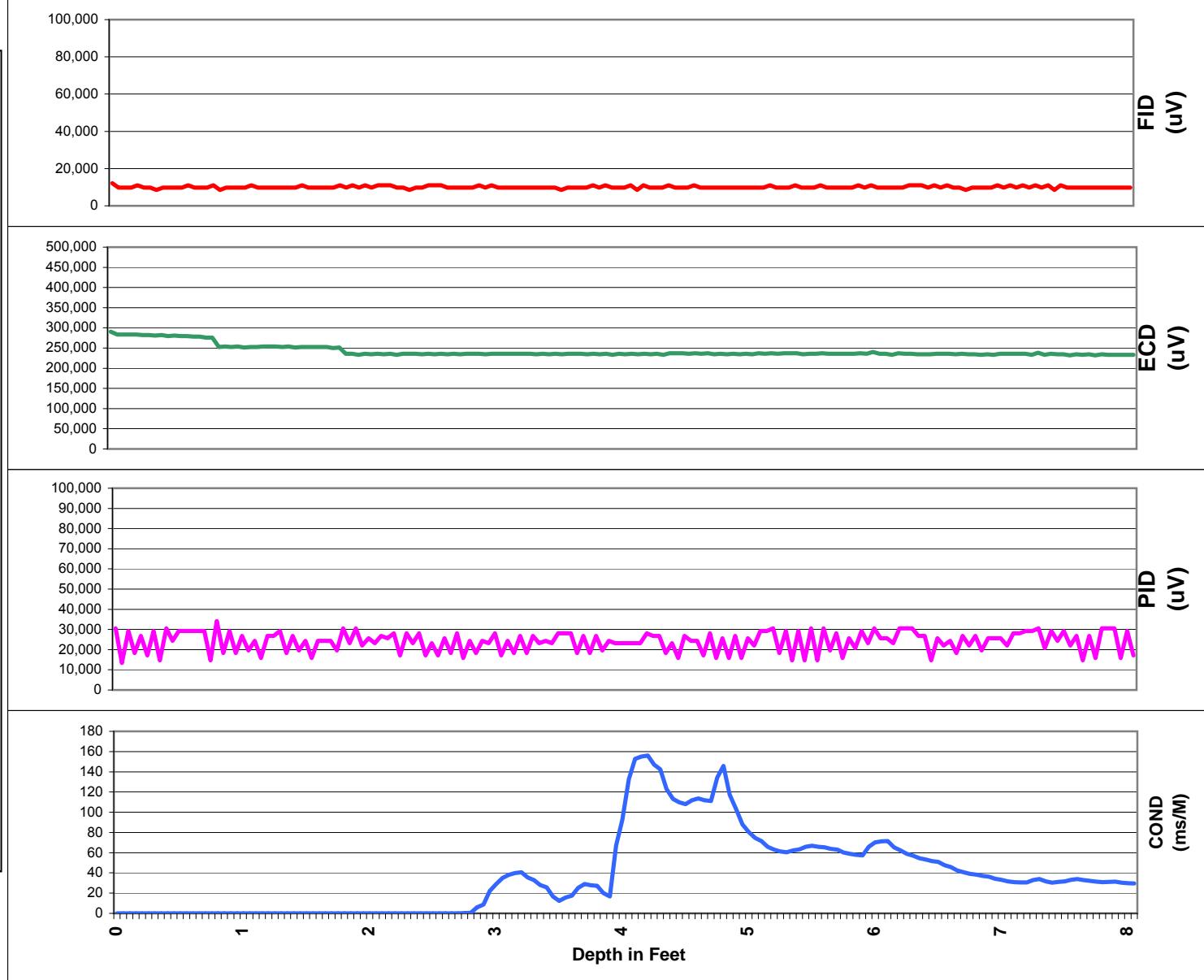
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**Staten Island, NY**



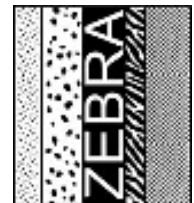
for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

Date: 6/14/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 29 of 0

**ZEBRA EC/MIP Summary Log, Point CDMMP4BR2**  
**Staten Island, NY**

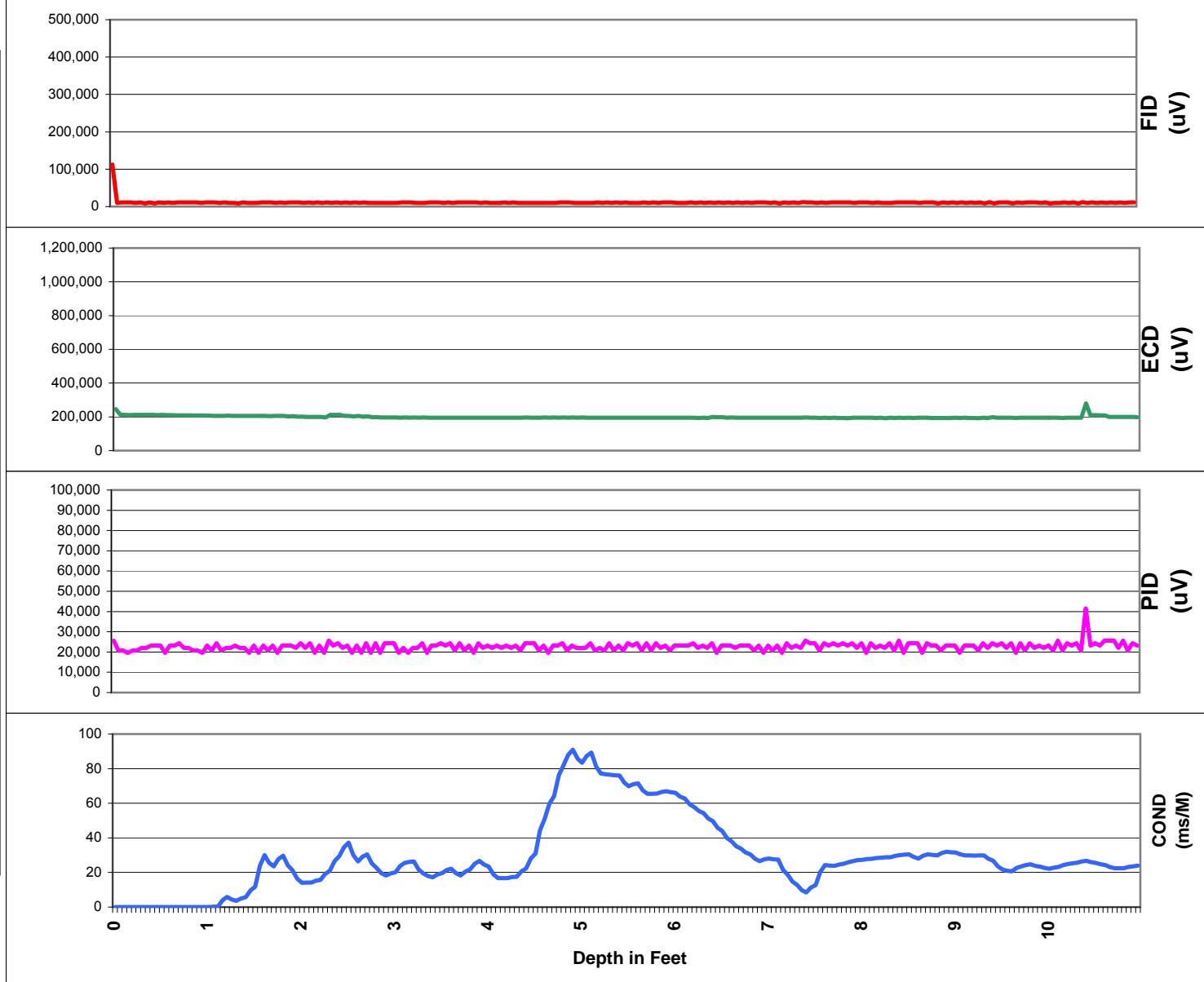


for: CDM  
by: Zebra Environmental  
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Date: 6/14/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 27 of 0

**ZEBRA EC/MIP Summary Log, Point CDMMP4BR3  
Staten Island, NY**

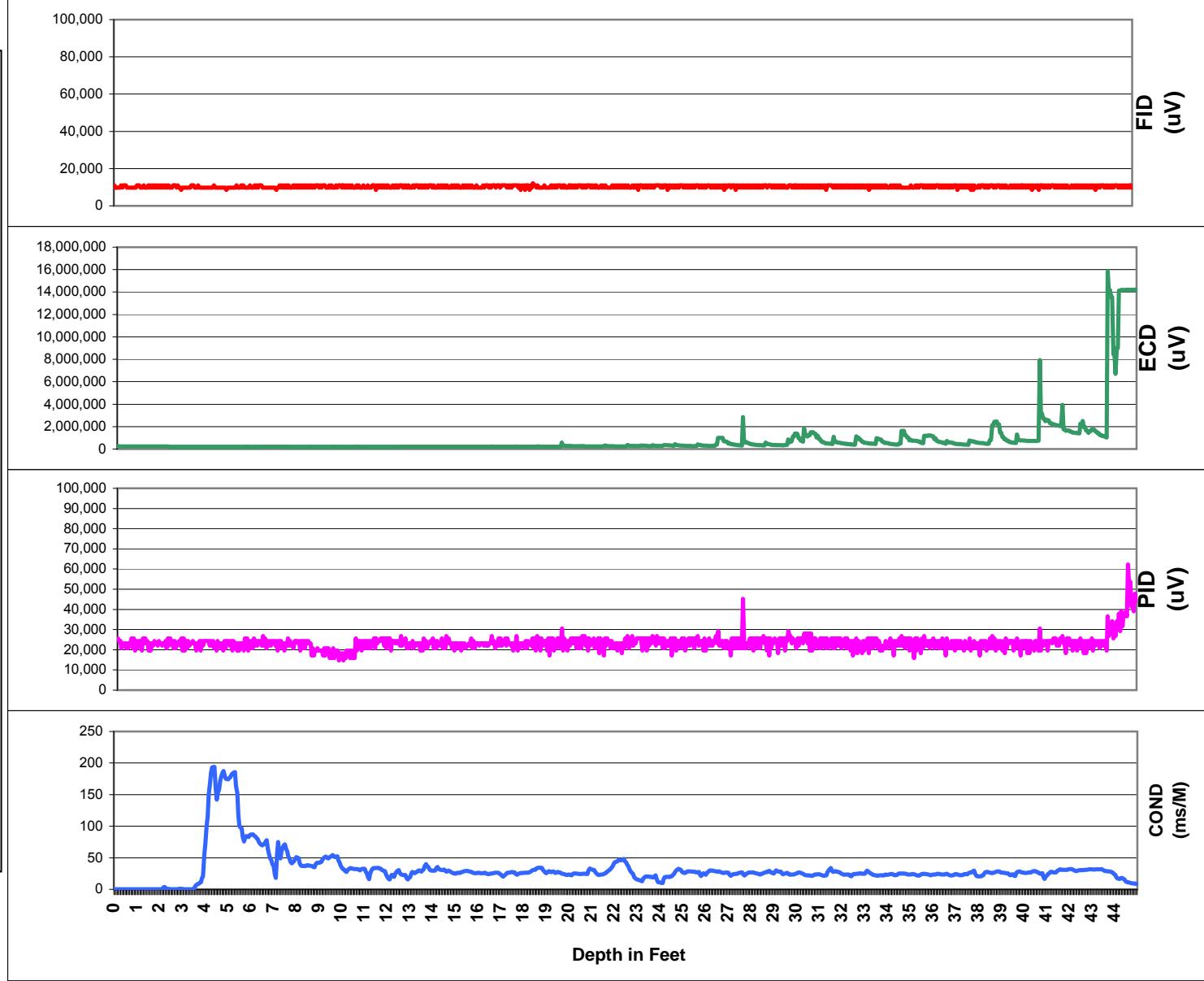


for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300



Date: 6/14/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 28 of 0

**ZEBRA EC/MIP Summary Log, Point CDMMIP4C**  
**Staten Island, NY**

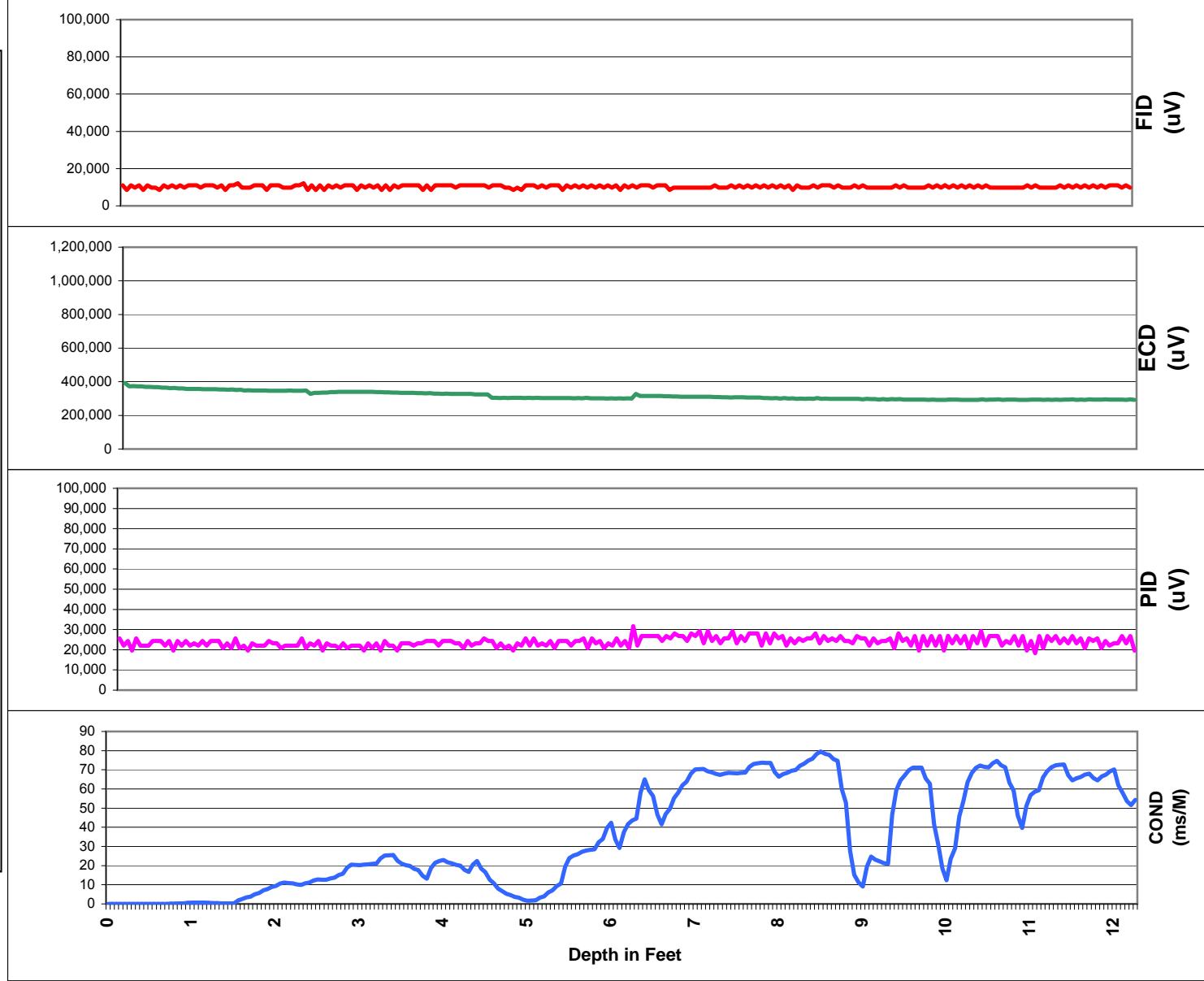


for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300



Date: 6/14/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 29 of 0

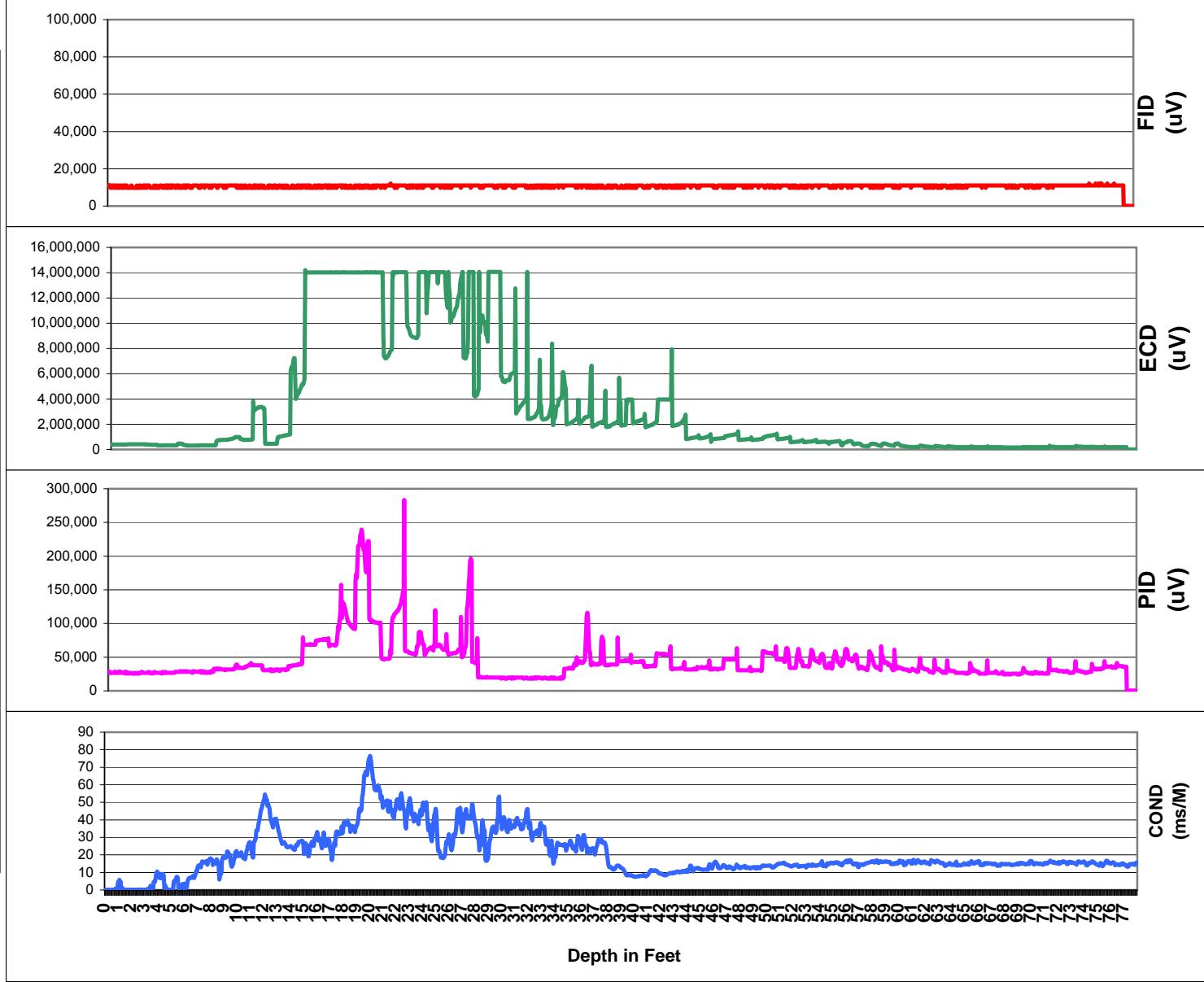
**ZEBRA EC/MIP Summary Log, Point CDMMIP4R**  
**Staten Island, NY**



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Date: 6/11/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 20 of 0

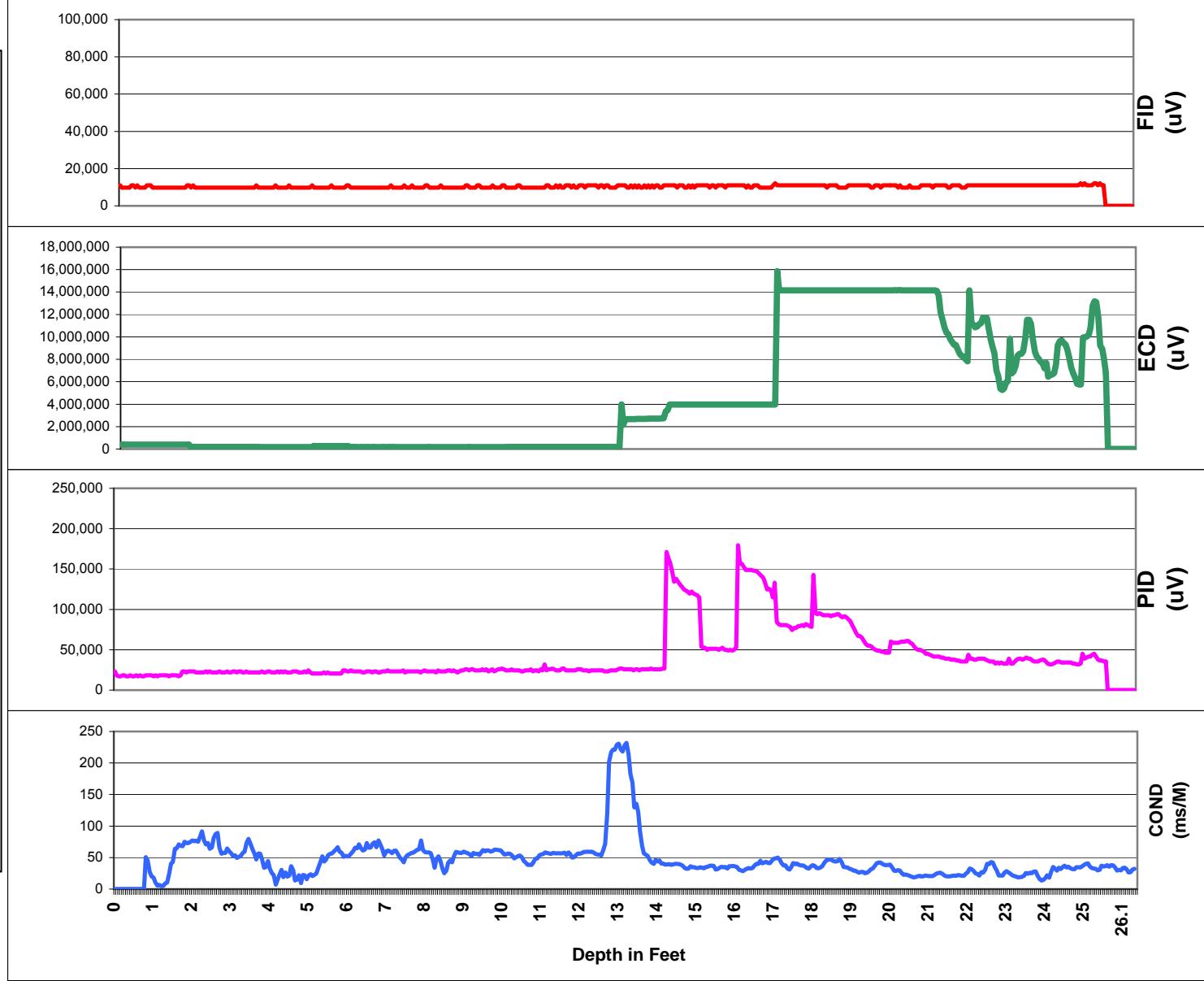
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**Staten Island, NY**



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Date: 5/17/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 5 of 0

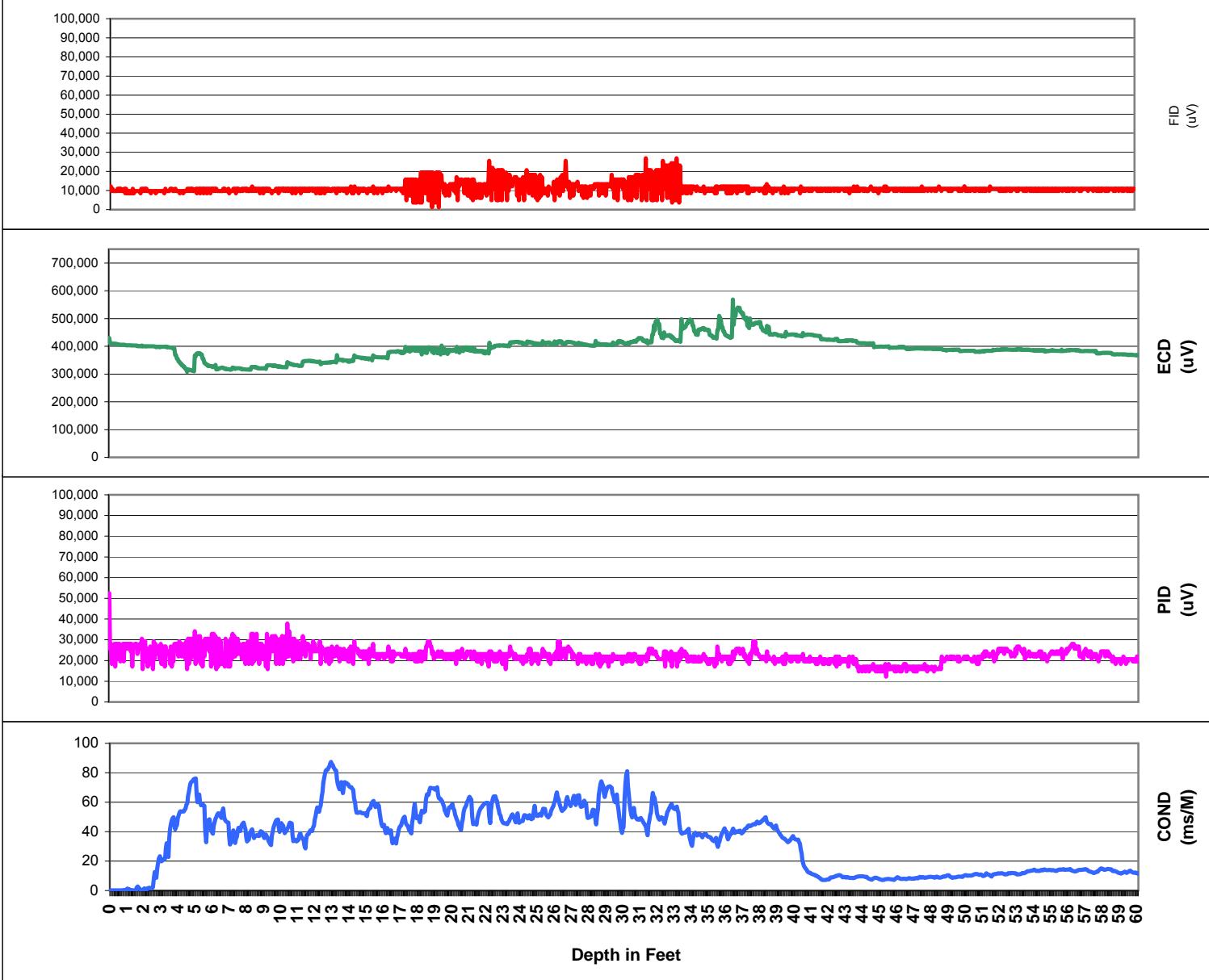
**ZEBRA EC/MIP Summary Log, Point CDMMIP5B**  
**Staten Island, NY**



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Date: 5/21/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 13 of 0

## ZEBRA EC/MIP Summary Log, Point CDMMIP5C

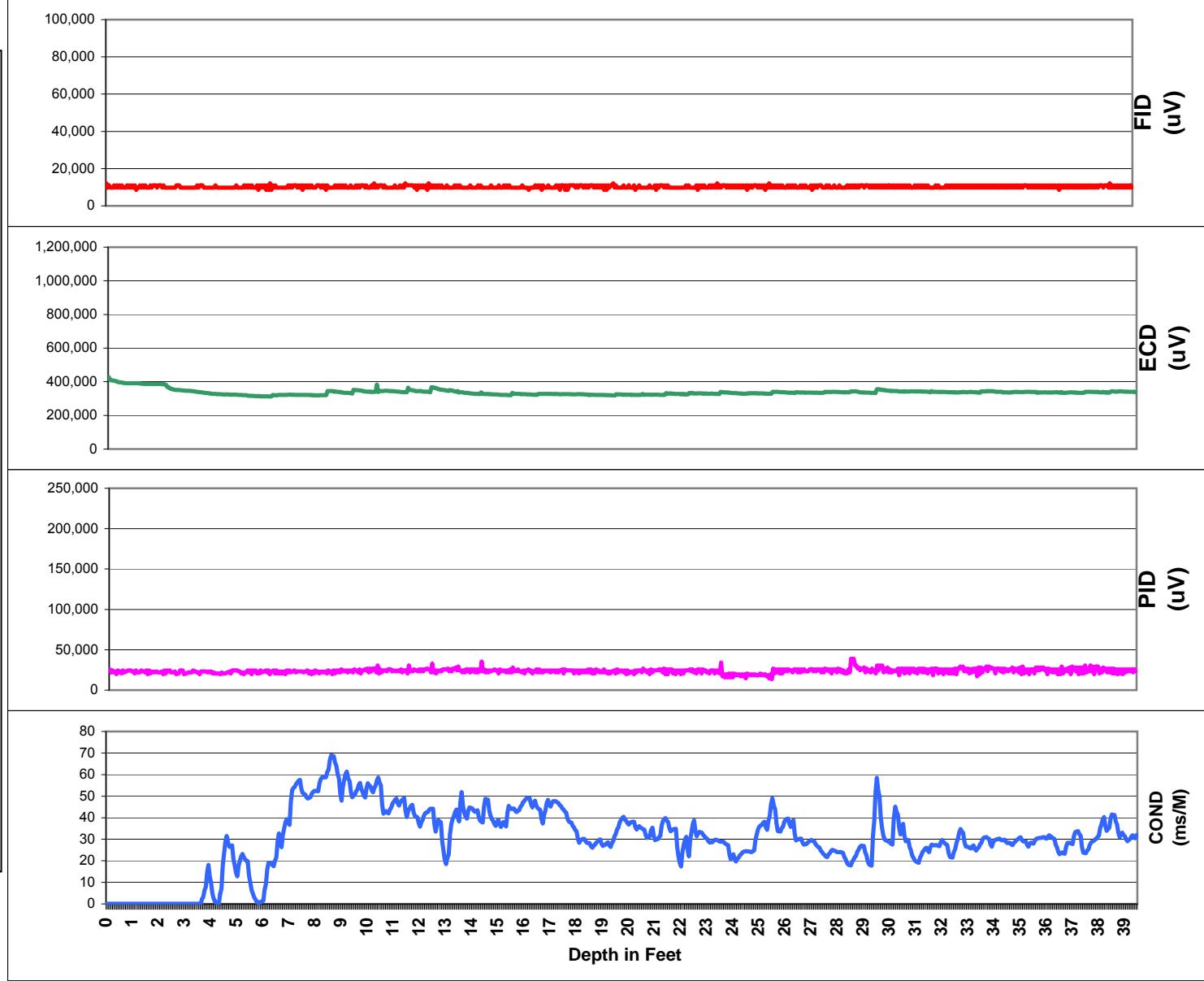


for: CDM  
by: Zebra Environmental  
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Date: 6/9/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 14 of 0

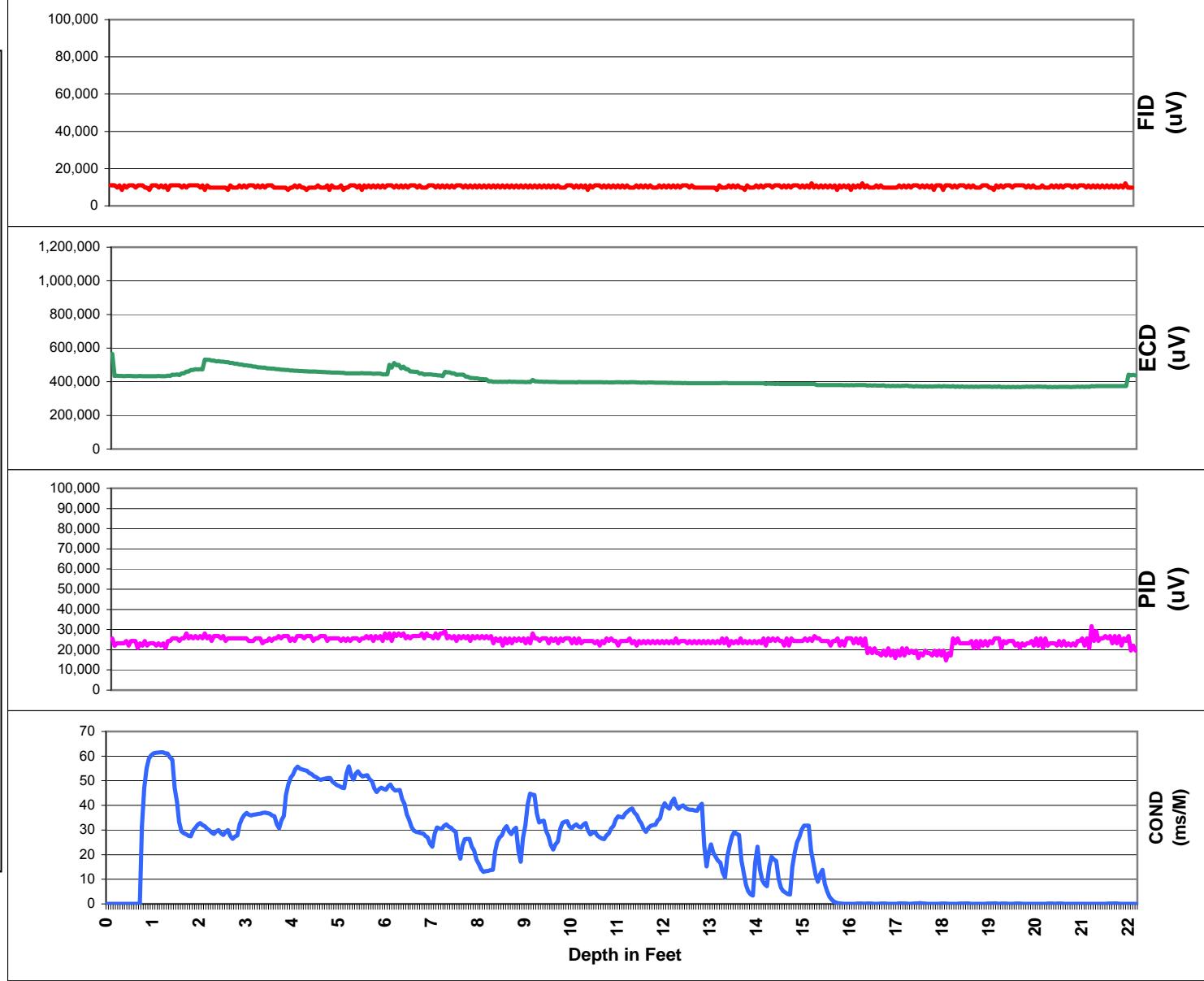
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**Staten Island, NY**



for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
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Date: 6/12/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 23 of 0

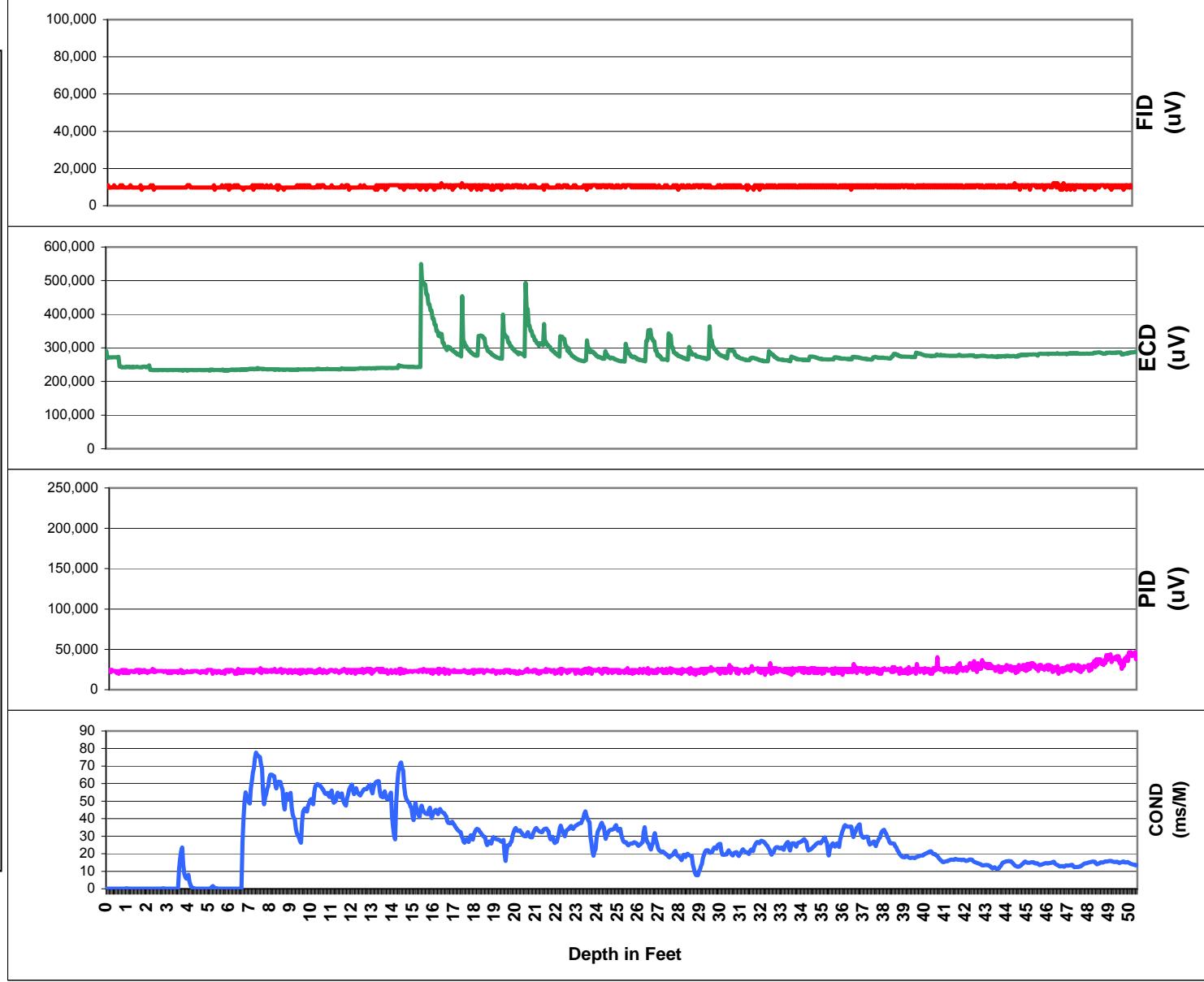
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**Staten Island, NY**



for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

Date: 6/12/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 22 of 0

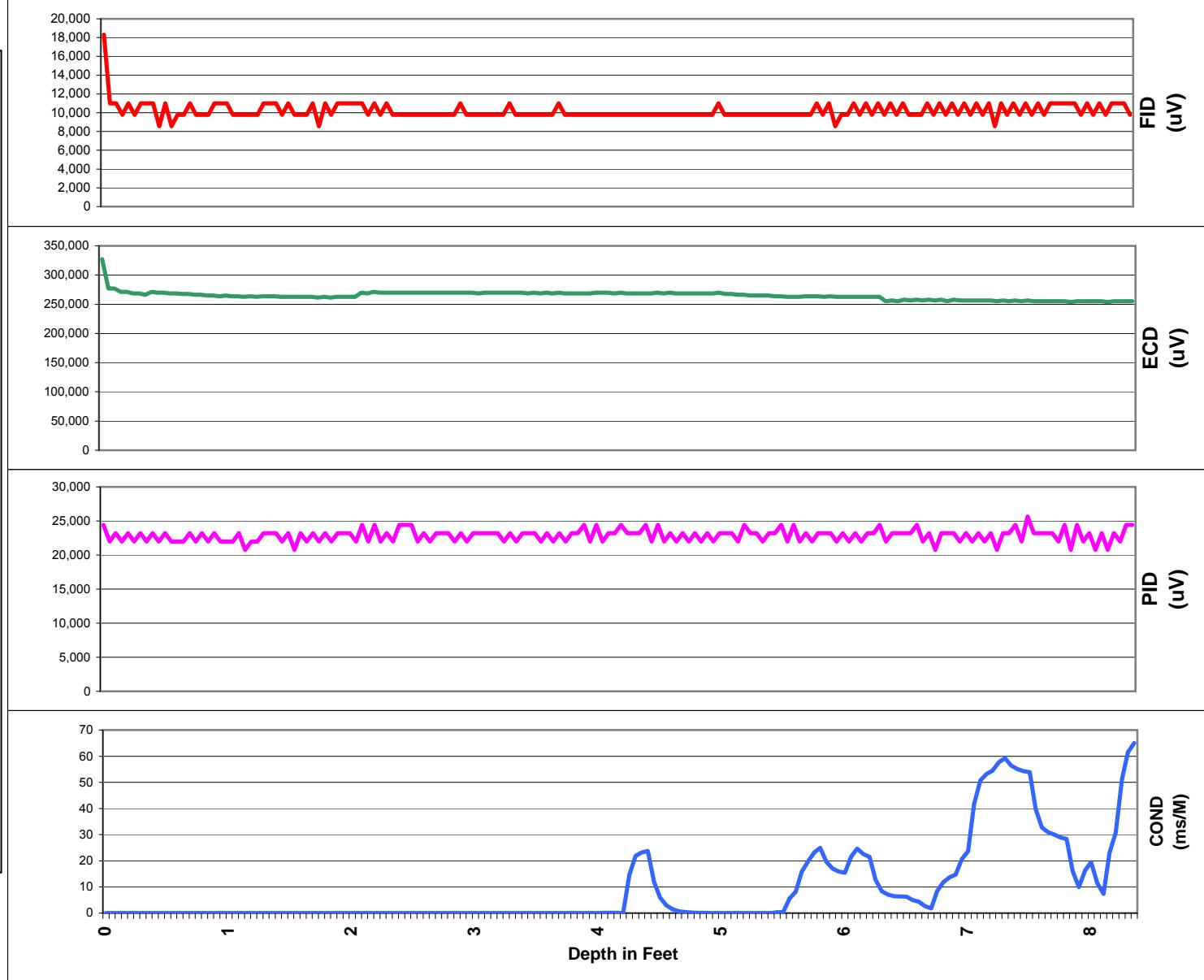
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**Staten Island, NY**



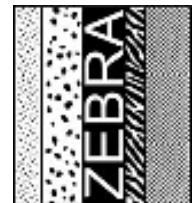
for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

Date: 6/14/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 26 of 0

**ZEBRA EC/MIP Summary Log, Point CDMMIP5ER**  
**Staten Island, NY**

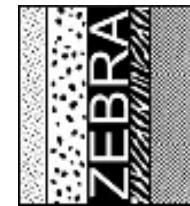
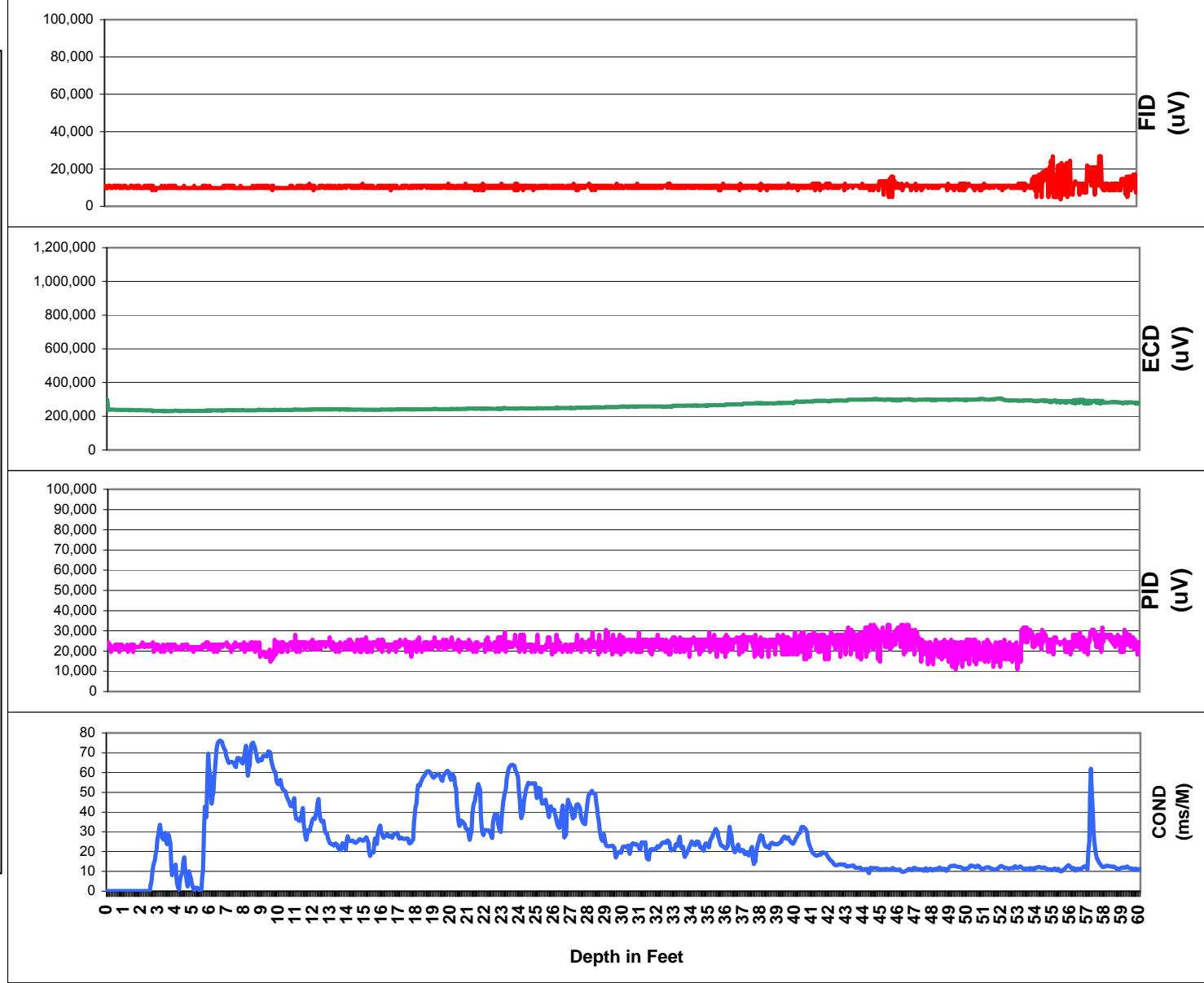


for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300



Date: 6/14/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 25 of 0

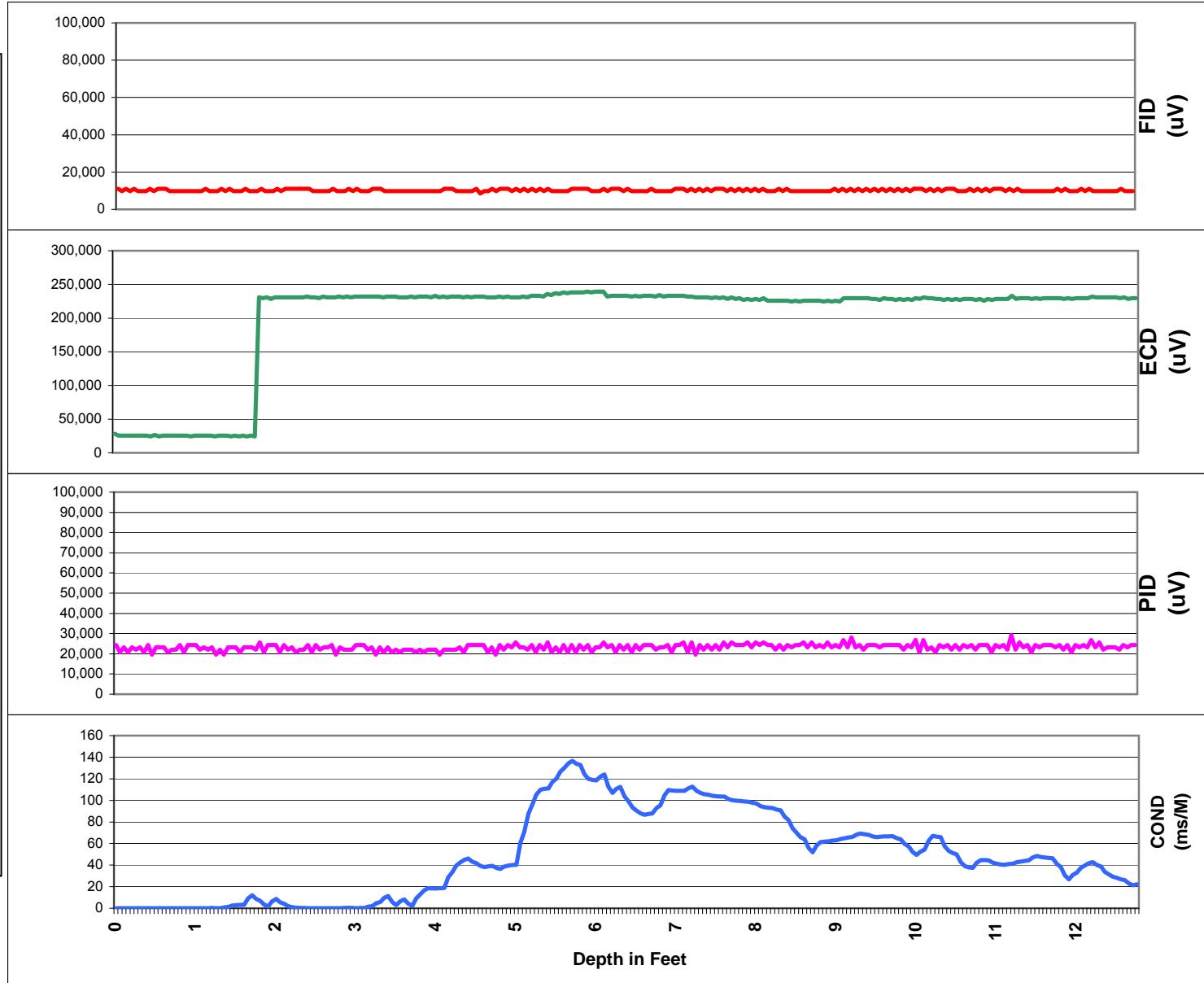
**ZEBRA EC/MIP Summary Log, Point CDMMIP5F**  
**Staten Island, NY**



for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

Date: 6/15/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 31 of 0

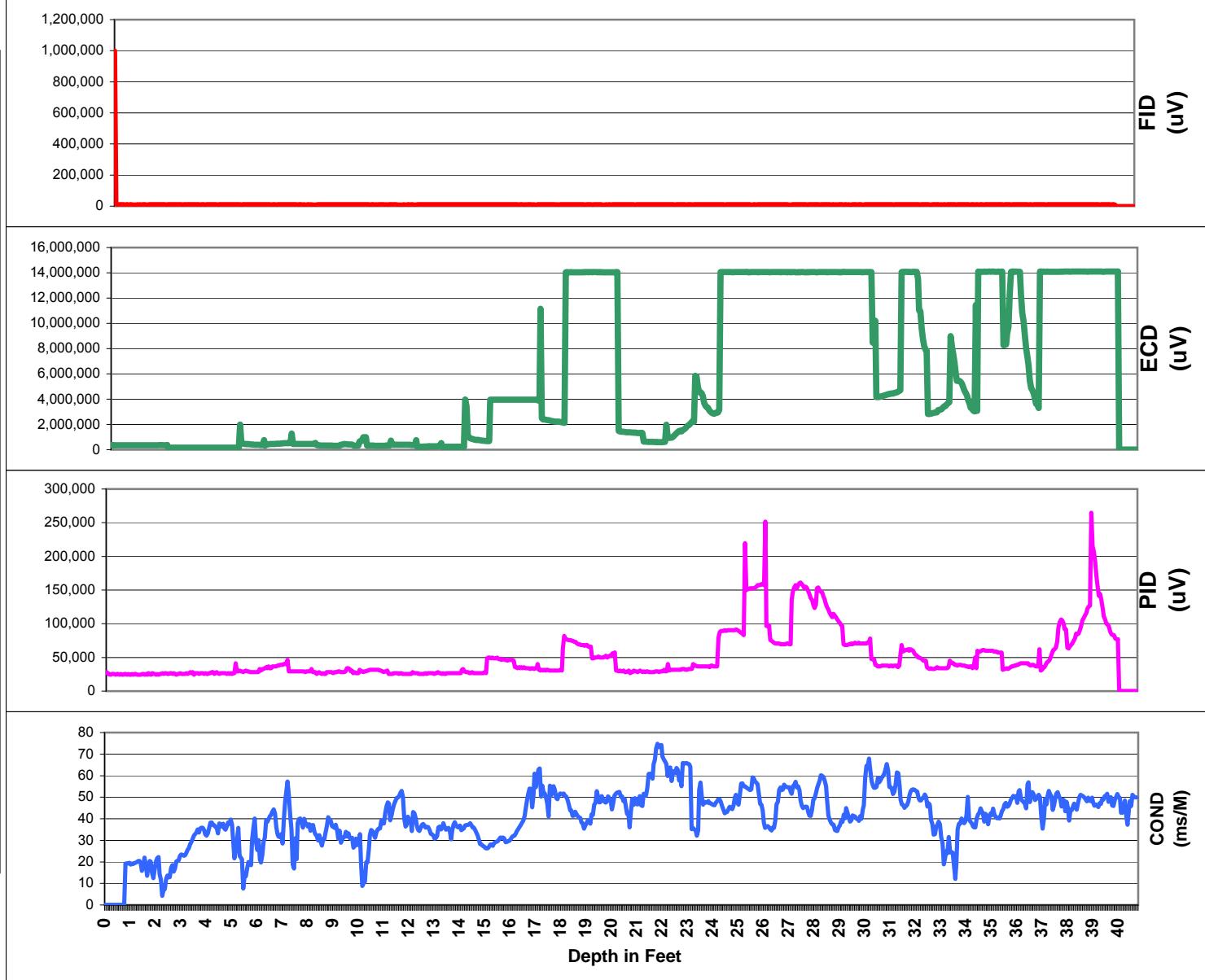
**ZEBRA EC/MIP Summary Log, Point CDMMIP5FR**  
**Staten Island, NY**



for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
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Date: 6/15/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 30 of 0

**ZEBRA EC/MIP Summary Log, Point CDMMIP6**  
**Staten Island, NY**

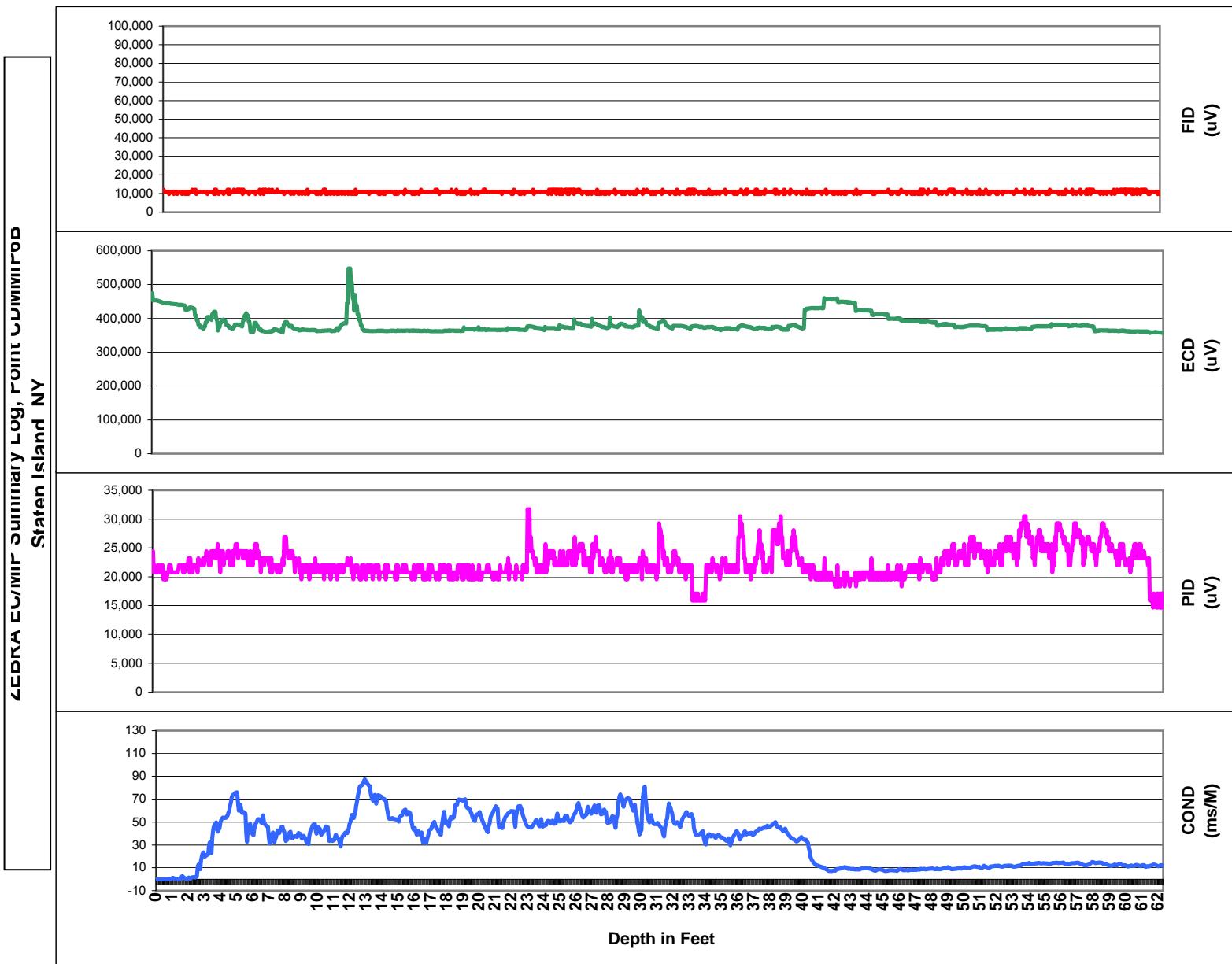


for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300



Date: 5/17/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 6 of 0

ZEBRA CTD/WTR Summary Log, Point 15 of 0

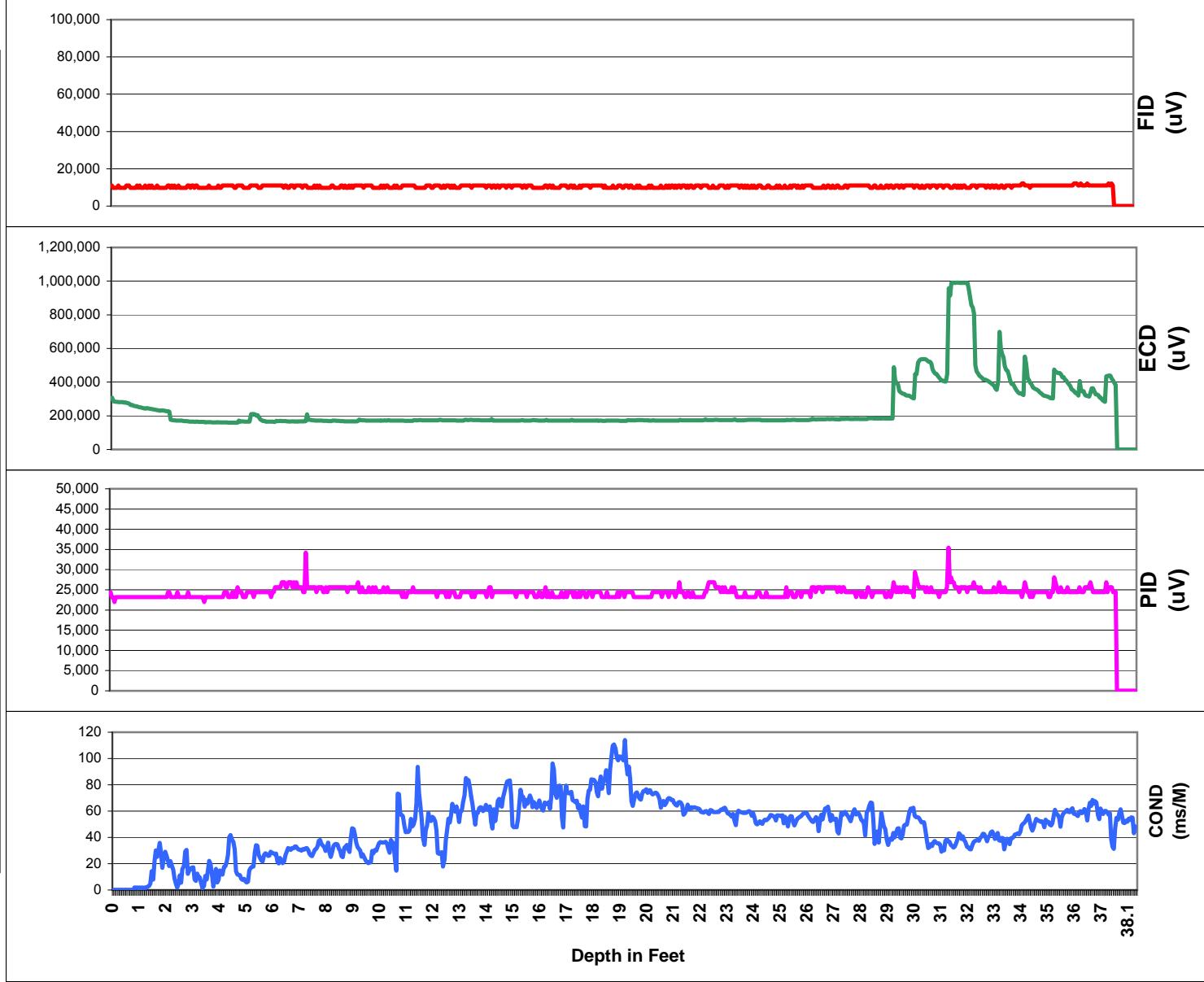


Date: 6/9/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 15 of 0

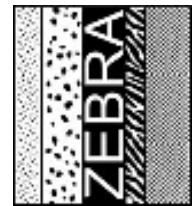
for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300



**ZEBRA EC/MIP Summary Log, Point CDMMIP7**  
**Staten Island, NY**

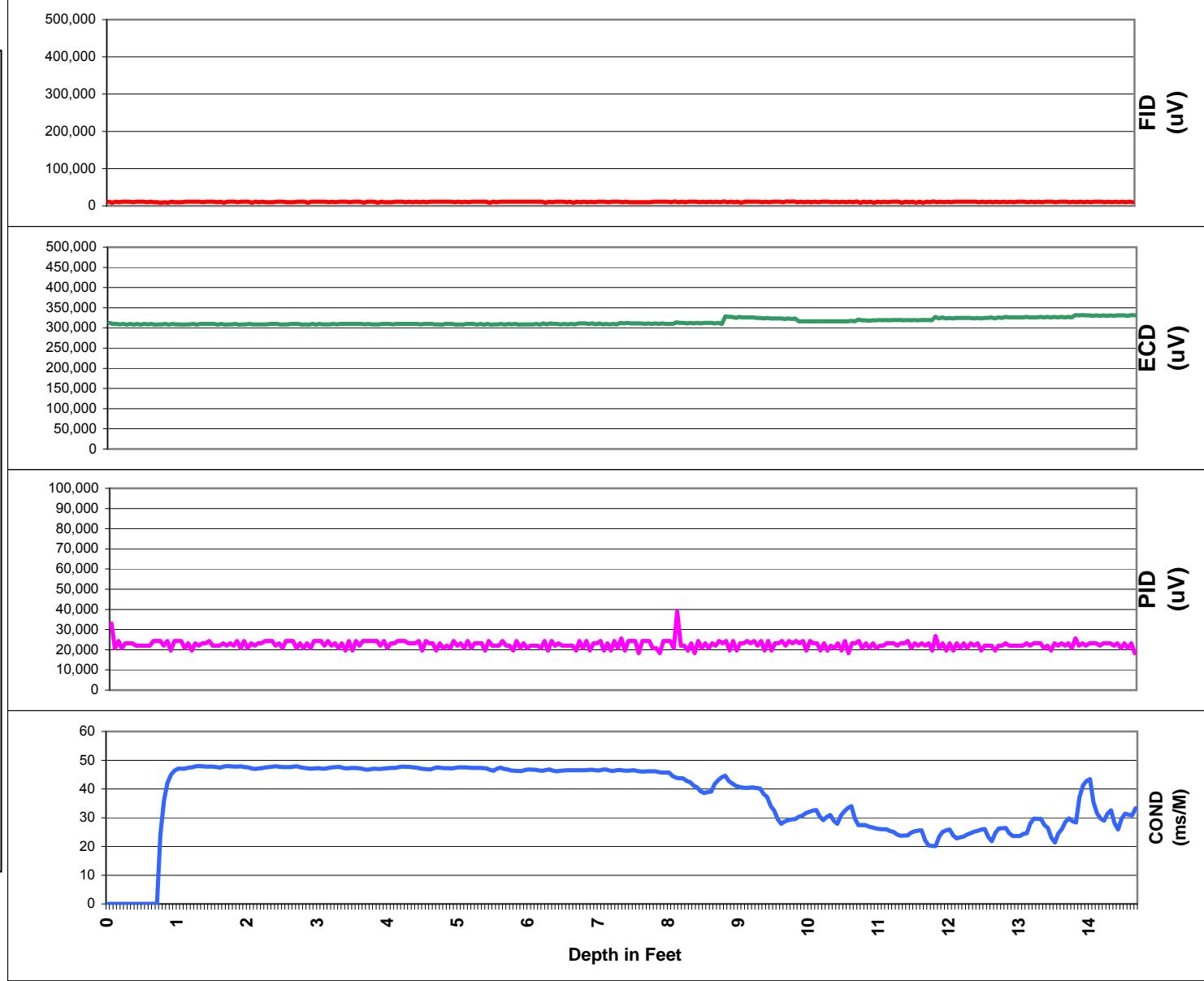


for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

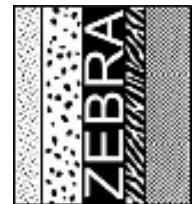


Date: 5/10/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 7 of 0

**ZEBRA EC/MIP Summary Log, Point CDMMIP7B**  
**Staten Island, NY**

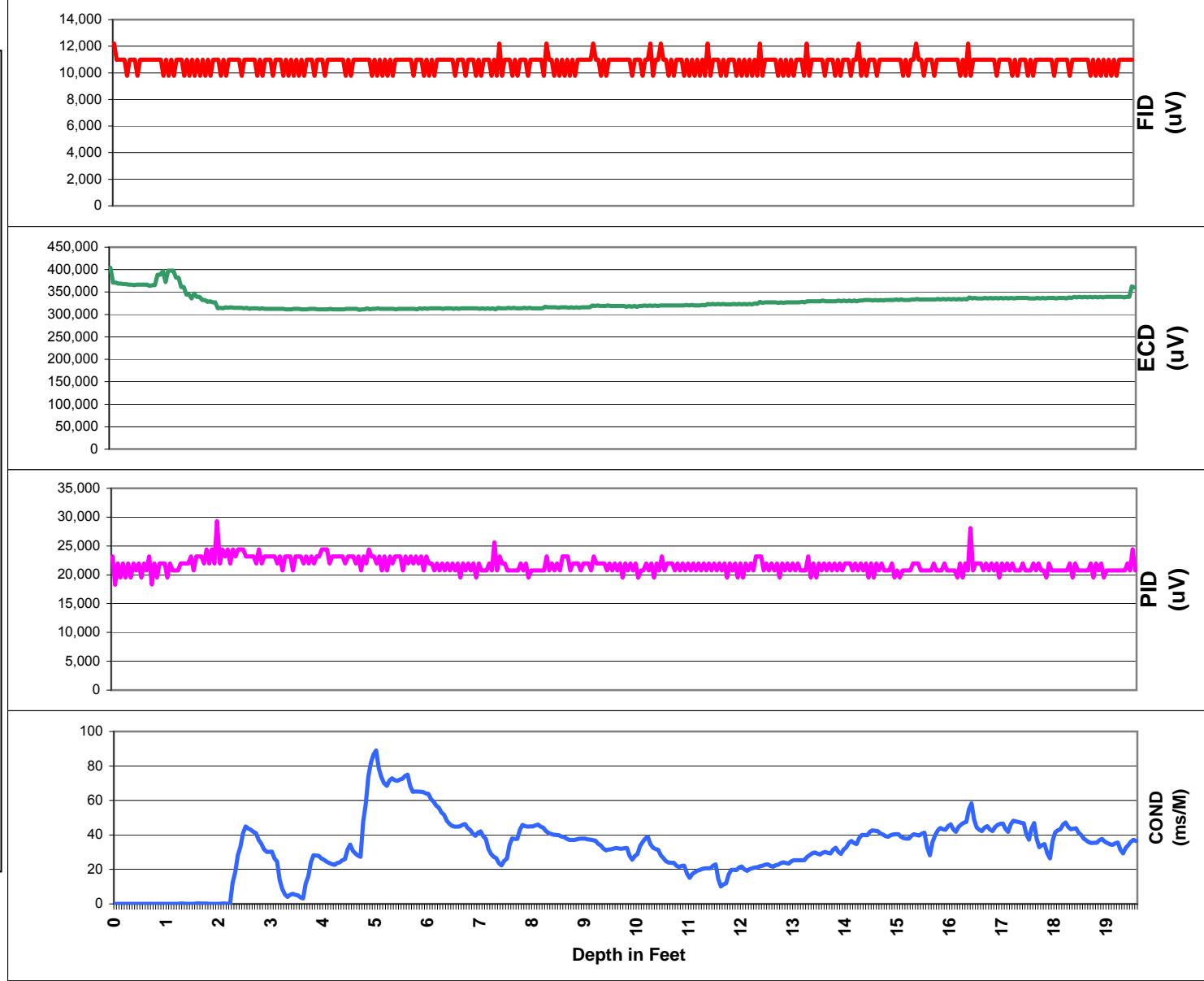


for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300



Date: 6/10/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 16 of 0

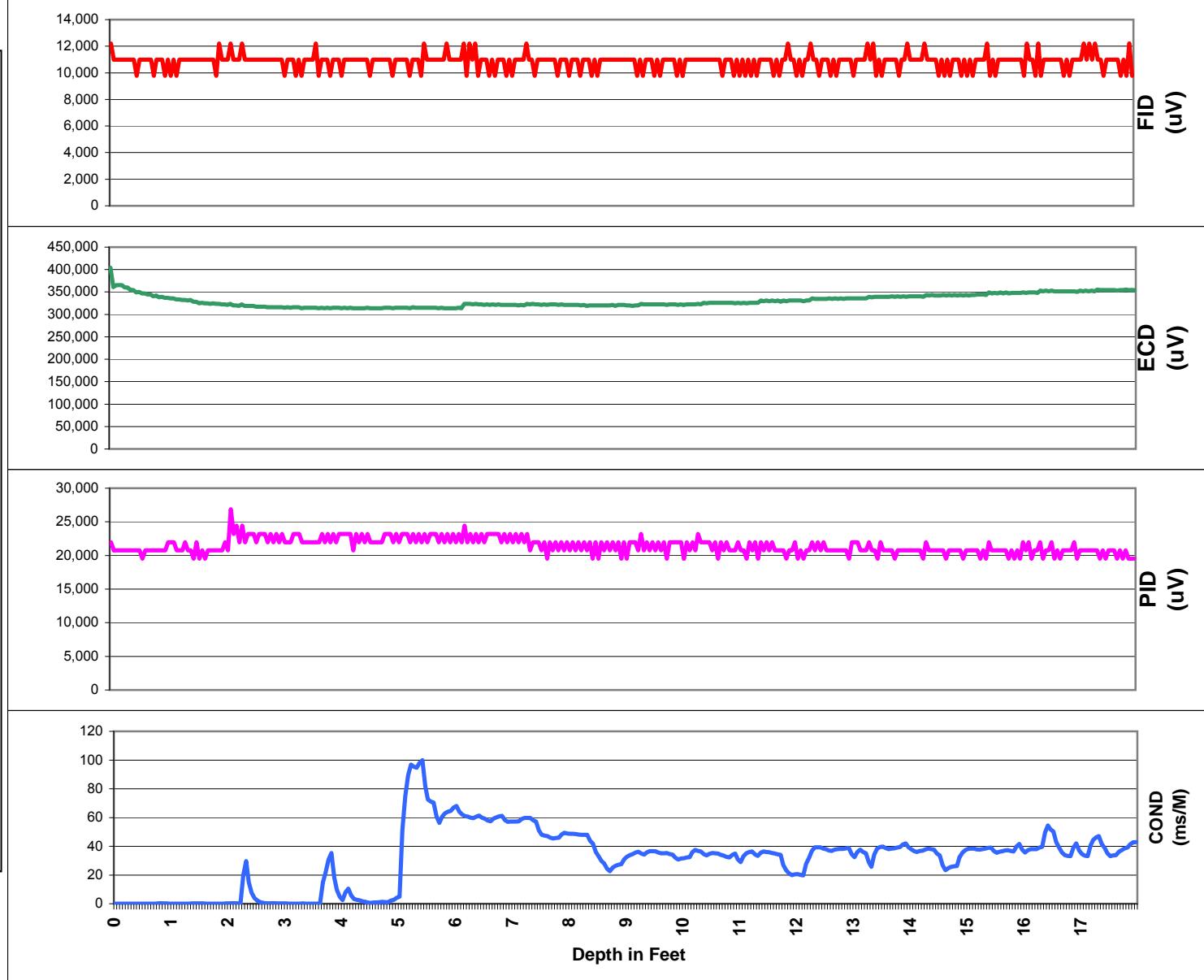
**ZEBRA EC/MIP Summary Log, Point CDMMIP7B1**  
**Staten Island, NY**



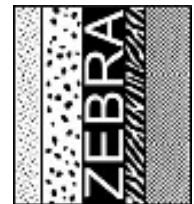
for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

Date: 6/10/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 17 of 0

**ZEBRA EC/MIP Summary Log, Point CDMMIP7B2**  
**Staten Island, NY**

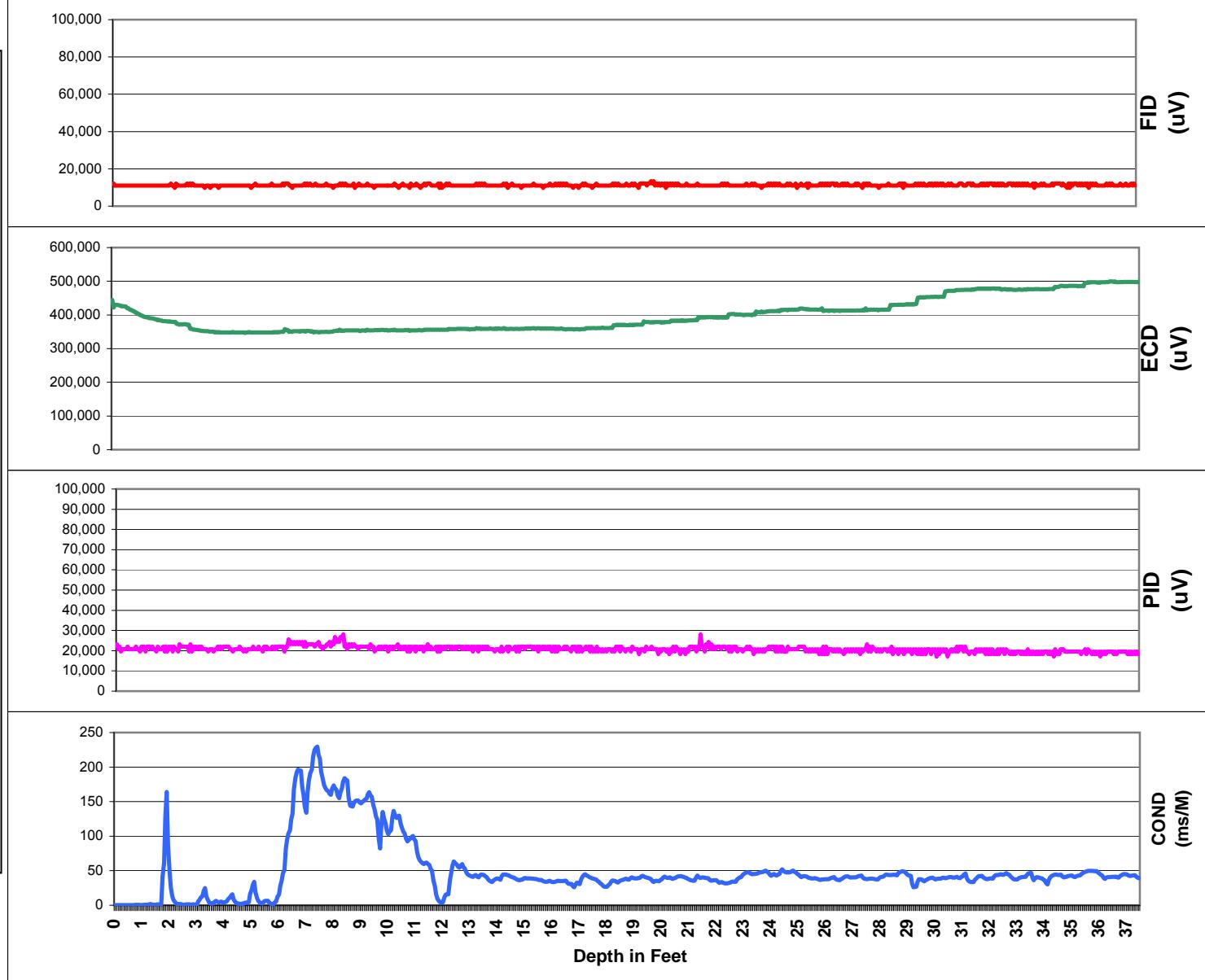


for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300



Date: 6/10/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 18 of 0

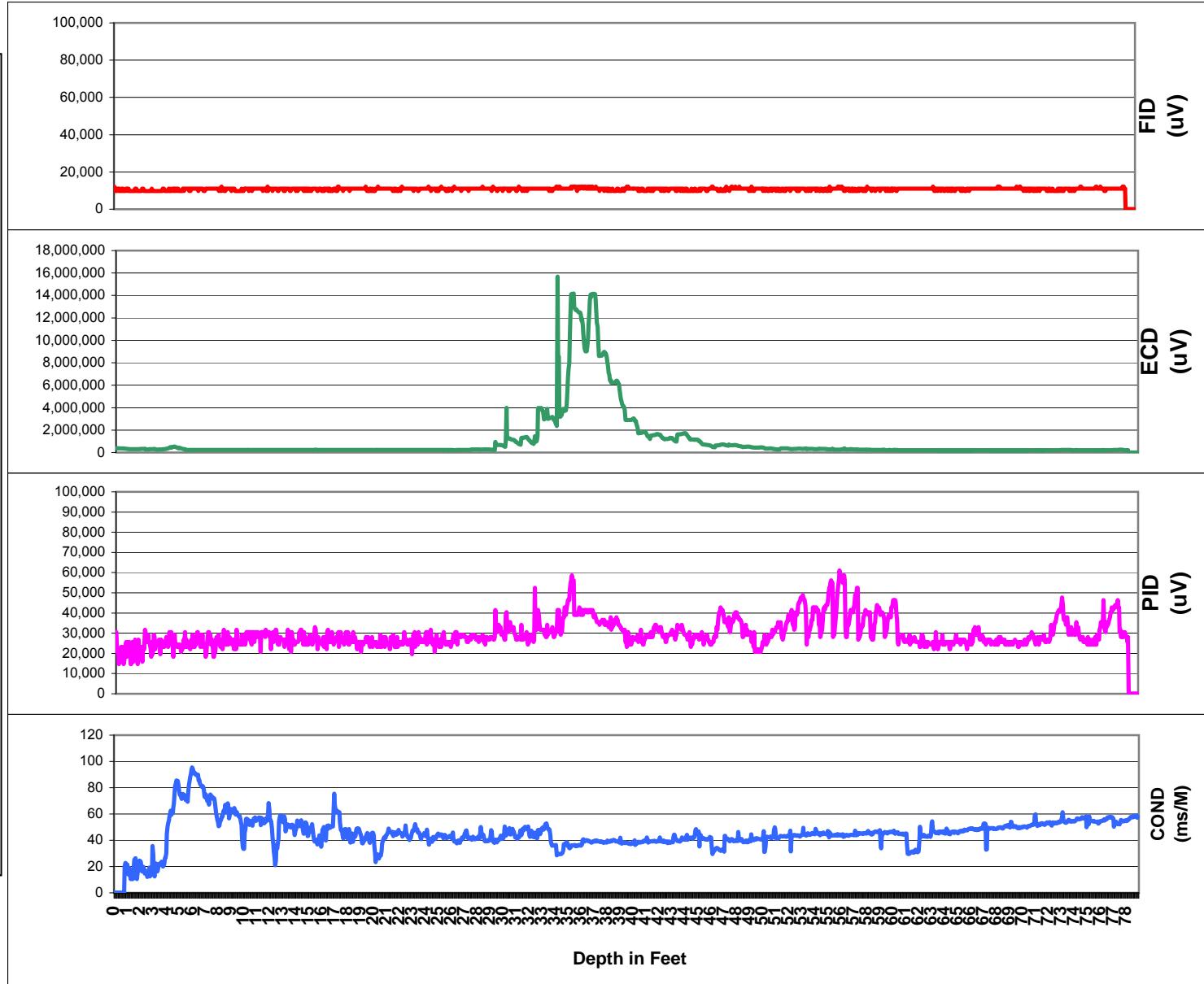
**ZEBRA EC/MIP Summary Log, Point CDMMIP7B3**  
**Staten Island, NY**



for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

Date: 6/10/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 19 of 0

**ZEBRA EC/MIP Summary Log, Point CDM/P2BB  
Staten Island, NY**

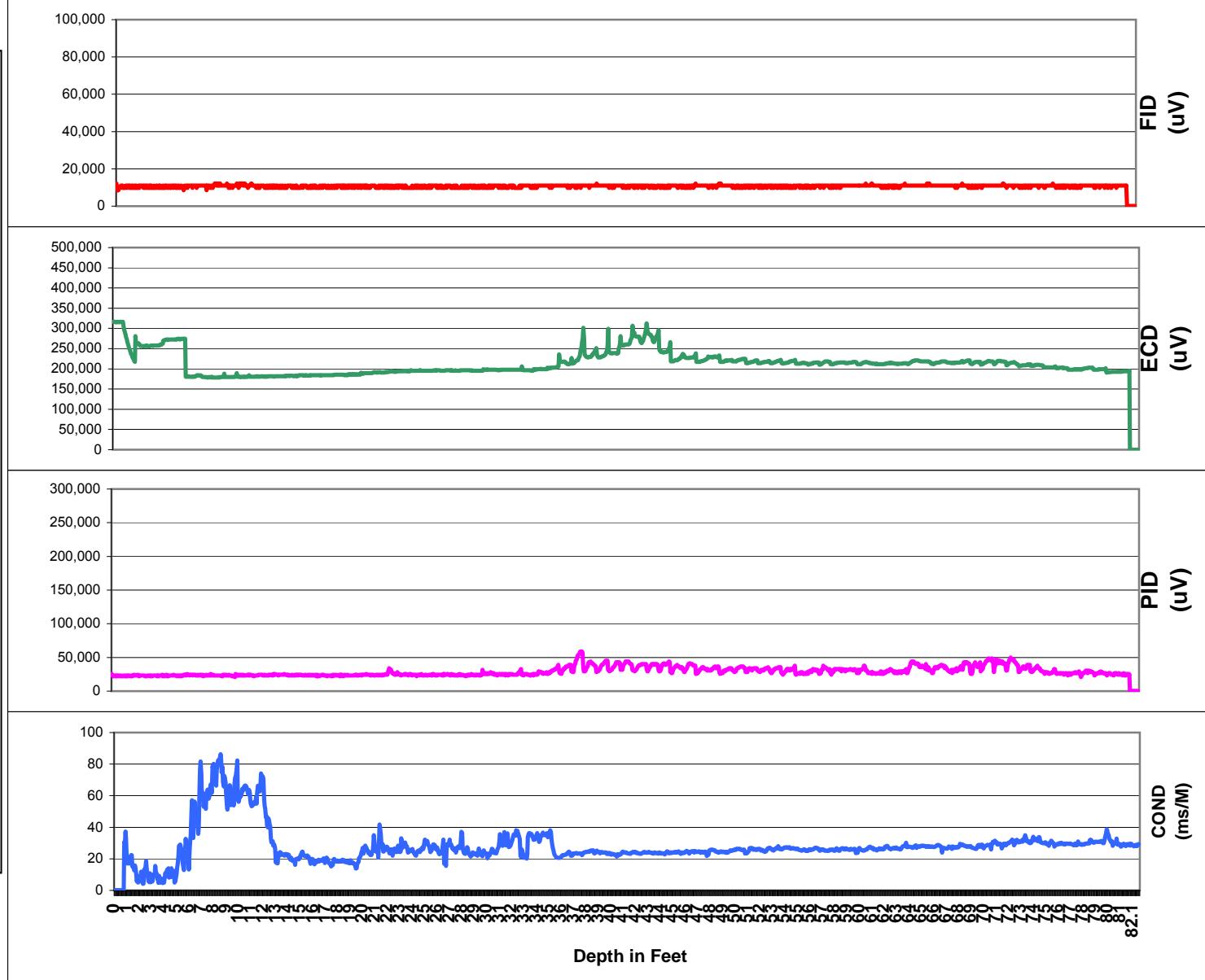


for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

**ZEBRA**

Date: 5/19/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 9 of 0

**ZEBRA EC/MIP Summary Log, Point CDMP3BBBB**  
**Staten Island, NY**



for: CDM  
by: Zebra Environmental  
30 No. Prospect Avenue  
Lynbrook, NY 11563  
(516) 596-6300

Date: 5/19/2011  
Proj. Name: Paul Miller  
Proj. #: DS18666  
Operators: John Diamond  
Point 10 of 0

**Appendix B**  
**Investigation Derived Waste Laboratory Data**

Report Date:  
29-Mar-12 15:53

- Final Report  
 Re-Issued Report  
 Revised Report



## Laboratory Report

CDM  
100 Fieldcrest Avenue  
6th Floor  
Edison, NJ 08837

Work Order: L0466  
Project : Paul Miller Site, IDW  
Project #: D004437-35

Attn: Seth Kellogg

Laboratory ID	Client Sample ID	Matrix	Date Sampled	Date Received
L0466-01	80230-IDW-AQ	Aqueous	08-Mar-12 11:50	09-Mar-12 09:05
L0466-02	80230-IDW-SOLID	Soil	08-Mar-12 11:00	09-Mar-12 09:05

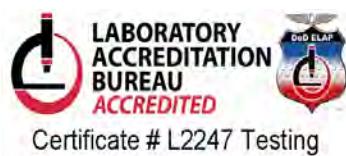
I attest that the information contained within the report has been reviewed for accuracy and checked against the quality control requirements for each method. The results relate only to the samples(s) as received. This report may not be reproduced, except in full, without written approval from Mitkem Laboratories.

All applicable NELAC or USEPA CLP requirements have been met.

Spectrum Analytical (Rhode Island) is accredited under the National Environmental Laboratory Approval Program (NELAP) and is certified by several States, as well as USEPA and US Department of Defense. The current list of our laboratory approvals and certifications is available on the Certifications page on our web site at [www.mitkem.com](http://www.mitkem.com).

Please contact the Laboratory or Technical Director at 401-732-3400 with any questions regarding the data contained in the laboratory report.

Department of Defense	N/A
Connecticut	PH-0153
Delaware	N/A
Maine	2007037
Massachusetts	M-RI907
New Hampshire	2631
New Jersey	RI001
New York	11522
North Carolina	581
Pennsylvania	68-00520
Rhode Island	LAI00301
USDA	P330-08-00023
USEPA - ISM	EP-W-09-039
USEPA - SOM	EP-W-11-033



Authorized by:

Yihai Ding  
Laboratory Director

## **REPORT NARRATIVE**

**Spectrum Analytical, Inc. Featuring Hanibal Technology, RI Division.**

**Client : CDM**

**Project: Paul Miller Site, IDW**

**Laboratory Workorder / SDG #: L0466**

**SW846 8260C, VOC by GC-MS**

### **I. SAMPLE RECEIPT**

No exceptions or unusual conditions were encountered unless a Sample Condition Notification Form, or other record of communication is included with the Sample Receipt Documentation.

### **II. HOLDING TIMES**

#### **A. Sample Preparation:**

All samples were prepared within the method-specified holding times with the following exceptions:

80230-IDW-AQ (L0466-01ADL) exceed by-6 Days

Please note the initial analysis was performed within method holding time. Dilution analysis matched the initial analysis well. Both results were included in this report.

#### **B. Sample Analysis:**

All samples were analyzed within the method-specified holding times.

### **III. METHODS**

Samples were analyzed following procedures in laboratory test code:  
SW846 8260C

### **IV. PREPARATION**

Aqueous Samples were prepared following procedures in laboratory test code: SW5030

Soil Samples were prepared following procedures in laboratory test code: SW5035

## V. INSTRUMENTATION

The following instrumentation was used

Instrument Code: V1

Instrument Type: GCMS-VOA

Description: HP5890 II / HP5972

Manufacturer: Hewlett-Packard

Model: 5890 / 5972

GC Column used: 30 m X 0.25 mm ID [1.40 um thickness] DB-624 capillary column.

Instrument Code: V5

Instrument Type: GCMS-VOA

Description: HP6890 / HP6890

Manufacturer: Hewlett-Packard

Model: 6890 / 6890

GC Column used: 30 m X 0.25 mm ID [1.40 um thickness] DB-624 capillary column.

Instrument Code: V6

Instrument Type: GCMS-VOA

Description: HP6890 / HP5973

Manufacturer: Hewlett-Packard

Model: 6890 / 5973

GC Column used: 30 m X 0.25 mm ID [1.40 um thickness] DB-624 capillary column.

## VI. ANALYSIS

### A. Calibration:

Calibrations met the method/SOP acceptance criteria.

### B. Blanks:

All method blanks were within the acceptance criteria.

**C. Surrogates:**

Surrogate standard percent recoveries were within the QC limits with the following exceptions. Please note that the acceptance criteria allow one surrogate recovery outside of the QC limits per fraction.

80230-IDW-SOLID (L0466-02C-TCLP), recovery is above criteria for Dibromofluoromethane at 116% with criteria of (85-115).

**D. Spikes:**

**1. Laboratory Control Spikes (LCS):**

Percent recoveries for lab control samples were within the QC limits with the following exceptions. Please note that most test procedures allow for several compounds outside of the QC limits for the LCS, although this may indicate a bias for this specific compound.

LCS-65144 in batch 65144, Percent Recovery is outside QC Limits, recovery is above criteria for Acetone at 160% with criteria of (20-160).

LCS-65160 in batch 65160, Percent Recovery is outside QC Limits, recovery is above criteria for 1,1,1-Trichloroethane at 144% with criteria of (65-130), 2,2-Dichloropropane at 152% with criteria of (70-135), Carbon tetrachloride at 155% with criteria of (65-140) and Iodomethane at 131% with criteria of (72-121).

**2. Matrix Spike / Matrix Spike Duplicate (MS/MSD):**

No client-requested MS/MSD analyses were included in this SDG.

**E. Internal Standards:**

Internal standard peak areas were within the QC limits.

**F. Dilutions:**

The following samples were analyzed at dilution:

80230-IDW-AQ (L0466-01ADL) : Dilution Factor: 10

**G. Samples:**

No other unusual occurrences were noted during sample analysis.

I certify that this data package is in compliance with the terms and conditions agreed to by the client and Spectrum, both technically and for completeness, except for the conditions noted above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or designated person, as verified by the following signature.

Signed: \_\_\_\_\_

A handwritten signature in black ink, appearing to read "T. J. H. P.", is placed over the line for the signature.

Date: \_\_\_\_\_ 3/29/2012 \_\_\_\_\_

## **REPORT NARRATIVE**

**Spectrum Analytical, Inc. Featuring Hanibal Technology, RI Division.**

**Client : CDM**

**Project: Paul Miller Site, IDW**

**Laboratory Workorder / SDG #: L0466**

**SW846 8270D, SVOA by GC-MS**

### **I. SAMPLE RECEIPT**

No exceptions or unusual conditions were encountered unless a Sample Condition Notification Form, or other record of communication is included with the Sample Receipt Documentation.

### **II. HOLDING TIMES**

#### **A. Sample Preparation:**

All samples were prepared within the method-specified holding times.

#### **B. Sample Analysis:**

All samples were analyzed within the method-specified holding times.

### **III. METHODS**

Samples were analyzed following procedures in laboratory test code:  
SW846 8270D

### **IV. PREPARATION**

Aqueous Samples were prepared following procedures in laboratory test code: SW3510

### **V. INSTRUMENTATION**

The following instrumentation was used

Instrument Code: S3  
Instrument Type: GCMS-SEMI  
Description: HP6890 / HP5973  
Manufacturer: Hewlett-Packard  
Model: 6890 / 5973  
GC Column used: 30 m X 0.25 mm ID [0.25 um thickness] Rxi-5sil MS capillary column.

Instrument Code: S6  
Instrument Type: GCMS-Semi  
Description: HP7890A  
Manufacturer: Agilent  
Model: 7890A/5973  
GC Column used: 30 m X 0.25 mm ID [0.25 um thickness] Rxi-5sil MS capillary column.

## VI. ANALYSIS

### A. Calibration:

Calibrations met the method/SOP acceptance criteria.

### B. Blanks:

All method blanks were within the acceptance criteria.

### C. Surrogates:

Surrogate standard percent recoveries were within the QC limits.

### D. Spikes:

#### 1. Laboratory Control Spikes (LCS):

Percent recoveries for lab control samples were within the QC limits with the following exceptions. Please note that most test procedures allow for several compounds outside of the QC limits for the LCS, although this may indicate a bias for this specific compound.

LCSD-65051 in batch 65051, Percent Recovery is outside QC Limits, recovery is above criteria for Hexachloroethane at 97% with criteria of (30-95).

**2. Matrix Spike / Matrix Spike Duplicate (MS/MSD):**

No client-requested MS/MSD analyses were included in this SDG.

**E. Internal Standards:**

Internal standard peak areas were within the QC limits.

**F. Dilutions:**

No sample in this SDG required analysis at dilution.

**G. Samples:**

No other unusual occurrences were noted during sample analysis.

I certify that this data package is in compliance with the terms and conditions agreed to by the client and Spectrum, both technically and for completeness, except for the conditions noted above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or designated person, as verified by the following signature.

Signed: \_\_\_\_\_



\_\_\_\_\_

Date: \_\_\_\_\_ 3/26/2012 \_\_\_\_\_

## **REPORT NARRATIVE**

**Spectrum Analytical, Inc. Featuring Hanibal Technology, RI Division.**

**Client : CDM**

**Project: Paul Miller Site, IDW**

**Laboratory Workorder / SDG #: L0466**

**SW846 8081B, Organochlorine Pesticides by GC-ECD**

### **I. SAMPLE RECEIPT**

No exceptions or unusual conditions were encountered unless a Sample Condition Notification Form, or other record of communication is included with the Sample Receipt Documentation.

### **II. HOLDING TIMES**

#### **A. Sample Preparation:**

All samples were prepared within the method-specified holding times.

#### **B. Sample Analysis:**

All samples were analyzed within the method-specified holding times.

### **III. METHODS**

Samples were analyzed following procedures in laboratory test code:  
SW846 8081B

### **IV. PREPARATION**

Aqueous Samples were prepared following procedures in laboratory test code: SW3510

### **V. INSTRUMENTATION**

The following instrumentation was used

Instrument Code: E5

Instrument Type: GC-ECD

Description: HP6890

Manufacturer: Hewlett-Packard

Model: 6890

GC Column used: 30 m X 0.53 mm ID [0.50 um thickness] CLPPest capillary column.

GC Column used: 30 m X 0.53 mm ID [0.42 um thickness] CLPPestII capillary column.

## VI. ANALYSIS

### A. Calibration:

Calibrations met the method/SOP acceptance criteria.

### B. Blanks:

All method blanks were within the acceptance criteria.

### C. Surrogates:

Surrogate standard percent recoveries were within the QC limits with the following exceptions. Please note that the acceptance criteria allow one surrogate recovery outside of the QC limits per fraction.

80230-IDW-AQ (L0466-01B), recovery is below criteria for Decachlorobiphenyl on rear column, at 29% with criteria of (30-135) and Decachlorobiphenyl on front column, at 29% with criteria of (30-135).

### D. Spikes:

#### 1. Laboratory Control Spikes (LCS):

Percent recoveries for lab control samples were within the QC limits.

#### 2. Matrix Spike / Matrix Spike Duplicate (MS/MSD):

No client-requested MS/MSD analyses were included in this SDG.

**E. Dilutions:**

No sample in this SDG required analysis at dilution.

**F. Samples:**

The lower concentration between the primary and confirmatory GC column concentrations is reported due to the presence of interferences unless otherwise indicated. P flags are assigned to compounds when D% between the two columns are greater than 40%.

No other unusual occurrences were noted during sample analysis.

I certify that this data package is in compliance with the terms and conditions agreed to by the client and Spectrum, both technically and for completeness, except for the conditions noted above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or designated person, as verified by the following signature.



Signed: \_\_\_\_\_

Date: \_\_\_\_\_ 3/28/2012

## **REPORT NARRATIVE**

**Spectrum Analytical, Inc. Featuring Hanibal Technology, RI Division.**

**Client : CDM**

**Project: Paul Miller Site, IDW**

**Laboratory Workorder / SDG #: L0466**

**SW846 8082A, PCB by GC-ECD**

### **I. SAMPLE RECEIPT**

No exceptions or unusual conditions were encountered unless a Sample Condition Notification Form, or other record of communication is included with the Sample Receipt Documentation.

### **II. HOLDING TIMES**

#### **A. Sample Preparation:**

All samples were prepared within the method-specified holding times.

#### **B. Sample Analysis:**

All samples were analyzed within the method-specified holding times.

### **III. METHODS**

Samples were analyzed following procedures in laboratory test code:  
SW846 8082A

### **IV. PREPARATION**

Aqueous Samples were prepared following procedures in laboratory test code: SW3510

Soil Samples were prepared following procedures in laboratory test code: SW3550

## **V. INSTRUMENTATION**

The following instrumentation was used

Instrument Code: E2

Instrument Type: GC-ECD

Description: HP5890 II +

Manufacturer: Hewlett-Packard

Model: 5890

GC Column used: 30 m X 0.53 mm ID [0.50 um thickness] CLPPest capillary column.

## **VI. ANALYSIS**

### **A. Calibration:**

Calibrations met the method/SOP acceptance criteria.

### **B. Blanks:**

All method blanks were within the acceptance criteria.

### **C. Surrogates:**

Surrogate standard percent recoveries were within the QC limits with the following exceptions. Please note that the acceptance criteria allow one surrogate recovery outside of the QC limits per fraction.

80230-IDW-AQ (L0466-01B), recovery is below criteria for Decachlorobiphenyl on rear column, at 29% with criteria of (40-135) and Decachlorobiphenyl on front column, at 27% with criteria of (40-135).

80230-IDW-SOLID (L0466-02A), recovery is below criteria for Decachlorobiphenyl on rear column, at 52% with criteria of (60-125) and Decachlorobiphenyl on front column, at 48% with criteria of (60-125).

### **D. Spikes:**

#### **1. Laboratory Control Spikes (LCS):**

Percent recoveries for lab control samples were within the QC limits.

**2. Matrix Spike / Matrix Spike Duplicate (MS/MSD):**

No client-requested MS/MSD analyses were included in this SDG.

**E. Dilutions:**

No sample in this SDG required analysis at dilution.

**F. Samples:**

The lower concentration between the primary and confirmatory GC column concentrations is reported due to the presence of interferences unless otherwise indicated. P flags are assigned to compounds when D% between the two columns are greater than 40%.

No other unusual occurrences were noted during sample analysis.

I certify that this data package is in compliance with the terms and conditions agreed to by the client and Spectrum, both technically and for completeness, except for the conditions noted above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or designated person, as verified by the following signature.



Signed: \_\_\_\_\_

Date: \_\_\_\_\_ 3/28/2012

## **REPORT NARRATIVE**

**Spectrum Analytical, Inc. Featuring Hanibal Technology, RI Division.**

**Client : Sevenson Environmental Services Inc.**

**Project: Roebling Steel**

**Laboratory Workorder / SDG #: L0467**

**SW846 8082A, PCB by GC-ECD**

### **I. SAMPLE RECEIPT**

No exceptions or unusual conditions were encountered unless a Sample Condition Notification Form, or other record of communication is included with the Sample Receipt Documentation.

### **II. HOLDING TIMES**

#### **A. Sample Preparation:**

All samples were prepared within the method-specified holding times.

#### **B. Sample Analysis:**

All samples were analyzed within the method-specified holding times.

### **III. METHODS**

Samples were analyzed following procedures in laboratory test code:  
SW846 8082A

### **IV. PREPARATION**

Soil Samples were prepared following procedures in laboratory test code: SW3550

### **V. INSTRUMENTATION**

The following instrumentation was used

Instrument Code: E2

Instrument Type: GC-ECD

Description: HP5890 II +

Manufacturer: Hewlett-Packard

Model: 5890

GC Column used: 30 m X 0.53 mm ID [0.50 um thickness] CLPPest capillary column.

## VI. ANALYSIS

### A. Calibration:

Calibrations met the method/SOP acceptance criteria.

### B. Blanks:

All method blanks were within the acceptance criteria.

### C. Surrogates:

Surrogate standard percent recoveries were within the QC limits with the following exceptions. Please note that the acceptance criteria allow one surrogate recovery outside of the QC limits per fraction.

RS-BF-A7-030812 (L0467-01A), recovery is below criteria for Decachlorobiphenyl on front column, at 54% with criteria of (60-125).

### D. Spikes:

#### 1. Laboratory Control Spikes (LCS):

Percent recoveries for lab control samples were within the QC limits.

#### 2. Matrix Spike / Matrix Spike Duplicate (MS/MSD):

No client-requested MS/MSD analyses were included in this SDG.

### E. Dilutions:

No sample in this SDG required analysis at dilution.

**F. Samples:**

The lower concentration between the primary and confirmatory GC column concentrations is reported due to the presence of interferences unless otherwise indicated. P flags are assigned to compounds when D% between the two columns are greater than 40%.

No other unusual occurrences were noted during sample analysis.

I certify that this data package is in compliance with the terms and conditions agreed to by the client and Spectrum, both technically and for completeness, except for the conditions noted above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or designated person, as verified by the following signature.



Signed: \_\_\_\_\_

Date: \_\_\_\_\_ 3/28/2012

## **REPORT NARRATIVE**

**Spectrum Analytical, Inc. Featuring Hanibal Technology, RI Division.**

**Client : CDM**

**Project: Paul Miller Site, IDW**

**Laboratory Workorder / SDG #: L0466**

**SW846 8151A, Chlorinated Herbicides by GC-ECD**

### **I. SAMPLE RECEIPT**

No exceptions or unusual conditions were encountered unless a Sample Condition Notification Form, or other record of communication is included with the Sample Receipt Documentation.

### **II. HOLDING TIMES**

#### **A. Sample Preparation:**

All samples were prepared within the method-specified holding times.

#### **B. Sample Analysis:**

All samples were analyzed within the method-specified holding times.

### **III. METHODS**

Samples were analyzed following procedures in laboratory test code:  
SW846 8151A

### **IV. PREPARATION**

Aqueous Samples were prepared following procedures in laboratory test code: SW3510

### **V. INSTRUMENTATION**

The following instrumentation was used

Instrument Code: E4

Instrument Type: GC-ECD

Description: HP6890

Manufacturer: Hewlett-Packard

Model: 6890

GC Column used: 30 m X 0.53 mm ID [0.50 um thickness] CLPPest capillary column.

GC Column used: 30 m X 0.53 mm ID [0.42 um thickness] CLPPestII capillary column.

## VI. ANALYSIS

### A. Calibration:

Calibrations met the method/SOP acceptance criteria.

### B. Blanks:

All method blanks were within the acceptance criteria.

### C. Surrogates:

Surrogate standard percent recoveries were within the QC limits.

### D. Spikes:

#### 1. Laboratory Control Spikes (LCS):

Percent recoveries for lab control samples were within the QC limits.

#### 2. Matrix Spike / Matrix Spike Duplicate (MS/MSD):

No client-requested MS/MSD analyses were included in this SDG.

### E. Dilutions:

No sample in this SDG required analysis at dilution.

### F. Samples:

The lower concentration between the primary and confirmatory GC column concentrations is reported due to the presence of

interferences unless otherwise indicated. P flags are assigned to compounds when D% between the two columns are greater than 40%.

No other unusual occurrences were noted during sample analysis.

I certify that this data package is in compliance with the terms and conditions agreed to by the client and Spectrum, both technically and for completeness, except for the conditions noted above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or designated person, as verified by the following signature.



Signed: \_\_\_\_\_

Date: \_\_\_\_\_ 3/28/2012

## **REPORT NARRATIVE**

**Spectrum Analytical, Inc. Featuring Hanibal Technology, RI Division.**

**Client : CDM**

**Project: Paul Miller Site, IDW**

**Laboratory Workorder / SDG #: L0466**

**SW846 6010C, SW846 7470A**

### **I. SAMPLE RECEIPT**

No exceptions or unusual conditions were encountered unless a Sample Condition Notification Form, or other record of communication is included with the Sample Receipt Documentation.

### **II. HOLDING TIMES**

#### **A. Sample Preparation:**

All samples were prepared within the method-specified holding times.

#### **B. Sample Analysis:**

All samples were analyzed within the method-specified holding times.

### **III. METHODS**

Samples were analyzed following procedures in laboratory test code:  
SW846 6010C, SW846 7470A

### **IV. PREPARATION**

Aqueous Samples were prepared following procedures in laboratory test code: SW3005

Aqueous Samples were prepared following procedures in laboratory test code: SW7470A

## **V. INSTRUMENTATION**

The following instrumentation was used to perform

Instrument Code: FIMS2

Instrument Type: CVAA

Description: FIMS

Manufacturer: Perkin-Elmer

Model: FIMS100

Instrument Code: OPTIMA2

Instrument Type: ICP

Description: Optima 3100 XL

Manufacturer: Perkin-Elmer

Model: 3100 XL

Instrument Code: OPTIMA3

Instrument Type: ICP

Description: Optima ICP-OES

Manufacturer: Perkin-Elmer

Model: 4300 DV

## **VI. ANALYSIS**

### **A. Calibration:**

Calibrations met the method/SOP acceptance criteria.

### **B. Blanks:**

All method blanks were within the acceptance criteria.

### **C. Spikes:**

#### **1. Laboratory Control Spikes (LCS):**

Percent recoveries for laboratory control samples were within the QC limits.

#### **2. Matrix spike (MS):**

A matrix spike was not performed on any sample in this SDG.

**D. Post Digestion Spike (PDS):**

A post-digestion spike was not performed on any sample in this SDG.

**E. Duplicate sample:**

A duplicate analysis was not performed on any sample in this SDG.

**F. Serial Dilution (SD):**

Serial Dilution analysis was performed on sample: 80230-IDW-SOLID (L0466-02ASD).

Percent RPD was within the QC limits.

**G. Samples:**

No other unusual occurrences were noted during sample analysis.

I certify that this data package is in compliance with the terms and conditions agreed to by the client and Spectrum RI, both technically and for completeness, except for the conditions noted above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or designated person, as verified by the following signature.

Signed: 

Date: 03/29/12

## **REPORT NARRATIVE**

**Spectrum Analytical, Inc.    Featuring Hanibal Technology, RI Division.**

**Client : CDM**

**Project: Paul Miller Site,    IDW**

**Laboratory Workorder / SDG #: L0466**

**SM 4500 H+ B, SW846 1010, SW846 7.3.3.2, SW846 7.3.4.2, SW846  
9045C**

### **I. SAMPLE RECEIPT**

No exceptions or unusual conditions were encountered unless a Sample Condition Notification Form, or other record of communication is included with the Sample Receipt Documentation.

### **II. HOLDING TIMES**

#### **A. Sample Preparation:**

All samples were prepared within the method-specified holding times.

#### **B. Sample Analysis:**

All samples were analyzed within the method-specified holding times.

### **III. METHODS**

Samples were analyzed following procedures in laboratory test code: SM 4500 H+ B, SW846 1010, SW846 7.3.3.2, SW846 7.3.4.2, SW846 9045C

### **IV. PREPARATION**

Aqueous Samples were prepared following procedures in laboratory test code: SM 4500 H+ B, SW846 1010, SW846 7.3.3.2, SW846 7.3.4.2,

## V. INSTRUMENTATION

The following instrumentation was used to perform

Instrument Code: FLASH1

Instrument Type: WC

Description: Flash Point

Manufacturer: Koehler

Model: K16200

Instrument Code: LACHAT1

Instrument Type: WC

Description: Flow Injection Analyzer

Manufacturer: Zellweger Analytics

Model: Quik-Chem 8000

Instrument Code: SPEC2

Instrument Type: SP

Description: Spectronic 20 Genesys

Manufacturer: Spectronic Instruments

Model: 4004-000

Instrument Code: WC01

Instrument Type: Probe

Description: pH Meter

Manufacturer: Thermo Electron Corporation

Model: Orion 520A+

## VI. ANALYSIS

### A. Calibration:

Calibrations met the method/SOP acceptance criteria.

### B. Blanks:

All method blanks were within the acceptance criteria.

### C. Spikes:

#### 1. Laboratory Control Spikes (LCS):

Percent recoveries for lab control samples were within the QC limits.

The lab control sample for reactive cyanide typically results in a percent recovery of approximately 10%. This indicates the analysis is only measuring the correct type of cyanide. The LCS is performed with a “total” cyanide spike, only a small portion of which is “reactive”. Recovery of significantly greater than this level in the analysis of the LCS indicates the test is over estimating the concentration of reactive cyanide.

## **2. Matrix Spike / Matrix Spike Duplicate (MS/MSD):**

No client-requested MS/MSD analyses were included in this SDG.

### **D. Duplicate sample:**

Duplicate analysis was performed on sample: 80230-IDW-AQ (L0466-01CDUP) for pH and 80230-IDW-SOLID (L0466-02BDUP) for pH and Flashpoint.

Percent RPD was within the QC limits for all analyses.

### **E. Dilutions:**

No sample required dilution in this SDG.

### **F. Samples:**

For Flashpoint analysis, the result of “No Flash” indicates that no flash was observed, or that non-ignitable vapors from the sample extinguished the test flame at the temperature indicated. Where non-flammable vapors from the sample extinguished the test flame below 140 degrees, the sample temperature was elevated to at least 140 degrees, and re-exposed to the test flame.

No other unusual occurrences were noted during sample analysis.

I certify that this data package is in compliance with the terms and conditions agreed to by the client and Spectrum RI, both technically and for completeness, except for the conditions noted above.

Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or designated person, as

verified by the following signature.

Signed: 

Date: 03/29/12

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/29/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result	Qual	RL	Units	DF	Date Analyzed	Batch ID
<b>SW846 8260C -- VOC by GC-MS</b>							
Dichlorodifluoromethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Chloromethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Vinyl chloride	13		5.0	ug/L		1 03/16/2012 20:35	65160
Bromomethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Chloroethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Trichlorofluoromethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,1-Dichloroethene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Acetone	3.3	J	5.0	ug/L		1 03/16/2012 20:35	65160
Iodomethane	2.2	J	5.0	ug/L		1 03/16/2012 20:35	65160
Carbon disulfide	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Methylene chloride	ND		5.0	ug/L		1 03/16/2012 20:35	65160
trans-1,2-Dichloroethene	3.9	J	5.0	ug/L		1 03/16/2012 20:35	65160
Methyl tert-butyl ether	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,1-Dichloroethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Vinyl acetate	ND		5.0	ug/L		1 03/16/2012 20:35	65160
2-Butanone	ND		5.0	ug/L		1 03/16/2012 20:35	65160
cis-1,2-Dichloroethene	510	E	5.0	ug/L		1 03/16/2012 20:35	65160
2,2-Dichloropropane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Bromochloromethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Chloroform	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,1,1-Trichloroethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,1-Dichloropropene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Carbon tetrachloride	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,2-Dichloroethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Benzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Trichloroethene	87		5.0	ug/L		1 03/16/2012 20:35	65160
1,2-Dichloropropane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Dibromomethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Bromodichloromethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
cis-1,3-Dichloropropene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
4-Methyl-2-pentanone	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Toluene	8.0		5.0	ug/L		1 03/16/2012 20:35	65160
trans-1,3-Dichloropropene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,1,2-Trichloroethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,3-Dichloropropane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Tetrachloroethene	1600	E	5.0	ug/L		1 03/16/2012 20:35	65160
2-Hexanone	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Dibromochloromethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,2-Dibromoethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160

**Qualifiers:** ND - Not Detected at the Reporting Limit

S - Spike Recovery outside accepted recovery limits

J - Analyte detected below quantitation limits

R - RPD outside accepted recovery limits

B - Analyte detected in the associated Method Blank

E - Value above quantitation range

DF - Dilution Factor

RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result	Qual	RL	Units	DF	Date Analyzed	Batch ID
<b>SW846 8260C -- VOC by GC-MS</b>							<b>SW8260_W</b>
Chlorobenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,1,1,2-Tetrachloroethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Ethylbenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
m,p-Xylene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
o-Xylene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Xylene (Total)	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Styrene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Bromoform	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Isopropylbenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,1,2,2-Tetrachloroethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Bromobenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,2,3-Trichloropropane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
n-Propylbenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
2-Chlorotoluene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,3,5-Trimethylbenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
4-Chlorotoluene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
tert-Butylbenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,2,4-Trimethylbenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
sec-Butylbenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
4-Isopropyltoluene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,3-Dichlorobenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,4-Dichlorobenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
n-Butylbenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,2-Dichlorobenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,2-Dibromo-3-chloropropane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,2,4-Trichlorobenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Hexachlorobutadiene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,2,3-Trichlorobenzene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Naphthalene	ND		5.0	ug/L		1 03/16/2012 20:35	65160
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Cyclohexane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Methyl acetate	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Methylcyclohexane	ND		5.0	ug/L		1 03/16/2012 20:35	65160
Surrogate: Dibromofluoromethane	110		85-115	%REC		1 03/16/2012 20:35	65160
Surrogate: 1,2-Dichloroethane-d4	102		70-120	%REC		1 03/16/2012 20:35	65160
Surrogate: Toluene-d8	91.9		85-120	%REC		1 03/16/2012 20:35	65160
Surrogate: Bromofluorobenzene	90.6		75-120	%REC		1 03/16/2012 20:35	65160

**Qualifiers:** ND - Not Detected at the Reporting Limit

J - Analyte detected below quantitation limits

B - Analyte detected in the associated Method Blank

DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits

R - RPD outside accepted recovery limits

E - Value above quantitation range

RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/29/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result	Qual	RL	Units	DF	Date Analyzed	Batch ID
<b>SW846 8260C -- VOC by GC-MS</b>							
Dichlorodifluoromethane	ND		50	ug/L		10 03/28/2012 13:57	65295
Chloromethane	ND		50	ug/L		10 03/28/2012 13:57	65295
Vinyl chloride	15	J	50	ug/L		10 03/28/2012 13:57	65295
Bromomethane	ND		50	ug/L		10 03/28/2012 13:57	65295
Chloroethane	ND		50	ug/L		10 03/28/2012 13:57	65295
Trichlorofluoromethane	ND		50	ug/L		10 03/28/2012 13:57	65295
1,1-Dichloroethene	ND		50	ug/L		10 03/28/2012 13:57	65295
Acetone	ND		50	ug/L		10 03/28/2012 13:57	65295
Iodomethane	9.1	BJ	50	ug/L		10 03/28/2012 13:57	65295
Carbon disulfide	ND		50	ug/L		10 03/28/2012 13:57	65295
Methylene chloride	ND		50	ug/L		10 03/28/2012 13:57	65295
trans-1,2-Dichloroethene	ND		50	ug/L		10 03/28/2012 13:57	65295
Methyl tert-butyl ether	ND		50	ug/L		10 03/28/2012 13:57	65295
1,1-Dichloroethane	ND		50	ug/L		10 03/28/2012 13:57	65295
Vinyl acetate	ND		50	ug/L		10 03/28/2012 13:57	65295
2-Butanone	ND		50	ug/L		10 03/28/2012 13:57	65295
cis-1,2-Dichloroethene	500		50	ug/L		10 03/28/2012 13:57	65295
2,2-Dichloropropane	ND		50	ug/L		10 03/28/2012 13:57	65295
Bromochloromethane	ND		50	ug/L		10 03/28/2012 13:57	65295
Chloroform	ND		50	ug/L		10 03/28/2012 13:57	65295
1,1,1-Trichloroethane	ND		50	ug/L		10 03/28/2012 13:57	65295
1,1-Dichloropropene	ND		50	ug/L		10 03/28/2012 13:57	65295
Carbon tetrachloride	ND		50	ug/L		10 03/28/2012 13:57	65295
1,2-Dichloroethane	ND		50	ug/L		10 03/28/2012 13:57	65295
Benzene	ND		50	ug/L		10 03/28/2012 13:57	65295
Trichloroethene	82		50	ug/L		10 03/28/2012 13:57	65295
1,2-Dichloropropane	ND		50	ug/L		10 03/28/2012 13:57	65295
Dibromomethane	ND		50	ug/L		10 03/28/2012 13:57	65295
Bromodichloromethane	ND		50	ug/L		10 03/28/2012 13:57	65295
cis-1,3-Dichloropropene	ND		50	ug/L		10 03/28/2012 13:57	65295
4-Methyl-2-pentanone	ND		50	ug/L		10 03/28/2012 13:57	65295
Toluene	9.2	J	50	ug/L		10 03/28/2012 13:57	65295
trans-1,3-Dichloropropene	ND		50	ug/L		10 03/28/2012 13:57	65295
1,1,2-Trichloroethane	ND		50	ug/L		10 03/28/2012 13:57	65295
1,3-Dichloropropane	ND		50	ug/L		10 03/28/2012 13:57	65295
Tetrachloroethene	1700		50	ug/L		10 03/28/2012 13:57	65295
2-Hexanone	ND		50	ug/L		10 03/28/2012 13:57	65295
Dibromochloromethane	ND		50	ug/L		10 03/28/2012 13:57	65295
1,2-Dibromoethane	ND		50	ug/L		10 03/28/2012 13:57	65295

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 8260C -- VOC by GC-MS</b>						<b>SW8260_W</b>
Chlorobenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,1,1,2-Tetrachloroethane	ND		50 ug/L		10 03/21/2012 15:12	65204
Ethylbenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
m,p-Xylene	ND		50 ug/L		10 03/21/2012 15:12	65204
o-Xylene	ND		50 ug/L		10 03/21/2012 15:12	65204
Xylene (Total)	ND		50 ug/L		10 03/21/2012 15:12	65204
Styrene	ND		50 ug/L		10 03/21/2012 15:12	65204
Bromoform	ND		50 ug/L		10 03/21/2012 15:12	65204
Isopropylbenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,1,2,2-Tetrachloroethane	ND		50 ug/L		10 03/21/2012 15:12	65204
Bromobenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,2,3-Trichloropropane	ND		50 ug/L		10 03/21/2012 15:12	65204
n-Propylbenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
2-Chlorotoluene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,3,5-Trimethylbenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
4-Chlorotoluene	ND		50 ug/L		10 03/21/2012 15:12	65204
tert-Butylbenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,2,4-Trimethylbenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
sec-Butylbenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
4-Isopropyltoluene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,3-Dichlorobenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,4-Dichlorobenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
n-Butylbenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,2-Dichlorobenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,2-Dibromo-3-chloropropane	ND		50 ug/L		10 03/21/2012 15:12	65204
1,2,4-Trichlorobenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
Hexachlorobutadiene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,2,3-Trichlorobenzene	ND		50 ug/L		10 03/21/2012 15:12	65204
Naphthalene	ND		50 ug/L		10 03/21/2012 15:12	65204
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		50 ug/L		10 03/21/2012 15:12	65204
Cyclohexane	ND		50 ug/L		10 03/21/2012 15:12	65204
Methyl acetate	ND		50 ug/L		10 03/21/2012 15:12	65204
Methylcyclohexane	ND		50 ug/L		10 03/21/2012 15:12	65204
Surrogate: Dibromofluoromethane	116	S	85-115 %REC		10 03/21/2012 15:12	65204
Surrogate: 1,2-Dichloroethane-d4	98.1		70-120 %REC		10 03/21/2012 15:12	65204
Surrogate: Toluene-d8	85.7		85-120 %REC		10 03/21/2012 15:12	65204
Surrogate: Bromofluorobenzene	105		75-120 %REC		10 03/21/2012 15:12	65204

**Qualifiers:** ND - Not Detected at the Reporting Limit

J - Analyte detected below quantitation limits

B - Analyte detected in the associated Method Blank

DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits

R - RPD outside accepted recovery limits

E - Value above quantitation range

RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-SOLID  
**Lab ID:** L0466-02

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:00

Analyses	Result	Qual	RL	Units	DF	Date Analyzed	Batch ID
<b>SW846 8260C -- VOC by GC-MS</b>							<b>SW8260_LOW_S</b>
Dichlorodifluoromethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Chloromethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Vinyl chloride	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Bromomethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Chloroethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Trichlorofluoromethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
1,1-Dichloroethene	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Acetone	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Iodomethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Carbon disulfide	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Methylene chloride	1.7	BJ	5.6	ug/Kg		1 03/16/2012 13:19	65144
trans-1,2-Dichloroethene	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Methyl tert-butyl ether	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
1,1-Dichloroethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Vinyl acetate	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
2-Butanone	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
cis-1,2-Dichloroethene	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
2,2-Dichloropropane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Bromochloromethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Chloroform	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
1,1,1-Trichloroethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
1,1-Dichloropropene	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Carbon tetrachloride	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
1,2-Dichloroethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Benzene	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Trichloroethene	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
1,2-Dichloropropane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Dibromomethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Bromodichloromethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
cis-1,3-Dichloropropene	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
4-Methyl-2-pentanone	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Toluene	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
trans-1,3-Dichloropropene	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
1,1,2-Trichloroethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
1,3-Dichloropropane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Tetrachloroethene	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
2-Hexanone	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
Dibromochloromethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144
1,2-Dibromoethane	ND		5.6	ug/Kg		1 03/16/2012 13:19	65144

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-SOLID  
**Lab ID:** L0466-02

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:00

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 8260C -- VOC by GC-MS</b>						<b>SW8260_LOW_S</b>
Chlorobenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,1,1,2-Tetrachloroethane	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Ethylbenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
m,p-Xylene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
o-Xylene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Xylene (Total)	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Styrene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Bromoform	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Isopropylbenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,1,2,2-Tetrachloroethane	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Bromobenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,2,3-Trichloropropane	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
n-Propylbenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
2-Chlorotoluene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,3,5-Trimethylbenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
4-Chlorotoluene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
tert-Butylbenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,2,4-Trimethylbenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
sec-Butylbenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
4-Isopropyltoluene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,3-Dichlorobenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,4-Dichlorobenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
n-Butylbenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,2-Dichlorobenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,2-Dibromo-3-chloropropane	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,2,4-Trichlorobenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Hexachlorobutadiene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,2,3-Trichlorobenzene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Naphthalene	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
1,1,2-Trichloro-1,2,2-trifluoroethane	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Cyclohexane	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Methyl acetate	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Methylcyclohexane	ND		5.6 ug/Kg		1 03/16/2012 13:19	65144
Surrogate: Dibromofluoromethane	106		65-132 %REC		1 03/16/2012 13:19	65144
Surrogate: 1,2-Dichloroethane-d4	105		65-128 %REC		1 03/16/2012 13:19	65144
Surrogate: Toluene-d8	90.8		85-115 %REC		1 03/16/2012 13:19	65144
Surrogate: Bromofluorobenzene	92.6		77-111 %REC		1 03/16/2012 13:19	65144

## SW846 8260C -- VOC by GC-MS

**SW8260\_W**

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

Client: CDM  
Client Sample ID: 80230-IDW-SOLID  
Lab ID: L0466-02

Project: Paul Miller Site, IDW  
Collection Date: 03/08/12 11:00

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 8260C -- VOC by GC-MS</b>						<b>SW8260_W</b>
Vinyl chloride -- TCLP	ND		5.0 ug/L	1	03/20/2012 17:26	65195
1,1-Dichloroethene -- TCLP	ND		5.0 ug/L	1	03/20/2012 17:26	65195
2-Butanone -- TCLP	ND		5.0 ug/L	1	03/20/2012 17:26	65195
Chloroform -- TCLP	ND		5.0 ug/L	1	03/20/2012 17:26	65195
Carbon tetrachloride -- TCLP	ND		5.0 ug/L	1	03/20/2012 17:26	65195
1,2-Dichloroethane -- TCLP	ND		5.0 ug/L	1	03/20/2012 17:26	65195
Benzene -- TCLP	ND		5.0 ug/L	1	03/20/2012 17:26	65195
Trichloroethene -- TCLP	ND		5.0 ug/L	1	03/20/2012 17:26	65195
Tetrachloroethene -- TCLP	ND		5.0 ug/L	1	03/20/2012 17:26	65195
Chlorobenzene -- TCLP	ND		5.0 ug/L	1	03/20/2012 17:26	65195
Surrogate: Dibromofluoromethane -- TCLP	116	S	85-115 %REC	1	03/20/2012 17:26	65195
Surrogate: 1,2-Dichloroethane-d4 -- TCLP	104		70-120 %REC	1	03/20/2012 17:26	65195
Surrogate: Toluene-d8 -- TCLP	88.2		85-120 %REC	1	03/20/2012 17:26	65195
Surrogate: Bromofluorobenzene -- TCLP	103		75-120 %REC	1	03/20/2012 17:26	65195

Qualifiers: ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hamibal Tech

Date: 03/28/2012

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

SW8260\_LOW\_S

SW846 8260C -- VOC by GC-MS

Analyte	Result	MDL	TestCode: SW8260_LOW_S			SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPD Limit	Qual
			Units: ug/Kg	RL	SPK value							
Dichlorodifluoromethane	ND	0.98	5.0									
Chloromethane	ND	0.80	5.0									
Vinyl chloride	ND	0.63	5.0									
Bromomethane	ND	1.1	5.0									
Chloroethane	ND	1.0	5.0									
Trichlorofluoromethane	ND	0.42	5.0									
1,1-Dichloroethene	ND	0.95	5.0									
Acetone	ND	1.6	5.0									
Iodomethane	ND	0.69	5.0									
Carbon disulfide	ND	0.30	5.0									
Methylene chloride	1.424	1.3	5.0									
trans-1,2-Dichloroethene	ND	0.53	5.0									
Methyl tert-butyl ether	ND	0.61	5.0									
1,1-Dichloroethane	ND	0.67	5.0									
Vinyl acetate	ND	0.37	5.0									
2-Butanone	ND	2.0	5.0									
cis-1,2-Dichloroethene	ND	0.75	5.0									
2,2-Dichloropropane	ND	0.29	5.0									
Bromo-chloromethane	ND	0.76	5.0									
Chloroform	ND	0.64	5.0									
1,1,1-Trichloroethane	ND	0.53	5.0									
1,1-Dichloropropene	ND	0.81	5.0									
Carbon tetrachloride	ND	0.33	5.0									
1,2-Dichloroethane	ND	0.54	5.0									
Benzene	ND	0.61	5.0									
Trichloroethene	ND	0.62	5.0									
1,2-Dichloropropane	ND	0.69	5.0									
Dibromomethane	ND	0.58	5.0									
Bromodichloromethane	ND	0.97	5.0									
cis-1,3-Dichloropropene	ND	0.67	5.0									
4-Methyl-2-pentanone	ND	0.73	5.0									
Toluene	ND	0.47	5.0									
trans-1,3-Dichloropropene	ND	0.68	5.0									
1,1,2-Trichloroethane	ND	0.48	5.0									
1,3-Dichloropropane	ND	0.87	5.0									
Tetrachloroethene	ND	0.62	5.0									

Qualifiers: ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

mm11.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

**ANALYTICAL QC SUMMARY REPORT**

**SW8260\_LOW\_S**  
**SW846 8260C .. VOC by GC-MS**

Sample ID:	MB-65144	SampType:	MBLK	TestCode:	SW8260_LOW_S	Prep Date:	03/16/12 8:16	Run ID:	V5_120316A				
Client ID:	MB-65144	Batch ID:	65144	Units:	ug/Kg	Analysis Date:	03/16/12 11:29	SeqNo:	1707523				
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
2-Hexanone		ND	0.83		5.0								
Dibromochloromethane		ND	0.65		5.0								
1,2-Dibromoethane		ND	0.74		5.0								
Chlorobenzene		ND	0.51		5.0								
1,1,1,2-Tetrachloroethane		ND	0.77		5.0								
Ethybenzene		ND	0.50		5.0								
m,p-Xylene		ND	1.6		5.0								
o-Xylene		ND	0.47		5.0								
Xylene (Total)		ND	0.47		5.0								
Styrene		ND	0.52		5.0								
Bromoform		ND	2.0		5.0								
Isopropylbenzene		ND	0.58		5.0								
1,1,2,2-Tetrachloroethane		ND	0.68		5.0								
Bromobenzene		ND	0.58		5.0								
1,2,3-Trichloropropane		ND	0.87		5.0								
n-Propylbenzene		ND	0.44		5.0								
2-Chlorotoluene		ND	0.74		5.0								
1,3,5-Trimethylbenzene		ND	0.61		5.0								
4-Chlorotoluene		ND	0.84		5.0								
tert-Butylbenzene		ND	0.52		5.0								
1,2,4-Trimethylbenzene		ND	0.57		5.0								
sec-Butylbenzene		ND	0.62		5.0								
4-Isopropyltoluene		ND	0.71		5.0								
1,3-Dichlorobenzene		ND	0.70		5.0								
1,4-Dichlorobenzene		ND	0.80		5.0								
n-Butylbenzene		ND	0.67		5.0								
1,2-Dichlorobenzene		ND	0.62		5.0								
1,2-Dibromo-3-chloropropane		ND	1.3		5.0								
1,2,4-Trichlorobenzene		ND	0.63		5.0								
Hexachlorobutadiene		ND	0.62		5.0								
1,2,3-Trichlorobenzene		ND	0.64		5.0								
Naphthalene		ND	0.78		5.0								
1,1,2-Trichloro-1,2,2-trifluoroethane		ND	3.0		5.0								
Cyclohexane		ND	1.7		5.0								
Methyl acetate		ND	1.4		5.0								
Methylcyclohexane		ND	1.8		5.0								

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mm11.12.11.A

**S - Recovery outside accepted recovery limits**  
**R - RPD outside accepted recovery limits**

**MDL - Method Detection Limit**  
**RL - Reporting Limit**

**B - Analyte detected in the associated Method Blank**

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

**ANALYTICAL QC SUMMARY REPORT**  
**SW8260\_LOW\_S**  
**SW846 8260C -- VOC by GC-MS**

Sample ID:	MB-65144	SampType:	MBLK	TestCode:	SW8260_LOW_S	Prep Date:	03/16/12 8:16	Run ID:	V5_120316A				
Client ID:	MB-65144	Batch ID:	65144	Units:	ug/Kg	Analysis Date:	03/16/12 11:29	SeqNo:	1707523				
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Surrogate:		51.45	5.0	50.00	0	103	65	132	0				
Dibromofluoromethane		51.40	5.0	50.00	0	103	65	128	0				
Surrogate: 1,2-Dichloroethane-d4		47.09	5.0	50.00	0	94.2	85	115	0				
Surrogate: Toluene-d8		47.02	5.0	50.00	0	94.0	77	111	0				
Bromofluorobenzene													

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mm11.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

**B - Analyte detected in the associated Method Blank**

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_LOW\_S**  
**SW846 8260C -- VOC by GC-MS**

Sample ID: LCS-65144	SampType: LCS	TestCode: SW8260_LOW_S	Prep Date: 03/16/12 8:16	Run ID: V5_120316A
Client ID: LCS-65144	Batch ID: 65144	Units: ug/Kg	Analysis Date: 03/16/12 10:34	SeqNo: 1707519
Analyte	Result	MDL	SPK value	SPK Ref Val
Dichlorodifluoromethane	44.92	0.98	5.0	50.00
Chloromethane	49.83	0.80	5.0	50.00
Vinyl chloride	50.17	0.63	5.0	50.00
Bromomethane	49.96	1.1	5.0	50.00
Chloroethane	51.51	1.0	5.0	50.00
Trichlorofluoromethane	49.49	0.42	5.0	50.00
1,1-Dichloroethene	55.30	0.95	5.0	50.00
Acetone	80.13	1.6	5.0	50.00
Iodomethane	51.76	0.69	5.0	50.00
Carbon disulfide	53.61	0.30	5.0	50.00
Methylene chloride	44.38	1.3	5.0	50.00
trans-1,2-Dichloroethene	52.96	0.53	5.0	50.00
Methyl tert-butyl ether	47.34	0.61	5.0	50.00
1,1-Dichloroethane	51.18	0.67	5.0	50.00
Vinyl acetate	48.79	0.37	5.0	50.00
2-Butanone	67.97	2.0	5.0	50.00
cis-1,2-Dichloroethene	51.30	0.75	5.0	50.00
2,2-Dichloropropane	53.39	0.29	5.0	50.00
Bromochloromethane	48.15	0.76	5.0	50.00
Chloroform	49.23	0.64	5.0	50.00
1,1,1-Trichloroethane	51.57	0.53	5.0	50.00
1,1-Dichloropropene	51.81	0.81	5.0	50.00
Carbon tetrachloride	50.09	0.33	5.0	50.00
1,2-Dichloroethane	47.09	0.54	5.0	50.00
Benzene	51.25	0.61	5.0	50.00
Trichloroethene	51.22	0.62	5.0	50.00
1,2-Dichloropropane	48.93	0.69	5.0	50.00
Dibromomethane	47.62	0.58	5.0	50.00
Bromodichloromethane	47.93	0.97	5.0	50.00
cis-1,3-Dichloropropene	49.85	0.67	5.0	50.00
4-Methyl-2-pentanone	55.28	0.73	5.0	50.00
Toluene	51.11	0.47	5.0	50.00
trans-1,3-Dichloropropene	49.70	0.68	5.0	50.00
1,1,2-Trichloroethane	47.87	0.48	5.0	50.00
1,3-Dichloropropane	49.30	0.87	5.0	50.00
Tetrachloroethene	50.71	0.62	5.0	50.00
2-Hexanone	65.60	0.83	5.0	50.00
Dibromochloromethane	46.38	0.65	5.0	50.00

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits

**MDL - Method Detection Limit**  
R - RPD outside accepted recovery limits

**RL - Reporting Limit**

**B - Analyte detected in the associated Method Blank**

mm1.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_LOW\_S**  
**SW846 8260C -- VOC by GC-MS**

Sample ID:	LCS-65144	SampType:	LCS	TestCode:	SW8260_LOW_S	Prep Date:	03/16/12 8:16	Run ID:	V5_120316A				
Client ID:	LCS-65144	Batch ID:	65144	Units:	ug/Kg	Analysis Date:	03/16/12 10:34	SeqNo:	1707519				
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
1,2-Dibromoethane	47.52	0.74	5.0	50.00	0	95.0	70	125	0	0	0	0	
Chlorobenzene	50.81	0.51	5.0	50.00	0	102	75	125	0	0	0	0	
1,1,1,2-Tetrachloroethane	49.41	0.77	5.0	50.00	0	98.8	75	125	0	0	0	0	
Ethylbenzene	53.06	0.50	5.0	50.00	0	106	75	125	0	0	0	0	
m,p-Xylene	107.8	1.6	5.0	100.0	0	108	80	125	0	0	0	0	
o-Xylene	51.75	0.47	5.0	50.00	0	104	75	125	0	0	0	0	
Xylene (Total)	159.5	0.47	5.0	150.0	0	106	83	125	0	0	0	0	
Styrene	52.35	0.52	5.0	50.00	0	105	75	125	0	0	0	0	
Bromoform	47.38	2.0	5.0	50.00	0	94.8	55	135	0	0	0	0	
Isopropylbenzene	51.40	0.58	5.0	50.00	0	103	75	130	0	0	0	0	
1,1,2,2-Tetrachloroethane	50.35	0.68	5.0	50.00	0	101	55	130	0	0	0	0	
Bromobenzene	52.09	0.58	5.0	50.00	0	104	65	120	0	0	0	0	
1,2,3-Trichloropropane	49.05	0.87	5.0	50.00	0	98.1	65	130	0	0	0	0	
n-Propylbenzene	53.99	0.44	5.0	50.00	0	108	65	135	0	0	0	0	
2-Chlorotoluene	53.07	0.74	5.0	50.00	0	106	70	130	0	0	0	0	
1,3,5-Trimethylbenzene	53.47	0.61	5.0	50.00	0	107	65	135	0	0	0	0	
4-Chlorotoluene	54.80	0.84	5.0	50.00	0	110	75	125	0	0	0	0	
tert-Butylbenzene	56.33	0.52	5.0	50.00	0	113	65	130	0	0	0	0	
1,2,4-Trimethylbenzene	51.37	0.57	5.0	50.00	0	103	65	135	0	0	0	0	
sec-Butylbenzene	53.16	0.62	5.0	50.00	0	106	65	130	0	0	0	0	
4-Isopropyltoluene	53.10	0.71	5.0	50.00	0	106	75	135	0	0	0	0	
1,3-Dichlorobenzene	52.99	0.70	5.0	50.00	0	106	70	125	0	0	0	0	
1,4-Dichlorobenzene	50.52	0.80	5.0	50.00	0	101	70	125	0	0	0	0	
n-Butylbenzene	55.42	0.67	5.0	50.00	0	111	65	140	0	0	0	0	
1,2-Dichlorobenzene	48.97	0.62	5.0	50.00	0	97.9	75	120	0	0	0	0	
1,2-Dibromo-3-chloropropane	44.77	1.3	5.0	50.00	0	89.5	40	135	0	0	0	0	
1,2,4-Trichlorobenzene	51.04	0.63	5.0	50.00	0	102	65	130	0	0	0	0	
Hexachlorobutadiene	49.39	0.62	5.0	50.00	0	98.8	55	140	0	0	0	0	
1,2,3-Trichlorobenzene	50.31	0.64	5.0	50.00	0	101	60	135	0	0	0	0	
Naphthalene	61.08	0.78	5.0	50.00	0	122	40	125	0	0	0	0	
1,1,2-Trichloro-1,2,2-trifluoroethane	52.75	3.0	5.0	50.00	0	105	70	130	0	0	0	0	
Cyclohexane	51.37	1.7	5.0	50.00	0	103	70	130	0	0	0	0	
Methyl acetate	46.89	1.4	5.0	50.00	0	93.8	70	130	0	0	0	0	
Methylcyclohexane	51.46	1.8	5.0	50.00	0	103	70	130	0	0	0	0	
Surrogate:	48.69	5.0	50.00	50.00	0	97.4	65	132	0	0	0	0	
Dibromofluoromethane													

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits

**S - Recovery outside accepted recovery limits**  
**R - RPD outside accepted recovery limits**

**MDL - Method Detection Limit**  
**RL - Reporting Limit**

**B - Analyte detected in the associated Method Blank**

mm11.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

**ANALYTICAL QC SUMMARY REPORT**  
SW8260\_LOW\_S  
SW846 8260C -- VOC by GC-MS

Sample ID:	LCS-65144	SampType:	LCS	Batch ID:	65144	TestCode:	SW8260_LOW_S	Units:	ug/Kg	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Client ID:	LCS-65144																	
Analyte		Result	MDL		RL													
Surrogate: 1,2-Dichloroethane-d4		46.11			5.0		50.00		0		92.2	65	128		0			
Surrogate: Toluene-d8		49.63			5.0		50.00		0		99.3	85	115		0			
Surrogate: Bromofluorobenzene		49.18			5.0		50.00		0		98.4	77	111		0			

Qualifiers:  
ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mm11.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**

**SW846 8260C -- VOC by GC-MS**

Sample ID:	MB-65080	SampType:	MBLK	TestCode:	SW8260_W	Prep Date:	03/20/12 9:05	Run ID:	V5_120320B				
Client ID:	MB-65080	Batch ID:	65195	Units:	ug/L	Analysis Date:	03/20/12 16:32	SeqNo:	1708675				
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Vinyl chloride -- TCLP		ND	0.50	5.0	0	0	0	0	0	0	0	0	
1,1-Dichloroethene -- TCLP		ND	0.39	5.0	0	0	0	0	0	0	0	0	
2-Butanone -- TCLP		ND	2.1	5.0	0	0	0	0	0	0	0	0	
Chloroform -- TCLP		ND	0.33	5.0	0	0	0	0	0	0	0	0	
Carbon tetrachloride -- TCLP		ND	0.54	5.0	0	0	0	0	0	0	0	0	
1,2-Dichloroethane -- TCLP		ND	0.41	5.0	0	0	0	0	0	0	0	0	
Benzene -- TCLP		ND	0.33	5.0	0	0	0	0	0	0	0	0	
Trichloroethene -- TCLP		ND	0.36	5.0	0	0	0	0	0	0	0	0	
Tetrachloroethene -- TCLP		ND	0.65	5.0	0	0	0	0	0	0	0	0	
Chlorobenzene -- TCLP		ND	0.26	5.0	0	0	0	0	0	0	0	0	
Surrogate:			56.66	5.0	50.00	0	113	85	115	0	0	0	
Dibromofluoromethane --													
TCLP													
Surrogate: 1,2-		49.19	5.0	50.00	0	98.4	70	120	0	0	0	0	
Dichloroethane-d4 -- TCLP													
Surrogate: Toluene-d8 --		44.02	5.0	50.00	0	88.0	85	120	0	0	0	0	
TCLP													
Surrogate:		51.99	5.0	50.00	0	104	75	120	0	0	0	0	
Bromofluorobenzene -- TCLP													

Qualifiers: ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mm.11.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**  
**SW846 8260C -- VOC by GC-MS**

Sample ID:	MB-65160	SampType:	MBLK	TestCode:	SW8260_W	Prep Date:	03/16/12 16:38	Run ID:	V1_120316B				
Client ID:	MB-65160	Batch ID:	65160	Units:	ug/L	Analysis Date:	03/16/12 19:13	SeqNo:	1708143				
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Dichlorodifluoromethane	ND	0.66			5.0								
Chloromethane	ND	0.26			5.0								
Vinyl chloride	ND	0.50			5.0								
Bromomethane	ND	0.80			5.0								
Chloroethane	ND	0.48			5.0								
Trichlorofluoromethane	ND	0.54			5.0								
1,1-Dichloroethene	ND	0.39			5.0								
Acetone	ND	2.2			5.0								
Iodomethane	ND	0.63			5.0								
Carbon disulfide	ND	0.34			5.0								
Methylene chloride	ND	0.41			5.0								
trans-1,2-Dichloroethene	ND	0.65			5.0								
Methyl tert-butyl ether	ND	0.24			5.0								
1,1-Dichloroethane	ND	0.25			5.0								
Vinyl acetate	ND	0.35			5.0								
2-Butanone	ND	2.1			5.0								
cis-1,2-Dichloroethene	ND	0.48			5.0								
2,2-Dichloropropane	ND	0.30			5.0								
Bromochloromethane	ND	0.43			5.0								
Chloroform	ND	0.33			5.0								
1,1,1-Trichloroethane	ND	0.50			5.0								
1,1-Dichloropropene	ND	0.50			5.0								
Carbon tetrachloride	ND	0.54			5.0								
1,2-Dichloroethane	ND	0.41			5.0								
Benzene	ND	0.33			5.0								
Trichloroethene	ND	0.36			5.0								
1,2-Dichloropropane	ND	0.61			5.0								
Dibromomethane	ND	0.49			5.0								
Bromodichloromethane	ND	0.26			5.0								
cis-1,3-Dichloropropene	ND	0.45			5.0								
4-Methyl-2-pentanone	ND	0.82			5.0								
Toluene	ND	0.32			5.0								
trans-1,3-Dichloropropene	ND	0.48			5.0								
1,1,2-Trichloroethane	ND	0.38			5.0								
1,3-Dichloropropane	ND	0.22			5.0								
Tetrachloroethene	ND	0.65			5.0								
2-Hexanone	ND	1.7			5.0								
Dibromochloromethane	ND	0.57			5.0								

**Qualifiers:** ND - Not Detected at the MDL

J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits

R - RPD outside accepted recovery limits

MDL - Method Detection Limit

RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**  
**SW846 8260C -- VOC by GC-MS**

Sample ID:	MB-65160	SampType:	MBLK	TestCode:	SW8260_W	Units:	ug/L	SPK value	SPK Ref Val	%REC	SPK LowLimit	SPK HighLimit	RPD Ref Val	%RPD	RPD Limit	Qual
Client ID:	MB-65160	Batch ID:	65160	Result	MDL	RL										
1,2-Dibromoethane	ND	0.50		5.0												
Chlorobenzene	ND	0.26		5.0												
1,1,1,2-Tetrachloroethane	ND	0.41		5.0												
Ethylbenzene	ND	0.35		5.0												
m,p-Xylene	ND	0.77		5.0												
o-Xylene	ND	0.36		5.0												
Xyrene (Total)	ND	0.36		5.0												
Styrene	ND	0.50		5.0												
Bromoform	ND	0.77		5.0												
Isopropylbenzene	ND	0.38		5.0												
1,1,2,2-Tetrachloroethane	ND	0.42		5.0												
Bromobenzene	ND	0.36		5.0												
1,2,3-Trichloropropane	ND	0.82		5.0												
n-Propylbenzene	ND	0.64		5.0												
2-Chlorotoluene	ND	0.54		5.0												
1,3,5-Trimethylbenzene	ND	0.45		5.0												
4-Chlorotoluene	ND	0.45		5.0												
tert-Butylbenzene	ND	0.37		5.0												
1,2,4-Trimethylbenzene	ND	0.40		5.0												
sec-Butylbenzene	ND	0.28		5.0												
4-Isopropyltoluene	ND	0.46		5.0												
1,3-Dichlorobenzene	ND	0.29		5.0												
1,4-Dichlorobenzene	ND	0.40		5.0												
n-Butylbenzene	ND	0.33		5.0												
1,2-Dichlorobenzene	ND	0.33		5.0												
1,2-Dibromo-3-chloropropane	ND	0.75		5.0												
1,2,4-Trichlorobenzene	ND	0.26		5.0												
Hexachlorobutadiene	ND	0.41		5.0												
1,2,3-Trichlorobenzene	ND	0.33		5.0												
Naphthalene	ND	0.80		5.0												
1,1,2-Trichloro-1,2,2-trifluoroethane	ND	0.82		5.0												
Cyclohexane	ND	0.71		5.0												
Methyl acetate	ND	0.29		5.0												
Methylcyclohexane	ND	0.76		5.0												
Surrogate:	54.08			50.00												
Dibromofluoromethane				5.0												

Qualifiers:

ND - Not Detected at the MDL

B - Analyte detected in the associated Method Blank

J - Analyte detected below quantitation limits

MDL - Method Detection Limit

R - RPD outside accepted recovery limits

RL - Reporting Limit

mm11.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**  
**SW846 8260C -- VOC by GC-MS**

Sample ID: <b>MB-65160</b>		SampType: <b>MBLK</b>	TestCode: <b>SW8260_W</b>			Prep Date: <b>03/16/12 16:38</b>	Run ID: <b>V1_120316B</b>				
Client ID: <b>MB-65160</b>		Batch ID: <b>65160</b>	Units: <b>ug/L</b>			Analysis Date: <b>03/16/12 19:13</b>	SeqNo: <b>1708143</b>				
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPD Limit	Qual
Surrogate: 1,2-Dichloroethane-d4	50.59	5.0	50.00	0	101	70	120	0			
Surrogate: Toluene-d8	49.98	5.0	50.00	0	100	85	120	0			
Surrogate: Bromofluorobenzene	47.49	5.0	50.00	0	95.0	75	120	0			
Sample ID: <b>MB-65195</b>		SampType: <b>MBLK</b>	TestCode: <b>SW8260_W</b>			Prep Date: <b>03/20/12 9:05</b>	Run ID: <b>V5_120320B</b>				
Client ID: <b>MB-65195</b>		Batch ID: <b>65195</b>	Units: <b>ug/L</b>			Analysis Date: <b>03/20/12 15:38</b>	SeqNo: <b>1708673</b>				
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPD Limit	Qual
Vinyl chloride	ND	0.50	1.0								
1,1-Dichloroethene	ND	0.39	1.0								
2-Butanone	ND	2.1	5.0								
Chloroform	ND	0.33	1.0								
Carbon tetrachloride	ND	0.54	1.0								
1,2-Dichloroethane	ND	0.41	1.0								
Benzene	ND	0.33	1.0								
Trichloroethene	ND	0.36	1.0								
Tetrachloroethene	ND	0.65	1.0								
Chlorobenzene	ND	0.26	1.0								
Surrogate: Dibromofluoromethane	55.87	5.0	50.00	0	112	85	115	0			
Surrogate: 1,2-Dichloroethane-d4	51.00	5.0	50.00	0	102	70	120	0			
Surrogate: Toluene-d8	44.81	5.0	50.00	0	89.6	85	120	0			
Surrogate: Bromofluorobenzene	51.73	5.0	50.00	0	103	75	120	0			

Qualifiers: ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mm11.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**  
**SW846 8260C -- VOC by GC-MS**

Sample ID:	MB-65204	SampType:	MBLK	TestCode:	SW8260_W	Prep Date:	03/21/12 7:34	Run ID:	V5_120321A				
Client ID:	MB-65204	Batch ID:	65204	Units:	ug/L	Analysis Date:	03/21/12 9:38	SeqNo:	1708818				
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Dichlorodifluoromethane	ND	0.66		1.0									
Chloromethane	ND	0.26		1.0									
Vinyl chloride	ND	0.50		1.0									
Bromomethane	ND	0.80		1.0									
Chloroethane	ND	0.48		1.0									
Trichlorofluoromethane	ND	0.54		1.0									
1,1-Dichloroethene	ND	0.39		1.0									
Acetone	ND	2.2		5.0									
Iodomethane	ND	0.63		1.0									
Carbon disulfide	ND	0.34		1.0									
Methylene chloride	ND	0.41		1.0									
trans-1,2-Dichloroethene	ND	0.65		1.0									
Methyl tert-butyl ether	ND	0.24		1.0									
1,1-Dichloroethane	ND	0.25		1.0									
Vinyl acetate	ND	0.35		1.0									
2-Butanone	ND	2.1		5.0									
cis-1,2-Dichloroethene	ND	0.48		1.0									
2,2-Dichloropropane	ND	0.30		1.0									
Bromochloromethane	ND	0.43		1.0									
Chloroform	ND	0.33		1.0									
1,1,1-Trichloroethane	ND	0.50		1.0									
1,1-Dichloropropene	ND	0.50		1.0									
Carbon tetrachloride	ND	0.54		1.0									
1,2-Dichloroethane	ND	0.41		1.0									
Benzene	ND	0.33		1.0									
Trichloroethene	ND	0.36		1.0									
1,2-Dichloropropane	ND	0.61		1.0									
Dibromomethane	ND	0.49		1.0									
Bromodichloromethane	ND	0.26		1.0									
cis-1,3-Dichloropropene	ND	0.45		1.0									
4-Methyl-2-pentanone	ND	0.82		5.0									
Toluene	ND	0.32		1.0									
trans-1,3-Dichloropropene	ND	0.48		1.0									
1,1,2-Trichloroethane	ND	0.38		1.0									
1,3-Dichloropropane	ND	0.22		1.0									
Tetrachloroethene	ND	0.65		1.0									
2-Hexanone	ND	1.7		5.0									
Dibromochloromethane	ND	0.57		1.0									

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits

**S - Recovery outside accepted recovery limits**  
**R - RPD outside accepted recovery limits**

**MDL - Method Detection Limit**  
**RL - Reporting Limit**

**B - Analyte detected in the associated Method Blank**

mm11.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**  
**SW846 8260C -- VOC by GC-MS**

Sample ID:	MB-65204	SampType:	MBLK	TestCode:	SW8260_W	Units:	ug/L	SPK value	SPK Ref Val	%RPD Ref Val	%RPD RPD Limit	Qual
Client ID:	MB-65204	Batch ID:	65204	MDL	RL							
Analyte		Result										
1,2-Dibromoethane	ND	0.50										
Chlorobenzene	ND	0.26										
1,1,1,2-Tetrachloroethane	ND	0.41										
Ethylbenzene	ND	0.35										
m,p-Xylene	ND	0.77										
o-Xylene	ND	0.36										
Xyrene (Total)	ND	0.36										
Styrene	ND	0.50										
Bromoform	ND	0.77										
Isopropylbenzene	ND	0.38										
1,1,2,2-Tetrachloroethane	ND	0.42										
Bromobenzene	ND	0.36										
1,2,3-Trichloropropane	ND	0.82										
n-Propylbenzene	ND	0.64										
2-Chlorotoluene	ND	0.54										
1,3,5-Trimethylbenzene	ND	0.45										
4-Chlorotoluene	ND	0.45										
tert-Butylbenzene	ND	0.37										
1,2,4-Trimethylbenzene	ND	0.40										
sec-Butylbenzene	ND	0.28										
4-Isopropyltoluene	ND	0.46										
1,3-Dichlorobenzene	ND	0.29										
1,4-Dichlorobenzene	ND	0.40										
n-Butylbenzene	ND	0.33										
1,2-Dichlorobenzene	ND	0.33										
1,2-Dibromo-3-chloropropane	ND	0.75										
1,2,4-Trichlorobenzene	ND	0.26										
Hexachlorobutadiene	ND	0.41										
1,2,3-Trichlorobenzene	ND	0.33										
Naphthalene	ND	0.80										
1,1,2-Trichloro-1,2,2-trifluoroethane	ND	0.82										
Cyclohexane	ND	0.71										
Methyl acetate	ND	0.29										
Methylcyclohexane	ND	0.76										
Surrogate:	57.65											
Dibromofluoromethane												S
<b>Qualifiers:</b>	ND - Not Detected at the MDL											
mm11.12.11.A	J - Analyte detected below quantitation limits											
<b>MDL - Method Detection Limit</b>	S - Recovery outside accepted recovery limits											
<b>RL - Reporting Limit</b>	R - RPD outside accepted recovery limits											

B - Analyte detected in the associated Method Blank

MDL - Method Detection Limit

R - Recovery outside accepted recovery limits

RL - Reporting Limit

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**

**SW846 8260C -- VOC by GC-MS**

Sample ID: MB-65204		SampType: MBLK	TestCode: SW8260_W	Prep Date: 03/21/12 7:34	Run ID: V5_120321A							
Client ID: MB-65204		Batch ID: 65204	Units: ug/L	Analysis Date: 03/21/12 9:38	SeqNo: 1708818							
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	SPK LowLimit	SPK HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Surrogate: 1,2-Dichloroethane-d4	48.29	5.0	50.00	0	96.6	70	120	120	0	0		
Surrogate: Toluene-d8	42.83	5.0	50.00	0	85.7	85	120	120	0	0		
Surrogate: Bromofluorobenzene	52.34	5.0	50.00	0	105	75	120	120	0	0		

Qualifiers:  
ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mm11.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**

**SW846 8260C -- VOC by GC-MS**

Sample ID: LCS-65160	SampType: LCS	TestCode: SW8260_W	Prep Date: 03/16/12 16:38	Analysis Date: 03/16/12 16:57	Run ID: V1_120316B
Client ID: LCS-65160	Batch ID: 65160	Units: ug/L	SPK Ref Val	%REC	RPD Ref Val
Analyte	Result	MDL	SPK value	LowLimit	HighLimit
Dichlorodifluoromethane	52.36	0.66	5.0	50.00	0
Chloromethane	47.90	0.26	5.0	50.00	0
Vinyl chloride	52.21	0.50	5.0	50.00	0
Bromomethane	60.91	0.80	5.0	50.00	0
Chloroethane	57.49	0.48	5.0	50.00	0
Trichlorofluoromethane	53.64	0.54	5.0	50.00	0
1,1-Dichloroethene	45.21	0.39	5.0	50.00	0
Acetone	50.96	2.2	5.0	50.00	0
Iodomethane	65.65	0.63	5.0	50.00	0
Carbon disulfide	54.12	0.34	5.0	50.00	0
Methylene chloride	48.22	0.41	5.0	50.00	0
trans-1,2-Dichloroethene	50.69	0.65	5.0	50.00	0
Methyl tert-butyl ether	59.94	0.24	5.0	50.00	0
1,1-Dichloroethane	58.23	0.25	5.0	50.00	0
Vinyl acetate	55.80	0.35	5.0	50.00	0
2-Butanone	57.23	2.1	5.0	50.00	0
cis-1,2-Dichloroethene	51.01	0.48	5.0	50.00	0
2,2-Dichloropropane	76.09	0.30	5.0	50.00	0
Bromochloromethane	63.36	0.43	5.0	50.00	0
Chloroform	53.76	0.33	5.0	50.00	0
1,1,1-Trichloroethane	71.98	0.50	5.0	50.00	0
1,1-Dichloropropene	53.95	0.50	5.0	50.00	0
Carbon tetrachloride	77.74	0.54	5.0	50.00	0
1,2-Dichloroethane	52.68	0.41	5.0	50.00	0
Benzene	52.88	0.33	5.0	50.00	0
Trichloroethene	51.59	0.36	5.0	50.00	0
1,2-Dichloropropane	52.01	0.61	5.0	50.00	0
Dibromomethane	55.18	0.49	5.0	50.00	0
Bromodichloromethane	53.67	0.26	5.0	50.00	0
cis-1,3-Dichloropropene	58.19	0.45	5.0	50.00	0
4-Methyl-2-pentanone	55.54	0.82	5.0	50.00	0
Toluene	52.09	0.32	5.0	50.00	0
trans-1,3-Dichloropropene	61.75	0.48	5.0	50.00	0
1,1,2-Trichloroethane	51.63	0.38	5.0	50.00	0
1,3-Dichloropropane	54.49	0.22	5.0	50.00	0
Tetrachloroethene	49.55	0.65	5.0	50.00	0
2-Hexanone	54.61	1.7	5.0	50.00	0
Dibromochloromethane	56.25	0.57	5.0	50.00	0

Qualifiers: ND - Not Detected at the MDL

J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits

R - RPD outside accepted recovery limits

MDL - Method Detection Limit

RL - Reporting Limit

B - Analyte detected in the associated Method Blank

mm11.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**

**SW846 8260C -- VOC by GC-MS**

Sample ID:	LCS-65160	SampType:	LCS	TestCode:	SW8260_W	Prep Date:	03/16/12 16:38	Run ID:	V1_120316B				
Client ID:	LCS-65160	Batch ID:	65160	Units:	ug/L	Analysis Date:	03/16/12 16:57	SeqNo:	1708142				
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
1,2-Dibromoethane	56.14	0.50	5.0	50.00	0	112	80	120	0	0	120	0	
Chlorobenzene	51.00	0.26	5.0	50.00	0	102	80	120	0	0	120	0	
1,1,1,2-Tetrachloroethane	52.88	0.41	5.0	50.00	0	106	80	130	0	0	130	0	
Ethylbenzene	53.85	0.35	5.0	50.00	0	108	75	125	0	0	125	0	
m,p-Xylene	106.6	0.77	5.0	100.0	0	107	75	130	0	0	130	0	
o-Xylene	53.53	0.36	5.0	50.00	0	107	80	120	0	0	120	0	
Xylene (Total)	160.1	0.36	5.0	150.0	0	107	81	121	0	0	121	0	
Styrene	55.20	0.50	5.0	50.00	0	110	65	135	0	0	135	0	
Bromoform	62.52	0.77	5.0	50.00	0	125	70	130	0	0	130	0	
Isopropylbenzene	52.50	0.38	5.0	50.00	0	105	75	125	0	0	125	0	
1,1,2,2-Tetrachloroethane	59.05	0.42	5.0	50.00	0	118	65	130	0	0	130	0	
Bromobenzene	54.68	0.36	5.0	50.00	0	109	75	125	0	0	125	0	
1,2,3-Trichloropropane	58.30	0.82	5.0	50.00	0	117	75	125	0	0	125	0	
n-Propylbenzene	60.20	0.64	5.0	50.00	0	120	70	130	0	0	130	0	
2-Chlorotoluene	54.60	0.54	5.0	50.00	0	109	75	125	0	0	125	0	
1,3,5-Trimethylbenzene	55.41	0.45	5.0	50.00	0	111	75	130	0	0	130	0	
4-Chlorotoluene	55.87	0.45	5.0	50.00	0	112	75	130	0	0	130	0	
tert-Butylbenzene	54.06	0.37	5.0	50.00	0	108	70	130	0	0	130	0	
1,2,4-Trimethylbenzene	57.12	0.40	5.0	50.00	0	114	75	130	0	0	130	0	
sec-Butylbenzene	55.49	0.28	5.0	50.00	0	111	70	125	0	0	125	0	
4-Isopropyltoluene	55.01	0.46	5.0	50.00	0	110	75	130	0	0	130	0	
1,3-Dichlorobenzene	54.13	0.29	5.0	50.00	0	108	75	125	0	0	125	0	
1,4-Dichlorobenzene	54.66	0.40	5.0	50.00	0	109	75	125	0	0	125	0	
n-Butylbenzene	54.46	0.33	5.0	50.00	0	109	70	135	0	0	135	0	
1,2-Dichlorobenzene	52.05	0.33	5.0	50.00	0	104	70	120	0	0	120	0	
1,2-Dibromo-3-chloropropane	51.24	0.75	5.0	50.00	0	102	50	130	0	0	130	0	
1,2,4-Trichlorobenzene	47.81	0.26	5.0	50.00	0	95.6	65	135	0	0	135	0	
Hexachlorobutadiene	45.02	0.41	5.0	50.00	0	90.0	50	140	0	0	140	0	
1,2,3-Trichlorobenzene	44.86	0.33	5.0	50.00	0	89.7	55	140	0	0	140	0	
Naphthalene	36.62	0.80	5.0	50.00	0	73.2	55	140	0	0	140	0	
1,1,2-Trichloro-1,2,2-trifluoroethane	53.59	0.82	5.0	50.00	0	107	70	130	0	0	130	0	
Cyclohexane	58.38	0.71	5.0	50.00	0	117	70	130	0	0	130	0	
Methyl acetate	53.76	0.29	5.0	50.00	0	108	70	130	0	0	130	0	
Methylcyclohexane	55.42	0.76	5.0	50.00	0	111	70	130	0	0	130	0	
Surrogate:	52.82	5.0	50.00	50.00	0	106	85	115	0	0	115	0	
Dibromofluoromethane													

**Qualifiers:** ND - Not Detected at the MDL  
 J - Analyte detected below quantitation limits  
 MM.11.2.11.A

**S - Recovery outside accepted recovery limits**  
**R - RPD outside accepted recovery limits**

**MDL - Method Detection Limit**  
**RL - Reporting Limit**

**B - Analyte detected in the associated Method Blank**

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**  
**SW846 8260C -- VOC by GC-MS**

Sample ID: LCS-65160		SampType: LCS	TestCode: SW8260_W			Prep Date: 03/16/12 16:38			Run ID: V1_120316B		
Client ID: LCS-65160		Batch ID: 65160	Units: ug/L			Analysis Date: 03/16/12 16:57			SeqNo: 1708142		
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPD Limit	Qual
Surrogate: 1,2-Dichloroethane-d4	54.78	5.0	50.00	0	110	70	120	0			
Surrogate: Toluene-d8	50.39	5.0	50.00	0	101	85	120	0			
Surrogate: Bromofluorobenzene	50.13	5.0	50.00	0	100	75	120	0			
Sample ID: LCS-65195		SampType: LCS	TestCode: SW8260_W			Prep Date: 03/20/12 9:05			Run ID: V5_120320B		
Client ID: LCS-65195		Batch ID: 65195	Units: ug/L			Analysis Date: 03/20/12 14:18			SeqNo: 1708671		
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPD Limit	Qual
Vinyl chloride	47.18	0.50	5.0	50.00	0	94.4	50	145	0		
1,1-Dichloroethene	49.38	0.39	5.0	50.00	0	98.8	70	130	0		
2-Butanone	51.45	2.1	5.0	50.00	0	103	30	150	0		
Chloroform	56.61	0.33	5.0	50.00	0	113	65	135	0		
Carbon tetrachloride	61.51	0.54	5.0	50.00	0	123	65	140	0		
1,2-Dichloroethane	60.51	0.41	5.0	50.00	0	121	70	130	0		
Benzene	54.05	0.33	5.0	50.00	0	108	80	120	0		
Trichloroethene	59.50	0.36	5.0	50.00	0	119	70	125	0		
Tetrachloroethene	46.21	0.65	5.0	50.00	0	92.4	45	150	0		
Chlorobenzene	51.02	0.26	5.0	50.00	0	102	80	120	0		
Surrogate: Bromomethylfluoromethane	54.88	5.0	50.00	0	110	85	115	0			
Surrogate: 1,2-Dichloroethane-d4	52.10	5.0	50.00	0	104	70	120	0			
Surrogate: Toluene-d8	46.83	5.0	50.00	0	93.7	85	120	0			
Surrogate: Bromofluorobenzene	54.21	5.0	50.00	0	108	75	120	0			

Qualifiers: ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mm1.11.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**

**SW846 8260C -- VOC by GC-MS**

Sample ID:	LCS-65204	SampType:	LCS	TestCode:	SW8260_W	Prep Date:	03/21/12 7:34	Run ID:	V5_120321A			
Client ID:	LCS-65204	Batch ID:	65204	Units:	ug/L	Analysis Date:	03/21/12 8:44	SeqNo:	1708817			
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Dichlorodifluoromethane	54.38	0.66	5.0	50.00	0	109	30	155	0	0	0	
Chloromethane	44.56	0.26	5.0	50.00	0	89.1	40	125	0	0	0	
Vinyl chloride	40.41	0.50	5.0	50.00	0	80.8	50	145	0	0	0	
Bromomethane	42.48	0.80	5.0	50.00	0	85.0	30	145	0	0	0	
Chloroethane	39.96	0.48	5.0	50.00	0	79.9	60	135	0	0	0	
Trichlorofluoromethane	56.56	0.54	5.0	50.00	0	113	60	145	0	0	0	
1,1-Dichloroethene	42.44	0.39	5.0	50.00	0	84.9	70	130	0	0	0	
Acetone	48.47	2.2	5.0	50.00	0	96.9	40	140	0	0	0	
Iodomethane	51.33	0.63	5.0	50.00	0	103	72	121	0	0	0	
Carbon disulfide	45.23	0.34	5.0	50.00	0	90.5	35	160	0	0	0	
Methylene chloride	46.42	0.41	5.0	50.00	0	92.8	55	140	0	0	0	
trans-1,2-Dichloroethene	48.74	0.65	5.0	50.00	0	97.5	60	140	0	0	0	
Methyl tert-butyl ether	54.88	0.24	5.0	50.00	0	110	65	125	0	0	0	
1,1-Dichloroethane	50.46	0.25	5.0	50.00	0	101	70	135	0	0	0	
Vinyl acetate	59.01	0.35	5.0	50.00	0	118	38	163	0	0	0	
2-Butanone	53.31	2.1	5.0	50.00	0	107	30	150	0	0	0	
cis-1,2-Dichloroethene	49.76	0.48	5.0	50.00	0	99.5	70	125	0	0	0	
2,2-Dichloropropane	55.52	0.30	5.0	50.00	0	111	70	135	0	0	0	
Bromochloromethane	55.88	0.43	5.0	50.00	0	112	65	130	0	0	0	
Chloroform	52.48	0.33	5.0	50.00	0	105	65	135	0	0	0	
1,1,1-Trichloroethane	57.66	0.50	5.0	50.00	0	115	65	130	0	0	0	
1,1-Dichloropropene	46.92	0.50	5.0	50.00	0	93.8	75	130	0	0	0	
Carbon tetrachloride	56.86	0.54	5.0	50.00	0	114	65	140	0	0	0	
1,2-Dichloroethane	62.08	0.41	5.0	50.00	0	124	70	130	0	0	0	
Benzene	47.90	0.33	5.0	50.00	0	95.8	80	120	0	0	0	
Trichloroethene	52.18	0.36	5.0	50.00	0	104	70	125	0	0	0	
1,2-Dichloropropane	51.10	0.61	5.0	50.00	0	102	75	125	0	0	0	
Dibromomethane	55.37	0.49	5.0	50.00	0	111	75	125	0	0	0	
Bromodichloromethane	54.89	0.26	5.0	50.00	0	110	75	120	0	0	0	
cis-1,3-Dichloropropene	53.36	0.45	5.0	50.00	0	107	70	130	0	0	0	
1,1,2-Trichloroethane	62.72	0.82	5.0	50.00	0	125	60	135	0	0	0	
Toluene	49.64	0.32	5.0	50.00	0	99.3	75	120	0	0	0	
trans-1,3-Dichloropropene	60.76	0.48	5.0	50.00	0	122	55	140	0	0	0	
1,3-Dichloropropane	44.02	0.22	5.0	50.00	0	88.0	75	125	0	0	0	
Tetrachloroethene	39.27	0.65	5.0	50.00	0	78.5	45	150	0	0	0	
2-Hexanone	42.54	1.7	5.0	50.00	0	85.1	55	130	0	0	0	
Dibromochloromethane	48.36	0.57	5.0	50.00	0	96.7	60	135	0	0	0	

**Qualifiers:** ND - Not Detected at the MDL  
 J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
 R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
 RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**

**SW846 8260C -- VOC by GC-MS**

Sample ID:	LCS-65204	SampType:	LCS	TestCode:	SW8260_W	Prep Date:	03/21/12 7:34	Run ID:	V5_120321A				
Client ID:	LCS-65204	Batch ID:	65204	Units:	ug/L	Analysis Date:	03/21/12 8:44	SeqNo:	1708817				
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
1,2-Dibromoethane	45.89	0.50	5.0	50.00	0	91.8	80	120	0				
Chlorobenzene	42.51	0.26	5.0	50.00	0	85.0	80	120	0				
1,1,1,2-Tetrachloroethane	46.05	0.41	5.0	50.00	0	92.1	80	130	0				
Ethylbenzene	42.25	0.35	5.0	50.00	0	84.5	75	125	0				
m,p-Xylene	84.82	0.77	5.0	100.0	0	84.8	75	130	0				
o-Xylene	40.67	0.36	5.0	50.00	0	81.3	80	120	0				
Xylene (Total)	125.5	0.36	5.0	150.0	0	83.7	81	121	0				
Styrene	44.45	0.50	5.0	50.00	0	88.9	65	135	0				
Bromoform	49.60	0.77	5.0	50.00	0	99.2	70	130	0				
Isopropylbenzene	43.64	0.38	5.0	50.00	0	87.3	75	125	0				
1,1,2,2-Tetrachloroethane	40.05	0.42	5.0	50.00	0	80.1	65	130	0				
Bromobenzene	40.42	0.36	5.0	50.00	0	80.8	75	125	0				
1,2,3-Trichloropropane	42.58	0.82	5.0	50.00	0	85.2	75	125	0				
n-Propylbenzene	41.61	0.64	5.0	50.00	0	83.2	70	130	0				
2-Chlorotoluene	41.46	0.54	5.0	50.00	0	82.9	75	125	0				
1,3,5-Trimethylbenzene	43.99	0.45	5.0	50.00	0	88.0	75	130	0				
4-Chlorotoluene	41.52	0.45	5.0	50.00	0	83.0	75	130	0				
tert-Butylbenzene	44.13	0.37	5.0	50.00	0	88.3	70	130	0				
1,2,4-Trimethylbenzene	43.25	0.40	5.0	50.00	0	86.5	75	130	0				
sec-Butylbenzene	43.40	0.28	5.0	50.00	0	86.8	70	125	0				
4-Isopropyltoluene	47.71	0.46	5.0	50.00	0	95.4	75	130	0				
1,3-Dichlorobenzene	44.56	0.29	5.0	50.00	0	89.1	75	125	0				
1,4-Dichlorobenzene	41.17	0.40	5.0	50.00	0	82.3	75	125	0				
n-Butylbenzene	45.77	0.33	5.0	50.00	0	91.5	70	135	0				
1,2-Dichlorobenzene	44.18	0.33	5.0	50.00	0	88.4	70	120	0				
1,2-Dibromo-3-chloropropane	50.36	0.75	5.0	50.00	0	101	50	130	0				
1,2,4-Trichlorobenzene	47.37	0.26	5.0	50.00	0	94.7	65	135	0				
Hexachlorobutadiene	52.84	0.41	5.0	50.00	0	106	50	140	0				
1,2,3-Trichlorobenzene	46.55	0.33	5.0	50.00	0	93.1	55	140	0				
Naphthalene	47.59	0.80	5.0	50.00	0	95.2	55	140	0				
1,1,2-Trichloro-1,2,2-trifluoroethane	48.23	0.82	5.0	50.00	0	96.5	70	130	0				
Cyclohexane	49.76	0.71	5.0	50.00	0	99.5	70	130	0				
Methyl acetate	51.92	0.29	5.0	50.00	0	104	70	130	0				
Methylcyclohexane	53.08	0.76	5.0	50.00	0	106	70	130	0				
Surrogate:	57.10		5.0	50.00	0	114	85	115	0				
Dibromofluoromethane													

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

**B - Analyte detected in the associated Method Blank**

mm11.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8260\_W**  
**SW846 8260C -- VOC by GC-MS**

Sample ID: LCS-65204		SampType: LCS	TestCode: SW8260_W								
Client ID:	LCS-65204	Batch ID: 65204	Units: ug/L								
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
Surrogate: 1,2-Dichloroethane-d4	52.15	5.0	50.00	0	104	70	120	0			
Surrogate: Toluene-d8	43.97	5.0	50.00	0	87.9	85	120	0			
Surrogate: Bromofluorobenzene	54.26	5.0	50.00	0	109	75	120	0			
Sample ID: LCSD-65195		SampType: LCSD	TestCode: SW8260_W								
Client ID:	LCSD-65195	Batch ID: 65195	Units: ug/L								
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
Vinyl chloride	45.43	0.50	5.0	50.00	0	90.9	50	145	47.18	3.78	40
1,1-Dichloroethene	48.71	0.39	5.0	50.00	0	97.4	70	130	49.38	1.37	40
2-Butanone	53.12	2.1	5.0	50.00	0	106	30	150	51.45	3.2	40
Chloroform	55.07	0.33	5.0	50.00	0	110	65	135	56.61	2.75	40
Carbon tetrachloride	62.40	0.54	5.0	50.00	0	125	65	140	61.51	1.44	40
1,2-Dichloroethane	57.21	0.41	5.0	50.00	0	114	70	130	60.51	5.6	40
Benzene	52.26	0.33	5.0	50.00	0	105	80	120	54.05	3.37	40
Trichloroethene	56.83	0.36	5.0	50.00	0	114	70	125	59.50	4.6	40
Tetrachloroethene	45.26	0.65	5.0	50.00	0	90.5	45	150	46.21	2.08	40
Chlorobenzene	48.02	0.26	5.0	50.00	0	96.0	80	120	51.02	6.04	40
Surrogate: Bromomethylfluoromethane	55.14	5.0	50.00	0	110	85	115	0	0	40	
Surrogate: 1,2-Dichloroethane-d4	47.60	5.0	50.00	0	95.2	70	120	0	0	40	
Surrogate: Toluene-d8	47.19	5.0	50.00	0	94.4	85	120	0	0	40	
Surrogate: Bromofluorobenzene	53.29	5.0	50.00	0	107	75	120	0	0	40	

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mm11.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/22/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result	Qual	RL	Units	DF	Date Analyzed	Batch ID
<b>SW846 8270D -- SVOA by GC-MS</b>							<b>SW8270_W</b>
Phenol	9.8	J	10	ug/L	1	03/13/2012 16:22	65051
Bis(2-chloroethyl)ether	ND		10	ug/L	1	03/13/2012 16:22	65051
2-Chlorophenol	ND		10	ug/L	1	03/13/2012 16:22	65051
2-Methylphenol	ND		10	ug/L	1	03/13/2012 16:22	65051
2,2'-oxybis(1-Chloropropane)	ND		10	ug/L	1	03/13/2012 16:22	65051
4-Methylphenol	ND		10	ug/L	1	03/13/2012 16:22	65051
N-Nitroso-di-n-propylamine	ND		10	ug/L	1	03/13/2012 16:22	65051
Hexachloroethane	ND		10	ug/L	1	03/13/2012 16:22	65051
Nitrobenzene	ND		10	ug/L	1	03/13/2012 16:22	65051
Isophorone	20		10	ug/L	1	03/13/2012 16:22	65051
2-Nitrophenol	ND		10	ug/L	1	03/13/2012 16:22	65051
2,4-Dimethylphenol	ND		10	ug/L	1	03/13/2012 16:22	65051
2,4-Dichlorophenol	ND		10	ug/L	1	03/13/2012 16:22	65051
Naphthalene	2.4	J	10	ug/L	1	03/13/2012 16:22	65051
4-Chloroaniline	ND		10	ug/L	1	03/13/2012 16:22	65051
Bis(2-chloroethoxy)methane	ND		10	ug/L	1	03/13/2012 16:22	65051
Hexachlorobutadiene	ND		10	ug/L	1	03/13/2012 16:22	65051
4-Chloro-3-methylphenol	ND		10	ug/L	1	03/13/2012 16:22	65051
2-Methylnaphthalene	ND		10	ug/L	1	03/13/2012 16:22	65051
Hexachlorocyclopentadiene	ND		10	ug/L	1	03/13/2012 16:22	65051
2,4,6-Trichlorophenol	ND		10	ug/L	1	03/13/2012 16:22	65051
2,4,5-Trichlorophenol	ND		20	ug/L	1	03/13/2012 16:22	65051
2-Chloronaphthalene	ND		10	ug/L	1	03/13/2012 16:22	65051
2-Nitroaniline	ND		20	ug/L	1	03/13/2012 16:22	65051
Dimethylphthalate	7.3	J	10	ug/L	1	03/13/2012 16:22	65051
Acenaphthylene	ND		10	ug/L	1	03/13/2012 16:22	65051
2,6-Dinitrotoluene	ND		10	ug/L	1	03/13/2012 16:22	65051
3-Nitroaniline	ND		20	ug/L	1	03/13/2012 16:22	65051
Acenaphthene	ND		10	ug/L	1	03/13/2012 16:22	65051
2,4-Dinitrophenol	ND		20	ug/L	1	03/13/2012 16:22	65051
4-Nitrophenol	ND		20	ug/L	1	03/13/2012 16:22	65051
Dibenzofuran	ND		10	ug/L	1	03/13/2012 16:22	65051
2,4-Dinitrotoluene	ND		10	ug/L	1	03/13/2012 16:22	65051
Diethylphthalate	1.4	J	10	ug/L	1	03/13/2012 16:22	65051
4-Chlorophenyl-phenylether	ND		10	ug/L	1	03/13/2012 16:22	65051
Fluorene	ND		10	ug/L	1	03/13/2012 16:22	65051
4-Nitroaniline	ND		20	ug/L	1	03/13/2012 16:22	65051
4,6-Dinitro-2-methylphenol	ND		20	ug/L	1	03/13/2012 16:22	65051
N-Nitrosodiphenylamine	ND		10	ug/L	1	03/13/2012 16:22	65051

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/22/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result	Qual	RL	Units	DF	Date Analyzed	Batch ID
<b>SW846 8270D -- SVOA by GC-MS</b>							<b>SW8270_W</b>
4-Bromophenyl-phenylether	ND		10	ug/L		1 03/13/2012 16:22	65051
Hexachlorobenzene	ND		10	ug/L		1 03/13/2012 16:22	65051
Pentachlorophenol	ND		20	ug/L		1 03/13/2012 16:22	65051
Phenanthrone	ND		10	ug/L		1 03/13/2012 16:22	65051
Anthracene	ND		10	ug/L		1 03/13/2012 16:22	65051
Carbazole	ND		10	ug/L		1 03/13/2012 16:22	65051
Di-n-butylphthalate	ND		10	ug/L		1 03/13/2012 16:22	65051
Fluoranthene	ND		10	ug/L		1 03/13/2012 16:22	65051
Pyrene	ND		10	ug/L		1 03/13/2012 16:22	65051
Butylbenzylphthalate	ND		10	ug/L		1 03/13/2012 16:22	65051
3,3'-Dichlorobenzidine	ND		10	ug/L		1 03/13/2012 16:22	65051
Benzo(a)anthracene	ND		10	ug/L		1 03/13/2012 16:22	65051
Chrysene	ND		10	ug/L		1 03/13/2012 16:22	65051
Bis(2-ethylhexyl)phthalate	3.0	J	10	ug/L		1 03/13/2012 16:22	65051
Di-n-octylphthalate	ND		10	ug/L		1 03/13/2012 16:22	65051
Benzo(b)fluoranthene	ND		10	ug/L		1 03/13/2012 16:22	65051
Benzo(k)fluoranthene	ND		10	ug/L		1 03/13/2012 16:22	65051
Benzo(a)pyrene	ND		10	ug/L		1 03/13/2012 16:22	65051
Indeno(1,2,3-cd)pyrene	ND		10	ug/L		1 03/13/2012 16:22	65051
Dibeno(a,h)anthracene	ND		10	ug/L		1 03/13/2012 16:22	65051
Benzo(g,h,i)perylene	ND		10	ug/L		1 03/13/2012 16:22	65051
1,1'-Biphenyl	ND		10	ug/L		1 03/13/2012 16:22	65051
Acetophenone	ND		10	ug/L		1 03/13/2012 16:22	65051
Atrazine	ND		10	ug/L		1 03/13/2012 16:22	65051
Benzaldehyde	ND		10	ug/L		1 03/13/2012 16:22	65051
Caprolactam	ND		10	ug/L		1 03/13/2012 16:22	65051
Surrogate: Nitrobenzene-d5	80.2		40-110	%REC		1 03/13/2012 16:22	65051
Surrogate: 2-Fluorobiphenyl	82.5		50-110	%REC		1 03/13/2012 16:22	65051
Surrogate: Terphenyl-d14	58.5		50-135	%REC		1 03/13/2012 16:22	65051
Surrogate: Phenol-d5	14.3		10-115	%REC		1 03/13/2012 16:22	65051
Surrogate: 2-Fluorophenol	28.0		20-110	%REC		1 03/13/2012 16:22	65051
Surrogate: 2,4,6-Tribromophenol	90.4		40-125	%REC		1 03/13/2012 16:22	65051

**Qualifiers:** ND - Not Detected at the Reporting Limit

J - Analyte detected below quantitation limits

B - Analyte detected in the associated Method Blank

DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits

R - RPD outside accepted recovery limits

E - Value above quantitation range

RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/22/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-SOLID  
**Lab ID:** L0466-02

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:00

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 8270D -- SVOA by GC-MS</b>						<b>SW8270_W</b>
1,4-Dichlorobenzene -- TCLP	ND		33 ug/L		1 03/16/2012 17:36	65098
2-Methylphenol -- TCLP	ND		33 ug/L		1 03/16/2012 17:36	65098
4-Methylphenol -- TCLP	ND		33 ug/L		1 03/16/2012 17:36	65098
Hexachloroethane -- TCLP	ND		33 ug/L		1 03/16/2012 17:36	65098
Nitrobenzene -- TCLP	ND		33 ug/L		1 03/16/2012 17:36	65098
Hexachlorobutadiene -- TCLP	ND		33 ug/L		1 03/16/2012 17:36	65098
2,4,6-Trichlorophenol -- TCLP	ND		33 ug/L		1 03/16/2012 17:36	65098
2,4,5-Trichlorophenol -- TCLP	ND		67 ug/L		1 03/16/2012 17:36	65098
2,4-Dinitrotoluene -- TCLP	ND		33 ug/L		1 03/16/2012 17:36	65098
Hexachlorobenzene -- TCLP	ND		33 ug/L		1 03/16/2012 17:36	65098
Pentachlorophenol -- TCLP	ND		67 ug/L		1 03/16/2012 17:36	65098
Pyridine -- TCLP	ND		67 ug/L		1 03/16/2012 17:36	65098
Surrogate: Nitrobenzene-d5 -- TCLP	71.8		40-110 %REC		1 03/16/2012 17:36	65098
Surrogate: 2-Fluorobiphenyl -- TCLP	76.2		50-110 %REC		1 03/16/2012 17:36	65098
Surrogate: Terphenyl-d14 -- TCLP	77.2		50-135 %REC		1 03/16/2012 17:36	65098
Surrogate: Phenol-d5 -- TCLP	61.8		10-115 %REC		1 03/16/2012 17:36	65098
Surrogate: 2-Fluorophenol -- TCLP	69.3		20-110 %REC		1 03/16/2012 17:36	65098
Surrogate: 2,4,6-Tribromophenol -- TCLP	93.2		40-125 %REC		1 03/16/2012 17:36	65098

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Tech

Date: 03/22/2012

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8270\_W**

**SW846 8270D -- SVOA by GC-MS**

Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	Prep Date:	03/11/12 11:00	Run ID: S3_120313B
									Analysis Date:	03/13/12 11:59	
									%RPD	Ref Val	%RPD Limit
Phenol	ND	0.75	10								
Bis(2-chloroethyl)ether	ND	0.75	10								
2-Chlorophenol	ND	0.61	10								
2-Methylphenol	ND	0.96	10								
2,2'-oxybis(1-Chloropropane)	ND	0.78	10								
4-Methylphenol	ND	1.4	10								
N-Nitroso-di-n-propylamine	ND	0.63	10								
Hexachloroethane	ND	0.55	10								
Nitrobenzene	ND	1.6	10								
Isophorone	ND	0.47	10								
2-Nitrophenol	ND	0.60	10								
2,4-Dimethylphenol	ND	1.8	10								
2,4-Dichlorophenol	ND	0.57	10								
Naphthalene	ND	0.96	10								
4-Chloraniline	ND	2.0	10								
Bis(2-chloroethoxy)methane	ND	1.1	10								
Hexachlorobutadiene	ND	0.75	10								
4-Chloro-3-methylphenol	ND	0.60	10								
2-Methylnaphthalene	ND	0.94	10								
Hexachlorocyclopentadiene	ND	1.0	10								
2,4,6-Trichlorophenol	ND	0.53	10								
2,4,5-Trichlorophenol	ND	0.26	20								
2-Chloronaphthalene	ND	0.81	10								
2-Nitroaniline	ND	0.71	20								
Dimethylphthalate	ND	0.37	10								
Acenaphthylene	ND	0.42	10								
2,6-Dinitrotoluene	ND	0.52	10								
3-Nitroaniline	ND	0.97	20								
Acenaphthene	ND	0.65	10								
2,4-Dinitrophenol	ND	3.5	20								
4-Nitrophenol	ND	0.53	20								
Dibenzofuran	ND	0.52	10								
2,4-Dinitrotoluene	ND	0.41	10								
Diethylphthalate	ND	0.45	10								
4-Chlorophenyl-phenylether	ND	0.41	10								
Fluorene	ND	0.44	10								

**Qualifiers:** ND - Not Detected at the MDL

J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits

R - RPD outside accepted recovery limits

MDL - Method Detection Limit

RL - Reporting Limit

B - Analyte detected in the associated Method Blank

mmi.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8270\_W**  
**SW846 8270D -- SVOA by GC-MS**

Sample ID: <b>MB-65051</b>	SampType: <b>MBLK</b>	Batch ID: <b>65051</b>	TestCode: <b>SW8270_W</b>	Units: <b>ug/L</b>	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Analyte	Result	MDL	RL										
4-Nitroaniline	ND	0.96	20										
4,6-Dinitro-2-methylphenol	ND	0.79	20										
N-Nitrosodiphenylamine	ND	1.1	10										
4-Bromophenylphenylether	ND	0.54	10										
Hexachlorobenzene	ND	0.44	10										
Pentachlorophenol	ND	1.7	20										
Phenanthrene	ND	0.45	10										
Anthracene	ND	0.48	10										
Carbazole	ND	0.64	10										
Di-n-butylphthalate	ND	0.48	10										
Fluoranthene	ND	0.33	10										
Pyrene	ND	0.44	10										
Butylbenzylphthalate	ND	0.32	10										
3,3'-Dichlorobenzidine	ND	1.7	10										
Benzo(a)anthracene	ND	0.40	10										
Chrysene	ND	0.42	10										
Bis(2-ethylhexyl)phthalate	ND	1.3	10										
Di-n-octylphthalate	ND	0.47	10										
Benzo(b)fluoranthene	ND	0.94	10										
Benzo(k)fluoranthene	ND	1.2	10										
Benzo(a)pyrene	ND	1.2	10										
Indeno(1,2,3-cd)pyrene	ND	0.38	10										
Dibenz(a,h)anthracene	ND	0.44	10										
Benzo(g,h,i)perylene	ND	0.39	10										
1,1'-Biphenyl	ND	0.65	10										
Acetophenone	ND	0.51	10										
Atrazine	ND	1.3	10										
Benzaldehyde	ND	0.51	10										
Caproactam	ND	1.1	10										
Surrogate: Nitrobenzene-d5	44.51	10											
Surrogate: 2-Fluorobiphenyl	42.87	10											
Surrogate: Terphenyl-d14	45.37	10											
Surrogate: Phenol-d5	35.58	10											
Surrogate: 2-Fluorophenol	41.10	10											
Surrogate: 2,4,6-Tribromophenol	61.05	10											

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mml.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8270\_W**  
**SW846 8270D -- SVOA by GC-MS**

Sample ID:	MB-65091	SampType:	MBLK	Batch ID:	65098	TestCode:	SW8270_W	Units:	ug/L	Result	MDL	RL	SPK value	SPK Ref Val	%REC	SPK Ref Val	%RPD	RPDLimit	Qual
1,4-Dichlorobenzene -- TCLP		ND	3.7	33															
2-Methylphenol -- TCLP		ND	3.2	33															
4-Methylphenol -- TCLP		ND	4.7	33															
Hexachloroethane -- TCLP		ND	1.8	33															
Nitrobenzene -- TCLP		ND	5.3	33															
Hexachlorobutadiene -- TCLP		ND	2.5	33															
2,4,6-Trichlorophenol -- TCLP		ND	1.8	33															
2,4,5-Trichlorophenol -- TCLP		ND	0.87	67															
2,4-Dinitrotoluene -- TCLP		ND	1.4	33															
Hexachlorobenzene -- TCLP		ND	1.5	33															
Pentachlorophenol -- TCLP		ND	5.7	67															
Pyidine -- TCLP		ND	1.9	67															
Surrogate: Nitrobenzene-d5 -- TCLP		125.0	33	166.7															
Surrogate: 2-Fluorobiphenyl -- TCLP		131.0	33	166.7															
Surrogate: Terphenyl-d14 -- TCLP		133.9	33	166.7															
Surrogate: Phenol-d5 -- TCLP		131.5	33	166.7															
Surrogate: 2-Fluorophenol -- TCLP		130.3	33	166.7															
Surrogate: 2,4,6-Tribromophenol -- TCLP		163.9	33	166.7															

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

mm11.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8270\_W**

**SW846 8270D -- SVOA by GC-MS**

Analyte	Result	MDL	Units: ug/L	TestCode: SW8270_W	SPK value	SPK Ref Val	%REC			RPD Ref Val	%RPD	RPDLimit	Qual
							Analysis Date:	03/13/12 12:19	Prep Date:	03/11/12 11:00	Run ID:	S3_120313B	SeqNo:
Phenol	30.81	0.75	10	50.00	0	61.6	0	115	0				
Bis(2-chloroethyl)ether	37.05	0.75	10	50.00	0	74.1	35	110	0				
2-Chlorophenol	34.79	0.61	10	50.00	0	69.6	35	105	0				
2-Methylphenol	31.54	0.96	10	50.00	0	63.1	40	110	0				
2,2'-oxybis(1-Chloropropane)	39.15	0.78	10	50.00	0	78.3	30	123	0				
4-Methylphenol	30.83	1.4	10	50.00	0	61.7	30	110	0				
N-Nitroso-di-n-propylamine	29.74	0.63	10	50.00	0	59.5	35	130	0				
Hexachloroethane	44.75	0.55	10	50.00	0	89.5	30	95	0				
Nitrobenzene	42.81	1.6	10	50.00	0	85.6	45	110	0				
Isophorone	34.98	0.47	10	50.00	0	70.0	50	110	0				
2-Nitrophenol	40.94	0.60	10	50.00	0	81.9	40	115	0				
2,4-Dimethylphenol	39.03	1.8	10	50.00	0	78.1	30	110	0				
2,4-Dichlorophenol	38.42	0.57	10	50.00	0	76.8	50	105	0				
Naphthalene	40.41	0.96	10	50.00	0	80.8	40	100	0				
4-Chloroaniline	36.42	2.0	10	50.00	0	72.8	15	110	0				
Bis(2-chloroethoxy)methane	34.03	1.1	10	50.00	0	68.1	45	105	0				
Hexachlorobutadiene	48.77	0.75	10	50.00	0	97.5	25	105	0				
4-Chloro-3-methylphenol	36.63	0.60	10	50.00	0	73.3	45	110	0				
2-Methylnaphthalene	37.12	0.94	10	50.00	0	74.2	45	105	0				
Hexachlorocyclopentadiene	52.24	1.0	10	50.00	0	104	27	147	0				
2,4,6-Trichlorophenol	39.78	0.53	10	50.00	0	79.6	50	115	0				
2,4,5-Trichlorophenol	39.05	0.26	20	50.00	0	78.1	50	110	0				
2-Chloronaphthalene	43.50	0.81	10	50.00	0	87.0	50	105	0				
2-Nitroaniline	46.74	0.71	20	50.00	0	93.5	50	115	0				
Dimethylphthalate	42.46	0.37	10	50.00	0	84.9	25	125	0				
Acenaphthylene	41.78	0.42	10	50.00	0	83.6	50	105	0				
2,6-Dinitrotoluene	39.87	0.52	10	50.00	0	79.7	50	115	0				
3-Nitroaniline	36.40	0.97	20	50.00	0	72.8	20	125	0				
Acenaphthene	41.80	0.65	10	50.00	0	83.6	45	110	0				
2,4-Dinitrophenol	64.45	3.5	20	50.00	0	129	15	140	0				
4-Nitrophenol	56.37	0.53	20	50.00	0	113	0	125	0				
Dibenzofuran	41.63	0.52	10	50.00	0	83.3	55	105	0				
2,4-Dinitrotoluene	41.38	0.41	10	50.00	0	82.8	50	120	0				
Diethylphthalate	44.26	0.45	10	50.00	0	88.5	40	120	0				
4-Chlorophenyl-phenylether	40.93	0.41	10	50.00	0	81.9	50	110	0				
Fluorene	44.14	0.44	10	50.00	0	88.3	50	110	0				
4-Nitroaniline	34.84	0.96	20	50.00	0	69.7	35	120	0				
4,6-Dinitro-2-methylphenol	47.32	0.79	20	50.00	0	94.6	40	130	0				

**Qualifiers:** ND - Not Detected at the MDL  
 mm1.12.11.A J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
 R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
 RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8270\_W**

**SW846 8270D -- SVOA by GC-MS**

Analyte	Result	MDL	Units: ug/L	TestCode: SW8270_W	SPK value	SPK Ref Val	%REC			RPD Ref Val	%RPD	RPDLimit	Qual
							Analysis Date:	03/13/12 12:19	Prep Date:	03/11/12 11:00	Run ID:	S3_120313B	SeqNo:
N-Nitrosodiphenylamine	39.31	1.1	10	50.00	0	78.6	50	110	0				
4-Bromophenylphenylether	43.73	0.54	10	50.00	0	87.5	50	115	0				
Hexachlorobenzene	47.48	0.44	10	50.00	0	95.0	50	110	0				
Pentachlorophenol	41.16	1.7	20	50.00	0	82.3	40	115	0				
Phenanthrene	41.09	0.45	10	50.00	0	82.2	50	115	0				
Anthracene	41.06	0.48	10	50.00	0	82.1	55	110	0				
Carbazole	39.14	0.64	10	50.00	0	78.3	50	115	0				
Di-n-butylphthalate	42.65	0.48	10	50.00	0	85.3	55	115	0				
Fluoranthene	40.37	0.33	10	50.00	0	80.7	55	115	0				
Pyrene	37.73	0.44	10	50.00	0	75.5	50	130	0				
Butylbenzylphthalate	39.76	0.32	10	50.00	0	79.5	45	115	0				
3,3'-Dichlorobenzidine	25.28	1.7	10	50.00	0	50.6	20	110	0				
Benz(a)anthracene	40.49	0.40	10	50.00	0	81.0	55	110	0				
Chrysene	42.19	0.42	10	50.00	0	84.4	55	110	0				
Bis(2-ethylhexyl)phthalate	40.83	1.3	10	50.00	0	81.7	40	125	0				
Di-n-octylphthalate	38.95	0.47	10	50.00	0	77.9	35	135	0				
Benz(b)fluoranthene	43.01	0.94	10	50.00	0	86.0	45	120	0				
Benz(k)fluoranthene	42.82	1.2	10	50.00	0	85.6	45	125	0				
Benz(a)pyrene	41.48	1.2	10	50.00	0	83.0	55	110	0				
Indeno(1,2,3-cd)pyrene	47.26	0.38	10	50.00	0	94.5	45	125	0				
Dibenz(a,h)anthracene	47.54	0.44	10	50.00	0	95.1	40	125	0				
Benz(g,h,i)perylene	47.73	0.39	10	50.00	0	95.5	40	125	0				
1,1'-Biphenyl	43.65	0.65	10	50.00	0	87.3	55	108	0				
Acetophenone	40.51	0.51	10	50.00	0	81.0	56	145	0				
Atrazine	58.40	1.3	10	50.00	0	117	52	175	0				
Benzaldehyde	23.13	0.51	10	50.00	0	46.3	10	133	0				
Caprodiactam	27.43	1.1	10	50.00	0	54.9	10	146	0				
Surrogate: Nitrobenzene-d5	44.63	10	50.00	0	89.3	40	110	0					
Surrogate: 2-Fluorobiphenyl	42.09	10	50.00	0	84.2	50	110	0					
Surrogate: Terphenyl-d14	36.78	10	50.00	0	73.6	50	135	0					
Surrogate: Phenol-d5	34.05	10	50.00	0	68.1	10	115	0					
Surrogate: 2-Fluorophenol	39.28	10	50.00	0	78.6	20	110	0					
Surrogate: 2,4,6-Tribromophenol	56.13	10	50.00	0	112	40	125	0					

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mmi.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8270\_W**  
**SW846 8270D -- SVOA by GC-MS**

Analyte	Result	MDL	Units: ug/L	TestCode: SW8270_W		SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD	Ref Val	%RPD	RPDLimit	Qual
				Prep Date:	Analysis Date:										
1,4-Dichlorobenzene	130.6	3.7	33	166.7	0	78.4	30	100	0	0	0	0	0	0	0
2-Methylphenol	115.6	3.2	33	166.7	0	69.4	40	110	0	0	0	0	0	0	0
4-Methylphenol	123.5	4.7	33	166.7	0	74.1	30	110	0	0	0	0	0	0	0
Hexachloroethane	104.1	1.8	33	166.7	0	62.4	30	95	0	0	0	0	0	0	0
Nitrobenzene	126.5	5.3	33	166.7	0	75.9	45	110	0	0	0	0	0	0	0
Hexachlorobutadiene	140.2	2.5	33	166.7	0	84.1	25	105	0	0	0	0	0	0	0
2,4,6-Trichlorophenol	142.4	1.8	33	166.7	0	85.4	50	115	0	0	0	0	0	0	0
2,4,5-Trichlorophenol	139.7	0.87	67	166.7	0	83.8	50	110	0	0	0	0	0	0	0
2,4-Dinitrotoluene	135.3	1.4	33	166.7	0	81.2	50	120	0	0	0	0	0	0	0
Hexachlorobenzene	148.7	1.5	33	166.7	0	89.2	50	110	0	0	0	0	0	0	0
Pentachlorophenol	167.7	5.7	67	166.7	0	101	40	115	0	0	0	0	0	0	0
Pyridine	140.7	1.9	67	166.7	0	84.4	10	106	0	0	0	0	0	0	0
Surrogate: Nitrobenzene-d5	119.8	33	166.7	0	71.9	40	110	0	0	0	0	0	0	0	0
Surrogate: 2-Fluorobiphenyl	126.3	33	166.7	0	75.8	50	110	0	0	0	0	0	0	0	0
Surrogate: Terphenyl-d14	127.6	33	166.7	0	76.6	50	135	0	0	0	0	0	0	0	0
Surrogate: Phenol-d5	122.6	33	166.7	0	73.6	10	115	0	0	0	0	0	0	0	0
Surrogate: 2-Fluorophenol	118.7	33	166.7	0	71.2	20	110	0	0	0	0	0	0	0	0
Surrogate: 2,4,6-Tribromophenol	156.5	33	166.7	0	93.9	40	125	0	0	0	0	0	0	0	0

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

mm11.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8270\_W**

**SW846 8270D -- SVOA by GC-MS**

Sample ID:	LCSD-65051	SampType:	LCSD	TestCode:	SW8270_W	Units:	ug/L	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Client ID:	LCSD-65051	Batch ID:	65051													
Analyte		Result	MDL		RL											
Phenol	33.27	0.75	1.0	50.00	0	66.5	0	115	30.81	7.66	40					
Bis(2-chloroethyl)ether	35.90	0.75	1.0	50.00	0	71.8	35	110	37.05	3.16	40					
2-Chlorophenol	37.25	0.61	1.0	50.00	0	74.5	35	105	34.79	6.81	40					
2-Methylphenol	34.02	0.96	1.0	50.00	0	68.0	40	110	31.54	7.57	40					
2,2'-oxybis(1-Chloropropane)	41.19	0.78	1.0	50.00	0	82.4	30	123	39.15	5.09	40					
4-Methylphenol	32.85	1.4	1.0	50.00	0	65.7	30	110	30.83	6.35	40					
N-Nitroso-di-n-propylamine	31.85	0.63	1.0	50.00	0	63.7	35	130	29.74	6.85	40					
Hexachloroethane	48.41	0.55	1.0	50.00	0	96.8	30	95	44.75	7.85	40					
Nitrobenzene	43.84	1.6	1.0	50.00	0	87.7	45	110	42.81	2.38	40					
Isophorone	35.66	0.47	1.0	50.00	0	71.3	50	110	34.98	1.92	40					
2-Nitrophenol	43.32	0.60	1.0	50.00	0	86.6	40	115	40.94	5.64	40					
2,4-Dimethylphenol	42.68	1.8	1.0	50.00	0	85.4	30	110	39.03	8.94	40					
2,4-Dichlorophenol	40.40	0.57	1.0	50.00	0	80.8	50	105	38.42	5.03	40					
Naphthalene	41.68	0.96	1.0	50.00	0	83.4	40	100	40.41	3.09	40					
4-Chloroaniline	34.23	2.0	1.0	50.00	0	68.5	15	110	36.42	6.19	40					
Bis(2-chloroethoxy)methane	35.49	1.1	1.0	50.00	0	71.0	45	105	34.03	4.2	40					
Hexachlorobutadiene	50.10	0.75	1.0	50.00	0	100	25	105	48.77	2.69	40					
4-Chloro-3-methylphenol	36.99	0.60	1.0	50.00	0	74.0	45	110	36.63	0.982	40					
2-Methylnaphthalene	38.62	0.94	1.0	50.00	0	77.2	45	105	37.12	3.96	40					
Hexachlorocyclopentadiene	57.38	1.0	1.0	50.00	0	115	27	147	52.24	9.38	40					
2,4,6-Trichlorophenol	42.24	0.53	1.0	50.00	0	84.5	50	115	39.78	5.99	40					
2,4,5-Trichlorophenol	40.83	0.26	2.0	50.00	0	81.7	50	110	39.05	4.46	40					
2-Chloronaphthalene	43.91	0.81	1.0	50.00	0	87.8	50	105	43.50	0.916	40					
2-Nitroaniline	46.35	0.71	2.0	50.00	0	92.7	50	115	46.74	0.841	40					
Dimethylphthalate	41.86	0.37	1.0	50.00	0	83.7	25	125	42.46	1.44	40					
Acenaphthylene	40.90	0.42	1.0	50.00	0	81.8	50	105	41.78	2.12	40					
2,6-Dinitrotoluene	38.59	0.52	1.0	50.00	0	77.2	50	115	39.87	3.26	40					
3-Nitroaniline	35.27	0.97	2.0	50.00	0	70.5	20	125	36.40	3.17	40					
Acenaphthene	40.69	0.65	1.0	50.00	0	81.4	45	110	41.80	2.7	40					
2,4-Dinitrophenol	67.82	3.5	2.0	50.00	0	136	15	140	64.45	5.11	40					
4-Nitrophenol	59.75	0.53	2.0	50.00	0	120	0	125	56.37	5.82	40					
Dibenzofuran	40.71	0.52	1.0	50.00	0	81.4	55	105	41.63	2.23	40					
2,4-Dinitrotoluene	39.70	0.41	1.0	50.00	0	79.4	50	120	41.38	4.14	40					
Diethylphthalate	42.36	0.45	1.0	50.00	0	84.7	40	120	44.26	4.39	40					
4-Chlorophenyl-phenylether	41.74	0.41	1.0	50.00	0	83.5	50	110	40.93	1.96	40					
Fluorene	43.58	0.44	1.0	50.00	0	87.2	50	110	44.14	1.28	40					
4-Nitroaniline	32.92	0.96	2.0	50.00	0	65.8	35	120	34.84	5.66	40					
4,6-Dinitro-2-methylphenol	50.01	0.79	2.0	50.00	0	100	40	130	47.32	5.51	40					

**Qualifiers:** ND - Not Detected at the MDL

mmi.12.11.A

J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits

R - RPD outside accepted recovery limits

MDL - Method Detection Limit

RL - Reporting Limit

**B** - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8270\_W**

**SW846 8270D -- SVOA by GC-MS**

Analyte	Result	MDL	Units: ug/L	TestCode: SW8270_W	SPK value	SPK Ref Val	%REC LowLimit HighLimit			%RPD Ref Val	%RPD RPDLimit	Qual
							Analysis Date:	03/13/12 12:40	Prep Date:	03/11/12 11:00	Run ID:	S3_120313B
N-Nitrosodiphenylamine	39.56	1.1	1.0	50.00	0	79.1	50	110	39.31	0.636	40	
4-Bromophenylphenylether	47.62	0.54	1.0	50.00	0	95.2	50	115	43.73	8.51	40	
Hexachlorobenzene	50.97	0.44	1.0	50.00	0	102	50	110	47.48	7.09	40	
Pentachlorophenol	47.64	1.7	20	50.00	0	95.3	40	115	41.16	14.6	40	
Phenanthrene	41.51	0.45	1.0	50.00	0	83.0	50	115	41.09	1.0	40	
Anthracene	41.01	0.48	1.0	50.00	0	82.0	55	110	41.06	0.138	40	
Carbazole	37.70	0.64	1.0	50.00	0	75.4	50	115	39.14	3.75	40	
Di-n-butylphthalate	43.10	0.48	1.0	50.00	0	86.2	55	115	42.65	1.06	40	
Fluoranthene	39.42	0.33	10	50.00	0	78.8	55	115	40.37	2.38	40	
Pyrene	40.19	0.44	1.0	50.00	0	80.4	50	130	37.73	6.32	40	
Butylbenzylphthalate	41.05	0.32	10	50.00	0	82.1	45	115	39.76	3.19	40	
3,3'-Dichlorobenzidine	23.75	1.7	10	50.00	0	47.5	20	110	25.28	6.24	40	
Benz(a)anthracene	41.56	0.40	1.0	50.00	0	83.1	55	110	40.49	2.59	40	
Chrysene	41.60	0.42	1.0	50.00	0	83.2	55	110	42.19	1.4	40	
Bis(2-ethylhexyl)phthalate	41.26	1.3	10	50.00	0	82.5	40	125	40.83	1.06	40	
Di-n-octylphthalate	42.04	0.47	10	50.00	0	84.1	35	135	38.95	7.62	40	
Benz(b)fluoranthene	39.60	0.94	10	50.00	0	79.2	45	120	43.01	8.25	40	
Benz(k)fluoranthene	49.77	1.2	10	50.00	0	99.5	45	125	42.82	15	40	
Benz(a)pyrene	42.47	1.2	10	50.00	0	84.9	55	110	41.48	2.35	40	
Indeno(1,2,3-cd)pyrene	46.23	0.38	10	50.00	0	92.5	45	125	47.26	2.21	40	
Dibenz(a,h)anthracene	47.87	0.44	10	50.00	0	95.7	40	125	47.54	0.683	40	
Benz(g,h,i)perylene	45.91	0.39	10	50.00	0	91.8	40	125	47.73	3.88	40	
1,1'-Biphenyl	43.64	0.65	10	50.00	0	87.3	55	108	43.65	0.0174	40	
Acetophenone	44.23	0.51	10	50.00	0	88.5	56	145	40.51	8.77	40	
Atrazine	50.69	1.3	10	50.00	0	101	52	175	58.40	14.2	40	
Benzaldehyde	25.09	0.51	10	50.00	0	50.2	10	133	23.13	8.15	40	
Caprodiactam	26.29	1.1	10	50.00	0	52.6	10	146	27.43	4.26	40	
Surrogate: Nitrobenzene-d5	44.48	10	50.00	0	89.0	40	110	0	0	0		
Surrogate: 2-Fluorobiphenyl	42.26	10	50.00	0	84.5	50	110	0	0	0		
Surrogate: Terphenyl-d14	40.23	10	50.00	0	80.5	50	135	0	0	0		
Surrogate: Phenol-d5	36.18	10	50.00	0	72.4	10	115	0	0	0		
Surrogate: 2-Fluorophenol	40.69	10	50.00	0	81.4	20	110	0	0	0		
Surrogate: 2,4,6-Tribromophenol	61.61	10	50.00	0	123	40	125	0	0	0		

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mmi.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8270\_W**

**SW846 8270D -- SVOA by GC-MS**

Sample ID: <b>LCSD-65098</b>		SampType: <b>LCSD</b>	TestCode: <b>SW8270_W</b>		Prep Date: <b>03/14/12 8:22</b>		Analysis Date: <b>03/16/12 17:16</b>		Run ID: <b>S6_120316A</b>					
Client ID:	Batch ID:	<b>65098</b>	Units:	<b>ug/L</b>	MDL	RL	SPK value	SPK Ref Val	%REC	%RPD	RPDLimit	RPD Val	%RPD	Qual
1,4-Dichlorobenzene	132.1	3.7	33	166.7	0	79.3	30	100	130.6	1.14	40			
2-Methylphenol	118.2	3.2	33	166.7	0	70.9	40	110	115.6	2.19	40			
4-Methylphenol	126.5	4.7	33	166.7	0	75.9	30	110	123.5	2.42	40			
Hexachloroethane	105.8	1.8	33	166.7	0	63.5	30	95	104.1	1.62	40			
Nitrobenzene	129.0	5.3	33	166.7	0	77.4	45	110	126.5	1.91	40			
Hexachlorobutadiene	140.3	2.5	33	166.7	0	84.2	25	105	140.2	0.042	40			
2,4,6-Trichlorophenol	144.1	1.8	33	166.7	0	86.5	50	115	142.4	1.18	40			
2,4,5-Trichlorophenol	142.0	0.87	67	166.7	0	85.2	50	110	139.7	1.67	40			
2,4-Dinitrotoluene	134.8	1.4	33	166.7	0	80.9	50	120	135.3	0.379	40			
Hexachlorobenzene	147.4	1.5	33	166.7	0	88.4	50	110	148.7	0.892	40			
Pentachlorophenol	169.3	5.7	67	166.7	0	102	40	115	167.7	0.953	40			
Pyridine	145.5	1.9	67	166.7	0	87.3	10	106	140.7	3.39	40			
Surrogate: Nitrobenzene-d5	123.8	33	166.7	0	74.3	40	110	0						
Surrogate: 2-Fluorobiphenyl	129.5	33	166.7	0	77.7	50	110	0						
Surrogate: Terphenyl-d14	127.8	33	166.7	0	76.7	50	135	0						
Surrogate: Phenol-d5	126.1	33	166.7	0	75.7	10	115	0						
Surrogate: 2-Fluorophenol	123.1	33	166.7	0	73.9	20	110	0						
Surrogate: 2,4,6-Tribromophenol	160.6	33	166.7	0	96.4	40	125	0						

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 8081B -- Organochlorine Pesticides by GC-ECD</b>						<b>SW8081_W</b>
alpha-BHC	ND		0.050 ug/L		1 03/12/2012 14:31	65046
beta-BHC	ND		0.050 ug/L		1 03/12/2012 14:31	65046
delta-BHC	ND		0.050 ug/L		1 03/12/2012 14:31	65046
gamma-BHC (Lindane)	ND		0.050 ug/L		1 03/12/2012 14:31	65046
Heptachlor	ND		0.050 ug/L		1 03/12/2012 14:31	65046
Aldrin	ND		0.050 ug/L		1 03/12/2012 14:31	65046
Heptachlor epoxide	ND		0.050 ug/L		1 03/12/2012 14:31	65046
Endosulfan I	ND		0.050 ug/L		1 03/12/2012 14:31	65046
Dieldrin	ND		0.10 ug/L		1 03/12/2012 14:31	65046
4,4'-DDE	ND		0.10 ug/L		1 03/12/2012 14:31	65046
Endrin	ND		0.10 ug/L		1 03/12/2012 14:31	65046
Endosulfan II	ND		0.10 ug/L		1 03/12/2012 14:31	65046
4,4'-DDD	ND		0.10 ug/L		1 03/12/2012 14:31	65046
Endosulfan sulfate	ND		0.10 ug/L		1 03/12/2012 14:31	65046
4,4'-DDT	ND		0.10 ug/L		1 03/12/2012 14:31	65046
Methoxychlor	ND		0.50 ug/L		1 03/12/2012 14:31	65046
Endrin ketone	ND		0.10 ug/L		1 03/12/2012 14:31	65046
Endrin aldehyde	ND		0.10 ug/L		1 03/12/2012 14:31	65046
alpha-Chlordane	ND		0.050 ug/L		1 03/12/2012 14:31	65046
gamma-Chlordane	ND		0.050 ug/L		1 03/12/2012 14:31	65046
Toxaphene	ND		5.0 ug/L		1 03/12/2012 14:31	65046
Surrogate: Tetrachloro-m-xylene	44.9		25-140 %REC		1 03/12/2012 14:31	65046
Surrogate: Decachlorobiphenyl	28.7	S	30-135 %REC		1 03/12/2012 14:31	65046

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-SOLID  
**Lab ID:** L0466-02

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:00

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 8081B -- Organochlorine Pesticides by GC-ECD</b>						
gamma-BHC (Lindane) -- TCLP	ND		0.17 ug/L		1 03/14/2012 13:58	65096
Heptachlor -- TCLP	ND		0.17 ug/L		1 03/14/2012 13:58	65096
Heptachlor epoxide -- TCLP	ND		0.17 ug/L		1 03/14/2012 13:58	65096
Endrin -- TCLP	ND		0.33 ug/L		1 03/14/2012 13:58	65096
Methoxychlor -- TCLP	ND		1.7 ug/L		1 03/14/2012 13:58	65096
Toxaphene -- TCLP	ND		17 ug/L		1 03/14/2012 13:58	65096
Chlordane (technical) -- TCLP	ND		8.3 ug/L		1 03/14/2012 13:58	65096
Surrogate: Tetrachloro-m-xylene -- TCLP	93.2		25-140 %REC		1 03/14/2012 13:58	65096
Surrogate: Decachlorobiphenyl -- TCLP	86.3		30-135 %REC		1 03/14/2012 13:58	65096

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hamibal Tech

Date: 03/28/2012

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8081\_W**

**SW846 8081B -- Organochlorine Pesticides by GC-ECD**

Sample ID: <b>MB-65046</b>	Samp Type: <b>MBLK</b>	TestCode: <b>SW8081_W</b>	Prep Date: <b>03/10/12 11:00</b>	Run ID: <b>E5_120312A</b>							
Client ID: <b>MB-65046</b>	Batch ID: <b>65046</b>	Units: <b>ug/L</b>	Analysis Date: <b>03/12/12 13:32</b>	SeqNo: <b>1704181</b>							
Analyte	Result	MDL	RL	SPK value	%REC	SPK Ref Val	%RPD	Ref Val	%RPD	RPDLimit	Qual
alpha-BHC	ND	0.0018	0.050								
beta-BHC	ND	0.0020	0.050								
delta-BHC	ND	0.0027	0.050								
gamma-BHC (Lindane)	ND	0.0019	0.050								
Heptachlor	ND	0.0039	0.050								
Aldrin	ND	0.0043	0.050								
Heptachlor epoxide	ND	0.0028	0.050								
Endosulfan I	ND	0.0029	0.050								
Dieldrin	ND	0.0056	0.10								
4,4'-DDE	ND	0.0056	0.10								
Endrin	ND	0.0035	0.10								
Endosulfan II	ND	0.0031	0.10								
4,4'-DDD	ND	0.0064	0.10								
Endosulfan sulfate	ND	0.0045	0.10								
4,4'-DDT	ND	0.0070	0.10								
Methoxychlor	ND	0.031	0.50								
Endrin ketone	ND	0.0046	0.10								
Endrin aldehyde	ND	0.015	0.10								
alpha-Chlordane	ND	0.0024	0.050								
gamma-Chlordane	ND	0.0026	0.050								
Toxaphene	ND	0.14	5.0								

Sample ID: <b>MB-65046</b>	Samp Type: <b>MBLK</b>	TestCode: <b>SW8081_W</b>	Prep Date: <b>03/10/12 11:00</b>	Run ID: <b>E5_120312B</b>							
Client ID: <b>MB-65046</b>	Batch ID: <b>65046</b>	Units: <b>ug/L</b>	Analysis Date: <b>03/12/12 13:32</b>	SeqNo: <b>1704212</b>							
Analyte	Result	MDL	RL	SPK value	%REC	SPK Ref Val	%RPD	Ref Val	%RPD	RPDLimit	Qual
Surrogate: Tetrachloro-m-xylene	0.5776	0.050	0.6000	0	96.3	25	140	0	0		
Surrogate: Decachlorobiphenyl	1.167	0.100	1.200	0	97.2	30	135	0	0		

**Qualifiers:** ND - Not Detected at the MDL

mml1.12.11.A

J - Analyte detected below quantitation limits

R - RPD outside accepted recovery limits

RL - Reporting Limit

S - Recovery outside accepted recovery limits

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8081\_W**  
**SW846 8081B -- Organochlorine Pesticides by GC-ECD**

Sample ID:	MB-65091	SampType:	MBLK	TestCode:	SW8081_W	Prep Date:	03/14/12 8:18	Run ID:	E5_120314A				
Client ID:	MB-65091	Batch ID:	65096	Units:	ug/L	Analysis Date:	03/14/12 13:14	SeqNo:	1705164				
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
gamma-BHC (lindane) -- TCLP		ND	0.00063	0.17									
Heptachlor -- TCLP		ND	0.013	0.17									
Heptachlor epoxide -- TCLP		ND	0.0093	0.17									
Endrin -- TCLP		ND	0.012	0.33									
Methoxychlor -- TCLP		ND	0.10	1.7									
Toxaphene -- TCLP		ND	0.47	17									
Chlordane (technical) -- TCLP		ND	0.21	8.3									
Surrogate: Tetrachloro-m-xylene -- TCLP		1.836	0.17	2.000	0	91.8	25	140	0				
Surrogate:		4.145	0.33	4.000	0	104	30	135	0				
<hr/>													
Sample ID:	LCS-65046	SampType:	LCS	TestCode:	SW8081_W	Prep Date:	03/10/12 11:00	Run ID:	E5_120312A				
Client ID:	LCS-65046	Batch ID:	65046	Units:	ug/L	Analysis Date:	03/12/12 13:47	SeqNo:	1704182				
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
beta-BHC		0.1897	0.0020	0.050	0.2000	0	94.8	65	125	0			
gamma-BHC (lindane)		0.2024	0.0019	0.050	0.2000	0	101	25	135	0			
Aldrin		0.1982	0.0043	0.050	0.2000	0	99.1	25	140	0			
Heptachlor epoxide		0.2034	0.0028	0.050	0.2000	0	102	60	130	0			
Dieldrin		0.4138	0.0056	0.10	0.4000	0	103	60	130	0			
4,4'-DDE		0.4044	0.0056	0.10	0.4000	0	101	35	140	0			
Endrin		0.4279	0.0035	0.10	0.4000	0	107	55	135	0			
Endosulfan II		0.3932	0.0031	0.10	0.4000	0	98.3	30	130	0			
4,4'-DDD		0.4030	0.0064	0.10	0.4000	0	101	25	150	0			
4,4'-DDT		0.3993	0.0070	0.10	0.4000	0	99.8	45	140	0			
Endrin ketone		0.4001	0.0046	0.10	0.4000	0	100	75	125	0			
alpha-Chlordane		0.1958	0.0024	0.050	0.2000	0	97.9	65	125	0			
gamma-Chlordane		0.2017	0.0026	0.050	0.2000	0	101	60	125	0			
Surrogate: Tetrachloro-m-xylene		0.5491	0.050	0.6000	0	91.5	25	140	0				

Qualifiers: ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mmi.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8081\_W**

**SW846 8081B -- Organochlorine Pesticides by GC-ECD**

Sample ID:	LCS-65046	SampType: LCS	TestCode: SW8081_W	Prep Date:	03/10/12 11:00	Run ID:	E5_120312B						
Client ID:	LCS-65046	Batch ID:	65046	Analysis Date:	03/12/12 13:47	SeqNo:	1704213						
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
alpha-BHC	0.1981	0.0018	0.050	0.2000	0	99.0	60	130	0				
delta-BHC	0.2048	0.0027	0.050	0.2000	0	102	45	135	0				
Heptachlor	0.1992	0.0039	0.050	0.2000	0	99.6	40	130	0				
Endosulfan I	0.1924	0.0029	0.050	0.2000	0	96.2	50	110	0				
Endosulfan sulfate	0.4192	0.0045	0.10	0.4000	0	105	55	135	0				
Methoxychlor	2.099	0.031	0.50	2.000	0	105	55	150	0				
Endrin aldehyde	0.4098	0.015	0.10	0.4000	0	102	55	135	0				
Surrogate:													
Decachlorobiphenyl													

Sample ID:	LCS-65096	SampType: LCS	TestCode: SW8081_W	Prep Date:	03/14/12 8:18	Run ID:	E5_120314A						
Client ID:	LCS-65096	Batch ID:	65096	Analysis Date:	03/14/12 13:28	SeqNo:	1705165						
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
gamma-BHC (lindane)	0.7070	0.0063	0.17	0.6667	0	106	25	135	0				
Heptachlor	0.6779	0.013	0.17	0.6667	0	102	40	130	0				
Heptachlor epoxide	0.7033	0.0093	0.17	0.6667	0	105	60	130	0				
Endrin	1.497	0.012	0.33	1.333	0	112	55	135	0				
Methoxychlor	7.014	0.10	1.7	6.667	0	105	55	150	0				
Surrogate: Tetrachloro-m-xylene	1.932	0.17	0.17	2.000	0	96.6	25	140	0				
Surrogate:													
Decachlorobiphenyl													

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8081\_W**  
**SW846 8081B -- Organochlorine Pesticides by GC-ECD**

Sample ID:	LCSD-65046	SampType: LCSD	TestCode: SW8081_W	Prep Date: 03/10/12 11:00			Run ID: E5_120312A					
Client ID:	LCSD-65046	Batch ID: 65046	Units: ug/L	Analysis Date: 03/12/12 14:02			SeqNo: 1704183					
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
gamma-BHC (lindane)		0.2114	0.0019	0.050	0.2000	0	106	25	135	0.2024	4.38	30
Aldrin		0.2063	0.0043	0.050	0.2000	0	103	25	140	0.1982	4.03	30
Heptachlor epoxide		0.2113	0.0028	0.050	0.2000	0	106	60	130	0.2034	3.81	30
Endosulfan I		0.2004	0.0029	0.050	0.2000	0	100	50	110	0.1939	3.29	30
Dieldrin		0.4288	0.0056	0.10	0.4000	0	107	60	130	0.4138	3.58	30
4,4'-DDE		0.4158	0.0056	0.10	0.4000	0	104	35	140	0.4044	2.79	30
Endrin		0.4441	0.0035	0.10	0.4000	0	111	55	135	0.4279	3.73	30
Endosulfan II		0.4185	0.0031	0.10	0.4000	0	105	30	130	0.3932	6.23	30
4,4'-DDD		0.4383	0.0064	0.10	0.4000	0	110	25	150	0.4030	8.4	30
4,4'-DDT		0.4289	0.0070	0.10	0.4000	0	107	45	140	0.3993	7.14	30
Methoxychlor		2.246	0.031	0.50	2.000	0	112	55	150	2.115	6.02	30
Endrin aldehyde		0.4357	0.015	0.10	0.4000	0	109	55	135	0.4140	5.13	30
alpha-Chlordane		0.2040	0.0024	0.050	0.2000	0	102	65	125	0.1958	4.12	30
gamma-Chlordane		0.2116	0.0026	0.050	0.2000	0	106	60	125	0.2017	4.8	30
Surrogate: Tetrachloro-m-xylene		0.5806	0.050	0.6000	0	96.8	25	140	0			

Sample ID:	LCSD-65046	SampType: LCSD	TestCode: SW8081_W	Prep Date: 03/10/12 11:00			Run ID: E5_120312B					
Client ID:	LCSD-65046	Batch ID: 65046	Units: ug/L	Analysis Date: 03/12/12 14:02			SeqNo: 1704214					
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
alpha-BHC		0.2076	0.0018	0.050	0.2000	0	104	60	130	0.1981	4.68	30
beta-BHC		0.2004	0.0020	0.050	0.2000	0	100	65	125	0.2146	6.84	30
delta-BHC		0.2155	0.0027	0.050	0.2000	0	108	45	135	0.2048	5.08	30
Heptachlor		0.2097	0.0039	0.050	0.2000	0	105	40	130	0.1992	5.14	30
Endosulfan sulfate		0.4460	0.0045	0.10	0.4000	0	112	55	135	0.4192	6.2	30
Endrin ketone		0.4233	0.0046	0.10	0.4000	0	106	75	125	0.4042	4.6	30
Surrogate:		1.277	0.10	1.200	0	106	30	135	0			
Decachlorobiphenyl												

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mmi.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8081\_W**

**SW846 8081B -- Organochlorine Pesticides by GC-ECD**

Sample ID:	LCSD-65096	SampType: LCSD	TestCode: SW8081_W	Prep Date:	03/14/12 8:18	Run ID:	E5_120314A					
Client ID:	LCSD-65096	Batch ID: 65096	Units: ug/L	Analysis Date:	03/14/12 13:43	SeqNo:	1705166					
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
gamma-BHC (lindane)	0.7081	0.0063	0.17	0.6667	0	106	25	135	0.7070	0.152	30	
Heptachlor	0.6968	0.013	0.17	0.6667	0	105	40	130	0.6779	2.75	30	
Heptachlor epoxide	0.7014	0.0093	0.17	0.6667	0	105	60	130	0.7033	0.281	30	
Endrin	1.508	0.012	0.33	1.333	0	113	55	135	1.497	0.728	30	
Methoxychlor	7.137	0.10	1.7	6.667	0	107	55	150	7.014	1.74	30	
Surrogate: Tetrachloro-m-xylene	1.923	0.17	2.000	0	96.2	25	140	0				

Sample ID:	LCSD-65096	SampType: LCSD	TestCode: SW8081_W	Prep Date:	03/14/12 8:18	Run ID:	E5_120314B					
Client ID:	LCSD-65096	Batch ID: 65096	Units: ug/L	Analysis Date:	03/14/12 13:43	SeqNo:	1705185					
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Surrogate: Decachlorobiphenyl	4.209	0.33	4.000	0	105	30	135	0				

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mmi1.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 8082A -- PCB by GC-ECD</b>						
Aroclor-1016	ND		1.0 ug/L		1 03/12/2012 14:05	65047
Aroclor-1221	ND		1.0 ug/L		1 03/12/2012 14:05	65047
Aroclor-1232	ND		1.0 ug/L		1 03/12/2012 14:05	65047
Aroclor-1242	ND		1.0 ug/L		1 03/12/2012 14:05	65047
Aroclor-1248	ND		1.0 ug/L		1 03/12/2012 14:05	65047
Aroclor-1254	ND		1.0 ug/L		1 03/12/2012 14:05	65047
Aroclor-1260	ND		1.0 ug/L		1 03/12/2012 14:05	65047
Surrogate: Tetrachloro-m-xylene	48.0		34-137 %REC		1 03/12/2012 14:05	65047
Surrogate: Decachlorobiphenyl	27.4	S	40-135 %REC		1 03/12/2012 14:05	65047

**Qualifiers:** ND - Not Detected at the Reporting Limit

S - Spike Recovery outside accepted recovery limits

J - Analyte detected below quantitation limits

R - RPD outside accepted recovery limits

B - Analyte detected in the associated Method Blank

E - Value above quantitation range

DF - Dilution Factor

RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-SOLID  
**Lab ID:** L0466-02

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:00

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 8082A -- PCB by GC-ECD</b>						<b>SW8082_S</b>
Aroclor-1016	ND		37 ug/Kg		1 03/12/2012 15:50	65049
Aroclor-1221	ND		37 ug/Kg		1 03/12/2012 15:50	65049
Aroclor-1232	ND		37 ug/Kg		1 03/12/2012 15:50	65049
Aroclor-1242	ND		37 ug/Kg		1 03/12/2012 15:50	65049
Aroclor-1248	ND		37 ug/Kg		1 03/12/2012 15:50	65049
Aroclor-1254	ND		37 ug/Kg		1 03/12/2012 15:50	65049
Aroclor-1260	ND		37 ug/Kg		1 03/12/2012 15:50	65049
Surrogate: Tetrachloro-m-xylene	58.0		34-147 %REC		1 03/12/2012 15:50	65049
Surrogate: Decachlorobiphenyl	48.4	S	60-125 %REC		1 03/12/2012 15:50	65049

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hannibal Tech

Date: 03/28/2012

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8082\_S**

**SW846 8082A -- PCB by GC-ECD**

Sample ID:	<b>MB-65049</b>	SampType:	<b>MBLK</b>	TestCode:	<b>SW8082_S</b>	Prep Date:	<b>03/10/12 11:00</b>	Run ID:	<b>E2_120312A</b>		
Client ID:	<b>MB-65049</b>	Batch ID:	<b>65049</b>	Units:	<b>ug/Kg</b>	Analysis Date:	<b>03/12/12 14:26</b>	SeqNo:	<b>1704094</b>		
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
Aroclor-1016	ND	2.5	33								
Aroclor-1221	ND	4.4	33								
Aroclor-1232	ND	2.4	33								
Aroclor-1242	ND	2.5	33								
Aroclor-1248	ND	3.8	33								
Aroclor-1254	ND	4.4	33								
Aroclor-1260	ND	1.8	33								
Surrogate: Tetrachloro-m-xylene	18.38		1.7	20.00	0	91.9	34	147	0	0	
Surrogate:	35.88		3.3	40.00	0	89.7	60	125	0	0	
Decachlorobiphenyl											

Sample ID:	<b>LCS-65049</b>	SampType:	<b>LCS</b>	TestCode:	<b>SW8082_S</b>	Prep Date:	<b>03/10/12 11:00</b>	Run ID:	<b>E2_120312A</b>		
Client ID:	<b>LCS-65049</b>	Batch ID:	<b>65049</b>	Units:	<b>ug/Kg</b>	Analysis Date:	<b>03/12/12 14:47</b>	SeqNo:	<b>1704095</b>		
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
Aroclor-1016	131.3	2.5	33	133.3	0	98.5	40	140	0	0	
Aroclor-1260	135.4	1.8	33	133.3	0	102	60	130	0	0	
Surrogate: Tetrachloro-m-xylene	18.86		1.7	20.00	0	94.3	34	147	0	0	
Surrogate:	36.67		3.3	40.00	0	91.7	60	125	0	0	
Decachlorobiphenyl											

Sample ID:	<b>LCSD-65049</b>	SampType:	<b>LCSD</b>	TestCode:	<b>SW8082_S</b>	Prep Date:	<b>03/10/12 11:00</b>	Run ID:	<b>E2_120312A</b>		
Client ID:	<b>LCSD-65049</b>	Batch ID:	<b>65049</b>	Units:	<b>ug/Kg</b>	Analysis Date:	<b>03/12/12 15:08</b>	SeqNo:	<b>1704096</b>		
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
Aroclor-1016	130.7	2.5	33	133.3	0	98.0	40	140	131.3	0.456	30
Aroclor-1260	135.1	1.8	33	133.3	0	101	60	130	135.4	0.207	30
Surrogate: Tetrachloro-m-xylene	18.65		1.7	20.00	0	93.3	34	147	0	0	
Surrogate:	36.75		3.3	40.00	0	91.9	60	125	0	0	
Decachlorobiphenyl											

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

mmi.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8082\_W**  
**SW846 8082A -- PCB by GC-ECD**

Sample ID:	MB-65047	SampType: MBLK	TestCode: SW8082_W	Prep Date:	03/10/12 11:00	Run ID:	E2_120312A
Client ID:	MB-65047	Batch ID: 65047	Units: ug/L	Analysis Date:	03/12/12 12:42	SeqNo:	1704089
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%RPD RPD Limit Qual
Aroclor-1016		ND	0.12	1.0			
Aroclor-1221		ND	0.095	1.0			
Aroclor-1232		ND	0.19	1.0			
Aroclor-1242		ND	0.030	1.0			
Aroclor-1248		ND	0.063	1.0			
Aroclor-1254		ND	0.20	1.0			
Aroclor-1260		ND	0.11	1.0			
Surrogate: Tetrachloro-m-xylene		0.5324	0.050	0.6000	0	88.7	34 137 0
Surrogate: Decachlorobiphenyl		0.9473	0.10	1.200	0	78.9	40 135 0

Sample ID:	LCS-65047	SampType: LCS	TestCode: SW8082_W	Prep Date:	03/10/12 11:00	Run ID:	E2_120312A
Client ID:	LCS-65047	Batch ID: 65047	Units: ug/L	Analysis Date:	03/12/12 13:03	SeqNo:	1704090
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%RPD RPD Limit Qual
Aroclor-1016		3.732	0.12	1.0	4.000	0	93.3 25 145 0
Aroclor-1260		3.868	0.11	1.0	4.000	0	96.7 30 145 0
Surrogate: Tetrachloro-m-xylene		0.5373	0.050	0.6000	0	89.5 34 137 0	
Surrogate: Decachlorobiphenyl		0.9982	0.10	1.200	0	83.2 40 135 0	

Sample ID:	LCSD-65047	SampType: LCSD	TestCode: SW8082_W	Prep Date:	03/10/12 11:00	Run ID:	E2_120312A
Client ID:	LCSD-65047	Batch ID: 65047	Units: ug/L	Analysis Date:	03/12/12 13:23	SeqNo:	1704091
Analyte		Result	MDL	RL	SPK value	SPK Ref Val	%RPD RPD Limit Qual
Aroclor-1016		3.918	0.12	1.0	4.000	0	97.9 25 145 30
Aroclor-1260		4.055	0.11	1.0	4.000	0	101 30 145 30
Surrogate: Tetrachloro-m-xylene		0.5559	0.050	0.6000	0	92.6 34 137 0	
Surrogate: Decachlorobiphenyl		1.049	0.10	1.200	0	87.4 40 135 0	

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mmi.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 8151A -- Chlorinated Herbicides by GC-ECD</b>						<b>SW8151_W</b>
Dalapon	ND		2.5 µg/L		1 03/13/2012 15:49	65050
Dicamba	ND		0.10 µg/L		1 03/13/2012 15:49	65050
MCPP	ND		100 µg/L		1 03/13/2012 15:49	65050
MCPA	ND		100 µg/L		1 03/13/2012 15:49	65050
Dichlorprop	ND		1.0 µg/L		1 03/13/2012 15:49	65050
2,4-D	ND		1.0 µg/L		1 03/13/2012 15:49	65050
2,4,5-TP (Silvex)	ND		0.10 µg/L		1 03/13/2012 15:49	65050
2,4,5-T	ND		0.10 µg/L		1 03/13/2012 15:49	65050
2,4-DB	ND		1.0 µg/L		1 03/13/2012 15:49	65050
Dinoseb	ND		0.50 µg/L		1 03/13/2012 15:49	65050
Surrogate: DCAA	51.3		23-139 %REC		1 03/13/2012 15:49	65050

**Qualifiers:** ND - Not Detected at the Reporting Limit

S - Spike Recovery outside accepted recovery limits

J - Analyte detected below quantitation limits

R - RPD outside accepted recovery limits

B - Analyte detected in the associated Method Blank

E - Value above quantitation range

DF - Dilution Factor

RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/28/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-SOLID  
**Lab ID:** L0466-02

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:00

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 8151A -- Chlorinated Herbicides by GC-ECD</b>						<b>SW8151_W</b>
2,4-D -- TCLP	ND		3.3 µg/L	1	03/14/2012 16:29	65097
2,4,5-TP (Silvex) -- TCLP	ND		0.33 µg/L	1	03/14/2012 16:29	65097
Surrogate: DCAA -- TCLP	56.1		23-139 %REC		1 03/14/2012 16:29	65097

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hannibal Tech

Date: 03/28/2012

## ANALYTICAL QC SUMMARY REPORT

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

**SW8151\_W**

**SW846 8151A -- Chlorinated Herbicides by GC-ECD**

Sample ID:	MB-65050	Samp Type:	MBLK	TestCode:	SW8151_W	Prep Date:	03/10/12 11:00	Run ID:	E4_120313A		
Client ID:	MB-65050	Batch ID:	65050	Units:	µg/L	Analysis Date:	03/13/12 14:50	SeqNo:	1704809		
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
2,4-D	ND	0.67	1.0								
2,4,5-TP (Silver)	ND	0.046	0.10								
Sample ID:	MB-65050	Samp Type:	MBLK	TestCode:	SW8151_W	Prep Date:	03/10/12 11:00	Run ID:	E4_120313B		
Client ID:	MB-65050	Batch ID:	65050	Units:	µg/L	Analysis Date:	03/13/12 14:50	SeqNo:	1704854		
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
Surrogate: DCAA	2.946		1.0	5.000	0	58.9	23	139	0		
Sample ID:	MB-65091	Samp Type:	MBLK	TestCode:	SW8151_W	Prep Date:	03/14/12 8:20	Run ID:	E4_120314A		
Client ID:	MB-65091	Batch ID:	65097	Units:	µg/L	Analysis Date:	03/14/12 15:30	SeqNo:	1705349		
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
2,4-D -- TC1P	ND	2.2	3.3								
2,4,5-TP (Silver) -- TC1P	ND	0.15	0.33								
Sample ID:	MB-65091	Samp Type:	MBLK	TestCode:	SW8151_W	Prep Date:	03/14/12 8:20	Run ID:	E4_120314B		
Client ID:	MB-65091	Batch ID:	65097	Units:	µg/L	Analysis Date:	03/14/12 15:30	SeqNo:	1705357		
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
Surrogate: DCAA -- TC1P	5.512		3.3	16.67	0	33.1	23	139	0		
Sample ID:	LCS-65050	Samp Type:	LCS	TestCode:	SW8151_W	Prep Date:	03/10/12 11:00	Run ID:	E4_120313A		
Client ID:	LCS-65050	Batch ID:	65050	Units:	µg/L	Analysis Date:	03/13/12 15:09	SeqNo:	1704974		
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD RPDLimit	Qual
Dalapon	4.314	0.46	2.5	12.50	0	34.5	10	100	0		
2,4-DB	4.562	0.34	1.0	5.000	0	91.2	26	114	0		

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

mmi.12.11.A

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8151\_W**

**SW846 8151A -- Chlorinated Herbicides by GC-ECD**

Sample ID: LCS-65050		SampType: LCS	TestCode: SW8151_W				Prep Date: 03/10/12 11:00	Run ID: E4_120313B				
Client ID:	LCS-65050	Batch ID: 65050	Units: µg/L				Analysis Date: 03/13/12 15:09	SeqNo: 1704977				
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Dicamba	0.4353	0.024	0.10	0.5000	0	87.1	10	107	0			
MCPP	396.2	40	100	500.0	0	79.2	51	105	0			
MCPA	428.9	55	100	500.0	0	85.8	45	103	0			
Dichlorprop	5.090	0.64	1.0	5.000	0	102	65	120	0			
2,4-D	4.461	0.67	1.0	5.000	0	89.2	27	113	0			
2,4,5-TP (Silvex)	0.4141	0.046	0.10	0.5000	0	82.8	38	116	0			
2,4,5-T	0.4068	0.049	0.10	0.5000	0	81.4	43	104	0			
Dinoseb	2.493	0.42	0.50	2.500	0	99.7	44	130	0			
Surrogate: DCAA	4.151	1.0	5.000	0	83.0	23	139	0				
Sample ID: LCS-65097		SampType: LCS	TestCode: SW8151_W				Prep Date: 03/14/12 8:20	Run ID: E4_120314B				
Client ID:	LCS-65097	Batch ID: 65097	Units: µg/L				Analysis Date: 03/14/12 15:50	SeqNo: 1705358				
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
2,4-D	7.868	2.2	3.3	16.67	0	47.2	27	113	0			
2,4,5-TP (Silvex)	1.019	0.15	0.33	1.667	0	61.2	38	116	0			
Surrogate: DCAA	9.213	3.3	16.67	0	55.3	23	139	0				
Sample ID: LCSD-65050		SampType: LCSD	TestCode: SW8151_W				Prep Date: 03/10/12 11:00	Run ID: E4_120313A				
Client ID:	LCSD-65050	Batch ID: 65050	Units: µg/L				Analysis Date: 03/13/12 15:29	SeqNo: 1704975				
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Daapon	4.138	0.46	2.5	12.50	0	33.1	10	100	4.314	4.16	30	

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
mm1.12.11.A

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW8151\_W**

**SW846 8151A -- Chlorinated Herbicides by GC-ECD**

Sample ID: LCSD-65050		SampType: LCSD	TestCode: SW8151_W		Units: µg/L		SPK value		SPK Ref Val		%REC	%LowLimit	%HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Client ID:	LCSD-65050	Batch ID: 65050	Result	MDL	RL	Analysis Date:	03/13/12 15:29	Prep Date:	03/10/12 11:00	Run ID:	E4_120313B	SeqNo:	1704978				
Dicamba	0.4113	0.024	0.10	0.5000	0	82.3	10	107	0.4353	5.67	30						
MCPP	396.0	40	100	500.0	0	79.2	51	105	396.2	0.0538	30						
MCPA	420.9	55	100	500.0	0	84.2	45	103	428.9	1.87	30						
Dichlorprop	5.024	0.64	1.0	5.000	0	100	65	120	5.090	1.3	30						
2,4-D	4.312	0.67	1.0	5.000	0	86.2	27	113	4.461	3.38	30						
2,4,5-TP (Silvex)	0.3544	0.046	0.10	0.5000	0	70.9	38	116	0.4141	15.5	30						
2,4,5-T	0.3335	0.049	0.10	0.5000	0	66.7	43	104	0.4068	19.8	30						
2,4-DB	3.651	0.34	1.0	5.000	0	73.0	26	114	4.599	23	30						
Dinoseb	2.499	0.42	0.50	2.500	0	100	44	130	2.493	0.221	30						
Surrogate: DCAA	4.057		1.0	5.000	0	81.1	23	139	0								
Sample ID: LCSD-65097		SampType: LCSD	TestCode: SW8151_W		Units: µg/L		SPK value		SPK Ref Val		%REC	%LowLimit	%HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Client ID:	LCSD-65097	Batch ID: 65097	Result	MDL	RL	Analysis Date:	03/14/12 16:09	Prep Date:	03/14/12 8:20	Run ID:	E4_120314B	SeqNo:	1705359				
2,4-D	10.21	2.2	3.3	16.67	0	61.3	27	113	7.868	25.9	30						
2,4,5-TP (Silvex)	1.120	0.15	0.33	1.667	0	67.2	38	116	1.019	9.37	30						
Surrogate: DCAA	11.27		3.3	16.67	0	67.6	23	139	0								

**Qualifiers:** ND - Not Detected at the MDL  
J - Analyte detected below quantitation limits  
S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
I - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

MDL - Method Detection Limit  
RL - Reporting Limit

mm11.12.11.A

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/27/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result	Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 6010C -- Metals by ICP</b>						
Arsenic	34		20 ug/L		1 03/20/2012 10:54	65140
Barium	650	B	200 ug/L		1 03/20/2012 10:54	65140
Cadmium	4.9	J	5.0 ug/L		1 03/20/2012 10:54	65140
Chromium	480		20 ug/L		1 03/20/2012 10:54	65140
Lead	310		10 ug/L		1 03/20/2012 10:54	65140
Selenium	ND		30 ug/L		1 03/20/2012 10:54	65140
Silver	ND		30 ug/L		1 03/20/2012 10:54	65140
<b>SW846 7470A -- Mercury by FIA</b>						
Mercury	0.37		0.20 µg/L		1 03/19/2012 12:38	65142

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/27/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-SOLID  
**Lab ID:** L0466-02

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:00

Analyses	Result Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 6010C -- Metals by ICP</b>					
Arsenic -- TCLP	ND	20 ug/L	1	03/16/2012 12:36	65121
Barium -- TCLP	630	200 ug/L	1	03/16/2012 12:36	65121
Cadmium -- TCLP	1.2 J	5.0 ug/L	1	03/16/2012 12:36	65121
Chromium -- TCLP	1.1 J	20 ug/L	1	03/16/2012 12:36	65121
Lead -- TCLP	36	10 ug/L	1	03/16/2012 12:36	65121
Selenium -- TCLP	14 J	30 ug/L	1	03/16/2012 12:36	65121
Silver -- TCLP	ND	30 ug/L	1	03/16/2012 12:36	65121
<b>SW846 7470A -- Mercury by FIA</b>					
Mercury -- TCLP	0.033 J	0.20 µg/L	1	03/19/2012 12:40	65142

**Qualifiers:** ND - Not Detected at the Reporting Limit

S - Spike Recovery outside accepted recovery limits

J - Analyte detected below quantitation limits

R - RPD outside accepted recovery limits

B - Analyte detected in the associated Method Blank

E - Value above quantitation range

DF - Dilution Factor

RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Tech

Date: 03/27/2012

## ANALYTICAL QC SUMMARY REPORT

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

**Sample ID:** **MB-65091**      SampType: **MBLK**      TestCode: **SW6010\_W**  
**Client ID:** **MB-65091**      Batch ID: **65121**      Units: ug/L

Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD	Ref Val	%RPD	RPDLimit	Qual
Arsenic -- TCLP	ND	4.3	20	0	0	0	0	0	0	0	0	0	
Barium -- TCLP	ND	1.1	200	0	0	0	0	0	0	0	0	0	
Cadmium -- TCLP	ND	0.89	5.0	0	0	0	0	0	0	0	0	0	
Chromium -- TCLP	ND	0.64	20	0	0	0	0	0	0	0	0	0	
Lead -- TCLP	ND	4.2	10	0	0	0	0	0	0	0	0	0	
Selenium -- TCLP	ND	1.2	30	0	0	0	0	0	0	0	0	0	
Silver -- TCLP	ND	6.9	30	0	0	0	0	0	0	0	0	0	

Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD	Ref Val	%RPD	RPDLimit	Qual
Arsenic	ND	4.3	20	0	0	0	0	0	0	0	0	0	
Barium	ND	1.1	200	0	0	0	0	0	0	0	0	0	
Cadmium	ND	0.89	5.0	0	0	0	0	0	0	0	0	0	
Chromium	ND	0.64	20	0	0	0	0	0	0	0	0	0	
Lead	ND	4.2	10	0	0	0	0	0	0	0	0	0	
Selenium	ND	1.2	30	0	0	0	0	0	0	0	0	0	
Silver	ND	6.9	30	0	0	0	0	0	0	0	0	0	

Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD	Ref Val	%RPD	RPDLimit	Qual
Arsenic	ND	4.3	20	0	0	0	0	0	0	0	0	0	
Barium	ND	1.1	200	0	0	0	0	0	0	0	0	0	
Cadmium	ND	0.89	5.0	0	0	0	0	0	0	0	0	0	
Chromium	ND	0.64	20	0	0	0	0	0	0	0	0	0	
Lead	ND	4.2	10	0	0	0	0	0	0	0	0	0	
Selenium	ND	1.2	30	0	0	0	0	0	0	0	0	0	
Silver	ND	6.9	30	0	0	0	0	0	0	0	0	0	

Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD	Ref Val	%RPD	RPDLimit	Qual
Arsenic	ND	4.3	20	0	0	0	0	0	0	0	0	0	
Barium	ND	1.1	200	0	0	0	0	0	0	0	0	0	
Cadmium	ND	0.89	5.0	0	0	0	0	0	0	0	0	0	
Chromium	ND	0.64	20	0	0	0	0	0	0	0	0	0	
Lead	ND	4.2	10	0	0	0	0	0	0	0	0	0	
Selenium	ND	1.2	30	0	0	0	0	0	0	0	0	0	
Silver	ND	6.9	30	0	0	0	0	0	0	0	0	0	

Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD	Ref Val	%RPD	RPDLimit	Qual
Arsenic	ND	4.3	20	0	0	0	0	0	0	0	0	0	
Barium	ND	1.1	200	0	0	0	0	0	0	0	0	0	
Cadmium	ND	0.89	5.0	0	0	0	0	0	0	0	0	0	
Chromium	ND	0.64	20	0	0	0	0	0	0	0	0	0	
Lead	ND	4.2	10	0	0	0	0	0	0	0	0	0	
Selenium	ND	1.2	30	0	0	0	0	0	0	0	0	0	
Silver	ND	6.9	30	0	0	0	0	0	0	0	0	0	

Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD	Ref Val	%RPD	RPDLimit	Qual
Arsenic	ND	4.3	20	0	0	0	0	0	0	0	0	0	
Barium	ND	1.1	200	0	0	0	0	0	0	0	0	0	
Cadmium	ND	0.89	5.0	0	0	0	0	0	0	0	0	0	
Chromium	ND	0.64	20	0	0	0	0	0	0	0	0	0	
Lead	ND	4.2	10	0	0	0	0	0	0	0	0	0	
Selenium	ND	1.2	30	0	0	0	0	0	0	0	0	0	
Silver	ND	6.9	30	0	0	0	0	0	0	0	0	0	

<b>Qualifiers:</b>	ND - Not Detected at the MDL	S - Recovery outside accepted recovery limits	MDL - Method Detection Limit
nnm1.12.11.A	J - Analyte detected below quantitation limits	R - RPD outside accepted recovery limits	RL - Reporting Limit
		Run ID: OPTIMA2_120316A	SeqNo: 1706598
		Run ID: OPTIMA2_120316A	SeqNo: 1708446
		Run ID: OPTIMA3_120320A	SeqNo: 1708443
		Run ID: OPTIMA3_120320A	SeqNo: 1708445

Date: 03/27/2012

Run ID: OPTIMA2\_120316A

Run ID: OPTIMA2\_120316A

Run ID: OPTIMA3\_120320A

Run ID: OPTIMA3\_120320A

Analysis Date: 03/16/12 12:12:12

Analysis Date: 03/19/12 9:45

Analysis Date: 03/20/12 9:43

Analysis Date: 03/16/12 12:16

RPD Ref Val %RPD RPDLimit Qual

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW6010\_W**  
**SW846 6010C -- Metals by ICP**

Sample ID: LCS-65121		SampType: LCS	TestCode: SW6010_W		Prep Date: 03/15/12 10:50		Run ID: OPTIMA2_120316A					
Client ID: LCS-65121		Batch ID: 65121	Units: ug/L		Analysis Date: 03/16/12 12:19		SeqNo: 1706600					
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Arsenic	459.7	4.3	20	455.0	0	101	80	120	0	0	0	
Barium	9208	1.1	200	9100	0	101	80	120	0	0	0	
Cadmium	227.0	0.89	5.0	227.0	0	100	80	120	0	0	0	
Chromium	893.4	0.64	20	910.0	0	98.2	80	120	0	0	0	
Lead	455.7	4.2	10	455.0	0	100	80	120	0	0	0	
Selenium	456.3	1.2	30	455.0	0	100	80	120	0	0	0	
Silver	1175	6.9	30	1130	0	104	80	120	0	0	0	
Sample ID: LCS-65140		SampType: LCS	TestCode: SW6010_W		Prep Date: 03/19/12 9:45		Run ID: OPTIMA3_120320A					
Client ID: LCS-65140		Batch ID: 65140	Units: ug/L		Analysis Date: 03/20/12 9:47		SeqNo: 1708447					
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Arsenic	500.0	4.3	20	455.0	0	110	80	120	0	0	0	
Barium	9369	1.1	200	9100	0	103	80	120	0	0	0	
Cadmium	235.3	0.89	5.0	227.0	0	104	80	120	0	0	0	
Chromium	932.6	0.64	20	910.0	0	102	80	120	0	0	0	
Lead	491.9	4.2	10	455.0	0	108	80	120	0	0	0	
Selenium	492.9	1.2	30	455.0	0	108	80	120	0	0	0	
Silver	1171	6.9	30	1130	0	104	80	120	0	0	0	
Sample ID: L0466-02ASD		SampType: SD	TestCode: SW6010_W		Prep Date: 03/15/12 10:50		Run ID: OPTIMA2_120316A					
Client ID: 80230-IDW-SOLID		Batch ID: 65121	Units: ug/L		Analysis Date: 03/16/12 12:40		SeqNo: 1706606					
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Arsenic -- TCLP	ND	22	100	0	0	0	0	0	0	0	0	
Barium -- TCLP	650.8	5.5	1000	0	0	0	0	0	634.0	2.62	10	J
Cadmium -- TCLP	ND	4.5	25	0	0	0	0	0	1.213	0	10	
Chromium -- TCLP	ND	3.2	100	0	0	0	0	0	1.105	0	10	
Lead - TCLP	34.79	21	50	0	0	0	0	0	35.76	2.75	10	J
Selenium -- TCLP	ND	60	150	0	0	0	0	0	14.08	0	10	
Silver -- TCLP	ND	35	150	0	0	0	0	0	0	0	10	

**Qualifiers:** ND - Not Detected at the MDL  
 J - Analyte detected below quantitation limits  
 mml 1.12.11.A S - Recovery outside accepted recovery limits  
 J - RPD outside accepted recovery limits

MDL - Method Detection Limit  
 RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW7470**

**SW846 7470A -- Mercury by FIA**

		TestCode: SW7470						TestCode: SW7470								
		SampType: MBLK	Batch ID: 65142	Units: µg/L	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Sample ID: <b>MB-65091</b>																
Client ID: <b>MB-65091</b>																
Analyte																
Mercury -- TCLP		ND	0.028	0.20				0	0	0	0	0	0	0		
		TestCode: SW7470						TestCode: SW7470								
		SampType: MBLK	Batch ID: 65142	Units: µg/L	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Sample ID: <b>MB-65142</b>																
Client ID: <b>MB-65142</b>																
Analyte																
Mercury		ND	0.028	0.20				0	0	0	0	0	0	0		
		TestCode: SW7470						TestCode: SW7470								
		SampType: LCS	Batch ID: 65142	Units: µg/L	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Sample ID: <b>LCS-65142</b>																
Client ID: <b>LCS-65142</b>																
Analyte																
Mercury		4.548	0.028	0.20				4.550	0	100	80	120	0	0		

**Qualifiers:** ND - Not Detected at the MDL  
 mm1.12.11.A J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
 R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
 RL - Reporting Limit

Run ID: **FIMS2\_120319A**  
 SeqNo: 1707809

Run ID: **FIMS2\_120319A**  
 SeqNo: 1707807

Run ID: **FIMS2\_120319A**  
 SeqNo: 1707808

Run ID: **FIMS2\_120319A**  
 SeqNo: 1707808

B - Analyte detected in the associated Method Blank

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/27/2012

**Client:** CDM  
**Client Sample ID:** 80230-IDW-AQ  
**Lab ID:** L0466-01

**Project:** Paul Miller Site, IDW  
**Collection Date:** 03/08/12 11:50

Analyses	Result Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SM 4500 H+ B -- pH VALUE</b>					<b>SM4500_H+</b>
pH	7.2	1.0 S.U.		103/12/2012 11:30	R65823
<b>SW846 1010 -- FLASHPOINT by Pensky-Martens Closed-Cup Method</b>					<b>SW1010_W</b>
Ignitability	NO FLASH @ 155	200 °F		103/09/2012 10:15	R65903
<b>SW846 7.3.3.2 -- Reactive Cyanide Released from Wastes</b>					<b>SW7.3.3.2_W</b>
Reactive Cyanide	ND	0.020 mg/L		103/16/2012 16:15	65115
<b>SW846 7.3.4.2 -- Reactive Sulfide Released from Wastes</b>					<b>SW7.3.4.2_W</b>
Reactive Sulfide	ND	0.030 mg/L		103/14/2012 15:42	65116

**Qualifiers:** ND - Not Detected at the Reporting Limit  
J - Analyte detected below quantitation limits  
B - Analyte detected in the associated Method Blank  
DF - Dilution Factor

S - Spike Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits  
E - Value above quantitation range  
RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

03/27/2012

**Client:** CDM

**Client Sample ID:** 80230-IDW-SOLID

**Project:** Paul Miller Site, IDW

**Lab ID:** L0466-02

**Collection Date:** 03/08/12 11:00

Analyses	Result Qual	RL Units	DF	Date Analyzed	Batch ID
<b>SW846 1010 -- FLASHPOINT by Pensky-Martens Closed-Cup Method</b>					
Ignitability	NO FLASH @ 140	200 °F		103/15/2011 12:00	R65903
<b>SW846 7.3.3.2 -- Reactive Cyanide Released from Wastes</b>					
Reactive Cyanide	ND	1.1 mg/Kg		103/16/2012 16:03	65113
<b>SW846 7.3.4.2 -- Reactive Sulfide Released from Wastes</b>					
Reactive Sulfide	ND	1.1 mg/Kg		103/14/2012 15:36	65114
<b>SW846 9045C -- Soil and Waste pH</b>					
pH	8.6	1.0 S.U.		103/15/2012 11:00	R65902

**Qualifiers:** ND - Not Detected at the Reporting Limit

S - Spike Recovery outside accepted recovery limits

J - Analyte detected below quantitation limits

R - RPD outside accepted recovery limits

B - Analyte detected in the associated Method Blank

E - Value above quantitation range

DF - Dilution Factor

RL - Reporting Limit

# Spectrum Analytical, Inc. Featuring Hannibal Tech

Date: 03/27/2012

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SM4500\_H+**

**SM 4500 H+ B -- pH VALUE**

Sample ID: <b>L0466-01CDUP</b>	SampType: <b>DUP</b>	TestCode: <b>SM4500_H+</b>	Prep Date: <b>03/12/12 11:30</b>	Run ID: <b>WC01_120312A</b>									
Client ID: <b>80230-IDW-AQ</b>	Batch ID: <b>R65823</b>	Units: <b>S.U.</b>	Analysis Date: <b>03/12/12 11:37</b>	SeqNo: <b>1703756</b>									
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD	Ref Val	%RPD	RPDLimit	Qual
pH	7.200	1.0	1.0	0	0	0	0	0	7.180	0.278	20		

 **Qualifiers:** ND - Not Detected at the MDL S - Recovery outside accepted recovery limits MDL - Method Detection Limit  
mml 1.12.11.A J - Analyte detected below quantitation limits R - RPD outside accepted recovery limits RL - Reporting Limit

B - Analyte detected in the associated Method Blank  
RL - Reporting Limit

mml 1.12.11.A B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW1010\_S**  
**SW846 1010 -- FLASHPOINT by Pensky-Martens Closed-Cup Method**

Sample ID: <b>L0466-02BDUP</b>	SampType: <b>DUP</b>	TestCode: <b>SW1010_S</b>	Prep Date: <b>03/15/11 13:15</b>	Run ID: <b>FLASH1_120315A</b>								
Client ID: <b>80230-IDW-SOLID</b>	Batch ID: <b>R65903</b>	Units: <b>°F</b>	Analysis Date: <b>03/15/11 13:30</b>	SeqNo: <b>1705570</b>								
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPD Limit	Qual
Ignitability	NO FLASH @ 140	140	200	0	0	0	0	0	0	0	20	

**Qualifiers:** ND - Not Detected at the MDL  
mml 1.12.11.A J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW7.3.3.2\_S**

**SW846 7.3.3.2 -- Reactive Cyanide Released from Wastes**

Sample ID:	SampType:	Batch ID:	TestCode:	Units:	SPK value	SPK Ref Val	Prep Date:	Analysis Date:	Run ID:	SeqNo:	RPD Ref Val	%RPD RPDLimit	Qual
Sample ID: <b>MB-65113</b>	SampType: <b>MBLK</b>	Batch ID: <b>65113</b>	TestCode: <b>SW7.3.3.2_S</b>	Units: <b>mg/Kg</b>			03/14/12 10:00	03/16/12 15:55	LACHAT1_120316B	1707106			
Client ID: <b>MB-65113</b>													
Analyte	Result	MDL	RL										
Reactive Cyanide	ND	1.0	1.0										
Sample ID:	SampType:	Batch ID:	TestCode:	Units:	SPK value	SPK Ref Val	Prep Date:	Analysis Date:	Run ID:	SeqNo:	RPD Ref Val	%RPD RPDLimit	Qual
Sample ID: <b>LCS-65113</b>	SampType: <b>LCS</b>	Batch ID: <b>65113</b>	TestCode: <b>SW7.3.3.2_S</b>	Units: <b>mg/Kg</b>			03/14/12 10:00	03/16/12 15:58	LACHAT1_120316B	1707107			
Client ID: <b>LCS-65113</b>													
Analyte	Result	MDL	RL										
Reactive Cyanide	ND	1.0	1.0		5.000	0	0	0			12.4	0	
Sample ID:	SampType:	Batch ID:	TestCode:	Units:	SPK value	SPK Ref Val	Prep Date:	Analysis Date:	Run ID:	SeqNo:	RPD Ref Val	%RPD RPDLimit	Qual
Sample ID: <b>LCSD-65113</b>	SampType: <b>LCSD</b>	Batch ID: <b>65113</b>	TestCode: <b>SW7.3.3.2_S</b>	Units: <b>mg/Kg</b>			03/14/12 10:00	03/16/12 16:00	LACHAT1_120316B	1707108			
Client ID: <b>LCSD-65113</b>													
Analyte	Result	MDL	RL										
Reactive Cyanide	ND	1.0	1.0		5.000	0	0	0			12.4	0	

**Qualifiers:** ND - Not Detected at the MDL  
 mm1.12.11.A J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
 R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
 RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW7.3.3.2\_W**  
**SW846 7.3.3.2 -- Reactive Cyanide Released from Wastes**

Sample ID:	SampType:	Batch ID:	TestCode:	Units:	SPK value	SPK Ref Val	Prep Date:	Analysis Date:	Run ID:	SeqNo:	%RPD	RPD Limit	Qual
Sample ID: <b>MB-65115</b>	<b>MBLK</b>						03/14/12 10:00						
Client ID: <b>MB-65115</b>		<b>65115</b>		<b>mg/L</b>			03/16/12 16:08						
Analyte		Result	MDL	RL				%REC	LowLimit	HighLimit			
Reactive Cyanide	ND	0.020	0.020										
Sample ID: <b>LCS-65115</b>	<b>LCS</b>		TestCode: <b>SW7.3.3.2_W</b>				03/14/12 10:00						
Client ID: <b>LCS-65115</b>		<b>65115</b>		<b>mg/L</b>			03/16/12 16:10						
Analyte		Result	MDL	RL				%REC	LowLimit	HighLimit			
Reactive Cyanide	ND	0.020	0.020		0.1000								
Sample ID: <b>LCSD-65115</b>	<b>LCSD</b>		TestCode: <b>SW7.3.3.2_W</b>				03/14/12 10:00						
Client ID: <b>LCSD-65115</b>		<b>65115</b>		<b>mg/L</b>			03/16/12 16:13						
Analyte		Result	MDL	RL				%REC	LowLimit	HighLimit			
Reactive Cyanide	ND	0.020	0.020		0.1000								

**Qualifiers:** ND - Not Detected at the MDL  
 mml 1.12.11.A J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
 R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
 RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

### SW7.3.4.2\_S

### SW846 7.3.4.2 -- Reactive Sulfide Released from Wastes

		TestCode: SW7.3.4.2_S						TestCode: SW7.3.4.2_S			TestCode: SW7.3.4.2_S					
		SampType: MBLK	Batch ID: 65114	Units: mg/Kg	MDL	Result	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Sample ID:	<b>MB-65114</b>															
Client ID:	<b>MB-65114</b>															
Analyte																
Reactive Sulfide		ND	1.0	1.0												
		SampType: LCS	Batch ID: 65114	Units: mg/Kg	MDL	Result	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Sample ID:	<b>LCS-65114</b>															
Client ID:	<b>LCS-65114</b>															
Analyte																
Reactive Sulfide		9.724	1.0	1.0				12.50	0		77.8	22.6	114	0		
		SampType: LCSD	Batch ID: 65114	Units: mg/Kg	MDL	Result	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD Ref Val	%RPD	RPDLimit	Qual
Sample ID:	<b>LCSD-65114</b>															
Client ID:	<b>LCSD-65114</b>															
Analyte																
Reactive Sulfide		9.778	1.0	1.0				12.50	0		78.2	22.6	114	9.724	0.553	20

**Qualifiers:** ND - Not Detected at the MDL  
 mm1.12.11.A J - Analyte detected below quantitation limits

S - Recovery outside accepted recovery limits  
 R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
 RL - Reporting Limit

B - Analyte detected in the associated Method Blank

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW7.3.4.2\_W**

**SW846 7.3.4.2 -- Reactive Sulfide Released from Wastes**

Sample ID:	Batch ID:	SampType:	TestCode:	Units:	Prep Date:	Analysis Date:	Run ID:
<b>MB-65116</b>	<b>65116</b>	<b>MBLK</b>	<b>SW7.3.4.2_W</b>	<b>mg/L</b>	<b>03/14/12 15:30</b>	<b>03/14/12 15:38</b>	<b>SPEC2_120314A</b>
Client ID:						SeqNo:	1705730
Analyte		Result	MDL	RL	SPK Ref Val	%REC LowLimit HighLimit	RPD Ref Val
Reactive Sulfide	ND	0 . 030	0 . 030				%RPD RPDLimit Qual
Sample ID:	Batch ID:	SampType:	TestCode:	Units:	Prep Date:	Analysis Date:	Run ID:
<b>LCS-65116</b>	<b>65116</b>	<b>LCS</b>	<b>SW7.3.4.2_W</b>	<b>mg/L</b>	<b>03/14/12 15:30</b>	<b>03/14/12 15:40</b>	<b>SPEC2_120314A</b>
Client ID:						SeqNo:	1705731
Analyte		Result	MDL	RL	SPK Ref Val	%REC LowLimit HighLimit	RPD Ref Val
Reactive Sulfide	0 . 3879	0 . 030	0 . 030	0 . 5000	0	77 . 6 22 . 6	114 0
Sample ID:	Batch ID:	SampType:	TestCode:	Units:	Prep Date:	Analysis Date:	Run ID:
<b>LCSD-65116</b>	<b>65116</b>	<b>LCSD</b>	<b>SW7.3.4.2_W</b>	<b>mg/L</b>	<b>03/14/12 15:30</b>	<b>03/14/12 15:41</b>	<b>SPEC2_120314A</b>
Client ID:						SeqNo:	1705732
Analyte		Result	MDL	RL	SPK Ref Val	%REC LowLimit HighLimit	RPD Ref Val
Reactive Sulfide	0 . 3890	0 . 030	0 . 030	0 . 5000	0	77 . 8 22 . 6	114 0 . 3879 0 . 277 20

**Qualifiers:** ND - Not Detected at the MDL S - Recovery outside accepted recovery limits MDL - Method Detection Limit  
mm1.12.11.A J - Analyte detected below quantitation limits R - RPD outside accepted recovery limits RL - Reporting Limit

B - Analyte detected in the associated Method Blank  
RL - Reporting Limit

**CLIENT:** CDM  
**Work Order:** L0466  
**Project:** Paul Miller Site, IDW

## ANALYTICAL QC SUMMARY REPORT

**SW9045\_S  
SW846 9045C -- Soil and Waste pH**

Sample ID: <b>L0466-02BDUP</b>	SampType: <b>DUP</b>	TestCode: <b>SW9045_S</b>	Prep Date: <b>03/15/12 11:00</b>	Run ID: <b>WC01_120315A</b>									
Client ID: <b>80230-IDW-SOLID</b>	Batch ID: <b>R65902</b>	Units: <b>S.U.</b>	Analysis Date: <b>03/15/12 11:05</b>	SeqNo: <b>1705566</b>									
Analyte	Result	MDL	RL	SPK value	SPK Ref Val	%REC	LowLimit	HighLimit	RPD	Ref Val	%RPD	RPDLimit	Qual
pH	8.570	1.0	1.0	0	0	0	0	0	8.570	0	20		

**Qualifiers:** ND - Not Detected at the MDL  
mml 1.12.11.A J - Analyte detected below quantitation limits  
S - Recovery outside accepted recovery limits  
R - RPD outside accepted recovery limits

MDL - Method Detection Limit  
RL - Reporting Limit

B - Analyte detected in the associated Method Blank

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

**WorkOrder: L0466**

**Client ID:** CDM\_NJ  
**Project:** Paul Miller Dry Cleaner Site Port Richmond, NY  
**WO Name:** Paul Miller Site, IDW  
**Location:** PAUL\_MILLER, D004437-35

**Case:** SDG:  
**PO:** D004437-35

**HC Due:** 03/28/12      **Report Level:** LEVEL\_2  
**Fax Due:**      **Special Program:**  
**Fax Report:**       **EDD:** EQUIIS\_4\_NYSDEC

**Comments:** no HC

Lab Samp ID	Client Sample ID	Collection Date	Date Rec'd	Matrix	Test Code	Samp / Lab Test Comments	HF	HT	MS	SEL	Storage
L0466-01A	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SW8260_W	/ 8260_OLM4, +TICs	Y	VOA			
L0466-01B	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SW8081_W	/	O2				
L0466-01B	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SW8082_W	/	O2				
L0466-01B	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SW8151_W	/	O2				
L0466-01B	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SW8270_W	/ OLM4_SVOA, +TICs	Y	O2			
L0466-01C	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SM4500_H+	/	O2				
L0466-01C	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SW1010_W	/	O2				
L0466-01C	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SW7.3.3.2_W	/ REACTIVITY	O2				
L0466-01C	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SW7.3.4.2_W	/ REACTIVITY	O2				
L0466-01D	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SW6010_W	/ RCRA8	Y	M3			
L0466-01D	80230-IDW-AQ	03/08/2012 11:50	03/09/2012	Aqueous	SW7470	/ RCRA8	M3				
L0466-02A	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	P_Moist	/	O2				
L0466-02A	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW6010_W	/ TCLP_METALS	Y	O2			
L0466-02A	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW7470	/ TCLP_METALS	O2				
L0466-02A	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW8081_W	/ TCLP	Y	O2			
L0466-02A	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW8082_S	/	O2				
L0466-02A	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW8151_W	/ TCLP	Y	O2			
L0466-02A	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW8270_W	/ TCLP	Y	O2			
L0466-02B	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW1010_S	/	O2				
L0466-02B	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW7.3.3.2_S	/ REACTIVITY	O2				
L0466-02B	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW7.3.4.2_S	/ REACTIVITY	O2				
L0466-02B	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW9045_S	/	O2				
L0466-02C	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW8260_W	/ TCLP_VOA	Y	VOA			

HF = Fraction logged in but all tests have been placed on hold

HT = Test logged in but has been placed on hold

# Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

WorkOrder: L0466

Client ID: CDM\_NJ  
Project: Paul Miller Dry Cleaner Site Port Richmond, NY  
WO Name: Paul Miller Site, DW  
Location: PAUL\_MILLER, D004437-35  
Comments: no HC

Lab Samp ID	Client Sample ID	Collection Date	Date Recv'd	Matrix	Test Code	Samp / Lab Test Comments	HF	HT	MS	SEL	Storage
L0466-02D	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW/8260_LOW_S	/ 8260_OLM4,+TICs			Y	Y	VOA
L0466-02D	80230-IDW-SOLID	03/08/2012 11:00	03/09/2012	Soil	SW/8260_MED_S	/ 8260_OLM4,+TICs			Y	Y	VOA

HF = Fraction logged in but all tests have been placed on hold

HT = Test logged in but has been placed on hold



SPECTRUM ANALYTICAL, INC.  
Featuring  
HANIBAL TECHNOLOGY

# CHAIN OF CUSTODY RECORD

Page 1 of 1

## Special Handling:

TAT- Indicate Date Needed:  
 All TAT's subject to laboratory approval.  
 Min. 24-hour notification needed for rushes.  
 Samples disposed of after 30 days unless otherwise instructed.

Report To: Seth Bellomy  
Conn Smith

110 Fieldcrest Ave #8 16th Floor  
 Edison NJ 08837

732-225-7003

Project Mgr.: S. Kelly

1=Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> 2=HCl 3=H<sub>2</sub>SO<sub>4</sub> 4=HNO<sub>3</sub> 5=NaOH 6=Ascorbic Acid 7=CH<sub>3</sub>OH  
 8=NaHSO<sub>4</sub> 9=\_\_\_\_\_ 10=\_\_\_\_\_ 11=\_\_\_\_\_

DW=Drinking Water GW=Groundwater WW=Wastewater  
 O=Oil SW= Surface Water SO=Soil SL=Sludge A=Air  
 X1=\_\_\_\_\_ X2=\_\_\_\_\_ X3=\_\_\_\_\_

G=Grab C=Composite

Lab Id:	Sample Id:	Date:	Time:	Type	Matrix
164460-51	80230-IDW-AQ	3/8/12	11:50	C	GW
164460-52	80230-IDW-Solid	3/8/12	11:50	C	SO

Invoice To: Sane

Project No.:

Site Name: Paul Miller

Location: Port Richmond

State: NY  
 Sampler(s): F. Robinson / E. Kelly

RQN: \_\_\_\_\_

List preservative code below:

# of VOA Vials	# of Amber Glass	# of Clear Glass	# of Plastic	Containers:	Analyses:	QA/QC Reporting Level
5	1	1	1	PCBs	PCBs	□ Level I
5	1	1	1	PCPs	PCPs	□ Level II
5	1	1	1	PCPs	PCPs	□ Level III
5	1	1	1	PCPs	PCPs	□ Level IV
5	1	1	1	PCPs	PCPs	Other _____

State specific reporting standards:

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Reinquished by:

Frank Ma

Received by:

Kedex

Condition upon receipt:  Iced  Ambient 40°C

Date: 3/9/12 Time: 9:05

## Spectrum Analytical, Inc. Featuring Hanibal Technology -- Rhode Island Division

Received By: <i>Vernon Bryant</i>	Page 01 of 01																														
Reviewed By: <i>Dr. Paul</i>	Log-in Date 03/09/2012																														
Work Order: L0466	Client Name: CDM																														
Project Name/Event: Paul Miller Dry Cleaner Site Port Richmond, NY / D004437-35																															
Remarks: (1/2) Please see associated sample/extract transfer logbook pages submitted with this data package.		<table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th rowspan="2">Lab Sample ID</th> <th colspan="5">Preservation (pH)</th> <th rowspan="2">VOA Matrix</th> <th rowspan="2">Soil HeadSpace or Air Bubble &gt; or equal to 1/4"</th> </tr> <tr> <th>HNO3</th> <th>H<sub>2</sub>SO<sub>4</sub></th> <th>HCl</th> <th>NaOH</th> <th>H<sub>3</sub>PO<sub>4</sub></th> </tr> </thead> <tbody> <tr> <td>L0466-01</td> <td>&lt;2</td> <td></td> <td></td> <td></td> <td></td> <td>H</td> <td></td> </tr> <tr> <td>L0466-02</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>US</td> <td></td> </tr> </tbody> </table>	Lab Sample ID	Preservation (pH)					VOA Matrix	Soil HeadSpace or Air Bubble > or equal to 1/4"	HNO3	H <sub>2</sub> SO <sub>4</sub>	HCl	NaOH	H <sub>3</sub> PO <sub>4</sub>	L0466-01	<2					H		L0466-02						US	
Lab Sample ID	Preservation (pH)					VOA Matrix	Soil HeadSpace or Air Bubble > or equal to 1/4"																								
	HNO3		H <sub>2</sub> SO <sub>4</sub>	HCl	NaOH			H <sub>3</sub> PO <sub>4</sub>																							
L0466-01	<2						H																								
L0466-02							US																								
1. Custody Seal(s)	Present / Absent																														
2. Custody Seal Nos.	<i>N/A</i>																														
3. Traffic Reports/ Chain of Custody Records (TR/COCs) or Packing Lists	Present / Absent																														
4. Airbill	AirBill / Sticker Present / Absent																														
5. Airbill No.	FedEx 8764 5236 7117																														
6. Sample Tags	Present / Absent																														
Sample Tag Numbers	Listed / Not Listed on Chain-of-Custody																														
7. Sample Condition	Intact / Broken/ Leaking																														
8. Cooler Temperature Indicator Bottle	Present / Absent																														
9. Cooler Temperature	4 °C																														
10. Does information on TR/COCs and sample tags agree?	Yes / No																														
11. Date Received at Laboratory	03/09/2012																														
12. Time Received	09:05																														
Sample Transfer																															
Fraction (1) TVOA/VOA	Fraction (2) SVOA/PEST/ARO																														
Area #	Area #																														
By	By																														
On	On																														
IR Temp Gun ID:MT-1	VOA Matrix Key: US = Unpreserved Soil      A = Air UA = Unpreserved Aqueous    H = HCl M = MeOH                      E = Encore N = NaHSO <sub>4</sub> F = Freeze																														
CoolantCondition: ICE																															
Preservative Name/Lot No:																															
See Sample Condition Notification/Corrective Action Form Yes / No Rad OK Yes / No																															

**Edward Lawler [Warwick]**

**From:** Kellogg, Seth [KelloggDS@cdmsmith.com]  
**Sent:** Tuesday, March 06, 2012 8:02 AM  
**To:** Edward Lawler [Warwick]; Robinson, Frank; Kulkusky, Edward  
**Subject:** FW: IDW sampling

Ed – we also need to run IDW characterization samples for Paul Miller. Can you send bottles for Friday?  
See parameters below.

Thanks,  
Seth

---

**From:** Gene Streiter [mailto:[streiterseacoast@aol.com](mailto:streiterseacoast@aol.com)]  
**Sent:** Monday, March 05, 2012 9:41 AM  
**To:** Kellogg, Seth  
**Subject:** RE: IDW sampling

You're sampling or I'm sampling?

For aqueous: RCRA Characteristics  
PCB  
Metals RCRA  
VOC-TCLVOA 10  
SVOC-TCLBNA 20  
Total Herb  
Total Pest

For soil: RCRA Characteristics  
PCB  
Full TCLP

---

**From:** Kellogg, Seth [mailto:[KelloggDS@cdmsmith.com](mailto:KelloggDS@cdmsmith.com)]  
**Sent:** Monday, March 05, 2012 8:14 AM  
**To:** Gene Streiter  
**Subject:** IDW sampling

Gene,

We're finishing up the work at Paul Miller this week. What analyses do you need for the IDW disposal?  
We're going to have 4 drums of water and 1 of soil/asphalt.

Thanks,  
Seth

*Seth Kellogg, P.G.*

Senior Project Manager  
CDM Smith | 110 Fieldcrest Avenue, #8 | 6th Floor | Edison, New Jersey [08837](tel:08837) | [kelloggds@cdmsmith.com](mailto:kelloggds@cdmsmith.com)  
(732) 590-4674 (Phone/Fax)  
(732) 354-8463 (cell)

**Last Page of Data Report**

# Norlite Corporation

628 South Saratoga Street  
Cohoes, New York 12047  
Phone: (518) 235-0401 Fax: (518) 235-0233



## WASTE STREAM PROFILE No. \_\_\_\_\_

Approved for Acceptance:  Yes  No

Reviewed by: \_\_\_\_\_ Date \_\_\_\_\_

Approved by: \_\_\_\_\_ Date \_\_\_\_\_

PLEASE ATTACH: All Material Safety Data Sheets (MSDS), Analysis Reports, Handling Precautions, Additional Hazard Information, Support Data & Comments.

### GENERATOR INFORMATION

Site Name Paul Miller site  
Address 1465 Forest Ave  
Staten Island NY  
Mailing Name NYSDEC/Paul Miller site  
Address 625 Broadway  
Albany NY 12233  
EPA I.D. # NYD 986 933 299  
Technical Contact \_\_\_\_\_  
Phone # \_\_\_\_\_ Fax # \_\_\_\_\_  
Shipping Contact \_\_\_\_\_  
Phone # \_\_\_\_\_ Fax # \_\_\_\_\_

Billing Name Innovative Recycling Tech Inc  
Address 690 N Queens Rd  
Lindenhurst NY 11757  
Contact John Ewer  
Phone # 516 816 4766 Fax # \_\_\_\_\_  
Transporter Name Freehold Container Inc  
Address \_\_\_\_\_  
EPA I.D. # NJD 054 126 164  
Contact J Ewer  
Phone # 516 816 4766 Fax # 631 225 3056

### WASTE DESCRIPTION

Generators Name For Waste Groundwater  
Process Generating Waste Site investigation of former dry cleaner

### SHIPPING INFORMATION

DOT Shipping Name Hazardous waste liquid n.o.s  
DOT Hazard Class 9 Packing Group III UN/NA No. 3082  
EPA Hazardous Waste Codes F002  
Estimated Volume 5 / year  
Container Type:  Bulk  Drum  Roll-Off

### CHEMICAL COMPOSITION - (Totals to 100%)

Water 99% % \_\_\_\_\_ %  
Tetrachloroethane 1% % \_\_\_\_\_ %  
\_\_\_\_\_ % \_\_\_\_\_ %  
\_\_\_\_\_ % \_\_\_\_\_ %  
\_\_\_\_\_ % \_\_\_\_\_ %  
\_\_\_\_\_ % \_\_\_\_\_ %  
\_\_\_\_\_ % \_\_\_\_\_ %  
\_\_\_\_\_ % \_\_\_\_\_ %

### TOTAL METALS

Antimony (Sb) NE ppm Lead (Pb) NE ppm  
Arsenic (As) \_\_\_\_\_ ppm Mercury (Hg) \_\_\_\_\_ ppm  
Barium (Ba) \_\_\_\_\_ ppm Nickel (Ni) \_\_\_\_\_ ppm  
Beryllium (Be) \_\_\_\_\_ ppm Selenium (Se) \_\_\_\_\_ ppm  
Cadmium (Cd) \_\_\_\_\_ ppm Silver (Ag) \_\_\_\_\_ ppm  
Chromium (Cr) \_\_\_\_\_ ppm Thallium (Tl) \_\_\_\_\_ ppm  
Copper (Cu) ✓ ppm Zinc (Zn) \_\_\_\_\_ ppm

### OTHER CHARACTERISTICS (Yes/No)

Reactive NO Infectious NO Explosive NO  
Biological NO Pyrophoric NO Radioactive NO

List Acute Hazardous Wastes as defined in 40CFR 261.33(e) or 6NYCRR 371.4(d)(5).  
List any Hazardous Constituents as defined by 40CFR 261 Appendix VIII. Describe  
any special handling requirements associated with this waste system.

### OTHER COMPONENTS

Herbicides 9 ppm Cyanide 9 ppm  
Pesticides ✓ ppm Sulfide 9 ppm  
Dioxins \_\_\_\_\_ ppm PBB \_\_\_\_\_ ppm

CERTIFICATION: I attest and certify that all information provided is complete and accurate. This low grade fuel is properly described with no willful omissions and that all known or suspected hazards have been disclosed and the low grade fuel is not a PCB waste that is defined in 40 CFR 761.3. Any changes or additional information obtained about this waste stream will be promptly conveyed to the Norlite Corporation.

Seth Kellogg on behalf of NYSDEC  
Print Name and Title

Seth Kellogg  
Signature

4/9/12  
Date

**VEXOR Technology, Inc.**  
 955 West Smith Road  
 Medina, Ohio 44256  
 Phone: 330-721-9773  
 FAX: 330-721-9438  
 EPA ID# OHD 077772895  
[www.vexortechology.com](http://www.vexortechology.com)

## MATERIAL CHARACTERIZATION

### For VEXOR Use Only

Approval # \_\_\_\_\_  
 Sample # \_\_\_\_\_  
 Sales Rep \_\_\_\_\_  
 Date Submitted \_\_\_\_\_

Email: mail@vexortechology.com

Generator <u>Paul Miller Site</u>	Bill To Name <u>Innovative Recycling Technologies</u>
Site Address <u>1465 Forest Avenue</u>	Site Address <u>690 No. Queens Avenue</u>
City <u>Staten Island</u>	State <u>NY</u> Zip <u>10302</u>
Phone <u>(631)225-3044</u>	Fax <u>_____</u>
EPA ID # <u>NYD986933299</u>	SIC Code <u>_____</u>
Technical Contact <u>_____</u>	Title <u>V. President</u>
Title <u>_____</u>	Email <u>jdull@irtwaste.com</u>
<b>MATERIAL DESCRIPTION</b>	

Name and Description of Material: Drill Cuttings & Asphalt

Process Generating Material: From site investigation of former dry cleaner U.S. EPA Hazardous Waste:  Yes  No

Proper DOT Shipping Name: Drill Cuttings & Asphalt Non-DOT Regulated Material

Method of Shipment:  Bulk  Drum  Tote  Cubic Yard Box  Other/Explain:

Est. Annual Volume: \_\_\_\_\_ Cu. Yds. \_\_\_\_\_ Tons \_\_\_\_\_ Gallons \_\_\_\_\_ 5 Drums \_\_\_\_\_ container material (metal, plastic, etc.)

Frequency:  One Time Only  Daily  Weekly  Monthly  Other/Explain Yearly Approximate Drum Weight

Special Handling Instructions: \_\_\_\_\_

Preferred Disposal Method:  Landfill  Waste to Energy  Recycling  Other Most cost effective

### MATERIAL PROPERTIES AT 78°

- a) Physical State:  Solid  Semi-Solid  Powder  Liquid  Phases
- b) Reactivity:  Water reactive  Acid Reactive  Alkaline Reactive  Oxidizer  Autosetting  None
- c) Flash Point, °F:  ≤72  >72-100  >100-140  >140-200  >200  N/A
- d) S.G./Density Solid
- e) pH:  ≤2  >2-6  >6-9  >9-12.5  ≥12.5  N/A
- f) Odor:  None  Mild  Strong: Describe \_\_\_\_\_
- g) Color: Varies
- h) Total Organic Halogen (TOX)  ppm  <1000 ppm  >1000 ppm\* If this material is considered "USED OIL" and is to be managed as a "USED OIL", please complete the "USED OIL" ADDENDUM and attach to this profile.
- i) PCB Content:  0 ppm  1-49 ppm\*  ≥50 ppm \* Supporting analysis and documentation required.

### MATERIAL COMPOSITION: List all components, must add up to 100%.

Constituent	Range % (wt-vol)	
	Min	Max
Soil	95	95
Asphalt	5	5
A combined total should equal 100%		

Above is based on:  Generator Knowledge  Analytical Data MSDS

Please attach analysis, TCLP information and appropriate MSDS sheets.

SAMPLE SUBMITTED WITH THIS PROFILE:  Yes  No

### For VEXOR Use Only

Evaluated by: \_\_\_\_\_  
 Approved - Treatment: \_\_\_\_\_  
 Rejected - Reason: \_\_\_\_\_  
 Date Completed: \_\_\_\_\_  
 Price: \_\_\_\_\_ /Unit \_\_\_\_\_  
 Approved By: \_\_\_\_\_ Date: \_\_\_\_\_

I hereby certify that to the best of my knowledge and belief, the information contained herein is a true and accurate description of the material being offered for disposal. Samples of this material submitted to VEXOR are representative of the material described in this profile. I further certify that by utilizing this profile, neither myself nor any other employee of the company will deliver for treatment, processing or recycling or attempt to deliver for same any material that is classified as toxic waste, hazardous water, medical or infectious waste or any other material that this facility is prohibited from accepting by law.

Authorized Representative Name (Printed) Seth Kellogg Company On behalf of NY SPEC

Authorized Representative Signature Seth Kellogg Title: On behalf of NY SPEC Date: 4/9/12

## Appendix C

### Full Data Tables

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-GW-MIP5-15 MIP5 05/24/2011	80230-GW-MIP5-15-DUP MIP5 05/24/2011	80230-GW-MIP5-28 MIP5 05/24/2011	80230-GW-MIP6-15 MIP6 05/23/2011	80230-GW-MIP6-33.5 MIP6 05/23/2011	80230-MW01-030612 MW01 03/06/2012	
Chemical Name	CAS#	New York State Class GA								
<b>Volatile Organic Compounds - SW8260</b>										
1,1,1,2-Tetrachloroethane	630-20-6	5		500 U	2000 U	5 U	5 U	5 U	5 U	
1,1,1-Trichloroethane	71-55-6	5		500 U	2000 U	5 U	5 U	5 U	5 U	
1,1,2,2-Tetrachloroethane	79-34-5	5		500 U	2000 U	5 U	5 U	5 U	5 U	
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1	5		500 U	2000 U	5 U	5 U	5 U	5 UJ	
1,1,2-Trichloroethane	79-00-5	1		500 U	2000 U	5 U	5 U	5 U	0.6 J	
1,1-Dichloroethane	75-34-3	5		500 U	2000 U	5 U	5 U	5 U	5 U	
1,1-Dichloroethene	75-35-4	5		500 U	2000 U	5 U	5 U	5 U	2.8 J	
1,1-Dichloropropene	563-58-6			500 U	2000 U	5 U	5 U	5 U	5 U	
1,2,3-Trichlorobenzene	87-61-6	5		500 U	2000 UJ	5 U	5 U	5 U	5 UJ	
1,2,3-Trichloropropane	96-18-4	0.04		500 U	2000 U	5 U	5 U	5 U	5 U	
1,2,4-Trichlorobenzene	120-82-1	5		500 U	2000 UJ	5 UJ	5 UJ	5 UJ	5 U	
1,2,4-Trimethylbenzene	95-63-6	5		500 U	2000 U	5 U	5 U	5 U	5 U	
1,2-Dibromo-3-Chloropropane	96-12-8	0.04		500 U	2000 U	5 U	5 U	5 U	5 UJ	
1,2-Dibromoethane (EDB)	106-93-4	0.0006		500 U	2000 U	5 U	5 U	5 U	5 U	
1,2-Dichlorobenzene	95-50-1	3		500 U	2000 U	5 U	5 U	5 U	5 U	
1,2-Dichloroethane	107-06-2	0.6		500 U	2000 U	5 U	5 U	5 U	5 U	
1,2-Dichloropropane	78-87-5	1		500 U	2000 U	5 U	5 U	5 U	5 U	
1,3,5-Trimethylbenzene	108-67-8	5		500 U	2000 U	5 U	5 U	5 U	5 U	
1,3-Dichlorobenzene	541-73-1	3		500 U	2000 U	5 U	5 U	5 U	5 U	
1,3-Dichloropropane	142-28-9	5		500 U	2000 U	5 U	5 U	5 U	5 U	
1,4-Dichlorobenzene	106-46-7	3		500 U	2000 U	5 U	5 U	5 U	5 U	
2,2-Dichloropropane	594-20-7	5		500 U	2000 U	5 UJ	5 UJ	5 UJ	5 UJ	
2-Butanone (MEK)	78-93-3			500 R	2000 R	5 R	5 R	5 R	5 R	
2-Chlorotoluene	95-49-8	5		500 U	2000 U	5 U	5 U	5 U	5 U	
2-Hexanone	591-78-6			500 U	2000 U	5 U	5 U	5 U	5 UJ	
4-Chlorotoluene	106-43-4	5		500 U	2000 U	5 U	5 U	5 U	5 U	
4-Methyl-2-Pentanone (MIBK)	108-10-1			500 U	2000 U	5 U	5 U	5 U	5 U	
Acetone	67-64-1			500 R	2000 UJ	5 U	5 U	5 U	5 R	
Benzene	71-43-2	1		500 U	2000 U	5 R	5 R	5 R	5 U	
Bromobenzene	108-86-1	5		500 U	2000 U	5 U	5 U	5 U	5 U	
Bromochloromethane	74-97-5	5		500 U	2000 U	5 U	5 U	5 U	5 U	
Bromodichloromethane	75-27-4			500 U	2000 U	5 U	5 U	5 U	5 U	
Bromoform	75-25-2			500 U	2000 U	5 U	5 U	5 U	5 U	
Bromomethane	74-83-9	5		500 U	2000 U	5 U	5 U	5 U	5 U	
Carbon Disulfide	75-15-0	60		500 U	2000 U	5 U	5 U	5 U	5 U	
Carbon Tetrachloride	56-23-5	5		500 U	2000 U	5 U	5 U	5 U	5 U	

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID Sample Location Sampling Date			80230-GW-MIP5-15 MIP5 05/24/2011		80230-GW-MIP5-15-DUP MIP5 05/24/2011		80230-GW-MIP5-28 MIP5 05/24/2011		80230-GW-MIP6-15 MIP6 05/23/2011		80230-GW-MIP6-33.5 MIP6 05/23/2011		80230-MW01-030612 MW01 03/06/2012	
Chemical Name	CAS#	New York State Class GA												
Chlorobenzene	108-90-7	5	500	U	2000	U	5	U	5	U	5	U	1.1	J
Chloroethane	75-00-3	5	500	U	2000	U	5	U	5	U	5	U	5	UJ
Chloroform	67-66-3	7	500	U	2000	U	5	U	5	U	5	U	5	U
Chloromethane	74-87-3	5	500	U	2000	U	5	U	5	U	5	U	5	UJ
cis-1,2-Dichloroethene	156-59-2	5	780		2000	U	1.7	J	21		11		4300	D
cis-1,3-Dichloropropene	10061-01-5		500	U	2000	U	5	U	5	U	5	U	5	U
Cyclohexane	110-82-7		500	U	2000	U	5	U	5	U	5	U	5	UJ
Dibromochloromethane	124-48-1		500	U	2000	U	5	U	5	U	5	U	5	U
Dibromomethane	74-95-3	5	500	U	2000	U	5	U	5	U	5	U	5	U
Dichlorodifluoromethane	75-71-8	5	500	UJ	2000	UJ	5	UJ	5	UJ	5	UJ	5	UJ
Ethylbenzene	100-41-4	5	500	U	2000	U	5	U	5	U	5	U	5	U
Hexachlorobutadiene	87-68-3	0.5	500	U	2000	UJ	5	U	5	U	5	U	5	U
Isopropylbenzene	98-82-8	5	500	U	2000	U	5	U	5	U	5	U	5	U
m,p-Xylene	179601-23-1		500	U	2000	U	5	U	5	U	5	U	5	U
Methyl Acetate	79-20-9		500	U	2000	UJ	5	UJ	5	UJ	5	UJ	5	UJ
Methyl Iodide	74-88-4	5	500	U	2000	U	5	U	5	U	5	U	5	UJ
Methyl Tert-Butyl Ether (MTBE)	1634-04-4		500	U	2000	U	5	U	1.7	J	5	U	5	U
Methylcyclohexane	108-87-2		500	U	2000	U	5	UJ	5	UJ	5	UJ	5	UJ
Methylene Chloride	75-09-2	5	500	U	2000	U	5	U	5	U	5	U	5	U
Naphthalene	91-20-3		500	U	2000	U	5	U	5	U	5	U	5	UJ
n-Butylbenzene	104-51-8	5	500	U	2000	U	5	U	5	U	5	U	5	U
n-Propylbenzene	103-65-1	5	500	U	2000	U	5	U	5	U	5	U	5	U
o-Xylene	95-47-6	5	500	U	2000	U	5	U	5	U	5	U	5	U
p-Isopropyltoluene	99-87-6	5	500	U	2000	U	5	U	5	U	5	U	5	U
sec-Butylbenzene	135-98-8	5	500	U	2000	U	5	U	5	U	5	U	5	U
Styrene	100-42-5	5	500	U	2000	U	5	U	5	U	5	U	5	U
tert-Butylbenzene	98-06-6	5	500	U	2000	U	5	U	5	U	5	U	5	U
Tetrachloroethene	127-18-4	5	32000	D	33000		150		2100	D	380	D	750	D
Toluene	108-88-3	5	500	U	2000	U	5	U	5	U	5	U	2	J
Total Xylenes	1330-20-7		500	U	2000	U	5	U	5	U	5	U	5	U
trans-1,2-Dichloroethene	156-60-5	5	500	U	2000	U	5	U	5	U	5	U	98	
trans-1,3-Dichloropropene	10061-02-6		500	U	2000	U	5	U	5	U	5	U	5	U
Trichloroethene	79-01-6	5	500	U	2000	U	2.3	J	9		8.7		130	
Trichlorofluoromethane	75-69-4	5	500	U	2000	U	5	U	5	U	5	U	5	U
Vinyl Acetate	108-05-4		500	U	2000	U	5	UJ	5	UJ	5	UJ	5	U
Vinyl Chloride	75-01-4	2	500	U	2000	U	5	U	5	U	5	U	1200	JD

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-GW-MIP5-15 MIP5 05/24/2011	80230-GW-MIP5-15-DUP MIP5 05/24/2011	80230-GW-MIP5-28 MIP5 05/24/2011	80230-GW-MIP6-15 MIP6 05/23/2011	80230-GW-MIP6-33.5 MIP6 05/23/2011	80230-MW01-030612 MW01 03/06/2012
Chemical Name	CAS#	New York State Class GA							
Ethane	74-84-0								310
Ethene	74-85-1								700
Methane	74-82-8								1400
<b>Semi-Volatile Organic Compounds - SW8270</b>									
1,1'-Biphenyl	92-52-4	5		10 U	10 U	10 U	10 U	10 U	
2,4,5-Trichlorophenol	95-95-4			20 U	20 U	20 U	20 U	20 U	
2,4,6-Trichlorophenol	88-06-2			10 U	10 U	10 U	10 U	10 U	
2,4-Dichlorophenol	120-83-2	1		10 U	10 U	10 U	10 U	10 U	
2,4-Dimethylphenol	105-67-9	1		10 U	10 U	10 U	10 U	10 U	
2,4-Dinitrophenol	51-28-5	1		20 UJ	20 UJ	20 UJ	20 UJ	20 UJ	
2,4-Dinitrotoluene	121-14-2	5		10 U	10 U	10 U	10 U	10 U	
2,6-Dinitrotoluene	606-20-2	5		10 U	10 U	10 U	10 U	10 U	
2-Chloronaphthalene	91-58-7			10 U	10 U	10 U	10 U	10 U	
2-Chlorophenol	95-57-8			10 U	10 U	10 U	10 U	10 U	
2-Methylnaphthalene	91-57-6			10 UJ	10 UJ	10 UJ	10 UJ	10 UJ	
2-Methylphenol	95-48-7			10 U	10 U	10 U	10 U	10 U	
2-Nitroaniline	88-74-4	5		20 U	20 U	20 U	20 U	20 U	
2-Nitrophenol	88-75-5			10 U	10 U	10 U	10 U	10 U	
3,3'-Dichlorobenzidine	91-94-1	5		10 U	10 U	10 U	10 U	10 U	
3-Nitroaniline	99-09-2	5		20 U	20 U	20 U	20 U	20 U	
4,6-Dinitro-2-Methylphenol	534-52-1			20 U	20 U	20 U	20 U	20 U	
4-Bromophenyl-Phenylether	101-55-3			10 U	10 U	10 U	10 U	10 U	
4-Chloro-3-Methylphenol	59-50-7			10 U	10 U	10 U	10 U	10 U	
4-Chloroaniline	106-47-8	5		10 U	10 U	10 U	10 U	10 U	
4-Chlorophenyl-Phenylether	7005-72-3			10 U	10 U	10 U	10 U	10 U	
4-Methylphenol	106-44-5			10 U	10 U	10 U	10 U	10 U	
4-Nitroaniline	100-01-6	5		20 U	20 U	20 U	20 U	20 U	
4-Nitrophenol	100-02-7			20 U	20 U	20 U	20 U	20 U	
Acenaphthene	83-32-9			10 U	10 U	10 U	10 U	10 U	
Acenaphthylene	208-96-8			10 U	10 U	10 U	10 U	10 U	
Acetophenone	98-86-2			10 U	10 U	10 U	10 U	10 U	
Anthracene	120-12-7			10 U	10 U	10 U	10 U	10 U	
Atrazine	1912-24-9	7.5		10 U	10 U	10 U	10 U	10 U	
Benzaldehyde	100-52-7			10 U	10 U	10 U	10 U	10 U	
Benzo(a)Anthracene	56-55-3			10 U	10 U	10 U	10 U	10 U	
Benzo(a)Pyrene	50-32-8	0		10 U	10 U	10 U	10 U	10 U	
Benzo(b)Fluoranthene	205-99-2			10 U	10 U	10 U	10 U	10 U	

**Appendix C-1**  
**Groundwater Sample Results**  
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**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-GW-MIP5-15 MIP5 05/24/2011	80230-GW-MIP5-15-DUP MIP5 05/24/2011	80230-GW-MIP5-28 MIP5 05/24/2011	80230-GW-MIP6-15 MIP6 05/23/2011	80230-GW-MIP6-33.5 MIP6 05/23/2011	80230-MW01-030612 MW01 03/06/2012
Chemical Name	CAS#	New York State Class GA							
Benzo(g,h,i)Perylene	191-24-2			10 U		10 U		10 U	
Benzo(k)Fluoranthene	207-08-9			10 U		10 U		10 U	
bis(2-Chloroethoxy)Methane	111-91-1	5		10 U		10 U		10 U	
bis(2-Chloroethyl)Ether	111-44-4	1		10 U		10 U		10 U	
Bis(2-Ethylhexyl)Phthalate	117-81-7	5		10 U		1.7 J		10 U	
bis-Chloroisopropyl ether	108-60-1	5		10 U		10 U		10 U	
Butylbenzylphthalate	85-68-7			10 U		10 U		10 U	
Caprolactam	105-60-2			10 UJ		10 UJ		10 UJ	
Carbazole	86-74-8			10 U		10 U		10 U	
Chrysene	218-01-9			10 U		10 U		10 U	
Dibenzo(a,h)Anthracene	53-70-3			10 U		10 U		10 U	
Dibenzofuran	132-64-9			10 U		10 U		10 U	
Diethylphthalate	84-66-2			1.1 J		10 U		10 U	
Dimethylphthalate	131-11-3			10 U		10 U		10 U	
di-n-butylphthalate	84-74-2	50		10 U		10 U		10 U	
di-n-octylphthalate	117-84-0			10 U		10 U		10 U	
Fluoranthene	206-44-0			10 U		10 U		10 U	
Fluorene	86-73-7			10 U		10 U		10 U	
Hexachlorobenzene	118-74-1	0.04		10 U		10 U		10 U	
Hexachlorobutadiene	87-68-3	0.5		10 U		10 U		10 U	
Hexachlorocyclopentadiene	77-47-4	5		10 U		10 U		10 U	
Hexachloroethane	67-72-1	5		10 U		10 U		10 U	
Indeno(1,2,3-cd)Pyrene	193-39-5			10 U		10 U		10 U	
Isophorone	78-59-1			10 U		10 U		10 U	
Naphthalene	91-20-3			10 U		10 U		10 U	
Nitrobenzene	99-95-3	0.4		10 U		10 U		10 U	
n-Nitroso-di-n-Propylamine	621-64-7			10 U		10 U		10 U	
n-Nitrosodiphenylamine	86-30-6			10 U		10 U		10 U	
Pentachlorophenol	87-86-5	1		20 U		20 U		20 U	
Phenanthrene	85-01-8			10 U		10 U		10 U	
Phenol	108-95-2	1		10 U		10 U		10 U	
Pyrene	129-00-0			10 U		10 U		10 U	
<b>Inorganics - SW6010</b>									
Aluminum	7429-90-5			516		489		492	
Antimony	7440-36-0	3		20 U		20 U		20 U	
Arsenic	7440-38-2	25		20 U		20 U		20 U	
Barium	7440-39-3	1000		81 B		79.2 B		92.6 BEJ	
								153 BEJ	
								114 BEJ	



**Appendix C-1**  
**Groundwater Sample Results**  
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			Sample ID MW02 03/06/2012	80230-MW03-030712 MW03 03/07/2012	80230-MW04-030612 MW04 03/06/2012	80230-MW08S-030712 MW08S 03/07/2012	80230-MW09D-030512 MW09D 03/05/2012	
Chemical Name	CAS#	New York State Class GA						
<b>Volatile Organic Compounds - SW8260</b>								
1,1,1,2-Tetrachloroethane	630-20-6	5		5 U	5 U	5 U	5 U	5 U
1,1,1-Trichloroethane	71-55-6	5		5 U	5 U	5 U	5 U	5 U
1,1,2,2-Tetrachloroethane	79-34-5	5		5 U	5 U	5 U	5 U	5 U
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1	5		5 UJ	5 UJ	5 UJ	5 UJ	5 UJ
1,1,2-Trichloroethane	79-00-5	1		5 U	5 U	5 U	1.1 J	5 U
1,1-Dichloroethane	75-34-3	5		5 U	5 U	5 U	5 U	5 U
1,1-Dichloroethene	75-35-4	5		5 U	5 U	5 U	5 U	5 U
1,1-Dichloropropene	563-58-6			5 U	5 U	5 U	5 U	5 U
1,2,3-Trichlorobenzene	87-61-6	5		5 UJ	5 U	5 U	5 UJ	5 UJ
1,2,3-Trichloropropane	96-18-4	0.04		5 U	5 U	5 U	5 U	5 U
1,2,4-Trichlorobenzene	120-82-1	5		5 U	5 U	5 U	5 U	5 U
1,2,4-Trimethylbenzene	95-63-6	5		5 U	5 U	5 U	5 U	5 U
1,2-Dibromo-3-Chloropropane	96-12-8	0.04		5 UJ	5 U	5 U	5 UJ	5 UJ
1,2-Dibromoethane (EDB)	106-93-4	0.0006		5 U	5 U	5 U	5 U	5 U
1,2-Dichlorobenzene	95-50-1	3		5 U	5 U	5 U	5 U	5 U
1,2-Dichloroethane	107-06-2	0.6		5 U	5 U	5 U	5 U	5 U
1,2-Dichloropropane	78-87-5	1		5 U	5 U	5 U	5 U	5 U
1,3,5-Trimethylbenzene	108-67-8	5		5 U	5 U	5 U	5 U	5 U
1,3-Dichlorobenzene	541-73-1	3		5 U	5 U	5 U	5 U	5 U
1,3-Dichloropropane	142-28-9	5		5 U	5 U	5 U	5 U	5 U
1,4-Dichlorobenzene	106-46-7	3		5 U	5 U	5 U	5 U	5 U
2,2-Dichloropropane	594-20-7	5		5 UJ	5 UJ	5 UJ	5 UJ	5 UJ
2-Butanone (MEK)	78-93-3			5 R	5 R	5 R	5 R	5 R
2-Chlorotoluene	95-49-8	5		5 U	5 U	5 U	5 U	5 U
2-Hexanone	591-78-6			5 UJ	5 UJ	5 UJ	5 UJ	5 UJ
4-Chlorotoluene	106-43-4	5		5 U	5 U	5 U	5 U	5 U
4-Methyl-2-Pentanone (MIBK)	108-10-1			5 U	5 U	5 U	5 U	5 U
Acetone	67-64-1			5 R	5 R	5 R	5 R	5 R
Benzene	71-43-2	1		5 U	5 U	5 U	5 U	5 U
Bromobenzene	108-86-1	5		5 U	5 U	5 U	5 U	5 U
Bromochloromethane	74-97-5	5		5 U	5 U	5 U	5 U	5 U
Bromodichloromethane	75-27-4			5 U	5 U	5 U	5 U	5 U
Bromoform	75-25-2			5 U	5 U	5 U	5 U	5 U
Bromomethane	74-83-9	5		5 U	5 U	5 U	5 U	5 U
Carbon Disulfide	75-15-0	60		5 U	5 U	5 U	5 U	5 U
Carbon Tetrachloride	56-23-5	5		5 U	5 U	5 U	5 U	5 U

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID			80230-MW02-030612	MW02	80230-MW03-030712	MW03	80230-MW04-030612	MW04	80230-MW08S-030712	MW08S	80230-MW09D-030512
Chemical Name	CAS#	New York State Class GA		03/06/2012		03/07/2012		03/06/2012		03/07/2012	
Chlorobenzene	108-90-7	5		5 U		5 U		5 U		5 U	
Chloroethane	75-00-3	5		5 UJ		5 UJ		5 UJ		5 UJ	
Chloroform	67-66-3	7		5 U		5 U		5 U	0.81 J	5 U	
Chloromethane	74-87-3	5	0.58 J		5 UJ		5 UJ		5 UJ		0.92 J
cis-1,2-Dichloroethene	156-59-2	5	22		38		5 U		26		5 U
cis-1,3-Dichloropropene	10061-01-5			5 U		5 U		5 U		5 U	
Cyclohexane	110-82-7			5 UJ		5 UJ		5 UJ		5 UJ	
Dibromochloromethane	124-48-1			5 U		5 U		5 U		5 U	
Dibromomethane	74-95-3	5		5 U		5 U		5 U		5 U	
Dichlorodifluoromethane	75-71-8	5		5 UJ		5 UJ		5 UJ		5 UJ	
Ethylbenzene	100-41-4	5		5 U		5 U		5 U		5 U	
Hexachlorobutadiene	87-68-3	0.5		5 U		5 U		5 U		5 U	
Isopropylbenzene	98-82-8	5		5 U		5 U		5 U		5 U	
m,p-Xylene	179601-23-1			5 U		5 U		5 U		5 U	
Methyl Acetate	79-20-9			5 UJ		5 UJ		5 UJ		5 UJ	
Methyl Iodide	74-88-4	5		5 UJ		5 U		5 U		5 UJ	
Methyl Tert-Butyl Ether (MTBE)	1634-04-4			5 U		5 U		5 U		5 U	
Methylcyclohexane	108-87-2			5 UJ		5 UJ		5 UJ		5 UJ	
Methylene Chloride	75-09-2	5		5 U		5 U		5 U		5 U	
Naphthalene	91-20-3			5 UJ		5 UJ		5 UJ		5 UJ	
n-Butylbenzene	104-51-8	5		5 U		5 U		5 U		5 U	
n-Propylbenzene	103-65-1	5		5 U		5 U		5 U		5 U	
o-Xylene	95-47-6	5		5 U		5 U		5 U		5 U	
p-Isopropyltoluene	99-87-6	5		5 U		5 U		5 U		5 U	
sec-Butylbenzene	135-98-8	5		5 U		5 U		5 U		5 U	
Styrene	100-42-5	5		5 U		5 U		5 U		5 U	
tert-Butylbenzene	98-06-6	5		5 U		5 U		5 U		5 U	
Tetrachloroethene	127-18-4	5		5 U		5 U		5 U		1300 D	
Toluene	108-88-3	5		5 U		5 U		5 U		5 U	
Total Xylenes	1330-20-7			5 U		5 U		5 U		5 U	
trans-1,2-Dichloroethene	156-60-5	5		5 U		5 U		5 U		5 U	
trans-1,3-Dichloropropene	10061-02-6			5 U		5 U		5 U		5 U	
Trichloroethene	79-01-6	5		5 U		5 U		5 U		10	
Trichlorofluoromethane	75-69-4	5		5 U		5 U		5 U		5 U	
Vinyl Acetate	108-05-4			5 U		5 U		5 U		5 U	
Vinyl Chloride	75-01-4	2		11 J		5 UJ		2.8 J		5 UJ	

**Appendix C-1**  
**Groundwater Sample Results**  
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**Appendix C-1**  
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**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
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			Sample ID 80230-MW02-030612	MW02 03/06/2012	Sample Location MW03 Sampling Date 03/07/2012	80230-MW04-030612	MW04 03/06/2012	80230-MW08S-030712	MW08S 03/07/2012	80230-MW09D-030512	MW09D 03/05/2012
Chemical Name	CAS#	New York State Class GA									
Beryllium	7440-41-7										
Cadmium	7440-43-9	5									
Calcium	7440-70-2										
Chromium	7440-47-3	50									
Cobalt	7440-48-4										
Copper	7440-50-8	200									
Iron	7439-89-6	300									
Lead	7439-92-1	25									
Magnesium	7439-95-4										
Manganese	7439-96-5	300									
Mercury	7439-97-6	0.7									
Nickel	7440-02-0	100									
Potassium	7440-09-7										
Selenium	7782-49-2	10									
Silver	7440-22-4	50									
Sodium	7440-23-5										
Thallium	7440-28-0										
Vanadium	7440-62-2										
Zinc	7440-66-6										
<b>Wet Chemistry (mg/L)</b>											
Sulfide - 4500-S F	18496-25-8		0.03 U		0.056		0.03 U		0.048		0.03 U
Alkalinity, Total (as CaCO <sub>3</sub> ) - A2320	ALK		300		600		420		170		210
Chloride - E300.0	16887-00-6	250	230		130		430		71		90
Nitrate-NO <sub>3</sub> - E300.0	14797-55-8	10	0.65		0.13 U		0.13 U		5.1		5.8
Nitrite-NO <sub>2</sub> - E300.0	14797-65-0	1	0.13 U		0.13 U		0.13 U		0.13 U		0.13 U
Sulfate - E300.0	14808-79-8		57		2.1 J		15		35		45
Hardness As CaCO <sub>3</sub> - SM2340B	CACOA-H		300		440		230		230		320
Total Dissolved Solids - SM2540C	TDS		800		910		1000		360		370
Total Suspended Solids - SM2540D	TSS		40		80		21		95		68
Ammonia as N - SM4500	7664-41-7	2	1.5 R		0.2 R		0.2 R		0.2 R		0.2 R
Nitrogen, Kjeldahl - SM4500-NORGC	KN		3.8 R		0.57 R		0.86 R		0.2 R		0.2 R
Total Organic Carbon - SM5310B	TOC		12		45		11		10 U		10 U

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
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			Sample ID MW09S 03/05/2012	80230-MW10D-030512 MW10D 03/05/2012	80230-MW10S-030512 MW10S 03/05/2012	80230-MW11D-030812 MW11D 03/08/2011	80230-MW11S-030812 MW11S 03/08/2011	
Chemical Name	CAS#	New York State Class GA						
<b>Volatile Organic Compounds - SW8260</b>								
1,1,1,2-Tetrachloroethane	630-20-6	5	5 U	5 U	5 U	5 U	5 U	
1,1,1-Trichloroethane	71-55-6	5	5 U	5 U	5 U	5 U	5 U	
1,1,2,2-Tetrachloroethane	79-34-5	5	5 U	5 U	5 U	5 U	5 U	
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1	5	5 UJ	5 U	5 UJ	5 UJ	5 UJ	
1,1,2-Trichloroethane	79-00-5	1	5 U	5 U	5 U	5 U	5 U	
1,1-Dichloroethane	75-34-3	5	5 U	5 U	5 U	5 U	5 U	
1,1-Dichloroethene	75-35-4	5	0.65 J	5 U	5 U	5 U	15 U	
1,1-Dichloropropene	563-58-6		5 U	5 U	5 U	5 U	5 U	
1,2,3-Trichlorobenzene	87-61-6	5	5 UJ	5 UJ	5 UJ	5 U	5 U	
1,2,3-Trichloropropane	96-18-4	0.04	5 U	5 U	5 U	5 U	5 U	
1,2,4-Trichlorobenzene	120-82-1	5	5 U	5 U	5 U	5 U	5 U	
1,2,4-Trimethylbenzene	95-63-6	5	5 U	5 U	5 U	5 U	5 U	
1,2-Dibromo-3-Chloropropane	96-12-8	0.04	5 UJ	5 UJ	5 UJ	5 U	5 U	
1,2-Dibromoethane (EDB)	106-93-4	0.0006	5 U	5 U	5 U	5 U	5 U	
1,2-Dichlorobenzene	95-50-1	3	5 U	5 U	5 U	5 U	2.2 J	
1,2-Dichloroethane	107-06-2	0.6	5 U	5 U	5 U	5 U	5 U	
1,2-Dichloropropane	78-87-5	1	5 U	5 U	5 U	5 U	5 U	
1,3,5-Trimethylbenzene	108-67-8	5	5 U	5 U	5 U	5 U	5 U	
1,3-Dichlorobenzene	541-73-1	3	5 U	5 U	5 U	5 U	5 U	
1,3-Dichloropropane	142-28-9	5	5 U	5 U	5 U	5 U	5 U	
1,4-Dichlorobenzene	106-46-7	3	5 U	5 U	5 U	5 U	5 U	
2,2-Dichloropropane	594-20-7	5	5 UJ	5 U	5 UJ	5 UJ	5 UJ	
2-Butanone (MEK)	78-93-3		5 R	5 R	5 R	5 R	5 R	
2-Chlorotoluene	95-49-8	5	5 U	5 U	5 U	5 U	5 U	
2-Hexanone	591-78-6		5 UJ	5 U	5 UJ	5 UJ	5 UJ	
4-Chlorotoluene	106-43-4	5	5 U	5 U	5 U	5 U	5 U	
4-Methyl-2-Pentanone (MIBK)	108-10-1		5 U	5 U	5 U	5 U	5 U	
Acetone	67-64-1		5 R	5 R	5 R	5 R	5 R	
Benzene	71-43-2	1	5 U	5 U	5 U	5 U	5 U	
Bromobenzene	108-86-1	5	5 U	5 U	5 U	5 U	5 U	
Bromochloromethane	74-97-5	5	5 U	5 U	5 U	5 U	5 U	
Bromodichloromethane	75-27-4		5 U	5 U	5 U	5 U	5 U	
Bromoform	75-25-2		5 U	5 U	5 U	5 U	5 U	
Bromomethane	74-83-9	5	5 U	5 U	5 U	5 U	5 U	
Carbon Disulfide	75-15-0	60	5 U	5 U	5 U	5 U	5 U	
Carbon Tetrachloride	56-23-5	5	5 U	5 U	5 U	5 U	5 U	

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**Groundwater Sample Results**  
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			Sample ID MW09S 03/05/2012	80230-MW10D-030512 MW10D 03/05/2012	80230-MW10S-030512 MW10S 03/05/2012	80230-MW11D-030812 MW11D 03/08/2011	80230-MW11S-030812 MW11S 03/08/2011
Chemical Name	CAS#	New York State Class GA					
Chlorobenzene	108-90-7	5		5 U	5 U	5 U	2.6 J
Chloroethane	75-00-3	5		5 UJ	5 U	5 UJ	5 UJ
Chloroform	67-66-3	7	0.66 J	5 U	5 U	5 U	5 U
Chloromethane	74-87-3	5	5 UJ	5 U	5 UJ	5 UJ	5 UJ
cis-1,2-Dichloroethene	156-59-2	5	17	5 U	140	5 U	11000 D
cis-1,3-Dichloropropene	10061-01-5		5 U	5 U	5 U	5 U	5 U
Cyclohexane	110-82-7			5 UJ	5 U	5 UJ	5 UJ
Dibromochloromethane	124-48-1			5 U	5 U	5 U	5 U
Dibromomethane	74-95-3	5	5 U	5 U	5 U	5 U	5 U
Dichlorodifluoromethane	75-71-8	5	5 UJ	5 U	5 UJ	5 UJ	5 UJ
Ethylbenzene	100-41-4	5	5 U	5 U	5 U	5 U	5 U
Hexachlorobutadiene	87-68-3	0.5	5 U	5 U	5 U	5 U	5 U
Isopropylbenzene	98-82-8	5	5 U	5 U	5 U	5 U	5 U
m,p-Xylene	179601-23-1			5 U	5 U	5 U	5 U
Methyl Acetate	79-20-9			5 UJ	5 U	5 UJ	5 UJ
Methyl Iodide	74-88-4	5	5 UJ	5 UJ	5 UJ	5 U	5 U
Methyl Tert-Butyl Ether (MTBE)	1634-04-4			5 U	5 U	0.76 J	5 U
Methylcyclohexane	108-87-2			5 UJ	5 UJ	5 UJ	5 UJ
Methylene Chloride	75-09-2	5	5 U	5 U	5 U	5 U	5 U
Naphthalene	91-20-3			5 UJ	5 UJ	5 UJ	5 UJ
n-Butylbenzene	104-51-8	5	5 U	5 U	5 U	5 U	5 U
n-Propylbenzene	103-65-1	5	5 U	5 U	5 U	5 U	5 U
o-Xylene	95-47-6	5	5 U	5 U	5 U	5 U	5 U
p-Isopropyltoluene	99-87-6	5	5 U	5 U	5 U	5 U	5 U
sec-Butylbenzene	135-98-8	5	5 U	5 U	5 U	5 U	5 U
Styrene	100-42-5	5	5 U	5 U	5 U	5 U	5 U
tert-Butylbenzene	98-06-6	5	5 U	5 U	5 U	5 U	5 U
Tetrachloroethene	127-18-4	5	34	5 U	6.1	5 U	9000 D
Toluene	108-88-3	5	5 U	5 U	5 U	5 U	5 U
Total Xylenes	1330-20-7			5 U	5 U	5 U	5 U
trans-1,2-Dichloroethene	156-60-5	5	5 U	5 U	5 U	5 U	8.9
trans-1,3-Dichloropropene	10061-02-6			5 U	5 U	5 U	5 U
Trichloroethene	79-01-6	5	16	5 U	2.6 J	5 U	2400 D
Trichlorofluoromethane	75-69-4	5	5 U	5 U	5 U	5 U	5 U
Vinyl Acetate	108-05-4			5 U	5 U	5 U	5 U
Vinyl Chloride	75-01-4	2	0.58 J	5 U	1.3 J	5 UJ	37 J

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID MW09S 03/05/2012	80230-MW10D-030512 MW10D 03/05/2012	80230-MW10S-030512 MW10S 03/05/2012	80230-MW11D-030812 MW11D 03/08/2011	80230-MW11S-030812 MW11S 03/08/2011
Chemical Name	CAS#	New York State Class GA					
Ethane	74-84-0		1.2 U	1.2 U	3.2	1.3 U	10
Ethene	74-85-1		1.5 U	1.5 U	1.5 U	1.6 U	1.6 U
Methane	74-82-8		2.1	0.6 U	45	0.61 U	170
<b>Semi-Volatile Organic Compounds - SW8270</b>							
1,1'-Biphenyl	92-52-4	5					
2,4,5-Trichlorophenol	95-95-4						
2,4,6-Trichlorophenol	88-06-2						
2,4-Dichlorophenol	120-83-2	1					
2,4-Dimethylphenol	105-67-9	1					
2,4-Dinitrophenol	51-28-5	1					
2,4-Dinitrotoluene	121-14-2	5					
2,6-Dinitrotoluene	606-20-2	5					
2-Chloronaphthalene	91-58-7						
2-Chlorophenol	95-57-8						
2-Methylnaphthalene	91-57-6						
2-Methylphenol	95-48-7						
2-Nitroaniline	88-74-4	5					
2-Nitrophenol	88-75-5						
3,3'-Dichlorobenzidine	91-94-1	5					
3-Nitroaniline	99-09-2	5					
4,6-Dinitro-2-Methylphenol	534-52-1						
4-Bromophenyl-Phenylether	101-55-3						
4-Chloro-3-Methylphenol	59-50-7						
4-Chloroaniline	106-47-8	5					
4-Chlorophenyl-Phenylether	7005-72-3						
4-Methylphenol	106-44-5						
4-Nitroaniline	100-01-6	5					
4-Nitrophenol	100-02-7						
Acenaphthene	83-32-9						
Acenaphthylene	208-96-8						
Acetophenone	98-86-2						
Anthracene	120-12-7						
Atrazine	1912-24-9	7.5					
Benzaldehyde	100-52-7						
Benzo(a)Anthracene	56-55-3						
Benzo(a)Pyrene	50-32-8	0					
Benzo(b)Fluoranthene	205-99-2						

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MW09S-030512 MW09S 03/05/2012	80230-MW10D-030512 MW10D 03/05/2012	80230-MW10S-030512 MW10S 03/05/2012	80230-MW11D-030812 MW11D 03/08/2011	80230-MW11S-030812 MW11S 03/08/2011
Chemical Name	CAS#	New York State Class GA						
Benzo(g,h,i)Perylene	191-24-2							
Benzo(k)Fluoranthene	207-08-9							
bis(2-Chloroethoxy)Methane	111-91-1	5						
bis(2-Chloroethyl)Ether	111-44-4	1						
Bis(2-Ethylhexyl)Phthalate	117-81-7	5						
bis-Chloroisopropyl ether	108-60-1	5						
Butylbenzylphthalate	85-68-7							
Caprolactam	105-60-2							
Carbazole	86-74-8							
Chrysene	218-01-9							
Dibenzo(a,h)Anthracene	53-70-3							
Dibenzofuran	132-64-9							
Diethylphthalate	84-66-2							
Dimethylphthalate	131-11-3							
di-n-butylphthalate	84-74-2	50						
di-n-octylphthalate	117-84-0							
Fluoranthene	206-44-0							
Fluorene	86-73-7							
Hexachlorobenzene	118-74-1	0.04						
Hexachlorobutadiene	87-68-3	0.5						
Hexachlorocyclopentadiene	77-47-4	5						
Hexachloroethane	67-72-1	5						
Indeno(1,2,3-cd)Pyrene	193-39-5							
Isophorone	78-59-1							
Naphthalene	91-20-3							
Nitrobenzene	98-95-3	0.4						
n-Nitroso-di-n-Propylamine	621-64-7							
n-Nitrosodiphenylamine	86-30-6							
Pentachlorophenol	87-86-5	1						
Phenanthrene	85-01-8							
Phenol	108-95-2	1						
Pyrene	129-00-0							
<b>Inorganics - SW6010</b>								
Aluminum	7429-90-5							
Antimony	7440-36-0	3						
Arsenic	7440-38-2	25						
Barium	7440-39-3	1000						

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID 80230-MW09S-030512	MW09S 03/05/2012	80230-MW10D-030512	MW10D 03/05/2012	80230-MW10S-030512	MW10S 03/05/2012	80230-MW11D-030812	MW11D 03/08/2011	80230-MW11S-030812	MW11S 03/08/2011
Chemical Name	CAS#	New York State Class GA										
Beryllium	7440-41-7											
Cadmium	7440-43-9	5										
Calcium	7440-70-2											
Chromium	7440-47-3	50										
Cobalt	7440-48-4											
Copper	7440-50-8	200										
Iron	7439-89-6	300										
Lead	7439-92-1	25										
Magnesium	7439-95-4											
Manganese	7439-96-5	300										
Mercury	7439-97-6	0.7										
Nickel	7440-02-0	100										
Potassium	7440-09-7											
Selenium	7782-49-2	10										
Silver	7440-22-4	50										
Sodium	7440-23-5											
Thallium	7440-28-0											
Vanadium	7440-62-2											
Zinc	7440-66-6											
<b>Wet Chemistry (mg/L)</b>												
Sulfide - 4500-S F	18496-25-8		0.083		0.03 U		0.046		0.096		0.095	
Alkalinity, Total (as CaCO <sub>3</sub> ) - A2320	ALK		230		130		1000		300		360	
Chloride - E300.0	16887-00-6	250	72		88		150		88		280	
Nitrate-NO <sub>3</sub> - E300.0	14797-55-8	10	3.3		4.2		0.13 U		4.5		0.32	
Nitrite-NO <sub>2</sub> - E300.0	14797-65-0	1	0.13 U		0.13 U		0.13 U		0.13 U		0.13 U	
Sulfate - E300.0	14808-79-8		44		48		61		47		94	
Hardness As CaCO <sub>3</sub> - SM2340B	CACOA-H		320		330		620		390		550	
Total Dissolved Solids - SM2540C	TDS		390		420		690		590		1000	
Total Suspended Solids - SM2540D	TSS		540		87		230		420		190	
Ammonia as N - SM4500	7664-41-7	2	0.2 R		0.2 R		0.2 R		0.2 R		0.2 R	
Nitrogen, Kjeldahl - SM4500-NORGC	KN		0.28 R		0.2 R		0.2 R		0.2 R		0.2 R	
Total Organic Carbon - SM5310B	TOC		10 U		10 U		5.7 J		10 U		2.9 J	

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID 80230-MW12S-030812	MW12S 03/08/2012	80230-MW12S-030812-DUP		MW12S 03/08/2012	80230-MW13D-030712		MW13D 03/07/2012	80230-MW13S-030712		MW13S 03/07/2012	80230-MW14S-030812		
Chemical Name	CAS#	New York State Class GA														
<b>Volatile Organic Compounds - SW8260</b>																
1,1,1,2-Tetrachloroethane	630-20-6	5		500 U			500 U		5 U		5 U		500 U			
1,1,1-Trichloroethane	71-55-6	5		500 U			500 U		5 U		5 U		500 U			
1,1,2,2-Tetrachloroethane	79-34-5	5		500 U			500 U		5 U		5 U		500 U			
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1	5		500 UJ			500 UJ		5 UJ		5 UJ		500 UJ			
1,1,2-Trichloroethane	79-00-5	1		500 U			500 U		5 U		5 U		500 U			
1,1-Dichloroethane	75-34-3	5		500 U			500 U		5 U		5 U		500 U			
1,1-Dichloroethene	75-35-4	5		500 U			500 U		5 U		9.4 U		500 U			
1,1-Dichloropropene	563-58-6			500 U			500 U		5 U		5 U		500 U			
1,2,3-Trichlorobenzene	87-61-6	5		500 U			500 U		5 U		5 UJ		500 U			
1,2,3-Trichloropropane	96-18-4	0.04		500 U			500 U		5 U		5 U		500 U			
1,2,4-Trichlorobenzene	120-82-1	5		500 U			500 U		5 U		5 U		500 U			
1,2,4-Trimethylbenzene	95-63-6	5		500 U			500 U		5 U		5 U		500 U			
1,2-Dibromo-3-Chloropropane	96-12-8	0.04		500 U			500 U		5 U		5 UJ		500 U			
1,2-Dibromoethane (EDB)	106-93-4	0.0006		500 U			500 U		5 U		5 U		500 U			
1,2-Dichlorobenzene	95-50-1	3		500 U			500 U		5 U		1.1 J		500 U			
1,2-Dichloroethane	107-06-2	0.6		500 U			500 U		5 U		5 U		500 U			
1,2-Dichloropropane	78-87-5	1		500 U			500 U		5 U		5 U		500 U			
1,3,5-Trimethylbenzene	108-67-8	5		500 U			500 U		5 U		5 U		500 U			
1,3-Dichlorobenzene	541-73-1	3		500 U			500 U		5 U		5 U		500 U			
1,3-Dichloropropane	142-28-9	5		500 U			500 U		5 U		5 U		500 U			
1,4-Dichlorobenzene	106-46-7	3		500 U			500 U		5 U		0.92 J		500 U			
2,2-Dichloropropane	594-20-7	5		500 UJ			500 UJ		5 UJ		5 UJ		500 UJ			
2-Butanone (MEK)	78-93-3			500 R			500 R		5 R		5 R		500 R			
2-Chlorotoluene	95-49-8	5		500 U			500 U		5 U		5 U		500 U			
2-Hexanone	591-78-6			500 UJ			500 UJ		5 UJ		5 UJ		500 UJ			
4-Chlorotoluene	106-43-4	5		500 U			500 U		5 U		5 U		500 U			
4-Methyl-2-Pentanone (MIBK)	108-10-1			500 U			500 U		5 U		5 U		500 U			
Acetone	67-64-1			500 R			500 R		5 R		5 R		500 R			
Benzene	71-43-2	1		500 U			500 U		5 U		5 U		500 U			
Bromobenzene	108-86-1	5		500 U			500 U		5 U		5 U		500 U			
Bromochloromethane	74-97-5	5		500 U			500 U		5 U		5 U		500 U			
Bromodichloromethane	75-27-4			500 U			500 U		5 U		5 U		500 U			
Bromoform	75-25-2			500 U			500 U		5 U		5 U		500 U			
Bromomethane	74-83-9	5		500 U			500 U		5 U		5 U		500 U			
Carbon Disulfide	75-15-0	60		500 U			500 U		5 U		5 U		500 U			
Carbon Tetrachloride	56-23-5	5		500 U			500 U		5 U		5 U		500 U			

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID			80230-MW12S-030812	80230-MW12S-030812-DUP		80230-MW13D-030712		80230-MW13S-030712		80230-MW14S-030812	
Chemical Name	CAS#	New York State Class GA	MW12S 03/08/2012	MW12S 03/08/2012	MW13D 03/07/2012	MW13S 03/07/2012	MW14S 03/08/2012				
Chlorobenzene	108-90-7	5	500 U	500 U	5 U	3.1 J	500 U				
Chloroethane	75-00-3	5	500 UJ	500 UJ	5 UJ	5 UJ	500 U				
Chloroform	67-66-3	7	500 U	500 U	5 U	5 U	500 U				
Chloromethane	74-87-3	5	500 UJ	500 UJ	5 UJ	5 UJ	500 U				
cis-1,2-Dichloroethene	156-59-2	5	350 J	380 J	5 U	7000 D	850				
cis-1,3-Dichloropropene	10061-01-5		500 U	500 U	5 U	5 U	500 U				
Cyclohexane	110-82-7		500 UJ	500 UJ	5 UJ	5 UJ	500 U				
Dibromochloromethane	124-48-1		500 U	500 U	5 U	5 U	500 U				
Dibromomethane	74-95-3	5	500 U	500 U	5 U	5 U	500 U				
Dichlorodifluoromethane	75-71-8	5	500 UJ	500 UJ	5 UJ	5 UJ	500 U				
Ethylbenzene	100-41-4	5	500 U	500 U	5 U	5 U	500 U				
Hexachlorobutadiene	87-68-3	0.5	500 U	500 U	5 U	5 U	500 U				
Isopropylbenzene	98-82-8	5	500 U	500 U	5 U	5 U	500 U				
m,p-Xylene	179601-23-1		500 U	500 U	5 U	5 U	500 U				
Methyl Acetate	79-20-9		500 UJ	500 UJ	5 UJ	5 UJ	500 U				
Methyl Iodide	74-88-4	5	500 U	500 U	5 U	5 UJ	500 U				
Methyl Tert-Butyl Ether (MTBE)	1634-04-4		500 U	500 U	5 U	3.1 J	500 U				
Methylcyclohexane	108-87-2		500 UJ	500 UJ	5 UJ	5 UJ	500 U				
Methylene Chloride	75-09-2	5	500 U	500 U	5 U	5 U	500 U				
Naphthalene	91-20-3		500 UJ	500 UJ	5 UJ	5 UJ	500 U				
n-Butylbenzene	104-51-8	5	500 U	500 U	5 U	5 U	500 U				
n-Propylbenzene	103-65-1	5	500 U	500 U	5 U	5 U	500 U				
o-Xylene	95-47-6	5	500 U	500 U	5 U	5 U	500 U				
p-Isopropyltoluene	99-87-6	5	500 U	500 U	5 U	5 U	500 U				
sec-Butylbenzene	135-98-8	5	500 U	500 U	5 U	5 U	500 U				
Styrene	100-42-5	5	500 U	500 U	5 U	5 U	500 U				
tert-Butylbenzene	98-06-6	5	500 U	500 U	5 U	5 U	500 U				
Tetrachloroethene	127-18-4	5	71000 D	72000 D	5 U	2500 D	100000 D				
Toluene	108-88-3	5	500 U	500 U	5 U	5 U	500 U				
Total Xylenes	1330-20-7		500 U	500 U	5 U	5 U	500 U				
trans-1,2-Dichloroethene	156-60-5	5	500 U	500 U	5 U	160	500 U				
trans-1,3-Dichloropropene	10061-02-6		500 U	500 U	5 U	5 U	500 U				
Trichloroethene	79-01-6	5	500 U	500 U	5 U	950 D	1300				
Trichlorofluoromethane	75-69-4	5	500 U	500 U	5 U	5 U	500 U				
Vinyl Acetate	108-05-4		500 U	500 U	5 U	5 U	500 U				
Vinyl Chloride	75-01-4	2	500 UJ	500 UJ	5 UJ	210 JD	500 U				

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**



**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID 80230-MW12S-030812	MW12S 03/08/2012	80230-MW12S-030812-DUP	MW12S 03/08/2012	80230-MW13D-030712	MW13D 03/07/2012	80230-MW13S-030712	MW13S 03/07/2012	80230-MW14S-030812	MW14S 03/08/2012
Chemical Name	CAS#	New York State Class GA										
Beryllium	7440-41-7											
Cadmium	7440-43-9	5										
Calcium	7440-70-2											
Chromium	7440-47-3	50										
Cobalt	7440-48-4											
Copper	7440-50-8	200										
Iron	7439-89-6	300										
Lead	7439-92-1	25										
Magnesium	7439-95-4											
Manganese	7439-96-5	300										
Mercury	7439-97-6	0.7										
Nickel	7440-02-0	100										
Potassium	7440-09-7											
Selenium	7782-49-2	10										
Silver	7440-22-4	50										
Sodium	7440-23-5											
Thallium	7440-28-0											
Vanadium	7440-62-2											
Zinc	7440-66-6											
<b>Wet Chemistry (mg/L)</b>												
Sulfide - 4500-S F	18496-25-8		0.034		0.033		0.03 U		0.03 U		0.1	
Alkalinity, Total (as CaCO <sub>3</sub> ) - A2320	ALK		200		210		330		270		350	
Chloride - E300.0	16887-00-6	250	98		97		91		250		290	
Nitrate-NO <sub>3</sub> - E300.0	14797-55-8	10	5.4		5.1		4.8		0.13 U		0.13 U	
Nitrite-NO <sub>2</sub> - E300.0	14797-65-0	1	0.12 J		0.13 J		0.13 U		0.13 U		0.13 U	
Sulfate - E300.0	14808-79-8		33		33		46		120		100	
Hardness As CaCO <sub>3</sub> - SM2340B	CACOA-H		290		300		340		560		760	
Total Dissolved Solids - SM2540C	TDS		570		590		520		730		1200	
Total Suspended Solids - SM2540D	TSS		120		100		190		24		180	
Ammonia as N - SM4500	7664-41-7	2	0.2 R		0.2 R		0.2 R		0.2 R		0.2 R	
Nitrogen, Kjeldahl - SM4500-NORGC	KN		0.2 R		0.2 R		0.2 R		0.2 R		0.2 R	
Total Organic Carbon - SM5310B	TOC		2 J		10 U		10 U		6.1 J		3.3 J	

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID 80230-MW15D-030712	MW15D 03/07/2012	Sample Location MW16S	Sampling Date 03/06/2012	PM-GWS-1A-110311	PM-GWS-1A-110311-DUP	PM-GWS-1A 11/03/2011	PM-GWS-1B-1122011
Chemical Name	CAS#	New York State Class GA								
<b>Volatile Organic Compounds - SW8260</b>										
1,1,1,2-Tetrachloroethane	630-20-6	5		5 U		5 U	5 U	5 U	5 U	
1,1,1-Trichloroethane	71-55-6	5		5 U		5 U	5 U	5 U	5 U	
1,1,2,2-Tetrachloroethane	79-34-5	5		5 U		5 U	5 U	5 U	5 U	
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1	5		5 UJ		5 UJ	5 U	5 U	5 U	
1,1,2-Trichloroethane	79-00-5	1		5 U		5 U	5 U	5 U	5 U	
1,1-Dichloroethane	75-34-3	5		5 U		5 U	5 U	5 U	5 U	
1,1-Dichloroethene	75-35-4	5		5 U		5 U	5 U	5 U	5 U	
1,1-Dichloropropene	563-58-6			5 U		5 U	5 U	5 U	5 U	
1,2,3-Trichlorobenzene	87-61-6	5		5 UJ		5 UJ	5 U	5 U	5 UJ	
1,2,3-Trichloropropane	96-18-4	0.04		5 U		5 U	5 U	5 U	5 UJ	
1,2,4-Trichlorobenzene	120-82-1	5		5 U		5 U	5 U	5 U	5 UJ	
1,2,4-Trimethylbenzene	95-63-6	5		5 U		5 U	5 U	5 U	5 U	
1,2-Dibromo-3-Chloropropane	96-12-8	0.04		5 UJ		5 UJ	5 U	5 U	5 U	
1,2-Dibromoethane (EDB)	106-93-4	0.0006		5 U		5 U	5 U	5 U	5 U	
1,2-Dichlorobenzene	95-50-1	3		5 U		5 U	5 U	5 U	5 U	
1,2-Dichloroethane	107-06-2	0.6		5 U		5 U	5 U	5 U	5 U	
1,2-Dichloropropane	78-87-5	1		5 U		5 U	5 U	5 U	5 U	
1,3,5-Trimethylbenzene	108-67-8	5		5 U		5 U	5 U	5 U	5 U	
1,3-Dichlorobenzene	541-73-1	3		5 U		5 U	5 U	5 U	5 U	
1,3-Dichloropropane	142-28-9	5		5 U		5 U	5 U	5 U	5 U	
1,4-Dichlorobenzene	106-46-7	3		5 U		5 U	5 U	5 U	5 U	
2,2-Dichloropropane	594-20-7	5		5 UJ		5 UJ	5 U	5 U	5 U	
2-Butanone (MEK)	78-93-3			5 R		5 R	5 U	5 U	5 R	
2-Chlorotoluene	95-49-8	5		5 U		5 U	5 U	5 U	5 U	
2-Hexanone	591-78-6			5 UJ		5 U	5 U	5 U	5 UJ	
4-Chlorotoluene	106-43-4	5		5 U		5 U	5 U	5 U	5 U	
4-Methyl-2-Pentanone (MIBK)	108-10-1			5 U		5 U	5 U	5 U	5 U	
Acetone	67-64-1			5 R		5 R	5 U	4.5 J	5 R	
Benzene	71-43-2	1		5 U		5 U	5 U	5 U	5 U	
Bromobenzene	108-86-1	5		5 U		5 U	5 U	5 U	5 U	
Bromochloromethane	74-97-5	5		5 U		5 U	5 U	5 U	5 U	
Bromodichloromethane	75-27-4			5 U		5 U	5 U	5 U	5 U	
Bromoform	75-25-2			5 U		5 U	5 U	5 U	5 U	
Bromomethane	74-83-9	5		5 U		5 U	5 U	5 U	5 UJ	
Carbon Disulfide	75-15-0	60		5 U		5 U	5 U	5 U	5 U	
Carbon Tetrachloride	56-23-5	5		5 U		5 U	5 U	5 U	5 U	

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Sample ID Sample Location Sampling Date			80230-MW15D-030712 MW15D 03/07/2012		80230-MW16S-030612 MW16S 03/06/2012		PM-GWS-1A-110311 PM-GWS-1A 11/03/2011		PM-GWS-1A-110311-DUP PM-GWS-1A 11/03/2011		PM-GWS-1B-1122011 PM-GWS-1B 11/02/2011	
Chemical Name	CAS#	New York State Class GA										
Chlorobenzene	108-90-7	5		5 U		5 U		5 U		5 U		
Chloroethane	75-00-3	5		5 UJ		5 UJ		5 U		5 U		
Chloroform	67-66-3	7		5 U		5 U		5 U		5 U		
Chloromethane	74-87-3	5		5 UJ		5 UJ		5 U		5 U		
cis-1,2-Dichloroethene	156-59-2	5		1.3 J		34		11		12		
cis-1,3-Dichloropropene	10061-01-5			5 U		5 U		5 U		5 U		
Cyclohexane	110-82-7			5 UJ		5 UJ		5 U		5 U		
Dibromochloromethane	124-48-1			5 U		5 U		5 U		5 U		
Dibromomethane	74-95-3	5		5 U		5 U		5 U		5 U		
Dichlorodifluoromethane	75-71-8	5		5 UJ		5 UJ		5 U		5 U		
Ethylbenzene	100-41-4	5		5 U		5 U		5 U		5 U		
Hexachlorobutadiene	87-68-3	0.5		5 U		5 U		5 U		5 U		
Isopropylbenzene	98-82-8	5		5 U		5 U		5 U		5 U		
m,p-Xylene	179601-23-1			5 U		5 U		5 U		5 U		
Methyl Acetate	79-20-9			5 UJ		5 UJ		5 U		5 U		
Methyl Iodide	74-88-4	5		5 UJ		5 UJ		5 U		5 U		
Methyl Tert-Butyl Ether (MTBE)	1634-04-4			5 U		5 U		5 U		5 U		
Methylcyclohexane	108-87-2			5 UJ		5 UJ		5 U		5 U		
Methylene Chloride	75-09-2	5		5 U		5 U		5 U		5 U		
Naphthalene	91-20-3			5 UJ		5 UJ		5 U		5 U		
n-Butylbenzene	104-51-8	5		5 U		5 U		5 U		5 U		
n-Propylbenzene	103-65-1	5		5 U		5 U		5 U		5 U		
o-Xylene	95-47-6	5		5 U		5 U		5 U		5 U		
p-Isopropyltoluene	99-87-6	5		5 U		5 U		5 U		5 U		
sec-Butylbenzene	135-98-8	5		5 U		5 U		5 U		5 U		
Styrene	100-42-5	5		5 U		5 U		5 U		5 U		
tert-Butylbenzene	98-06-6	5		5 U		5 U		5 U		5 U		
Tetrachloroethene	127-18-4	5		5		2200 D		53		57		
Toluene	108-88-3	5		5 U		5 U		2.2 J		2.5 J		
Total Xylenes	1330-20-7			5 U		5 U		5 U		5 U		
trans-1,2-Dichloroethene	156-60-5	5		5 U		5 U		5 U		5 U		
trans-1,3-Dichloropropene	10061-02-6			5 U		5 U		5 U		5 U		
Trichloroethene	79-01-6	5		5 U		64		5.2		5.5		
Trichlorofluoromethane	75-69-4	5		5 U		5 U		5 U		5 U		
Vinyl Acetate	108-05-4			5 U		5 U		5 U		5 U		
Vinyl Chloride	75-01-4	2		5 UJ		5 UJ		5 U		5 U		

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID 80230-MW15D-030712	MW15D 03/07/2012	80230-MW16S-030612	MW16S 03/06/2012	PM-GWS-1A-110311 PM-GWS-1A 11/03/2011	PM-GWS-1A-110311-DUP PM-GWS-1A 11/03/2011	PM-GWS-1B-1122011 PM-GWS-1B 11/02/2011
Chemical Name	CAS#	New York State Class GA							
Benzo(g,h,i)Perylene	191-24-2								
Benzo(k)Fluoranthene	207-08-9								
bis(2-Chloroethoxy)Methane	111-91-1	5							
bis(2-Chloroethyl)Ether	111-44-4	1							
Bis(2-Ethylhexyl)Phthalate	117-81-7	5							
bis-Chloroisopropyl ether	108-60-1	5							
Butylbenzylphthalate	85-68-7								
Caprolactam	105-60-2								
Carbazole	86-74-8								
Chrysene	218-01-9								
Dibenzo(a,h)Anthracene	53-70-3								
Dibenzofuran	132-64-9								
Diethylphthalate	84-66-2								
Dimethylphthalate	131-11-3								
di-n-butylphthalate	84-74-2	50							
di-n-octylphthalate	117-84-0								
Fluoranthene	206-44-0								
Fluorene	86-73-7								
Hexachlorobenzene	118-74-1	0.04							
Hexachlorobutadiene	87-68-3	0.5							
Hexachlorocyclopentadiene	77-47-4	5							
Hexachloroethane	67-72-1	5							
Indeno(1,2,3-cd)Pyrene	193-39-5								
Isophorone	78-59-1								
Naphthalene	91-20-3								
Nitrobenzene	98-95-3	0.4							
n-Nitroso-di-n-Propylamine	621-64-7								
n-Nitrosodiphenylamine	86-30-6								
Pentachlorophenol	87-86-5	1							
Phenanthrene	85-01-8								
Phenol	108-95-2	1							
Pyrene	129-00-0								
<b>Inorganics - SW6010</b>									
Aluminum	7429-90-5								
Antimony	7440-36-0	3							
Arsenic	7440-38-2	25							
Barium	7440-39-3	1000							

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID 80230-MW15D-030712	MW15D 03/07/2012	80230-MW16S-030612	MW16S 03/06/2012	PM-GWS-1A-110311 PM-GWS-1A 11/03/2011	PM-GWS-1A-110311-DUP PM-GWS-1A 11/03/2011	PM-GWS-1B-1122011 PM-GWS-1B 11/02/2011
Chemical Name	CAS#	New York State Class GA							
Beryllium	7440-41-7								
Cadmium	7440-43-9	5							
Calcium	7440-70-2								
Chromium	7440-47-3	50							
Cobalt	7440-48-4								
Copper	7440-50-8	200							
Iron	7439-89-6	300							
Lead	7439-92-1	25							
Magnesium	7439-95-4								
Manganese	7439-96-5	300							
Mercury	7439-97-6	0.7							
Nickel	7440-02-0	100							
Potassium	7440-09-7								
Selenium	7782-49-2	10							
Silver	7440-22-4	50							
Sodium	7440-23-5								
Thallium	7440-28-0								
Vanadium	7440-62-2								
Zinc	7440-66-6								
<b>Wet Chemistry (mg/L)</b>									
Sulfide - 4500-S F	18496-25-8		0.11		0.038				
Alkalinity, Total (as CaCO <sub>3</sub> ) - A2320	ALK		310		400				
Chloride - E300.0	16887-00-6	250	99		480				
Nitrate-NO <sub>3</sub> - E300.0	14797-55-8	10	5		1.9				
Nitrite-NO <sub>2</sub> - E300.0	14797-65-0	1	0.13 U		0.13 U				
Sulfate - E300.0	14808-79-8		47		52				
Hardness As CaCO <sub>3</sub> - SM2340B	CACOA-H		360		600				
Total Dissolved Solids - SM2540C	TDS		400		1200				
Total Suspended Solids - SM2540D	TSS		260		60				
Ammonia as N - SM4500	7664-41-7	2	0.2 R		0.21 R				
Nitrogen, Kjeldahl - SM4500-NORGC	KN		0.2 R		0.27 R				
Total Organic Carbon - SM5310B	TOC		10 U		10 U				

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID PM-GWS-2-1122011	PM-GWS-5-110311	PM-GWS-6-110311	PM-GWS-7-1122011	
Chemical Name	CAS#	New York State Class GA	PM-GWS-2 11/02/2011	PM-GWS-5 11/03/2011	PM-GWS-6 11/03/2011	PM-GWS-7 11/02/2011	
<b>Volatile Organic Compounds - SW8260</b>							
1,1,1,2-Tetrachloroethane	630-20-6	5	5 U	5 U	5 U	5 U	
1,1,1-Trichloroethane	71-55-6	5	5 U	5 U	5 U	5 U	
1,1,2,2-Tetrachloroethane	79-34-5	5	5 U	5 U	5 U	5 U	
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1	5	5 U	5 U	5 U	5 U	
1,1,2-Trichloroethane	79-00-5	1	5 U	5 U	5 U	5 U	
1,1-Dichloroethane	75-34-3	5	5 U	5 U	5 U	5 U	
1,1-Dichloroethene	75-35-4	5	5 U	5 U	5 U	5 U	
1,1-Dichloropropene	563-58-6		5 U	5 U	5 U	5 U	
1,2,3-Trichlorobenzene	87-61-6	5	5 UJ	5 UJ	5 U	5 U	
1,2,3-Trichloropropane	96-18-4	0.04	5 U	5 U	5 UJ	5 UJ	
1,2,4-Trichlorobenzene	120-82-1	5	5 UJ	5 UJ	5 UJ	5 UJ	
1,2,4-Trimethylbenzene	95-63-6	5	5 U	5 U	5 U	5 U	
1,2-Dibromo-3-Chloropropane	96-12-8	0.04	5 U	5 U	5 U	5 U	
1,2-Dibromoethane (EDB)	106-93-4	0.0006	5 U	5 U	5 U	5 U	
1,2-Dichlorobenzene	95-50-1	3	5 U	5 U	5 U	5 U	
1,2-Dichloroethane	107-06-2	0.6	5 U	5 U	5 U	5 U	
1,2-Dichloropropane	78-87-5	1	5 U	5 U	5 U	5 U	
1,3,5-Trimethylbenzene	108-67-8	5	5 U	5 U	5 U	5 U	
1,3-Dichlorobenzene	541-73-1	3	5 U	5 U	5 U	5 U	
1,3-Dichloropropane	142-28-9	5	5 U	5 U	5 U	5 U	
1,4-Dichlorobenzene	106-46-7	3	5 U	5 U	5 U	5 U	
2,2-Dichloropropane	594-20-7	5	5 U	5 U	5 U	5 U	
2-Butanone (MEK)	78-93-3		5 R	5 R	5 R	140 J	
2-Chlorotoluene	95-49-8	5	5 U	5 U	5 U	5 U	
2-Hexanone	591-78-6		5 UJ	5 U	5 U	5 U	
4-Chlorotoluene	106-43-4	5	5 U	5 U	5 U	5 U	
4-Methyl-2-Pentanone (MIBK)	108-10-1		5 U	5 U	5 U	5 U	
Acetone	67-64-1		5 R	5 R	8.3 J	89 J	
Benzene	71-43-2	1	5 U	5 U	5 U	5 U	
Bromobenzene	108-86-1	5	5 U	5 U	5 U	5 U	
Bromochloromethane	74-97-5	5	5 U	5 U	5 U	5 U	
Bromodichloromethane	75-27-4		5 U	5 U	5 U	5 U	
Bromoform	75-25-2		5 U	5 UJ	5 UJ	5 UJ	
Bromomethane	74-83-9	5	5 UJ	5 UJ	5 UJ	5 UJ	
Carbon Disulfide	75-15-0	60	5 U	5 U	5 U	2.1 J	
Carbon Tetrachloride	56-23-5	5	5 U	5 U	5 U	5 U	

**Appendix C-1  
Groundwater Sample Results  
Former Paul Miller Dry Cleaners Site  
Port Richmond, Richmond County, New York**

Sample ID			PM-GWS-2-1122011		PM-GWS-5-110311		PM-GWS-6-110311		PM-GWS-7-1122011
Sample Location			PM-GWS-2 11/02/2011		PM-GWS-5 11/03/2011		PM-GWS-6 11/03/2011		PM-GWS-7 11/02/2011
Chemical Name	CAS#	New York State Class GA							
Chlorobenzene	108-90-7	5		5 U		5 U		5 U	5 U
Chloroethane	75-00-3	5		5 U		5 U		5 U	5 U
Chloroform	67-66-3	7		5 U		5 U		5 U	5 U
Chloromethane	74-87-3	5		5 U		5 U		5 U	5 U
cis-1,2-Dichloroethene	156-59-2	5		90 J		15 J		12 J	27 J
cis-1,3-Dichloropropene	10061-01-5			5 UJ		5 UJ		5 UJ	5 UJ
Cyclohexane	110-82-7			5 U		5 U		5 U	5 U
Dibromochloromethane	124-48-1			5 U		5 U		5 U	5 U
Dibromomethane	74-95-3	5		5 U		5 UJ		5 U	5 U
Dichlorodifluoromethane	75-71-8	5		5 UJ		5 UJ		5 UJ	5 UJ
Ethylbenzene	100-41-4	5		5 U		5 U		5 U	5 U
Hexachlorobutadiene	87-68-3	0.5		5 UJ		5 UJ		5 U	5 U
Isopropylbenzene	98-82-8	5		5 U		5 U		5 U	5 U
m,p-Xylene	179601-23-1			5 U		5 U		5 U	5 U
Methyl Acetate	79-20-9			5 U		5 U		5 U	5 U
Methyl Iodide	74-88-4	5		5 U		5 U		5 U	5 U
Methyl Tert-Butyl Ether (MTBE)	1634-04-4			5 U		5 U		5 U	5 U
Methylcyclohexane	108-87-2			5 UJ		5 UJ		5 UJ	5 UJ
Methylene Chloride	75-09-2	5		5 U		5 U		5 U	5 U
Naphthalene	91-20-3			5 UJ		5 UJ		5 U	5 U
n-Butylbenzene	104-51-8	5		5 U		5 UJ		5 U	5 U
n-Propylbenzene	103-65-1	5		5 U		5 U		5 U	5 U
o-Xylene	95-47-6	5		5 U		5 U		5 U	5 U
p-Isopropyltoluene	99-87-6	5		5 U		5 U		5 U	5 U
sec-Butylbenzene	135-98-8	5		5 U		5 U		5 U	5 U
Styrene	100-42-5	5		5 U		5 U		5 U	5 U
tert-Butylbenzene	98-06-6	5		5 UJ		5 U		5 U	5 U
Tetrachloroethene	127-18-4	5		150 J		51		11 J	18 J
Toluene	108-88-3	5		5 U		5 U		5 U	5 U
Total Xylenes	1330-20-7			5 U		5 U		5 U	5 U
trans-1,2-Dichloroethene	156-60-5	5		5 U		5 U		5 U	5 U
trans-1,3-Dichloropropene	10061-02-6			5 U		5 U		5 U	5 U
Trichloroethene	79-01-6	5		16		14		4.3 J	3.1 J
Trichlorofluoromethane	75-69-4	5		5 U		5 U		5 U	5 U
Vinyl Acetate	108-05-4			5 U		5 U		5 U	5 U
Vinyl Chloride	75-01-4	2		2.1 J		5 U		5 U	5 U

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID PM-GWS-2-1122011	PM-GWS-5-110311	PM-GWS-6-110311	PM-GWS-7-1122011
Chemical Name	CAS#	New York State Class GA	PM-GWS-2 11/02/2011	PM-GWS-5 11/03/2011	PM-GWS-6 11/03/2011	PM-GWS-7 11/02/2011
Ethane	74-84-0					
Ethene	74-85-1					
Methane	74-82-8					
<b>Semi-Volatile Organic Compounds - SW8270</b>						
1,1'-Biphenyl	92-52-4	5				
2,4,5-Trichlorophenol	95-95-4					
2,4,6-Trichlorophenol	88-06-2					
2,4-Dichlorophenol	120-83-2	1				
2,4-Dimethylphenol	105-67-9	1				
2,4-Dinitrophenol	51-28-5	1				
2,4-Dinitrotoluene	121-14-2	5				
2,6-Dinitrotoluene	606-20-2	5				
2-Chloronaphthalene	91-58-7					
2-Chlorophenol	95-57-8					
2-Methylnaphthalene	91-57-6					
2-Methylphenol	95-48-7					
2-Nitroaniline	88-74-4	5				
2-Nitrophenol	88-75-5					
3,3'-Dichlorobenzidine	91-94-1	5				
3-Nitroaniline	99-09-2	5				
4,6-Dinitro-2-Methylphenol	534-52-1					
4-Bromophenyl-Phenylether	101-55-3					
4-Chloro-3-Methylphenol	59-50-7					
4-Chloroaniline	106-47-8	5				
4-Chlorophenyl-Phenylether	7005-72-3					
4-Methylphenol	106-44-5					
4-Nitroaniline	100-01-6	5				
4-Nitrophenol	100-02-7					
Acenaphthene	83-32-9					
Acenaphthylene	208-96-8					
Acetophenone	98-86-2					
Anthracene	120-12-7					
Atrazine	1912-24-9	7.5				
Benzaldehyde	100-52-7					
Benzo(a)Anthracene	56-55-3					
Benzo(a)Pyrene	50-32-8	0				
Benzo(b)Fluoranthene	205-99-2					

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

		Sample ID Sample Location Sampling Date	PM-GWS-2-1122011 PM-GWS-2 11/02/2011	PM-GWS-5-110311 PM-GWS-5 11/03/2011	PM-GWS-6-110311 PM-GWS-6 11/03/2011	PM-GWS-7-1122011 PM-GWS-7 11/02/2011
Chemical Name	CAS#	New York State Class GA				
Benzo(g,h,i)Perylene	191-24-2					
Benzo(k)Fluoranthene	207-08-9					
bis(2-Chloroethoxy)Methane	111-91-1	5				
bis(2-Chloroethyl)Ether	111-44-4	1				
Bis(2-Ethylhexyl)Phthalate	117-81-7	5				
bis-Chloroisopropyl ether	108-60-1	5				
Butylbenzylphthalate	85-68-7					
Caprolactam	105-60-2					
Carbazole	86-74-8					
Chrysene	218-01-9					
Dibenzo(a,h)Anthracene	53-70-3					
Dibenzofuran	132-64-9					
Diethylphthalate	84-66-2					
Dimethylphthalate	131-11-3					
di-n-butylphthalate	84-74-2	50				
di-n-octylphthalate	117-84-0					
Fluoranthene	206-44-0					
Fluorene	86-73-7					
Hexachlorobenzene	118-74-1	0.04				
Hexachlorobutadiene	87-68-3	0.5				
Hexachlorocyclopentadiene	77-47-4	5				
Hexachloroethane	67-72-1	5				
Indeno(1,2,3-cd)Pyrene	193-39-5					
Isophorone	78-59-1					
Naphthalene	91-20-3					
Nitrobenzene	98-95-3	0.4				
n-Nitroso-di-n-Propylamine	621-64-7					
n-Nitrosodiphenylamine	86-30-6					
Pentachlorophenol	87-86-5	1				
Phenanthrene	85-01-8					
Phenol	108-95-2	1				
Pyrene	129-00-0					
<b>Inorganics - SW6010</b>						
Aluminum	7429-90-5					
Antimony	7440-36-0	3				
Arsenic	7440-38-2	25				
Barium	7440-39-3	1000				

**Appendix C-1**  
**Groundwater Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID PM-GWS-2-1122011	PM-GWS-5-110311	PM-GWS-6-110311	PM-GWS-7-1122011
Chemical Name	CAS#	New York State Class GA	PM-GWS-2 11/02/2011	PM-GWS-5 11/03/2011	PM-GWS-6 11/03/2011	PM-GWS-7 11/02/2011
Beryllium	7440-41-7					
Cadmium	7440-43-9	5				
Calcium	7440-70-2					
Chromium	7440-47-3	50				
Cobalt	7440-48-4					
Copper	7440-50-8	200				
Iron	7439-89-6	300				
Lead	7439-92-1	25				
Magnesium	7439-95-4					
Manganese	7439-96-5	300				
Mercury	7439-97-6	0.7				
Nickel	7440-02-0	100				
Potassium	7440-09-7					
Selenium	7782-49-2	10				
Silver	7440-22-4	50				
Sodium	7440-23-5					
Thallium	7440-28-0					
Vanadium	7440-62-2					
Zinc	7440-66-6					
<b>Wet Chemistry (mg/L)</b>						
Sulfide - 4500-S F	18496-25-8					
Alkalinity, Total (as CaCO <sub>3</sub> ) - A2320	ALK					
Chloride - E300.0	16887-00-6	250				
Nitrate-NO <sub>3</sub> - E300.0	14797-55-8	10				
Nitrite-NO <sub>2</sub> - E300.0	14797-65-0	1				
Sulfate - E300.0	14808-79-8					
Hardness As CaCO <sub>3</sub> - SM2340B	CACOA-H					
Total Dissolved Solids - SM2540C	TDS					
Total Suspended Solids - SM2540D	TSS					
Ammonia as N - SM4500	7664-41-7	2				
Nitrogen, Kjeldahl - SM4500-NORGC	KN					
Total Organic Carbon - SM5310B	TOC					

Sample ID	80230-FB-030512	Lab Sample Number	L0441-04	Sampling Date	03/05/2012	Sample Type	FB	80230-FB-030612	L0441-10	80230-FB-030712	L0441-17	80230-FB-030812	L0441-24	80230-FB-110523	K0909-08
Chemical	CAS#														05/23/2011
<b>Volatile Organic Compounds</b>															
1,1,1,2-Tetrachloroethane	630-20-6		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1,1-Trichloroethane	71-55-6		5 U		5 U		5 U		5 U		5 UJ		5 U		5 U
1,1,2,2-Tetrachloroethane	79-34-5		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1,2-Trichloroethane	79-00-5		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1-Dichloroethane	75-34-3		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1-Dichloroethene	75-35-4		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1-Dichloropropene	563-58-6		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2,3-Trichlorobenzene	87-61-6		5 UJ		5 U		5 U		5 U		5 U		5 U		5 UJ
1,2,3-Trichloropropane	96-18-4		5 U		5 U		5 U		5 U		5 UJ		5 U		5 U
1,2,4-Trichlorobenzene	120-82-1		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2,4-Trimethylbenzene	95-63-6		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2-Dibromo-3-Chloropropane	96-12-8		5 UJ		5 U		5 U		5 U		5 U		5 U		5 UJ
1,2-Dibromoethane (Edb)	106-93-4		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2-Dichlorobenzene	95-50-1		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2-Dichloroethane	107-06-2		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2-Dichloropropane	78-87-5		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,3,5-Trimethylbenzene	108-67-8		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,3-Dichlorobenzene	541-73-1		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,3-Dichloropropane	142-28-9		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,4-Dichlorobenzene	106-46-7		5 U		5 U		5 U		5 U		5 U		5 U		5 U
2,2-Dichloropropane	594-20-7		5 U		5 U		5 U		5 U		5 UJ		5 U		5 UJ
2-Butanone (Mek)	78-93-3		5 R		5 R		5 R		5 R		5 R		5 R		5 U
2-Chlorotoluene	95-49-8		5 U		5 U		5 U		5 U		5 U		5 U		5 U
2-Hexanone	591-78-6		5 U		5 U		5 U		5 U		5 U		5 U		5 U
4-Chlorotoluene	106-43-4		5 U		5 U		5 U		5 U		5 U		5 U		5 U
4-Methyl-2-Pentanone (Mibk)	108-10-1		5 U		5 U		5 U		5 U		5 U		5 U		5 U
Acetone	67-64-1		5 R		5 R		5 R		5 R		5 R		5 U		5 R

	Sample ID	80230-FB-030512 L0441-04 03/05/2012 FB	80230-FB-030612 L0441-10 03/06/2012 FB	80230-FB-030712 L0441-17 03/07/2012 FB	80230-FB-030812 L0441-24 03/08/2012 FB	80230-FB-110523 K0909-08 05/23/2011 EB
Benzene	71-43-2	5 U	5 U	5 U	5 U	5 U
Bromobenzene	108-86-1	5 U	5 U	5 U	5 U	5 U
Bromochloromethane	74-97-5	5 U	5 U	5 U	5 U	5 U
Bromodichloromethane	75-27-4	5 U	5 U	5 U	5 U	5 U
Bromoform	75-25-2	5 U	5 U	5 U	5 U	5 U
Bromomethane	74-83-9	5 U	5 U	5 U	5 U	5 U
Carbon Disulfide	75-15-0	5 U	5 U	5 U	5 U	5 U
Carbon Tetrachloride	56-23-5	5 U	5 U	5 U	5 UJ	5 U
Chlorobenzene	108-90-7	5 U	5 U	5 U	5 U	5 U
Chloroethane	75-00-3	5 U	5 U	5 U	5 U	5 U
Chloroform	67-66-3	5 U	5 U	5 U	5 U	5 U
Chloromethane	74-87-3	5 U	5 U	5 U	5 U	5 U
Cis-1,2-Dichloroethene	156-59-2	5 U	5 U	5 U	5 U	5 U
Cis-1,3-Dichloropropene	10061-01-5	5 U	5 U	5 U	5 U	5 U
Cyclohexane	110-82-7	5 U	5 U	5 U	5 U	5 U
Dibromochloromethane	124-48-1	5 U	5 U	5 U	5 U	5 U
Dibromomethane	74-95-3	5 U	5 U	5 U	5 U	5 U
Dichlorodifluoromethane	75-71-8	5 U	5 U	5 U	5 U	5 U
Ethylbenzene	100-41-4	5 U	5 U	5 U	5 U	5 U
Hexachlorobutadiene	87-68-3	5 U	5 U	5 U	5 U	5 U
Isopropylbenzene	98-82-8	5 U	5 U	5 U	5 U	5 U
M,P-Xylene	179601-23-1	5 U	5 U	5 U	5 U	5 U
Methyl Acetate	79-20-9	5 U	5 U	5 U	5 U	5 U
Methyl Iodide	74-88-4	5 UJ	5 U	5 U	5 U	5 U
Methyl Tert-Butyl Ether (Mtbe)	1634-04-4	5 U	5 U	5 U	5 U	5 U
Methylcyclohexane	108-87-2	5 U	5 U	5 U	5 U	5 U
Methylene Chloride	75-09-2	3.9 U	3.3 U	10 U	5 U	5 U
Naphthalene	91-20-3	5 UJ	5 U	5 U	5 U	5 UJ
N-Butylbenzene	104-51-8	5 U	5 U	5 U	5 U	5 U
N-Propylbenzene	103-65-1	5 U	5 U	5 U	5 U	5 U

	Sample ID	80230-FB-030512	80230-FB-030612	80230-FB-030712	80230-FB-030812	80230-FB-110523
	Lab Sample Number	L0441-04	L0441-10	L0441-17	L0441-24	K0909-08
	Sampling Date	03/05/2012	03/06/2012	03/07/2012	03/08/2012	05/23/2011
	Sample Type	FB	FB	FB	FB	EB
O-Xylene	95-47-6	5 U	5 U	5 U	5 U	5 U
P-Isopropyltoluene	99-87-6	5 U	5 U	5 U	5 U	5 U
Sec-Butylbenzene	135-98-8	5 U	5 U	5 U	5 U	5 U
Styrene	100-42-5	5 U	5 U	5 U	5 U	5 U
Tert-Butylbenzene	98-06-6	5 U	5 U	5 U	5 U	5 U
Tetrachloroethene	127-18-4	5 U	3.1 J	5 U	4.5 J	5 U
Toluene	108-88-3	5 U	5 U	5 U	5 U	5 U
Total Xylenes	1330-20-7	5 U	5 U	5 U	5 U	5 U
Trans-1,2-Dichloroethene	156-60-5	5 U	5 U	5 U	5 U	5 U
Trans-1,3-Dichloropropene	10061-02-6	5 U	5 U	5 U	5 U	5 U
Trichloroethene	79-01-6	5 U	5 U	5 U	5 U	5 U
Trichlorofluoromethane	75-69-4	5 U	5 U	5 U	5 U	5 U
Vinyl Acetate	108-05-4	5 U	5 U	5 U	5 U	5 U
Vinyl Chloride	75-01-4	5 U	5 U	5 U	5 U	5 U
Semi-Volatile Organic Compounds						
1,1'-Biphenyl	92-52-4					10 U
2,4,5-Trichlorophenol	95-95-4					20 U
2,4,6-Trichlorophenol	88-06-2					10 U
2,4-Dichlorophenol	120-83-2					10 U
2,4-Dimethylphenol	105-67-9					10 U
2,4-Dinitrophenol	51-28-5					20 UJ
2,4-Dinitrotoluene	121-14-2					10 U
2,6-Dinitrotoluene	606-20-2					10 U
2-Chloronaphthalene	91-58-7					10 U
2-Chlorophenol	95-57-8					10 U
2-Methylnaphthalene	91-57-6					10 UJ
2-Methylphenol	95-48-7					10 U
2-Nitroaniline	88-74-4					20 U
2-Nitrophenol	88-75-5					10 U
3,3'-Dichlorobenzidine	91-94-1					10 U

	Sample ID	80230-FB-030512 L0441-04 03/05/2012 FB	80230-FB-030612 L0441-10 03/06/2012 FB	80230-FB-030712 L0441-17 03/07/2012 FB	80230-FB-030812 L0441-24 03/08/2012 FB	80230-FB-110523 K0909-08 05/23/2011 EB
3-Nitroaniline	99-09-2					20 U
4,6-Dinitro-2-Methylphenol	534-52-1					20 U
4-Bromophenyl-Phenylether	101-55-3					10 U
4-Chloro-3-Methylphenol	59-50-7					10 U
4-Chloroaniline	106-47-8					10 U
4-Chlorophenyl-Phenylether	7005-72-3					10 U
4-Methylphenol	106-44-5					10 U
4-Nitroaniline	100-01-6					20 U
4-Nitrophenol	100-02-7					20 U
Acenaphthene	83-32-9					10 U
Acenaphthylene	208-96-8					10 U
Acetophenone	98-86-2					10 U
Anthracene	120-12-7					10 U
Atrazine	1912-24-9					10 U
Benzaldehyde	100-52-7					10 U
Benzo(A)Anthracene	56-55-3					10 U
Benzo(A)Pyrene	50-32-8					10 U
Benzo(B)Fluoranthene	205-99-2					10 U
Benzo(G,H,I)Perylene	191-24-2					10 U
Benzo(K)Fluoranthene	207-08-9					10 U
Bis(2-Chloroethoxy)Methane	111-91-1					10 U
Bis(2-Chloroethyl) Ether	111-44-4					10 U
Bis(2-Ethylhexyl)Phthalate	117-81-7					10 U
Bis-Chloroisopropyl Ether	108-60-1					10 U
Butylbenzylphthalate	85-68-7					10 U
Caprolactam	105-60-2					10 U
Carbazole	86-74-8					10 U
Chrysene	218-01-9					10 U
Dibenzo(A,H)Anthracene	53-70-3					10 U
Dibenzofuran	132-64-9					10 U

	Sample ID	80230-FB-030512	80230-FB-030612	80230-FB-030712	80230-FB-030812	80230-FB-110523
	Lab Sample Number	L0441-04	L0441-10	L0441-17	L0441-24	K0909-08
	Sampling Date	03/05/2012	03/06/2012	03/07/2012	03/08/2012	05/23/2011
	Sample Type	FB	FB	FB	FB	EB
Diethylphthalate	84-66-2					10 U
Dimethylphthalate	131-11-3					10 U
Di-N-Butylphthalate	84-74-2					10 U
Di-N-Octylphthalate	117-84-0					10 U
Fluoranthene	206-44-0					10 U
Fluorene	86-73-7					10 U
Hexachlorobenzene	118-74-1					10 U
Hexachlorobutadiene	87-68-3					10 U
Hexachlorocyclopentadiene	77-47-4					10 U
Hexachloroethane	67-72-1					10 U
Indeno(1,2,3-Cd)Pyrene	193-39-5					10 U
Isophorone	78-59-1					10 U
Naphthalene	91-20-3					10 U
Nitrobenzene	98-95-3					10 U
N-Nitroso-Di-N-Propylamine	621-64-7					10 U
N-Nitrosodiphenylamine	86-30-6					10 U
Pentachlorophenol	87-86-5					20 U
Phenanthrene	85-01-8					10 U
Phenol	108-95-2					10 U
Pyrene	129-00-0					10 U
Methane, Ethane, Ethene						
Ethane	74-84-0	1.2 U	120	1.3 U	1.2 U	
Ethene	74-85-1	1.5 U	1.6 U	1.6 U	1.5 U	
Methane	74-82-8	0.56 U	2.8	2.2	1.8	
Pesticides						
4,4'-DDD	72-54-8					0.1 U
4,4'-DDE	72-55-9					0.1 U
4,4'-DDT	50-29-3					0.1 U
Aldrin	309-00-2					0.05 U
Alpha-Bhc	319-84-6					0.05 U

	Sample ID	80230-FB-030512	80230-FB-030612	80230-FB-030712	80230-FB-030812	80230-FB-110523
	Lab Sample Number	L0441-04	L0441-10	L0441-17	L0441-24	K0909-08
	Sampling Date	03/05/2012	03/06/2012	03/07/2012	03/08/2012	05/23/2011
	Sample Type	FB	FB	FB	FB	EB
Alpha-Chlordane	5103-71-9					0.05 U
Beta-Bhc	319-85-7					0.05 U
Delta-Bhc	319-86-8					0.05 U
Dieldrin	60-57-1					0.1 U
Endosulfan I	959-98-8					0.05 U
Endosulfan II	33213-65-9					0.1 U
Endosulfan Sulfate	1031-07-8					0.1 U
Endrin	72-20-8					0.1 U
Endrin Aldehyde	7421-93-4					0.1 U
Endrin Ketone	53494-70-5					0.1 U
Gamma-Bhc (Lindane)	58-89-9					0.05 U
Gamma-Chlordane	5103-74-2					0.05 U
Heptachlor	76-44-8					0.05 U
Heptachlor Epoxide	1024-57-3					0.05 U
Methoxychlor	72-43-5					0.5 U
Toxaphene	8001-35-2					5 U
<b>Polychlorinated Organic Compounds</b>						
Aroclor 1016	12674-11-2					1 U
Aroclor 1221	11104-28-2					1 U
Aroclor 1232	11141-16-5					1 U
Aroclor 1242	53469-21-9					1 U
Aroclor 1248	12672-29-6					1 U
Aroclor 1254	11097-69-1					1 U
Aroclor 1260	11096-82-5					1 U
<b>Inorganics</b>						
Aluminum	7429-90-5					200 U
Antimony	7440-36-0					20 U
Arsenic	7440-38-2					20 U
Barium	7440-39-3					200 UEJ
Beryllium	7440-41-7					5 U

	Sample ID Lab Sample Number	80230-FB-030512 L0441-04 03/05/2012 FB	80230-FB-030612 L0441-10 03/06/2012 FB	80230-FB-030712 L0441-17 03/07/2012 FB	80230-FB-030812 L0441-24 03/08/2012 FB	80230-FB-110523 K0909-08 05/23/2011 EB
Cadmium	7440-43-9					5   U
Calcium	7440-70-2					800   U
Chromium	7440-47-3					20   U
Cobalt	7440-48-4					50   U
Copper	7440-50-8					30   U
Iron	7439-89-6					200   U
Lead	7439-92-1					10   U
Magnesium	7439-95-4					500   U
Manganese	7439-96-5					50   U
Mercury - SW7470	7439-97-6					0.2   U
Nickel	7440-02-0					50   U
Potassium	7440-09-7					1000   U
Selenium	7782-49-2					30   U
Silver	7440-22-4					30   U
Sodium	7440-23-5					159   B
Thallium	7440-28-0					20   U
Vanadium	7440-62-2					50   U
Zinc	7440-66-6					11.4   B

Sample ID	80230-FB-110524	Lab Sample Number	K0918-06	Sampling Date	05/24/2011	Sample Type	EB	80230-FB-110525	K0940-03	80230-FB-110526	K0941-01	80230-FB-110616	K1072-04	80230-FB-110617	K1072-01
Chemical	CAS#														
<b>Volatile Organic Compounds</b>															
1,1,1,2-Tetrachloroethane	630-20-6		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1,1-Trichloroethane	71-55-6		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1,2,2-Tetrachloroethane	79-34-5		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1,2-Trichloroethane	79-00-5		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1-Dichloroethane	75-34-3		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1-Dichloroethene	75-35-4		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,1-Dichloropropene	563-58-6		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2,3-Trichlorobenzene	87-61-6		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2,3-Trichloropropane	96-18-4		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2,4-Trichlorobenzene	120-82-1		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2,4-Trimethylbenzene	95-63-6		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2-Dibromo-3-Chloropropane	96-12-8		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2-Dibromoethane (Edb)	106-93-4		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2-Dichlorobenzene	95-50-1		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2-Dichloroethane	107-06-2		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,2-Dichloropropane	78-87-5		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,3,5-Trimethylbenzene	108-67-8		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,3-Dichlorobenzene	541-73-1		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,3-Dichloropropane	142-28-9		5 U		5 U		5 U		5 U		5 U		5 U		5 U
1,4-Dichlorobenzene	106-46-7		5 U		5 U		5 U		5 U		5 U		5 U		5 U
2,2-Dichloropropane	594-20-7		5 U		5 U		5 U		5 U		5 U		5 U		5 U
2-Butanone (Mek)	78-93-3		5 R		5 R		5 R		5 R		5 R		5 R		5 R
2-Chlorotoluene	95-49-8		5 U		5 U		5 U		5 U		5 U		5 U		5 U
2-Hexanone	591-78-6		5 U		5 U		5 U		5 U		5 U		5 U		5 U
4-Chlorotoluene	106-43-4		5 U		5 U		5 U		5 U		5 U		5 U		5 U
4-Methyl-2-Pentanone (Mibk)	108-10-1		5 U		5 U		5 U		5 U		5 U		5 U		5 U
Acetone	67-64-1		5 R		5 R		5 R		5 R		5 R		6.8 J		3.4 J

	Sample ID	80230-FB-110524 K0918-06 05/24/2011 EB	80230-FB-110525 K0940-03 05/25/2011 EB	80230-FB-110526 K0941-01 05/26/2011 EB	80230-FB-110616 K1072-04 06/16/2011 EB	80230-FB-110617 K1072-01 06/17/2011 EB
Benzene	71-43-2	5 U	5 U	5 U	5 U	5 U
Bromobenzene	108-86-1	5 U	5 U	5 U	5 U	5 U
Bromochloromethane	74-97-5	5 U	5 U	5 U	5 U	5 U
Bromodichloromethane	75-27-4	5 U	5 U	5 U	5 U	5 U
Bromoform	75-25-2	5 U	5 U	5 U	5 U	5 U
Bromomethane	74-83-9	5 U	5 U	5 U	5 U	5 U
Carbon Disulfide	75-15-0	5 U	5 U	5 U	5 U	5 U
Carbon Tetrachloride	56-23-5	5 U	5 U	5 U	5 U	5 U
Chlorobenzene	108-90-7	5 U	5 U	5 U	5 U	5 U
Chloroethane	75-00-3	5 U	5 U	5 U	5 U	5 U
Chloroform	67-66-3	5 U	5 U	5 U	5 U	5 U
Chloromethane	74-87-3	5 U	5 U	5 U	5 U	5 U
Cis-1,2-Dichloroethene	156-59-2	5 U	5 U	5 U	5 U	5 U
Cis-1,3-Dichloropropene	10061-01-5	5 U	5 U	5 U	5 U	5 U
Cyclohexane	110-82-7	5 U	5 U	5 U	5 U	5 U
Dibromochloromethane	124-48-1	5 U	5 U	5 U	5 U	5 U
Dibromomethane	74-95-3	5 U	5 U	5 U	5 U	5 U
Dichlorodifluoromethane	75-71-8	5 UJ				
Ethylbenzene	100-41-4	5 U	5 U	5 U	5 U	5 U
Hexachlorobutadiene	87-68-3	5 U	5 U	5 U	5 U	5 U
Isopropylbenzene	98-82-8	5 U	5 U	5 U	5 U	5 U
M,P-Xylene	179601-23-1	5 U	5 U	5 U	5 U	5 U
Methyl Acetate	79-20-9	5 U	5 U	5 U	5 U	5 U
Methyl Iodide	74-88-4	5 U	5 U	5 U	5 U	5 U
Methyl Tert-Butyl Ether (Mtbe)	1634-04-4	5 U	5 U	5 U	5 U	5 U
Methylcyclohexane	108-87-2	5 U	5 U	5 U	5 U	5 U
Methylene Chloride	75-09-2	5 U	5 U	5 U	3.1 J	5 U
Naphthalene	91-20-3	5 U	5 U	5 U	5 U	5 U
N-Butylbenzene	104-51-8	5 U	5 U	5 U	5 U	5 U
N-Propylbenzene	103-65-1	5 U	5 U	5 U	5 U	5 U

	Sample ID	80230-FB-110524	80230-FB-110525	80230-FB-110526	80230-FB-110616	80230-FB-110617
	Lab Sample Number	K0918-06	K0940-03	K0941-01	K1072-04	K1072-01
	Sampling Date	05/24/2011	05/25/2011	05/26/2011	06/16/2011	06/17/2011
	Sample Type	EB	EB	EB	EB	EB
O-Xylene	95-47-6	5 U	5 U	5 U	5 U	5 U
P-Isopropyltoluene	99-87-6	5 U	5 U	5 U	5 U	5 U
Sec-Butylbenzene	135-98-8	5 U	5 U	5 U	5 U	5 U
Styrene	100-42-5	5 U	5 U	5 U	5 U	5 U
Tert-Butylbenzene	98-06-6	5 U	5 U	5 U	5 U	5 U
Tetrachloroethene	127-18-4	5 U	5 UJ	5 UJ	5 U	5 UJ
Toluene	108-88-3	5 U	5 U	5 U	5 U	5 U
Total Xylenes	1330-20-7	5 U	5 U	5 U	5 U	5 U
Trans-1,2-Dichloroethene	156-60-5	5 U	5 U	5 U	5 U	5 U
Trans-1,3-Dichloropropene	10061-02-6	5 U	5 U	5 U	5 U	5 U
Trichloroethene	79-01-6	5 U	5 U	5 U	5 U	5 U
Trichlorofluoromethane	75-69-4	5 U	5 U	5 U	5 U	5 U
Vinyl Acetate	108-05-4	5 U	5 U	5 U	5 U	5 U
Vinyl Chloride	75-01-4	5 U	5 U	5 U	5 U	5 U
Semi-Volatile Organic Compounds						
1,1'-Biphenyl	92-52-4	10 U				
2,4,5-Trichlorophenol	95-95-4	20 U				
2,4,6-Trichlorophenol	88-06-2	10 U				
2,4-Dichlorophenol	120-83-2	10 U				
2,4-Dimethylphenol	105-67-9	10 U	10 U	10 U	10 UJ	10 UJ
2,4-Dinitrophenol	51-28-5	20 UJ				
2,4-Dinitrotoluene	121-14-2	10 U				
2,6-Dinitrotoluene	606-20-2	10 U				
2-Chloronaphthalene	91-58-7	10 U				
2-Chlorophenol	95-57-8	10 U				
2-Methylnaphthalene	91-57-6	10 UJ	10 UJ	10 UJ	10 U	10 U
2-Methylphenol	95-48-7	10 U				
2-Nitroaniline	88-74-4	20 U				
2-Nitrophenol	88-75-5	10 U				
3,3'-Dichlorobenzidine	91-94-1	10 U				

	Sample ID	80230-FB-110524	80230-FB-110525	80230-FB-110526	80230-FB-110616	80230-FB-110617
	Lab Sample Number	K0918-06	K0940-03	K0941-01	K1072-04	K1072-01
	Sampling Date	05/24/2011	05/25/2011	05/26/2011	06/16/2011	06/17/2011
	Sample Type	EB	EB	EB	EB	EB
3-Nitroaniline	99-09-2	20 U				
4,6-Dinitro-2-Methylphenol	534-52-1	20 U				
4-Bromophenyl-Phenylether	101-55-3	10 U				
4-Chloro-3-Methylphenol	59-50-7	10 U				
4-Chloroaniline	106-47-8	10 U				
4-Chlorophenyl-Phenylether	7005-72-3	10 U				
4-Methylphenol	106-44-5	10 U				
4-Nitroaniline	100-01-6	20 U				
4-Nitrophenol	100-02-7	20 U				
Acenaphthene	83-32-9	10 U				
Acenaphthylene	208-96-8	10 U				
Acetophenone	98-86-2	10 U				
Anthracene	120-12-7	10 U				
Atrazine	1912-24-9	10 U				
Benzaldehyde	100-52-7	10 U				
Benzo(A)Anthracene	56-55-3	10 U				
Benzo(A)Pyrene	50-32-8	10 U				
Benzo(B)Fluoranthene	205-99-2	10 U				
Benzo(G,H,I)Perylene	191-24-2	10 U				
Benzo(K)Fluoranthene	207-08-9	10 U				
Bis(2-Chloroethoxy)Methane	111-91-1	10 U				
Bis(2-Chloroethyl) Ether	111-44-4	10 U				
Bis(2-Ethylhexyl)Phthalate	117-81-7	10 U	7.5 BJ	5.9 BJ	10 U	1.6 J
Bis-Chloroisopropyl Ether	108-60-1	10 U				
Butylbenzylphthalate	85-68-7	10 U				
Caprolactam	105-60-2	10 U	10 UJ	10 UJ	10 U	10 UJ
Carbazole	86-74-8	10 U				
Chrysene	218-01-9	10 U				
Dibenzo(A,H)Anthracene	53-70-3	10 U				
Dibenzofuran	132-64-9	10 U				

	Sample ID Lab Sample Number Sampling Date Sample Type	80230-FB-110524 K0918-06 05/24/2011 EB	80230-FB-110525 K0940-03 05/25/2011 EB	80230-FB-110526 K0941-01 05/26/2011 EB	80230-FB-110616 K1072-04 06/16/2011 EB	80230-FB-110617 K1072-01 06/17/2011 EB
Diethylphthalate	84-66-2	10 U				
Dimethylphthalate	131-11-3	10 U				
Di-N-Butylphthalate	84-74-2	10 U				
Di-N-Octylphthalate	117-84-0	10 U				
Fluoranthene	206-44-0	10 U				
Fluorene	86-73-7	10 U				
Hexachlorobenzene	118-74-1	10 U				
Hexachlorobutadiene	87-68-3	10 U				
Hexachlorocyclopentadiene	77-47-4	10 U				
Hexachloroethane	67-72-1	10 U				
Indeno(1,2,3-Cd)Pyrene	193-39-5	10 U				
Isophorone	78-59-1	10 U				
Naphthalene	91-20-3	10 U				
Nitrobenzene	98-95-3	10 U	10 UJ	10 UJ	10 U	10 U
N-Nitroso-Di-N-Propylamine	621-64-7	10 U				
N-Nitrosodiphenylamine	86-30-6	10 U				
Pentachlorophenol	87-86-5	20 U				
Phenanthrene	85-01-8	10 U				
Phenol	108-95-2	10 U				
Pyrene	129-00-0	10 U				
Methane, Ethane, Ethene						
Ethane	74-84-0					
Ethene	74-85-1					
Methane	74-82-8					
Pesticides						
4,4'-DDD	72-54-8	0.1 U	0.1 U	0.1 U		
4,4'-DDE	72-55-9	0.1 U	0.1 U	0.1 U		
4,4'-DDT	50-29-3	0.1 U	0.1 U	0.1 U		
Aldrin	309-00-2	0.05 U	0.05 U	0.05 U		
Alpha-Bhc	319-84-6	0.05 U	0.05 U	0.05 U		

	Sample ID Lab Sample Number	80230-FB-110524 K0918-06 05/24/2011 EB	80230-FB-110525 K0940-03 05/25/2011 EB	80230-FB-110526 K0941-01 05/26/2011 EB	80230-FB-110616 K1072-04 06/16/2011 EB	80230-FB-110617 K1072-01 06/17/2011 EB
Alpha-Chlordane	5103-71-9	0.05 U	0.05 U	0.05 U		
Beta-Bhc	319-85-7	0.05 U	0.05 U	0.05 U		
Delta-Bhc	319-86-8	0.05 U	0.05 U	0.05 U		
Dieldrin	60-57-1	0.1 U	0.1 U	0.1 U		
Endosulfan I	959-98-8	0.05 U	0.05 U	0.05 U		
Endosulfan II	33213-65-9	0.1 U	0.1 U	0.1 U		
Endosulfan Sulfate	1031-07-8	0.1 U	0.1 U	0.1 U		
Endrin	72-20-8	0.1 U	0.1 U	0.1 U		
Endrin Aldehyde	7421-93-4	0.1 U	0.1 U	0.1 U		
Endrin Ketone	53494-70-5	0.1 U	0.1 U	0.1 U		
Gamma-Bhc (Lindane)	58-89-9	0.05 U	0.05 U	0.05 U		
Gamma-Chlordane	5103-74-2	0.05 U	0.05 U	0.05 U		
Heptachlor	76-44-8	0.05 U	0.05 U	0.05 U		
Heptachlor Epoxide	1024-57-3	0.05 U	0.05 U	0.05 U		
Methoxychlor	72-43-5	0.5 U	0.5 U	0.5 U		
Toxaphene	8001-35-2	5 U	5 U	5 U		
<b>Polychlorinated Organic Compounds</b>						
Aroclor 1016	12674-11-2	1 U	1 U	1 U		
Aroclor 1221	11104-28-2	1 U	1 U	1 U		
Aroclor 1232	11141-16-5	1 U	1 U	1 U		
Aroclor 1242	53469-21-9	1 U	1 U	1 U		
Aroclor 1248	12672-29-6	1 U	1 U	1 U		
Aroclor 1254	11097-69-1	1 U	1 U	1 U		
Aroclor 1260	11096-82-5	1 U	1 U	1 U		
<b>Inorganics</b>						
Aluminum	7429-90-5	200 U	200 U	200 U	200 U	
Antimony	7440-36-0	20 U	20 U	20 U	20 U	
Arsenic	7440-38-2	20 U	20 U	20 U	20 U	
Barium	7440-39-3	200 U	200 U	200 U	1.7 B	
Beryllium	7440-41-7	5 U	5 U	5 U	5 U	

	Sample ID Lab Sample Number Sampling Date Sample Type	80230-FB-110524 K0918-06 05/24/2011 EB	80230-FB-110525 K0940-03 05/25/2011 EB	80230-FB-110526 K0941-01 05/26/2011 EB	80230-FB-110616 K1072-04 06/16/2011 EB	80230-FB-110617 K1072-01 06/17/2011 EB
Cadmium	7440-43-9	5 U	5 U	5 U	5 U	
Calcium	7440-70-2	800 U	172 B	267 B	221 B	
Chromium	7440-47-3	20 U	20 U	20 U	20 U	
Cobalt	7440-48-4	50 U	50 U	50 U	50 U	
Copper	7440-50-8	30 U	30 U	30 U	30 U	
Iron	7439-89-6	200 U	200 U	200 U	200 U	
Lead	7439-92-1	10 U	10 U	10 U	10 U	
Magnesium	7439-95-4	500 U	500 U	500 U	500 U	
Manganese	7439-96-5	50 U	50 U	50 U	50 U	
Mercury - SW7470	7439-97-6	0.2 U	0.2 U	0.2 U	0.2 U	
Nickel	7440-02-0	50 U	50 U	50 U	50 U	
Potassium	7440-09-7	1000 U	1000 U	1000 U	1000 U	
Selenium	7782-49-2	30 U	30 U	30 U	30 U	
Silver	7440-22-4	30 U	30 U	30 U	30 U	
Sodium	7440-23-5	271 B	1000 U	1000 U	227 B	
Thallium	7440-28-0	20 U	20 U	20 U	20 U	
Vanadium	7440-62-2	50 U	50 U	50 U	50 U	
Zinc	7440-66-6	50 U	50 U	25.8 B	13.2 B	

Sample ID	80230-FB-110624	Lab Sample Number	K1157-01	Sampling Date	06/24/2011	Sample Type	EB	80230-FB-AQ-110523	K0909-09	80230-FB-AQ-110524A	K0909-12	80230-TB-030512	L0441-01	80230-TB-030612	L0441-07
Chemical	CAS#														
Volatile Organic Compounds															
1,1,1,2-Tetrachloroethane	630-20-6		5 U		5 U		5 U		5 U		5 U		5 U		
1,1,1-Trichloroethane	71-55-6		5 U		5 U		5 U		5 U		5 U		5 U		
1,1,2,2-Tetrachloroethane	79-34-5		5 U		5 U		5 U		5 U		5 U		5 U		
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5 U		5 U		5 U		5 U		5 U		5 U		
1,1,2-Trichloroethane	79-00-5		5 U		5 U		5 U		5 U		5 U		5 U		
1,1-Dichloroethane	75-34-3		5 U		5 U		5 U		5 U		5 U		5 U		
1,1-Dichloroethene	75-35-4		5 U		5 U		5 U		5 U		5 U		5 U		
1,1-Dichloropropene	563-58-6		5 U		5 U		5 U		5 U		5 U		5 U		
1,2,3-Trichlorobenzene	87-61-6		5 U		5 U		5 U		5 U		5 U		5 U		
1,2,3-Trichloropropane	96-18-4		5 U		5 U		5 U		5 U		5 U		5 U		
1,2,4-Trichlorobenzene	120-82-1		5 U		5 U		5 U		5 U		5 U		5 U		
1,2,4-Trimethylbenzene	95-63-6		5 U		5 U		5 U		5 U		5 U		5 U		
1,2-Dibromo-3-Chloropropane	96-12-8		5 U		5 U		5 U		5 U		5 U		5 U		
1,2-Dibromoethane (Edb)	106-93-4		5 U		5 U		5 U		5 U		5 U		5 U		
1,2-Dichlorobenzene	95-50-1		5 U		5 U		5 U		5 U		5 U		5 U		
1,2-Dichloroethane	107-06-2		5 U		5 U		5 U		5 U		5 U		5 U		
1,2-Dichloropropane	78-87-5		5 U		5 U		5 U		5 U		5 U		5 U		
1,3,5-Trimethylbenzene	108-67-8		5 U		5 U		5 U		5 U		5 U		5 U		
1,3-Dichlorobenzene	541-73-1		5 U		5 U		5 U		5 U		5 U		5 U		
1,3-Dichloropropane	142-28-9		5 U		5 U		5 U		5 U		5 U		5 U		
1,4-Dichlorobenzene	106-46-7		5 U		5 U		5 U		5 U		5 U		5 U		
2,2-Dichloropropane	594-20-7		5 U		5 U		5 U		5 U		5 U		5 U		
2-Butanone (Mek)	78-93-3		5 R		5 R		5 R		5 R		5 R		5 R		
2-Chlorotoluene	95-49-8		5 U		5 U		5 U		5 U		5 U		5 U		
2-Hexanone	591-78-6		5 UJ		5 U		5 U		5 U		5 U		5 U		
4-Chlorotoluene	106-43-4		5 U		5 U		5 U		5 U		5 U		5 U		
4-Methyl-2-Pentanone (Mibk)	108-10-1		5 U		5 U		5 U		5 U		5 U		5 U		
Acetone	67-64-1		2.6 J		5 U		5 U		5 U		5 R		5 R		

	Sample ID	80230-FB-110624	80230-FB-AQ-110523	80230-FB-AQ-110524A	80230-TB-030512	80230-TB-030612
	Lab Sample Number	K1157-01	K0909-09	K0909-12	L0441-01	L0441-07
	Sampling Date	06/24/2011	05/23/2011	05/24/2011	03/05/2012	03/06/2012
	Sample Type	EB	EB	EB	TB	TB
Benzene	71-43-2	5 U	5 R	5 R	5 U	5 U
Bromobenzene	108-86-1	5 U	5 U	5 U	5 U	5 U
Bromochloromethane	74-97-5	5 U	5 U	5 U	5 U	5 U
Bromodichloromethane	75-27-4	5 U	5 U	5 U	5 U	5 U
Bromoform	75-25-2	5 U	5 U	5 U	5 U	5 U
Bromomethane	74-83-9	5 U	5 U	5 U	5 U	5 U
Carbon Disulfide	75-15-0	5 U	5 U	5 U	5 U	5 U
Carbon Tetrachloride	56-23-5	5 U	5 U	5 U	5 U	5 U
Chlorobenzene	108-90-7	5 U	5 U	5 U	5 U	5 U
Chloroethane	75-00-3	5 U	5 U	5 U	5 U	5 U
Chloroform	67-66-3	5 U	5 U	5 U	5 U	5 U
Chloromethane	74-87-3	5 U	5 U	5 U	5 U	5 U
Cis-1,2-Dichloroethene	156-59-2	5 U	5 U	5 U	5 U	5 U
Cis-1,3-Dichloropropene	10061-01-5	5 U	5 U	5 U	5 U	5 U
Cyclohexane	110-82-7	5 U	5 U	5 U	5 U	5 U
Dibromochloromethane	124-48-1	5 U	5 U	5 U	5 U	5 U
Dibromomethane	74-95-3	5 U	5 U	5 U	5 U	5 U
Dichlorodifluoromethane	75-71-8	5 U	5 UJ	5 UJ	5 U	5 U
Ethylbenzene	100-41-4	5 U	5 U	5 U	5 U	5 U
Hexachlorobutadiene	87-68-3	5 U	5 U	5 U	5 U	5 U
Isopropylbenzene	98-82-8	5 U	5 U	5 U	5 U	5 U
M,P-Xylene	179601-23-1	5 U	5 U	5 U	5 U	5 U
Methyl Acetate	79-20-9	5 U	5 UJ	5 UJ	5 U	5 U
Methyl Iodide	74-88-4	5 U	5 U	5 U	5 U	5 U
Methyl Tert-Butyl Ether (Mtbe)	1634-04-4	5 U	5 U	5 U	5 U	5 U
Methylcyclohexane	108-87-2	5 U	5 U	5 U	5 U	5 U
Methylene Chloride	75-09-2	1.9 U	5 U	5 U	5 U	5 U
Naphthalene	91-20-3	5 U	5 U	5 U	5 U	5 U
N-Butylbenzene	104-51-8	5 U	5 U	5 U	5 U	5 U
N-Propylbenzene	103-65-1	5 U	5 U	5 U	5 U	5 U

	Sample ID	80230-FB-110624	80230-FB-AQ-110523	80230-FB-AQ-110524A	80230-TB-030512	80230-TB-030612
	Lab Sample Number	K1157-01	K0909-09	K0909-12	L0441-01	L0441-07
	Sampling Date	06/24/2011	05/23/2011	05/24/2011	03/05/2012	03/06/2012
	Sample Type	EB	EB	EB	TB	TB
O-Xylene	95-47-6	5 U	5 U	5 U	5 U	5 U
P-Isopropyltoluene	99-87-6	5 U	5 U	5 U	5 U	5 U
Sec-Butylbenzene	135-98-8	5 U	5 U	5 U	5 U	5 U
Styrene	100-42-5	5 U	5 U	5 U	5 U	5 U
Tert-Butylbenzene	98-06-6	5 U	5 U	5 U	5 U	5 U
Tetrachloroethene	127-18-4	5 U	1.4 J	1.6 J	5 U	5 U
Toluene	108-88-3	5 U	5 U	5 U	5 U	5 U
Total Xylenes	1330-20-7	5 U	5 U	5 U	5 U	5 U
Trans-1,2-Dichloroethene	156-60-5	5 U	5 U	5 U	5 U	5 U
Trans-1,3-Dichloropropene	10061-02-6	5 U	5 U	5 U	5 U	5 U
Trichloroethene	79-01-6	5 U	5 U	5 U	5 U	5 U
Trichlorofluoromethane	75-69-4	5 U	5 U	5 U	5 U	5 U
Vinyl Acetate	108-05-4	5 U	5 U	5 U	5 U	5 U
Vinyl Chloride	75-01-4	5 U	5 U	5 U	5 U	5 U
Semi-Volatile Organic Compounds						
1,1'-Biphenyl	92-52-4	10 U	10 U	10 U		
2,4,5-Trichlorophenol	95-95-4	20 U	20 U	20 U		
2,4,6-Trichlorophenol	88-06-2	10 U	10 U	10 U		
2,4-Dichlorophenol	120-83-2	10 U	10 U	10 U		
2,4-Dimethylphenol	105-67-9	10 U	10 U	10 U		
2,4-Dinitrophenol	51-28-5	20 U	20 UJ	20 UJ		
2,4-Dinitrotoluene	121-14-2	10 U	10 U	10 U		
2,6-Dinitrotoluene	606-20-2	10 U	10 U	10 U		
2-Chloronaphthalene	91-58-7	10 U	10 U	10 U		
2-Chlorophenol	95-57-8	10 U	10 U	10 U		
2-Methylnaphthalene	91-57-6	10 U	10 UJ	10 UJ		
2-Methylphenol	95-48-7	10 U	10 U	10 U		
2-Nitroaniline	88-74-4	20 U	20 U	20 U		
2-Nitrophenol	88-75-5	10 U	10 U	10 U		
3,3'-Dichlorobenzidine	91-94-1	10 U	10 U	10 U		

	Sample ID	80230-FB-110624	80230-FB-AQ-110523	80230-FB-AQ-110524A	80230-TB-030512	80230-TB-030612
	Lab Sample Number	K1157-01	K0909-09	K0909-12	L0441-01	L0441-07
	Sampling Date	06/24/2011	05/23/2011	05/24/2011	03/05/2012	03/06/2012
	Sample Type	EB	EB	EB	TB	TB
3-Nitroaniline	99-09-2	20 U	20 U	20 U		
4,6-Dinitro-2-Methylphenol	534-52-1	20 U	20 U	20 U		
4-Bromophenyl-Phenylether	101-55-3	10 U	10 U	10 U		
4-Chloro-3-Methylphenol	59-50-7	10 U	10 U	10 U		
4-Chloroaniline	106-47-8	10 U	10 U	10 U		
4-Chlorophenyl-Phenylether	7005-72-3	10 U	10 U	10 U		
4-Methylphenol	106-44-5	10 U	10 U	10 U		
4-Nitroaniline	100-01-6	20 U	20 U	20 U		
4-Nitrophenol	100-02-7	20 U	20 U	20 U		
Acenaphthene	83-32-9	10 U	10 U	10 U		
Acenaphthylene	208-96-8	10 U	10 U	10 U		
Acetophenone	98-86-2	10 U	10 U	10 U		
Anthracene	120-12-7	10 U	10 U	10 U		
Atrazine	1912-24-9	10 U	10 U	10 U		
Benzaldehyde	100-52-7	10 U	10 U	10 U		
Benzo(A)Anthracene	56-55-3	10 U	10 U	10 U		
Benzo(A)Pyrene	50-32-8	10 U	10 U	10 U		
Benzo(B)Fluoranthene	205-99-2	10 U	10 U	10 U		
Benzo(G,H,I)Perylene	191-24-2	10 U	10 U	10 U		
Benzo(K)Fluoranthene	207-08-9	10 U	10 U	10 U		
Bis(2-Chloroethoxy)Methane	111-91-1	10 U	10 U	10 U		
Bis(2-Chloroethyl) Ether	111-44-4	10 U	10 U	10 U		
Bis(2-Ethylhexyl)Phthalate	117-81-7	10 U	10 U	10 U		
Bis-Chloroisopropyl Ether	108-60-1	10 U	10 U	10 U		
Butylbenzylphthalate	85-68-7	10 U	10 U	10 U		
Caprolactam	105-60-2	10 U	10 U	10 U		
Carbazole	86-74-8	10 U	10 U	10 U		
Chrysene	218-01-9	10 U	10 U	10 U		
Dibenzo(A,H)Anthracene	53-70-3	10 U	10 U	10 U		
Dibenzofuran	132-64-9	10 U	10 U	10 U		

	Sample ID Lab Sample Number	80230-FB-110624 K1157-01 06/24/2011 EB	80230-FB-AQ-110523 K0909-09 05/23/2011 EB	80230-FB-AQ-110524A K0909-12 05/24/2011 EB	80230-TB-030512 L0441-01 03/05/2012 TB	80230-TB-030612 L0441-07 03/06/2012 TB
Diethylphthalate	84-66-2	10 U	10 U	10 U		
Dimethylphthalate	131-11-3	10 U	10 U	10 U		
Di-N-Butylphthalate	84-74-2	10 U	10 U	10 U		
Di-N-Octylphthalate	117-84-0	10 U	10 U	10 U		
Fluoranthene	206-44-0	10 U	10 U	10 U		
Fluorene	86-73-7	10 U	10 U	10 U		
Hexachlorobenzene	118-74-1	10 U	10 U	10 U		
Hexachlorobutadiene	87-68-3	10 U	10 U	10 U		
Hexachlorocyclopentadiene	77-47-4	10 U	10 U	10 U		
Hexachloroethane	67-72-1	10 U	10 U	10 U		
Indeno(1,2,3-Cd)Pyrene	193-39-5	10 U	10 U	10 U		
Isophorone	78-59-1	10 U	10 U	10 U		
Naphthalene	91-20-3	10 U	10 U	10 U		
Nitrobenzene	98-95-3	10 U	10 U	10 U		
N-Nitroso-Di-N-Propylamine	621-64-7	10 U	10 U	10 U		
N-Nitrosodiphenylamine	86-30-6	10 U	10 U	10 U		
Pentachlorophenol	87-86-5	20 U	20 U	20 U		
Phenanthrene	85-01-8	10 U	10 U	10 U		
Phenol	108-95-2	10 U	10 U	10 U		
Pyrene	129-00-0	10 U	10 U	10 U		
Methane, Ethane, Ethene						
Ethane	74-84-0					
Ethene	74-85-1					
Methane	74-82-8					
Pesticides						
4,4'-DDD	72-54-8					
4,4'-DDE	72-55-9					
4,4'-DDT	50-29-3					
Aldrin	309-00-2					
Alpha-Bhc	319-84-6					

	Sample ID	80230-FB-110624	80230-FB-AQ-110523	80230-FB-AQ-110524A	80230-TB-030512	80230-TB-030612
	Lab Sample Number	K1157-01	K0909-09	K0909-12	L0441-01	L0441-07
	Sampling Date	06/24/2011	05/23/2011	05/24/2011	03/05/2012	03/06/2012
	Sample Type	EB	EB	EB	TB	TB
Alpha-Chlordane	5103-71-9					
Beta-Bhc	319-85-7					
Delta-Bhc	319-86-8					
Dieldrin	60-57-1					
Endosulfan I	959-98-8					
Endosulfan II	33213-65-9					
Endosulfan Sulfate	1031-07-8					
Endrin	72-20-8					
Endrin Aldehyde	7421-93-4					
Endrin Ketone	53494-70-5					
Gamma-Bhc (Lindane)	58-89-9					
Gamma-Chlordane	5103-74-2					
Heptachlor	76-44-8					
Heptachlor Epoxide	1024-57-3					
Methoxychlor	72-43-5					
Toxaphene	8001-35-2					
<b>Polychlorinated Organic Compounds</b>						
Aroclor 1016	12674-11-2					
Aroclor 1221	11104-28-2					
Aroclor 1232	11141-16-5					
Aroclor 1242	53469-21-9					
Aroclor 1248	12672-29-6					
Aroclor 1254	11097-69-1					
Aroclor 1260	11096-82-5					
<b>Inorganics</b>						
Aluminum	7429-90-5		200 U	200 U		
Antimony	7440-36-0		20 U	20 U		
Arsenic	7440-38-2		20 U	20 U		
Barium	7440-39-3		200 UEJ	1.1 BEJ		
Beryllium	7440-41-7		5 U	5 U		

	Sample ID Lab Sample Number	80230-FB-110624 K1157-01 06/24/2011 EB	80230-FB-AQ-110523 K0909-09 05/23/2011 EB	80230-FB-AQ-110524A K0909-12 05/24/2011 EB	80230-TB-030512 L0441-01 03/05/2012 TB	80230-TB-030612 L0441-07 03/06/2012 TB
Cadmium	7440-43-9			5   U	5   U	
Calcium	7440-70-2		800   U	800   U		
Chromium	7440-47-3		20   U	20   U		
Cobalt	7440-48-4		50   U	50   U		
Copper	7440-50-8		30   U	30   U		
Iron	7439-89-6		200   U	200   U		
Lead	7439-92-1		10   U	10   U		
Magnesium	7439-95-4		500   U	500   U		
Manganese	7439-96-5		50   U	50   U		
Mercury - SW7470	7439-97-6		0.2   U	0.2   U		
Nickel	7440-02-0		50   U	50   U		
Potassium	7440-09-7		1000   U	1000   U		
Selenium	7782-49-2		30   U	30   U		
Silver	7440-22-4		30   U	30   U		
Sodium	7440-23-5		165   B	163   B		
Thallium	7440-28-0		20   U	20   U		
Vanadium	7440-62-2		50   U	50   U		
Zinc	7440-66-6		17.4   B	16   B		

Sample ID	80230-TB-030712	Lab Sample Number	L0441-13	Sampling Date	03/07/2012	Sample Type	TB	80230-TB-030812	L0441-20	PM-FB-AQ-1122011	K2297-05	PM-FB-SO-1112011	K2296-02	PM-FB-SO-1122011	K2297-04
Chemical	CAS#														
<b>Volatile Organic Compounds</b>															
1,1,1,2-Tetrachloroethane	630-20-6		5 U		5 U		5 U		5 U		5 U		5 U		
1,1,1-Trichloroethane	71-55-6		5 U		5 U		5 U		5 U		5 U		5 U		
1,1,2,2-Tetrachloroethane	79-34-5		5 U		5 U		5 U		5 U		5 U		5 U		
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5 U		5 U		5 U		5 U		5 U		5 U		
1,1,2-Trichloroethane	79-00-5		5 U		5 U		5 U		5 U		5 U		5 U		
1,1-Dichloroethane	75-34-3		5 U		5 U		5 U		5 U		5 U		5 U		
1,1-Dichloroethene	75-35-4		5 U		5 U		5 U		5 U		5 U		5 U		
1,1-Dichloropropene	563-58-6		5 U		5 U		5 U		5 U		5 U		5 U		
1,2,3-Trichlorobenzene	87-61-6		5 UJ		5 U		5 UJ		5 UJ		5 UJ		5 UJ		
1,2,3-Trichloropropane	96-18-4		5 U		5 U		5 U		5 U		5 U		5 U		
1,2,4-Trichlorobenzene	120-82-1		5 U		5 U		5 U		5 U		5 U		5 U		
1,2,4-Trimethylbenzene	95-63-6		5 U		5 U		5 U		5 U		5 U		5 U		
1,2-Dibromo-3-Chloropropane	96-12-8		5 UJ		5 U		5 U		5 U		5 U		5 U		
1,2-Dibromoethane (Edb)	106-93-4		5 U		5 U		5 U		5 U		5 U		5 U		
1,2-Dichlorobenzene	95-50-1		5 U		5 U		5 U		5 U		5 U		5 U		
1,2-Dichloroethane	107-06-2		5 U		5 U		5 U		5 U		5 U		5 U		
1,2-Dichloropropane	78-87-5		5 U		5 U		5 U		5 U		5 U		5 U		
1,3,5-Trimethylbenzene	108-67-8		5 U		5 U		5 U		5 U		5 U		5 U		
1,3-Dichlorobenzene	541-73-1		5 U		5 U		5 U		5 U		5 U		5 U		
1,3-Dichloropropane	142-28-9		5 U		5 U		5 U		5 U		5 U		5 U		
1,4-Dichlorobenzene	106-46-7		5 U		5 U		5 U		5 U		5 U		5 U		
2,2-Dichloropropane	594-20-7		5 U		5 UJ		5 U		5 U		5 U		5 U		
2-Butanone (Mek)	78-93-3		5 R		5 R		3 J		5 R		3.2 J				
2-Chlorotoluene	95-49-8		5 U		5 U		5 U		5 U		5 U		5 U		
2-Hexanone	591-78-6		5 U		5 UJ		5 U		5 U		5 UJ		5 U		
4-Chlorotoluene	106-43-4		5 U		5 U		5 U		5 U		5 U		5 U		
4-Methyl-2-Pentanone (Mibk)	108-10-1		5 U		5 U		5 U		5 U		5 U		5 U		
Acetone	67-64-1		5 R		5 R		6.8 J		5 R		6.9 J				

	Sample ID	80230-TB-030712	80230-TB-030812	PM-FB-AQ-1122011	PM-FB-SO-1112011	PM-FB-SO-1122011
	Lab Sample Number	L0441-13	L0441-20	K2297-05	K2296-02	K2297-04
	Sampling Date	03/07/2012	03/08/2012	11/02/2011	11/01/2011	11/02/2011
	Sample Type	TB	TB	FB	FB	FB
Benzene	71-43-2	5 U	5 U	5 U	5 U	5 U
Bromobenzene	108-86-1	5 U	5 U	5 U	5 U	5 U
Bromochloromethane	74-97-5	5 U	5 U	5 U	5 U	5 U
Bromodichloromethane	75-27-4	5 U	5 U	5 U	5 U	5 U
Bromoform	75-25-2	5 U	5 U	5 UJ	5 U	5 UJ
Bromomethane	74-83-9	5 U	5 U	5 U	5 U	5 U
Carbon Disulfide	75-15-0	5 U	5 U	5 U	5 U	5 U
Carbon Tetrachloride	56-23-5	5 U	5 U	5 U	5 U	5 U
Chlorobenzene	108-90-7	5 U	5 U	5 U	5 U	5 U
Chloroethane	75-00-3	5 U	5 UJ	5 U	5 U	5 U
Chloroform	67-66-3	5 U	5 U	5 U	5 U	5 U
Chloromethane	74-87-3	5 U	5 U	5 U	5 U	5 U
Cis-1,2-Dichloroethene	156-59-2	5 U	5 U	5 U	5 U	5 U
Cis-1,3-Dichloropropene	10061-01-5	5 U	5 U	5 U	5 U	5 U
Cyclohexane	110-82-7	5 U	5 U	5 U	5 U	5 U
Dibromochloromethane	124-48-1	5 U	5 U	5 U	5 U	5 U
Dibromomethane	74-95-3	5 U	5 U	5 UJ	5 U	5 UJ
Dichlorodifluoromethane	75-71-8	5 U	5 UJ	5 U	5 U	5 U
Ethylbenzene	100-41-4	5 U	5 U	5 U	5 U	5 U
Hexachlorobutadiene	87-68-3	5 U	5 U	5 UJ	5 UJ	5 UJ
Isopropylbenzene	98-82-8	5 U	5 U	5 U	5 U	5 U
M,P-Xylene	179601-23-1	5 U	5 U	5 U	5 U	5 U
Methyl Acetate	79-20-9	5 U	5 U	5 U	5 U	5 U
Methyl Iodide	74-88-4	5 UJ	5 U	5 U	5 U	5 U
Methyl Tert-Butyl Ether (Mtbe)	1634-04-4	5 U	5 U	5 U	5 U	5 U
Methylcyclohexane	108-87-2	5 U	5 U	5 U	5 U	5 U
Methylene Chloride	75-09-2	5 U	5 U	5 U	5 U	5 U
Naphthalene	91-20-3	5 UJ	5 UJ	5 UJ	5 UJ	5 UJ
N-Butylbenzene	104-51-8	5 U	5 U	5 UJ	5 U	5 UJ
N-Propylbenzene	103-65-1	5 U	5 U	5 U	5 U	5 U

	Sample ID	80230-TB-030712	80230-TB-030812	PM-FB-AQ-1122011	PM-FB-SO-1112011	PM-FB-SO-1122011
	Lab Sample Number	L0441-13	L0441-20	K2297-05	K2296-02	K2297-04
	Sampling Date	03/07/2012	03/08/2012	11/02/2011	11/01/2011	11/02/2011
	Sample Type	TB	TB	FB	FB	FB
O-Xylene	95-47-6	5 U	5 U	5 U	5 U	5 U
P-Isopropyltoluene	99-87-6	5 U	5 U	5 U	5 U	5 U
Sec-Butylbenzene	135-98-8	5 U	5 U	5 U	5 U	5 U
Styrene	100-42-5	5 U	5 U	5 U	5 U	5 U
Tert-Butylbenzene	98-06-6	5 U	5 U	5 U	5 U	5 U
Tetrachloroethene	127-18-4	5 U	5 U	5 U	5 U	5 U
Toluene	108-88-3	5 U	5 U	5 U	5 U	5 U
Total Xylenes	1330-20-7	5 U	5 U	5 U	5 U	5 U
Trans-1,2-Dichloroethene	156-60-5	5 U	5 U	5 U	5 U	5 U
Trans-1,3-Dichloropropene	10061-02-6	5 U	5 U	5 U	5 U	5 U
Trichloroethene	79-01-6	5 U	5 U	5 U	5 U	5 U
Trichlorofluoromethane	75-69-4	5 U	5 U	5 U	5 U	5 U
Vinyl Acetate	108-05-4	5 U	5 U	5 U	5 U	5 U
Vinyl Chloride	75-01-4	5 U	5 UJ	5 U	5 U	5 U
Semi-Volatile Organic Compounds						
1,1'-Biphenyl	92-52-4					
2,4,5-Trichlorophenol	95-95-4					
2,4,6-Trichlorophenol	88-06-2					
2,4-Dichlorophenol	120-83-2					
2,4-Dimethylphenol	105-67-9					
2,4-Dinitrophenol	51-28-5					
2,4-Dinitrotoluene	121-14-2					
2,6-Dinitrotoluene	606-20-2					
2-Chloronaphthalene	91-58-7					
2-Chlorophenol	95-57-8					
2-Methylnaphthalene	91-57-6					
2-Methylphenol	95-48-7					
2-Nitroaniline	88-74-4					
2-Nitrophenol	88-75-5					
3,3'-Dichlorobenzidine	91-94-1					

	Sample ID	80230-TB-030712	80230-TB-030812	PM-FB-AQ-1122011	PM-FB-SO-1112011	PM-FB-SO-1122011
	Lab Sample Number	L0441-13	L0441-20	K2297-05	K2296-02	K2297-04
	Sampling Date	03/07/2012	03/08/2012	11/02/2011	11/01/2011	11/02/2011
	Sample Type	TB	TB	FB	FB	FB
3-Nitroaniline	99-09-2					
4,6-Dinitro-2-Methylphenol	534-52-1					
4-Bromophenyl-Phenylether	101-55-3					
4-Chloro-3-Methylphenol	59-50-7					
4-Chloroaniline	106-47-8					
4-Chlorophenyl-Phenylether	7005-72-3					
4-Methylphenol	106-44-5					
4-Nitroaniline	100-01-6					
4-Nitrophenol	100-02-7					
Acenaphthene	83-32-9					
Acenaphthylene	208-96-8					
Acetophenone	98-86-2					
Anthracene	120-12-7					
Atrazine	1912-24-9					
Benzaldehyde	100-52-7					
Benzo(A)Anthracene	56-55-3					
Benzo(A)Pyrene	50-32-8					
Benzo(B)Fluoranthene	205-99-2					
Benzo(G,H,I)Perylene	191-24-2					
Benzo(K)Fluoranthene	207-08-9					
Bis(2-Chloroethoxy)Methane	111-91-1					
Bis(2-Chloroethyl) Ether	111-44-4					
Bis(2-Ethylhexyl)Phthalate	117-81-7					
Bis-Chloroisopropyl Ether	108-60-1					
Butylbenzylphthalate	85-68-7					
Caprolactam	105-60-2					
Carbazole	86-74-8					
Chrysene	218-01-9					
Dibenzo(A,H)Anthracene	53-70-3					
Dibenzofuran	132-64-9					

	Sample ID	80230-TB-030712	80230-TB-030812	PM-FB-AQ-1122011	PM-FB-SO-1112011	PM-FB-SO-1122011
	Lab Sample Number	L0441-13	L0441-20	K2297-05	K2296-02	K2297-04
	Sampling Date	03/07/2012	03/08/2012	11/02/2011	11/01/2011	11/02/2011
	Sample Type	TB	TB	FB	FB	FB
Diethylphthalate	84-66-2					
Dimethylphthalate	131-11-3					
Di-N-Butylphthalate	84-74-2					
Di-N-Octylphthalate	117-84-0					
Fluoranthene	206-44-0					
Fluorene	86-73-7					
Hexachlorobenzene	118-74-1					
Hexachlorobutadiene	87-68-3					
Hexachlorocyclopentadiene	77-47-4					
Hexachloroethane	67-72-1					
Indeno(1,2,3-Cd)Pyrene	193-39-5					
Isophorone	78-59-1					
Naphthalene	91-20-3					
Nitrobenzene	98-95-3					
N-Nitroso-Di-N-Propylamine	621-64-7					
N-Nitrosodiphenylamine	86-30-6					
Pentachlorophenol	87-86-5					
Phenanthrene	85-01-8					
Phenol	108-95-2					
Pyrene	129-00-0					
Methane, Ethane, Ethene						
Ethane	74-84-0					
Ethene	74-85-1					
Methane	74-82-8					
Pesticides						
4,4'-DDD	72-54-8					
4,4'-DDE	72-55-9					
4,4'-DDT	50-29-3					
Aldrin	309-00-2					
Alpha-Bhc	319-84-6					

	Sample ID	80230-TB-030712	80230-TB-030812	PM-FB-AQ-1122011	PM-FB-SO-1112011	PM-FB-SO-1122011
	Lab Sample Number	L0441-13	L0441-20	K2297-05	K2296-02	K2297-04
	Sampling Date	03/07/2012	03/08/2012	11/02/2011	11/01/2011	11/02/2011
	Sample Type	TB	TB	FB	FB	FB
Alpha-Chlordane	5103-71-9					
Beta-Bhc	319-85-7					
Delta-Bhc	319-86-8					
Dieldrin	60-57-1					
Endosulfan I	959-98-8					
Endosulfan II	33213-65-9					
Endosulfan Sulfate	1031-07-8					
Endrin	72-20-8					
Endrin Aldehyde	7421-93-4					
Endrin Ketone	53494-70-5					
Gamma-Bhc (Lindane)	58-89-9					
Gamma-Chlordane	5103-74-2					
Heptachlor	76-44-8					
Heptachlor Epoxide	1024-57-3					
Methoxychlor	72-43-5					
Toxaphene	8001-35-2					
<b>Polychlorinated Organic Compounds</b>						
Aroclor 1016	12674-11-2					
Aroclor 1221	11104-28-2					
Aroclor 1232	11141-16-5					
Aroclor 1242	53469-21-9					
Aroclor 1248	12672-29-6					
Aroclor 1254	11097-69-1					
Aroclor 1260	11096-82-5					
<b>Inorganics</b>						
Aluminum	7429-90-5					
Antimony	7440-36-0					
Arsenic	7440-38-2					
Barium	7440-39-3					
Beryllium	7440-41-7					

	Sample ID	80230-TB-030712	80230-TB-030812	PM-FB-AQ-1122011	PM-FB-SO-1112011	PM-FB-SO-1122011
	Lab Sample Number	L0441-13	L0441-20	K2297-05	K2296-02	K2297-04
	Sampling Date	03/07/2012	03/08/2012	11/02/2011	11/01/2011	11/02/2011
	Sample Type	TB	TB	FB	FB	FB
Cadmium	7440-43-9					
Calcium	7440-70-2					
Chromium	7440-47-3					
Cobalt	7440-48-4					
Copper	7440-50-8					
Iron	7439-89-6					
Lead	7439-92-1					
Magnesium	7439-95-4					
Manganese	7439-96-5					
Mercury - SW7470	7439-97-6					
Nickel	7440-02-0					
Potassium	7440-09-7					
Selenium	7782-49-2					
Silver	7440-22-4					
Sodium	7440-23-5					
Thallium	7440-28-0					
Vanadium	7440-62-2					
Zinc	7440-66-6					

Sample ID		PM-TB-110311	PM-TB-1112011		PM-TB-1122011		TB-20110524A		TB-20110524B	
Lab Sample Number	K2297-13	K2296-01 11/01/2011 TB		K2297-06 11/02/2011 TB		K0918-02 05/24/2011 TB		K0909-11 05/24/2011 TB		
Sampling Date										
Sample Type										
<b>Chemical</b>										
<b>Volatile Organic Compounds</b>										
1,1,1,2-Tetrachloroethane	630-20-6	5 U		5 U		5 U		5 U		5 U
1,1,1-Trichloroethane	71-55-6	5 U		5 U		5 U		5 U		5 U
1,1,2,2-Tetrachloroethane	79-34-5	5 U		5 U		5 U		5 U		5 U
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1	5 U		5 U		5 U		5 U		5 U
1,1,2-Trichloroethane	79-00-5	5 U		5 U		5 U		5 U		5 U
1,1-Dichloroethane	75-34-3	5 U		5 U		5 U		5 U		5 U
1,1-Dichloroethene	75-35-4	5 U		5 U		5 U		5 U		5 U
1,1-Dichloropropene	563-58-6	5 U		5 U		5 U		5 U		5 U
1,2,3-Trichlorobenzene	87-61-6	5 U		5 UJ		5 UJ		5 U		5 U
1,2,3-Trichloropropane	96-18-4	5 UJ		5 U		5 U		5 U		5 U
1,2,4-Trichlorobenzene	120-82-1	5 U		5 U		5 U		5 U		5 U
1,2,4-Trimethylbenzene	95-63-6	5 U		5 U		5 U		5 U		5 U
1,2-Dibromo-3-Chloropropane	96-12-8	5 U		5 U		5 U		5 U		5 U
1,2-Dibromoethane (Edb)	106-93-4	5 U		5 U		5 U		5 U		5 U
1,2-Dichlorobenzene	95-50-1	5 U		5 U		5 U		5 U		5 U
1,2-Dichloroethane	107-06-2	5 U		5 U		5 U		5 U		5 U
1,2-Dichloropropane	78-87-5	5 U		5 U		5 U		5 U		5 U
1,3,5-Trimethylbenzene	108-67-8	5 U		5 U		5 U		5 U		5 U
1,3-Dichlorobenzene	541-73-1	5 U		5 U		5 U		5 U		5 U
1,3-Dichloropropane	142-28-9	5 U		5 U		5 U		5 U		5 U
1,4-Dichlorobenzene	106-46-7	5 U		5 U		5 U		5 U		5 U
2,2-Dichloropropane	594-20-7	5 U		5 U		5 U		5 U		5 U
2-Butanone (Mek)	78-93-3	5 R		5 R		5 R		5 R		5 R
2-Chlorotoluene	95-49-8	5 U		5 UJ		5 UJ		5 U		5 U
2-Hexanone	591-78-6	5 U		5 UJ		5 UJ		5 U		5 U
4-Chlorotoluene	106-43-4	5 U		5 U		5 U		5 U		5 U
4-Methyl-2-Pentanone (Mibk)	108-10-1	5 U		5 U		5 U		5 U		5 U
Acetone	67-64-1	5 R		5 R		5 R		5 R		5 R

	Sample ID Lab Sample Number Sampling Date Sample Type	PM-TB-110311 K2297-13 11/03/2011 TB	PM-TB-1112011 K2296-01 11/01/2011 TB	PM-TB-1122011 K2297-06 11/02/2011 TB	TB-20110524A K0918-02 05/24/2011 TB	TB-20110524B K0909-11 05/24/2011 TB
Benzene	71-43-2	5 U	5 U	5 U	5 U	5 R
Bromobenzene	108-86-1	5 U	5 U	5 U	5 U	5 U
Bromochloromethane	74-97-5	5 U	5 U	5 U	5 U	5 U
Bromodichloromethane	75-27-4	5 U	5 U	5 U	5 U	5 U
Bromoform	75-25-2	5 UJ	5 U	5 U	5 U	5 U
Bromomethane	74-83-9	5 U	5 U	5 U	5 U	5 U
Carbon Disulfide	75-15-0	5 U	5 U	5 U	5 U	5 U
Carbon Tetrachloride	56-23-5	5 U	5 U	5 U	5 U	5 U
Chlorobenzene	108-90-7	5 U	5 U	5 U	5 U	5 U
Chloroethane	75-00-3	5 U	5 U	5 U	5 U	5 U
Chloroform	67-66-3	5 U	5 U	5 U	5 U	5 U
Chloromethane	74-87-3	5 U	5 U	5 U	5 U	5 U
Cis-1,2-Dichloroethene	156-59-2	5 U	5 U	5 U	5 U	5 U
Cis-1,3-Dichloropropene	10061-01-5	5 U	5 U	5 U	5 U	5 U
Cyclohexane	110-82-7	5 U	5 U	5 U	5 U	5 U
Dibromochloromethane	124-48-1	5 U	5 U	5 U	5 U	5 U
Dibromomethane	74-95-3	5 U	5 U	5 U	5 U	5 U
Dichlorodifluoromethane	75-71-8	5 U	5 U	5 U	5 UJ	5 UJ
Ethylbenzene	100-41-4	5 U	5 U	5 U	5 U	5 U
Hexachlorobutadiene	87-68-3	5 U	5 UJ	5 UJ	5 U	5 U
Isopropylbenzene	98-82-8	5 U	5 U	5 U	5 U	5 U
M,P-Xylene	179601-23-1	5 U	5 U	5 U	5 U	5 U
Methyl Acetate	79-20-9	5 U	5 U	5 U	5 U	5 UJ
Methyl Iodide	74-88-4	5 U	5 U	5 U	5 U	5 U
Methyl Tert-Butyl Ether (Mtbe)	1634-04-4	5 U	5 U	5 U	5 U	5 U
Methylcyclohexane	108-87-2	5 U	5 U	5 U	5 U	5 U
Methylene Chloride	75-09-2	5 U	5 U	5 U	5 U	5 U
Naphthalene	91-20-3	5 U	5 UJ	5 UJ	5 U	5 U
N-Butylbenzene	104-51-8	5 U	5 U	5 U	5 U	5 U
N-Propylbenzene	103-65-1	5 U	5 U	5 U	5 U	5 U

	Sample ID Lab Sample Number	PM-TB-110311 K2297-13 11/03/2011 TB	PM-TB-1112011 K2296-01 11/01/2011 TB	PM-TB-1122011 K2297-06 11/02/2011 TB	TB-20110524A K0918-02 05/24/2011 TB	TB-20110524B K0909-11 05/24/2011 TB
O-Xylene	95-47-6	5 U	5 U	5 U	5 U	5 U
P-Isopropyltoluene	99-87-6	5 U	5 U	5 U	5 U	5 U
Sec-Butylbenzene	135-98-8	5 U	5 U	5 U	5 U	5 U
Styrene	100-42-5	5 U	5 U	5 U	5 U	5 U
Tert-Butylbenzene	98-06-6	5 U	5 U	5 U	5 U	5 U
Tetrachloroethene	127-18-4	5 UJ	5 UJ	5 UJ	5 U	5 U
Toluene	108-88-3	5 U	5 U	5 U	5 U	5 U
Total Xylenes	1330-20-7	5 U	5 U	5 U	5 U	5 U
Trans-1,2-Dichloroethene	156-60-5	5 U	5 U	5 U	5 U	5 U
Trans-1,3-Dichloropropene	10061-02-6	5 U	5 U	5 U	5 U	5 U
Trichloroethene	79-01-6	5 U	5 U	5 U	5 U	5 U
Trichlorofluoromethane	75-69-4	5 U	5 U	5 U	5 U	5 U
Vinyl Acetate	108-05-4	5 U	5 U	5 U	5 U	5 U
Vinyl Chloride	75-01-4	5 U	5 U	5 U	5 U	5 U
Semi-Volatile Organic Compounds						
1,1'-Biphenyl	92-52-4					
2,4,5-Trichlorophenol	95-95-4					
2,4,6-Trichlorophenol	88-06-2					
2,4-Dichlorophenol	120-83-2					
2,4-Dimethylphenol	105-67-9					
2,4-Dinitrophenol	51-28-5					
2,4-Dinitrotoluene	121-14-2					
2,6-Dinitrotoluene	606-20-2					
2-Chloronaphthalene	91-58-7					
2-Chlorophenol	95-57-8					
2-Methylnaphthalene	91-57-6					
2-Methylphenol	95-48-7					
2-Nitroaniline	88-74-4					
2-Nitrophenol	88-75-5					
3,3'-Dichlorobenzidine	91-94-1					

	Sample ID	PM-TB-110311	PM-TB-1112011	PM-TB-1122011	TB-20110524A	TB-20110524B
	Lab Sample Number	K2297-13	K2296-01	K2297-06	K0918-02	K0909-11
	Sampling Date	11/03/2011	11/01/2011	11/02/2011	05/24/2011	05/24/2011
	Sample Type	TB	TB	TB	TB	TB
3-Nitroaniline	99-09-2					
4,6-Dinitro-2-Methylphenol	534-52-1					
4-Bromophenyl-Phenylether	101-55-3					
4-Chloro-3-Methylphenol	59-50-7					
4-Chloroaniline	106-47-8					
4-Chlorophenyl-Phenylether	7005-72-3					
4-Methylphenol	106-44-5					
4-Nitroaniline	100-01-6					
4-Nitrophenol	100-02-7					
Acenaphthene	83-32-9					
Acenaphthylene	208-96-8					
Acetophenone	98-86-2					
Anthracene	120-12-7					
Atrazine	1912-24-9					
Benzaldehyde	100-52-7					
Benzo(A)Anthracene	56-55-3					
Benzo(A)Pyrene	50-32-8					
Benzo(B)Fluoranthene	205-99-2					
Benzo(G,H,I)Perylene	191-24-2					
Benzo(K)Fluoranthene	207-08-9					
Bis(2-Chloroethoxy)Methane	111-91-1					
Bis(2-Chloroethyl) Ether	111-44-4					
Bis(2-Ethylhexyl)Phthalate	117-81-7					
Bis-Chloroisopropyl Ether	108-60-1					
Butylbenzylphthalate	85-68-7					
Caprolactam	105-60-2					
Carbazole	86-74-8					
Chrysene	218-01-9					
Dibenzo(A,H)Anthracene	53-70-3					
Dibenzofuran	132-64-9					

	Sample ID	PM-TB-110311	PM-TB-1112011	PM-TB-1122011	TB-20110524A	TB-20110524B
	Lab Sample Number	K2297-13	K2296-01	K2297-06	K0918-02	K0909-11
	Sampling Date	11/03/2011	11/01/2011	11/02/2011	05/24/2011	05/24/2011
	Sample Type	TB	TB	TB	TB	TB
Diethylphthalate	84-66-2					
Dimethylphthalate	131-11-3					
Di-N-Butylphthalate	84-74-2					
Di-N-Octylphthalate	117-84-0					
Fluoranthene	206-44-0					
Fluorene	86-73-7					
Hexachlorobenzene	118-74-1					
Hexachlorobutadiene	87-68-3					
Hexachlorocyclopentadiene	77-47-4					
Hexachloroethane	67-72-1					
Indeno(1,2,3-Cd)Pyrene	193-39-5					
Isophorone	78-59-1					
Naphthalene	91-20-3					
Nitrobenzene	98-95-3					
N-Nitroso-Di-N-Propylamine	621-64-7					
N-Nitrosodiphenylamine	86-30-6					
Pentachlorophenol	87-86-5					
Phenanthrene	85-01-8					
Phenol	108-95-2					
Pyrene	129-00-0					
Methane, Ethane, Ethene						
Ethane	74-84-0					
Ethene	74-85-1					
Methane	74-82-8					
Pesticides						
4,4'-DDD	72-54-8					
4,4'-DDE	72-55-9					
4,4'-DDT	50-29-3					
Aldrin	309-00-2					
Alpha-Bhc	319-84-6					

	Sample ID Lab Sample Number	PM-TB-110311 K2297-13 11/03/2011 TB	PM-TB-1112011 K2296-01 11/01/2011 TB	PM-TB-1122011 K2297-06 11/02/2011 TB	TB-20110524A K0918-02 05/24/2011 TB	TB-20110524B K0909-11 05/24/2011 TB
Alpha-Chlordane	5103-71-9					
Beta-Bhc	319-85-7					
Delta-Bhc	319-86-8					
Dieldrin	60-57-1					
Endosulfan I	959-98-8					
Endosulfan II	33213-65-9					
Endosulfan Sulfate	1031-07-8					
Endrin	72-20-8					
Endrin Aldehyde	7421-93-4					
Endrin Ketone	53494-70-5					
Gamma-Bhc (Lindane)	58-89-9					
Gamma-Chlordane	5103-74-2					
Heptachlor	76-44-8					
Heptachlor Epoxide	1024-57-3					
Methoxychlor	72-43-5					
Toxaphene	8001-35-2					
<b>Polychlorinated Organic Compounds</b>						
Aroclor 1016	12674-11-2					
Aroclor 1221	11104-28-2					
Aroclor 1232	11141-16-5					
Aroclor 1242	53469-21-9					
Aroclor 1248	12672-29-6					
Aroclor 1254	11097-69-1					
Aroclor 1260	11096-82-5					
<b>Inorganics</b>						
Aluminum	7429-90-5					
Antimony	7440-36-0					
Arsenic	7440-38-2					
Barium	7440-39-3					
Beryllium	7440-41-7					

	Sample ID Lab Sample Number	PM-TB-110311 K2297-13 11/03/2011 TB	PM-TB-1112011 K2296-01 11/01/2011 TB	PM-TB-1122011 K2297-06 11/02/2011 TB	TB-20110524A K0918-02 05/24/2011 TB	TB-20110524B K0909-11 05/24/2011 TB
Cadmium	7440-43-9					
Calcium	7440-70-2					
Chromium	7440-47-3					
Cobalt	7440-48-4					
Copper	7440-50-8					
Iron	7439-89-6					
Lead	7439-92-1					
Magnesium	7439-95-4					
Manganese	7439-96-5					
Mercury - SW7470	7439-97-6					
Nickel	7440-02-0					
Potassium	7440-09-7					
Selenium	7782-49-2					
Silver	7440-22-4					
Sodium	7440-23-5					
Thallium	7440-28-0					
Vanadium	7440-62-2					
Zinc	7440-66-6					

Sample ID	TRIP BLANK	
Lab Sample Number	K0909-06	
Sampling Date	05/23/2011	
Sample Type	TB	
Chemical	CAS#	
<b>Volatile Organic Compounds</b>		
1,1,1,2-Tetrachloroethane	630-20-6	5 U
1,1,1-Trichloroethane	71-55-6	5 U
1,1,2,2-Tetrachloroethane	79-34-5	5 U
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1	5 U
1,1,2-Trichloroethane	79-00-5	5 U
1,1-Dichloroethane	75-34-3	5 U
1,1-Dichloroethene	75-35-4	5 U
1,1-Dichloropropene	563-58-6	5 U
1,2,3-Trichlorobenzene	87-61-6	5 U
1,2,3-Trichloropropane	96-18-4	5 U
1,2,4-Trichlorobenzene	120-82-1	5 U
1,2,4-Trimethylbenzene	95-63-6	5 U
1,2-Dibromo-3-Chloropropane	96-12-8	5 U
1,2-Dibromoethane (Edb)	106-93-4	5 U
1,2-Dichlorobenzene	95-50-1	5 U
1,2-Dichloroethane	107-06-2	5 U
1,2-Dichloropropane	78-87-5	5 U
1,3,5-Trimethylbenzene	108-67-8	5 U
1,3-Dichlorobenzene	541-73-1	5 U
1,3-Dichloropropane	142-28-9	5 U
1,4-Dichlorobenzene	106-46-7	5 U
2,2-Dichloropropane	594-20-7	5 U
2-Butanone (Mek)	78-93-3	5 R
2-Chlorotoluene	95-49-8	5 U
2-Hexanone	591-78-6	5 U
4-Chlorotoluene	106-43-4	5 U
4-Methyl-2-Pentanone (Mibk)	108-10-1	5 U
Acetone	67-64-1	5 U

	Sample ID Lab Sample Number	TRIP BLANK K0909-06 05/23/2011 TB
Benzene	71-43-2	5 R
Bromobenzene	108-86-1	5 U
Bromo(chloromethane)	74-97-5	5 U
Bromodichloromethane	75-27-4	5 U
Bromoform	75-25-2	5 U
Bromomethane	74-83-9	5 U
Carbon Disulfide	75-15-0	5 U
Carbon Tetrachloride	56-23-5	5 U
Chlorobenzene	108-90-7	5 U
Chloroethane	75-00-3	5 U
Chloroform	67-66-3	5 U
Chloromethane	74-87-3	5 U
Cis-1,2-Dichloroethene	156-59-2	5 U
Cis-1,3-Dichloropropene	10061-01-5	5 U
Cyclohexane	110-82-7	5 U
Dibromochloromethane	124-48-1	5 U
Dibromomethane	74-95-3	5 U
Dichlorodifluoromethane	75-71-8	5 UJ
Ethylbenzene	100-41-4	5 U
Hexachlorobutadiene	87-68-3	5 U
Isopropylbenzene	98-82-8	5 U
M,P-Xylene	179601-23-1	5 U
Methyl Acetate	79-20-9	5 UJ
Methyl Iodide	74-88-4	5 U
Methyl Tert-Butyl Ether (Mtbe)	1634-04-4	5 U
Methylcyclohexane	108-87-2	5 U
Methylene Chloride	75-09-2	5 U
Naphthalene	91-20-3	5 U
N-Butylbenzene	104-51-8	5 U
N-Propylbenzene	103-65-1	5 U

	Sample ID Lab Sample Number	TRIP BLANK K0909-06 05/23/2011 TB
O-Xylene	95-47-6	5 U
P-Isopropyltoluene	99-87-6	5 U
Sec-Butylbenzene	135-98-8	5 U
Styrene	100-42-5	5 U
Tert-Butylbenzene	98-06-6	5 U
Tetrachloroethene	127-18-4	5 U
Toluene	108-88-3	5 U
Total Xylenes	1330-20-7	5 U
Trans-1,2-Dichloroethene	156-60-5	5 U
Trans-1,3-Dichloropropene	10061-02-6	5 U
Trichloroethene	79-01-6	5 U
Trichlorofluoromethane	75-69-4	5 U
Vinyl Acetate	108-05-4	5 U
Vinyl Chloride	75-01-4	5 U
Semi-Volatile Organic Compounds		
1,1'-Biphenyl	92-52-4	
2,4,5-Trichlorophenol	95-95-4	
2,4,6-Trichlorophenol	88-06-2	
2,4-Dichlorophenol	120-83-2	
2,4-Dimethylphenol	105-67-9	
2,4-Dinitrophenol	51-28-5	
2,4-Dinitrotoluene	121-14-2	
2,6-Dinitrotoluene	606-20-2	
2-Chloronaphthalene	91-58-7	
2-Chlorophenol	95-57-8	
2-Methylnaphthalene	91-57-6	
2-Methylphenol	95-48-7	
2-Nitroaniline	88-74-4	
2-Nitrophenol	88-75-5	
3,3'-Dichlorobenzidine	91-94-1	

Sample ID	TRIP BLANK
Lab Sample Number	K0909-06
Sampling Date	05/23/2011
Sample Type	TB
3-Nitroaniline	99-09-2
4,6-Dinitro-2-Methylphenol	534-52-1
4-Bromophenyl-Phenylether	101-55-3
4-Chloro-3-Methylphenol	59-50-7
4-Chloroaniline	106-47-8
4-Chlorophenyl-Phenylether	7005-72-3
4-Methylphenol	106-44-5
4-Nitroaniline	100-01-6
4-Nitrophenol	100-02-7
Acenaphthene	83-32-9
Acenaphthylene	208-96-8
Acetophenone	98-86-2
Anthracene	120-12-7
Atrazine	1912-24-9
Benzaldehyde	100-52-7
Benzo(A)Anthracene	56-55-3
Benzo(A)Pyrene	50-32-8
Benzo(B)Fluoranthene	205-99-2
Benzo(G,H,I)Perylene	191-24-2
Benzo(K)Fluoranthene	207-08-9
Bis(2-Chloroethoxy)Methane	111-91-1
Bis(2-Chloroethyl) Ether	111-44-4
Bis(2-Ethylhexyl)Phthalate	117-81-7
Bis-Chloroisopropyl Ether	108-60-1
Butylbenzylphthalate	85-68-7
Caprolactam	105-60-2
Carbazole	86-74-8
Chrysene	218-01-9
Dibenzo(A,H)Anthracene	53-70-3
Dibenzofuran	132-64-9

Sample ID	TRIP BLANK
Lab Sample Number	K0909-06
Sampling Date	05/23/2011
Sample Type	TB
Diethylphthalate	84-66-2
Dimethylphthalate	131-11-3
Di-N-Butylphthalate	84-74-2
Di-N-Octylphthalate	117-84-0
Fluoranthene	206-44-0
Fluorene	86-73-7
Hexachlorobenzene	118-74-1
Hexachlorobutadiene	87-68-3
Hexachlorocyclopentadiene	77-47-4
Hexachloroethane	67-72-1
Indeno(1,2,3-Cd)Pyrene	193-39-5
Isophorone	78-59-1
Naphthalene	91-20-3
Nitrobenzene	98-95-3
N-Nitroso-Di-N-Propylamine	621-64-7
N-Nitrosodiphenylamine	86-30-6
Pentachlorophenol	87-86-5
Phenanthrene	85-01-8
Phenol	108-95-2
Pyrene	129-00-0
<b>Methane, Ethane, Ethene</b>	
Ethane	74-84-0
Ethene	74-85-1
Methane	74-82-8
<b>Pesticides</b>	
4,4'-DDD	72-54-8
4,4'-DDE	72-55-9
4,4'-DDT	50-29-3
Aldrin	309-00-2
Alpha-Bhc	319-84-6

Sample ID	TRIP BLANK		
Lab Sample Number	K0909-06		
Sampling Date	05/23/2011		
Sample Type	TB		
Alpha-Chlordane	5103-71-9		
Beta-Bhc	319-85-7		
Delta-Bhc	319-86-8		
Dieldrin	60-57-1		
Endosulfan I	959-98-8		
Endosulfan II	33213-65-9		
Endosulfan Sulfate	1031-07-8		
Endrin	72-20-8		
Endrin Aldehyde	7421-93-4		
Endrin Ketone	53494-70-5		
Gamma-Bhc (Lindane)	58-89-9		
Gamma-Chlordane	5103-74-2		
Heptachlor	76-44-8		
Heptachlor Epoxide	1024-57-3		
Methoxychlor	72-43-5		
Toxaphene	8001-35-2		
<b>Polychlorinated Organic Compounds</b>			
Aroclor 1016	12674-11-2		
Aroclor 1221	11104-28-2		
Aroclor 1232	11141-16-5		
Aroclor 1242	53469-21-9		
Aroclor 1248	12672-29-6		
Aroclor 1254	11097-69-1		
Aroclor 1260	11096-82-5		
<b>Inorganics</b>			
Aluminum	7429-90-5		
Antimony	7440-36-0		
Arsenic	7440-38-2		
Barium	7440-39-3		
Beryllium	7440-41-7		

Sample ID	Lab Sample Number	Sampling Date	TRIP BLANK
			K0909-06
			05/23/2011
			TB
Cadmium	7440-43-9		
Calcium	7440-70-2		
Chromium	7440-47-3		
Cobalt	7440-48-4		
Copper	7440-50-8		
Iron	7439-89-6		
Lead	7439-92-1		
Magnesium	7439-95-4		
Manganese	7439-96-5		
Mercury - SW7470	7439-97-6		
Nickel	7440-02-0		
Potassium	7440-09-7		
Selenium	7782-49-2		
Silver	7440-22-4		
Sodium	7440-23-5		
Thallium	7440-28-0		
Vanadium	7440-62-2		
Zinc	7440-66-6		

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP1-16.5 MIP1 05/24/2011	80230-MIP1B-14 MIP1B 05/25/2011	80230-MIP1C-26 MIP1C 05/26/2011	80230-MIP1D-37.5 MIP1D 05/26/2011
Chemical Name	CAS#	Part 375 Unrestricted					
<b>Volatile Organic Compounds (µg/kg)</b>							
1,1,1,2-Tetrachloroethane	630-20-6		5.2 U	5.4 U	5.3 U	5.3 U	
1,1,1-Trichloroethane	71-55-6	680	5.2 U	5.4 U	5.3 U	5.3 U	
1,1,2,2-Tetrachloroethane	79-34-5		5.2 U	5.4 U	5.3 U	5.3 U	
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5.2 U	5.4 U	5.3 U	5.3 U	
1,1,2-Trichloroethane	79-00-5		5.2 U	5.4 U	5.3 U	5.3 U	
1,1-Dichloroethane	75-34-3	270	5.2 U	5.4 U	5.3 U	5.3 U	
1,1-Dichloroethene	75-35-4	330	5.2 U	5.4 U	5.3 U	5.3 U	
1,1-Dichloropropene	563-58-6		5.2 U	5.4 U	5.3 U	5.3 U	
1,2,3-Trichlorobenzene	87-61-6		5.2 U	5.4 U	5.3 U	5.3 U	
1,2,3-Trichloropropane	96-18-4		5.2 U	5.4 U	5.3 U	5.3 U	
1,2,4-Trichlorobenzene	120-82-1		5.2 U	5.4 U	5.3 U	5.3 U	
1,2,4-Trimethylbenzene	95-63-6	3600	5.2 U	5.4 U	5.3 U	5.3 U	
1,2-Dibromo-3-Chloropropane	96-12-8		5.2 UJ	5.4 U	5.3 U	5.3 U	
1,2-Dibromoethane (EDB)	106-93-4		5.2 U	5.4 U	5.3 U	5.3 U	
1,2-Dichlorobenzene	95-50-1	1100	5.2 U	5.4 U	5.3 U	5.3 U	
1,2-Dichloroethane	107-06-2	20	5.2 U	5.4 U	5.3 U	5.3 U	
1,2-Dichloropropane	78-87-5		5.2 U	5.4 U	5.3 U	5.3 U	
1,3,5-Trimethylbenzene	108-67-8	8400	5.2 U	5.4 U	5.3 U	5.3 U	
1,3-Dichlorobenzene	541-73-1	2400	5.2 U	5.4 U	5.3 U	5.3 U	
1,3-Dichloropropane	142-28-9		5.2 U	5.4 U	5.3 U	5.3 U	
1,4-Dichlorobenzene	106-46-7	1800	5.2 U	5.4 U	5.3 U	5.3 U	
2,2-Dichloropropane	594-20-7		5.2 U	5.4 U	5.3 U	5.3 U	
2-Butanone (MEK)	78-93-3	120	5.2 R	5.4 R	5.3 R	5.3 R	
2-Chlorotoluene	95-49-8		5.2 U	5.4 U	5.3 U	5.3 U	
2-Hexanone	591-78-6		5.2 U	5.4 U	5.3 U	5.3 U	
4-Chlorotoluene	106-43-4		5.2 U	5.4 U	5.3 U	5.3 U	
4-Methyl-2-Pentanone (MIBK)	108-10-1		5.2 U	5.4 U	5.3 U	5.3 U	
Acetone	67-64-1	50	5.2 R	16 R	5.3 R	5.3 R	
Benzene	71-43-2	60	5.2 U	5.4 U	5.3 U	5.3 U	
Bromobenzene	108-86-1		5.2 U	5.4 U	5.3 U	5.3 U	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP1-16.5	80230-MIP1B-14	80230-MIP1C-26	80230-MIP1D-37.5
			Sample Location	MIP1 05/24/2011	MIP1B 05/25/2011	MIP1C 05/26/2011	MIP1D 05/26/2011
Bromochloromethane	74-97-5			5.2 U	5.4 U	5.3 U	5.3 U
Bromodichloromethane	75-27-4			5.2 U	5.4 U	5.3 U	5.3 U
Bromoform	75-25-2			5.2 U	5.4 U	5.3 U	5.3 U
Bromomethane	74-83-9			5.2 U	5.4 U	5.3 U	5.3 U
Carbon Disulfide	75-15-0			5.2 U	5.4 U	5.3 U	5.3 U
Carbon Tetrachloride	56-23-5	760		5.2 U	5.4 U	5.3 U	5.3 U
Chlorobenzene	108-90-7	1100		5.2 U	5.4 U	5.3 U	5.3 U
Chloroethane	75-00-3			5.2 U	5.4 U	5.3 U	5.3 U
Chloroform	67-66-3	370		5.2 U	5.4 U	5.3 U	5.3 U
Chloromethane	74-87-3			5.2 U	5.4 U	5.3 U	5.3 U
cis-1,2-Dichloroethene	156-59-2	250		5.2 U	1.3 J	5.3 U	5.3 U
cis-1,3-Dichloropropene	10061-01-5			5.2 U	5.4 U	5.3 U	5.3 U
Cyclohexane	110-82-7			5.2 U	5.4 U	5.3 U	5.3 U
Dibromochloromethane	124-48-1			5.2 U	5.4 U	5.3 U	5.3 U
Dibromomethane	74-95-3			5.2 U	5.4 U	5.3 U	5.3 U
Dichlorodifluoromethane	75-71-8			5.2 U	5.4 U	5.3 U	5.3 U
Ethylbenzene	100-41-4	1000		5.2 U	5.4 U	5.3 U	5.3 U
Hexachlorobutadiene	87-68-3			5.2 U	5.4 U	5.3 U	5.3 U
Isopropylbenzene	98-82-8			5.2 U	5.4 U	5.3 U	5.3 U
m,p-Xylene	179601-23-1	260		5.2 U	5.4 U	5.3 U	5.3 U
Methyl Acetate	79-20-9			5.2 U	5.4 U	5.3 U	5.3 U
Methyl Iodide	74-88-4			5.2 U	5.4 U	5.3 U	5.3 U
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	930		5.2 U	5.4 U	5.3 U	5.3 U
Methylcyclohexane	108-87-2			5.2 U	5.4 U	5.3 U	5.3 U
Methylene Chloride	75-09-2	50		2.4 J	5.4 U	5.3 U	5.3 U
Naphthalene	91-20-3	12000		5.2 UJ	5.4 U	5.3 U	5.3 U
n-Butylbenzene	104-51-8	12000		5.2 U	5.4 U	5.3 U	5.3 U
n-Propylbenzene	103-65-1	3900		5.2 U	5.4 U	5.3 U	5.3 U
o-Xylene	95-47-6	260		5.2 U	5.4 U	5.3 U	5.3 U
p-Isopropyltoluene	99-87-6			5.2 U	5.4 U	5.3 U	5.3 U
sec-Butylbenzene	135-98-8	11000		5.2 U	5.4 U	5.3 U	5.3 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP1-16.5 MIP1 05/24/2011	80230-MIP1B-14 MIP1B 05/25/2011	80230-MIP1C-26 MIP1C 05/26/2011	80230-MIP1D-37.5 MIP1D 05/26/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Styrene	100-42-5			5.2 U	5.4 U	5.3 U	5.3 U
tert-Butylbenzene	98-06-6	5900		5.2 U	5.4 U	5.3 U	5.3 U
Tetrachloroethene	127-18-4	1300		8.1	5.4 U	5.3 U	5.3 U
Toluene	108-88-3	700		5.2 U	5.4 U	5.3 U	5.3 U
Total Xylenes	1330-20-7	260		5.2 U	5.4 U	5.3 U	5.3 U
trans-1,2-Dichloroethene	156-60-5	190		5.2 U	5.4 U	5.3 U	5.3 U
trans-1,3-Dichloropropene	10061-02-6			5.2 U	5.4 U	5.3 U	5.3 U
Trichloroethene	79-01-6	470		5.2 U	5.4 U	5.3 U	5.3 U
Trichlorofluoromethane	75-69-4			5.2 U	5.4 U	5.3 U	5.3 U
Vinyl Acetate	108-05-4			5.2 U	5.4 U	5.3 U	5.3 U
Vinyl Chloride	75-01-4	20		5.2 U	5.4 U	5.3 U	5.3 U
<b>Semi-Volatile Organic Compounds (µg/kg)</b>							
1,1'-Biphenyl	92-52-4			380 U	360 U	370 U	350 U
2,4,5-Trichlorophenol	95-95-4			780 U	740 U	750 U	720 U
2,4,6-Trichlorophenol	88-06-2			380 U	360 U	370 U	350 U
2,4-Dichlorophenol	120-83-2			380 U	360 U	370 U	350 U
2,4-Dimethylphenol	105-67-9			380 U	360 U	370 U	350 U
2,4-Dinitrophenol	51-28-5			780 UJ	740 UJ	750 UJ	720 UJ
2,4-Dinitrotoluene	121-14-2			380 U	360 U	370 U	350 U
2,6-Dinitrotoluene	606-20-2			380 U	360 U	370 U	350 U
2-Chloronaphthalene	91-58-7			380 U	360 U	370 U	350 U
2-Chlorophenol	95-57-8			380 U	360 U	370 U	350 U
2-Methylnaphthalene	91-57-6			380 UJ	360 UJ	370 UJ	350 UJ
2-Methylphenol	95-48-7	330		380 U	360 U	370 U	350 U
2-Nitroaniline	88-74-4			780 U	740 U	750 U	720 U
2-Nitrophenol	88-75-5			380 U	360 U	370 U	350 U
3,3'-Dichlorobenzidine	91-94-1			380 U	360 U	370 U	350 U
3-Nitroaniline	99-09-2			780 U	740 U	750 U	720 U
4,6-Dinitro-2-Methylphenol	534-52-1			780 U	740 U	750 U	720 U
4-Bromophenyl-Phenylether	101-55-3			380 U	360 U	370 U	350 U
4-Chloro-3-Methylphenol	59-50-7			380 U	360 U	370 U	350 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP1-16.5	80230-MIP1B-14	80230-MIP1C-26	80230-MIP1D-37.5
			Sample Location	MIP1 05/24/2011	MIP1B 05/25/2011	MIP1C 05/26/2011	MIP1D 05/26/2011
4-Chloroaniline	106-47-8			380 U	360 U	370 U	350 U
4-Chlorophenyl-Phenylether	7005-72-3			380 U	360 U	370 U	350 U
4-Methylphenol	106-44-5	330		380 U	360 U	370 U	350 U
4-Nitroaniline	100-01-6			780 U	740 U	750 U	720 U
4-Nitrophenol	100-02-7			780 U	740 U	750 U	720 U
Acenaphthene	83-32-9	20000		380 U	360 U	370 U	350 U
Acenaphthylene	208-96-8	100000		380 U	360 U	370 U	350 U
Acetophenone	98-86-2			380 U	360 U	370 U	350 U
Anthracene	120-12-7	100000		380 U	360 U	370 U	350 U
Atrazine	1912-24-9			380 U	360 U	370 U	350 U
Benzaldehyde	100-52-7			380 U	360 U	370 U	350 U
Benzo(a)Anthracene	56-55-3	1000		380 U	360 U	370 U	350 U
Benzo(a)Pyrene	50-32-8	1000		380 U	360 U	370 U	350 U
Benzo(b)Fluoranthene	205-99-2	1000		380 U	360 U	370 U	350 U
Benzo(g,h,i)Perylene	191-24-2	100000		380 U	360 U	370 U	350 U
Benzo(k)Fluoranthene	207-08-9	800		380 U	360 U	370 U	350 U
bis(2-Chloroethoxy)Methane	111-91-1			380 U	360 U	370 U	350 U
bis(2-Chloroethyl) Ether	111-44-4			380 U	360 U	370 U	350 U
bis(2-Ethylhexyl)Phthalate	117-81-7			360 J	360 U	370 U	350 U
bis-Chloroisopropyl Ether	108-60-1			380 U	360 U	370 U	350 U
Butylbenzylphthalate	85-68-7			380 U	360 U	370 U	350 U
Caprolactam	105-60-2			380 U	360 U	370 U	350 U
Carbazole	86-74-8			380 U	360 U	370 U	350 U
Chrysene	218-01-9	1000		380 U	360 U	370 U	350 U
Dibenzo(a,h)Anthracene	53-70-3	330		380 U	360 U	370 U	350 U
Dibenzofuran	132-64-9	7000		380 U	360 U	370 U	350 U
Diethylphthalate	84-66-2			380 U	360 U	370 U	350 U
Dimethylphthalate	131-11-3			380 U	360 U	370 U	350 U
di-n-Butylphthalate	84-74-2			380 U	360 U	370 U	350 U
di-n-Octylphthalate	117-84-0			380 U	360 U	370 U	350 U
Fluoranthene	206-44-0	100000		380 U	360 U	50 J	55 J

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP1-16.5 MIP1 05/24/2011	80230-MIP1B-14 MIP1B 05/25/2011	80230-MIP1C-26 MIP1C 05/26/2011	80230-MIP1D-37.5 MIP1D 05/26/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Fluorene	86-73-7	30000	380 U	360 U	370 U	350 U	
Hexachlorobenzene	118-74-1	330	380 U	360 U	370 U	350 U	
Hexachlorobutadiene	87-68-3		380 U	360 U	370 U	350 U	
Hexachlorocyclopentadiene	77-47-4		380 U	360 U	370 U	350 U	
Hexachloroethane	67-72-1		380 U	360 U	370 U	350 U	
Indeno(1,2,3-cd)Pyrene	193-39-5	500	380 U	360 U	370 U	350 U	
Isophorone	78-59-1		380 U	360 U	370 U	350 U	
Naphthalene	91-20-3	12000	380 U	360 U	370 U	350 U	
Nitrobenzene	98-95-3		380 U	360 UJ	370 UJ	350 UJ	
n-Nitroso-di-n-Propylamine	621-64-7		380 U	360 U	370 U	350 U	
n-Nitrosodiphenylamine	86-30-6		380 U	360 U	370 U	350 U	
Pentachlorophenol	87-86-5	800	780 U	740 U	750 U	720 U	
Phenanthrene	85-01-8	100000	380 U	360 U	370 U	350 U	
Phenol	108-95-2	330	380 U	360 U	370 U	350 U	
Pyrene	129-00-0	100000	380 U	360 U	37 J	45 J	
<b>Pesticides (µg/kg)</b>							
4,4'-DDD	72-54-8	3.3		3.7 U			3.6 U
4,4'-DDE	72-55-9	3.3		3.7 U			3.6 U
4,4'-DDT	50-29-3	3.3		3.7 U			3.6 U
Aldrin	309-00-2	5		1.9 U			1.8 U
Alpha-BHC	319-84-6	20		1.9 U			1.8 U
Alpha-Chlordane	5103-71-9	94		1.9 U			1.8 U
Beta-BHC	319-85-7	36		1.9 U			1.8 U
Delta-BHC	319-86-8	40		1.9 U			1.8 U
Dieldrin	60-57-1	5		3.7 U			3.6 U
Endosulfan I	959-98-8	2400		1.9 U			1.8 U
Endosulfan II	33213-65-9	2400		3.7 U			3.6 U
Endosulfan Sulfate	1031-07-8	2400		3.7 U			3.6 U
Endrin	72-20-8	14		3.7 U			3.6 U
Endrin Aldehyde	7421-93-4			3.7 U			3.6 U
Endrin Ketone	53494-70-5			3.7 U			3.6 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP1-16.5 MIP1 05/24/2011	80230-MIP1B-14 MIP1B 05/25/2011	80230-MIP1C-26 MIP1C 05/26/2011	80230-MIP1D-37.5 MIP1D 05/26/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Gamma-BHC (Lindane)	58-89-9	100		1.9 U			1.8 U
Gamma-Chlordane	5103-74-2			1.9 U			1.8 U
Heptachlor	76-44-8	42		1.9 U			1.8 U
Heptachlor Epoxide	1024-57-3			1.9 U			1.8 U
Methoxychlor	72-43-5			19 U			18 U
Toxaphene	8001-35-2			190 U			180 U
<b>Polychlorinated Biphenyls (µg/kg)</b>							
Aroclor 1016	12674-11-2	100		37 U			36 U
Aroclor 1221	11104-28-2	100		37 U			36 U
Aroclor 1232	11141-16-5	100		37 U			36 U
Aroclor 1242	53469-21-9	100		37 U			36 U
Aroclor 1248	12672-29-6	100		37 U			36 U
Aroclor 1254	11097-69-1	100		37 U			36 U
Aroclor 1260	11096-82-5	100		37 U			36 U
<b>Inorganics (mg/kg)</b>							
Aluminum	7429-90-5			10800			5510
Antimony	7440-36-0			0.75 UNJ			0.35 BNJ
Arsenic	7440-38-2	13		2.4 *J			2.4 *J
Barium	7440-39-3	350		81.2			69.3 *J
Beryllium	7440-41-7	7.2		0.86			0.59
Cadmium	7440-43-9	2.5		0.18 B			0.2 B
Calcium	7440-70-2			2900			947
Chromium	7440-47-3	30		26.3 *J			16.3
Cobalt	7440-48-4			11.2 *J			6.4 *E
Copper	7440-50-8	50		30.3 *J			16.4 *J
Iron	7439-89-6			15600 *J			16200 *J
Lead	7439-92-1	63		24.4			6.2
Magnesium	7439-95-4			5820 *J			3230
Manganese	7439-96-5	1600		194			336 *J
Mercury	7439-97-6	0.18		0.005 B			
Nickel	7440-02-0	30		31			14.7

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID 80230-MIP1-16.5	MIP1 05/24/2011	80230-MIP1B-14 MIP1B 05/25/2011	80230-MIP1C-26 MIP1C 05/26/2011	80230-MIP1D-37.5 MIP1D 05/26/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Potassium	7440-09-7			1950	*J		1170
Selenium	7782-49-2	3.9		0.51	B		0.57
Silver	7440-22-4	2		1.1	U		1.1
Sodium	7440-23-5			720	EJ		90.3
Thallium	7440-28-0			2.8			1.3
Vanadium	7440-62-2			39.3	J*		23.8
Zinc	7440-66-6	109		61.3	*J		30.2

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP2-30 MIP2 05/24/2011	80230-MIP2B-40 MIP2B 05/26/2011	80230-MIP2C-35 MIP2C 06/16/2011	80230-MIP2C-35ME MIP2C 06/16/2011
Chemical Name	CAS#	Part 375 Unrestricted					
<b>Volatile Organic Compounds (µg/kg)</b>							
1,1,1,2-Tetrachloroethane	630-20-6		5.2 U	5.8 U	6.2 U	370 U	
1,1,1-Trichloroethane	71-55-6	680	5.2 U	5.8 U	6.2 U	370 U	
1,1,2,2-Tetrachloroethane	79-34-5		5.2 U	5.8 U	6.2 U	370 U	
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5.2 U	5.8 U	6.2 U	370 U	
1,1,2-Trichloroethane	79-00-5		5.2 U	5.8 U	6.2 U	370 U	
1,1-Dichloroethane	75-34-3	270	5.2 U	5.8 U	6.2 U	370 U	
1,1-Dichloroethene	75-35-4	330	5.2 U	5.8 U	6.2 U	370 U	
1,1-Dichloropropene	563-58-6		5.2 U	5.8 U	6.2 U	370 U	
1,2,3-Trichlorobenzene	87-61-6		5.2 U	5.8 U	6.2 UJ	370 UJ	
1,2,3-Trichloropropane	96-18-4		5.2 U	5.8 U	6.2 U	370 U	
1,2,4-Trichlorobenzene	120-82-1		5.2 U	5.8 U	6.2 UJ	370 UJ	
1,2,4-Trimethylbenzene	95-63-6	3600	5.2 U	5.8 U	6.2 U	370 U	
1,2-Dibromo-3-Chloropropane	96-12-8		5.2 UJ	5.8 U	6.2 U	370 UJ	
1,2-Dibromoethane (EDB)	106-93-4		5.2 U	5.8 U	6.2 U	370 U	
1,2-Dichlorobenzene	95-50-1	1100	5.2 U	5.8 U	6.2 U	370 U	
1,2-Dichloroethane	107-06-2	20	5.2 U	5.8 U	6.2 U	370 U	
1,2-Dichloropropane	78-87-5		5.2 U	5.8 U	3.9 J	370 U	
1,3,5-Trimethylbenzene	108-67-8	8400	5.2 U	5.8 U	6.2 U	370 U	
1,3-Dichlorobenzene	541-73-1	2400	5.2 U	5.8 U	6.2 U	370 U	
1,3-Dichloropropane	142-28-9		5.2 U	5.8 U	6.2 U	370 U	
1,4-Dichlorobenzene	106-46-7	1800	5.2 U	5.8 U	6.2 U	370 U	
2,2-Dichloropropane	594-20-7		5.2 U	5.8 U	6.2 UJ	370 UJ	
2-Butanone (MEK)	78-93-3	120	5.2 R	5.8 R	6.2 R	370 R	
2-Chlorotoluene	95-49-8		5.2 U	5.8 U	6.2 U	370 U	
2-Hexanone	591-78-6		5.2 U	5.8 U	6.2 UJ	370 UJ	
4-Chlorotoluene	106-43-4		5.2 U	5.8 U	6.2 U	370 U	
4-Methyl-2-Pentanone (MIBK)	108-10-1		5.2 U	5.8 U	6.2 U	370 U	
Acetone	67-64-1	50	5.2 R	4.3 J	6.2 R	370 R	
Benzene	71-43-2	60	5.2 U	5.8 U	6.2 U	370 U	
Bromobenzene	108-86-1		5.2 U	5.8 U	6.2 U	370 U	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Sample ID Sample Location Sampling Date	80230-MIP2-30	80230-MIP2B-40	80230-MIP2C-35	80230-MIP2C-35ME
			MIP2 05/24/2011	MIP2B 05/26/2011	MIP2C 06/16/2011	MIP2C 06/16/2011
Bromochloromethane	74-97-5	Part 375 Unrestricted	5.2 U	5.8 U	6.2 UJ	370 UJ
Bromodichloromethane	75-27-4		5.2 U	5.8 U	6.2 U	370 U
Bromoform	75-25-2		5.2 U	5.8 U	6.2 UJ	370 UJ
Bromomethane	74-83-9		5.2 U	5.8 U	6.2 U	370 U
Carbon Disulfide	75-15-0		5.2 U	5.8 U	6.2 U	370 U
Carbon Tetrachloride	56-23-5	760	5.2 U	5.8 U	6.2 U	370 U
Chlorobenzene	108-90-7	1100	5.2 U	5.8 U	6.2 U	370 U
Chloroethane	75-00-3		5.2 U	5.8 U	6.2 U	370 U
Chloroform	67-66-3	370	5.2 U	5.8 U	6.2 U	370 U
Chloromethane	74-87-3		5.2 U	5.8 U	6.2 U	370 U
cis-1,2-Dichloroethene	156-59-2	250	1.5 J	5.8 U	55	370 U
cis-1,3-Dichloropropene	10061-01-5		5.2 U	5.8 U	6.2 UJ	370 UJ
Cyclohexane	110-82-7		5.2 U	5.8 U	6.2 U	370 U
Dibromochloromethane	124-48-1		5.2 U	5.8 U	6.2 U	370 U
Dibromomethane	74-95-3		5.2 U	5.8 U	6.2 U	370 U
Dichlorodifluoromethane	75-71-8		5.2 U	5.8 U	6.2 U	370 UJ
Ethylbenzene	100-41-4	1000	5.2 U	5.8 U	6.2 U	370 U
Hexachlorobutadiene	87-68-3		5.2 U	5.8 U	6.2 UJ	370 UJ
Isopropylbenzene	98-82-8		5.2 U	5.8 U	6.2 U	370 U
m,p-Xylene	179601-23-1	260	5.2 U	5.8 U	6.2 U	370 U
Methyl Acetate	79-20-9		5.2 U	5.8 U	6.2 UJ	370 UJ
Methyl Iodide	74-88-4		5.2 U	5.8 U	6.2 U	370 U
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	930	5.2 U	5.8 U	6.2 U	370 U
Methylcyclohexane	108-87-2		5.2 U	5.8 U	6.2 U	370 U
Methylene Chloride	75-09-2	50	2.6 J	5.8 U	6.2 U	370 U
Naphthalene	91-20-3	12000	5.2 UJ	5.8 U	6.2 UJ	370 UJ
n-Butylbenzene	104-51-8	12000	5.2 U	5.8 U	6.2 UJ	370 UJ
n-Propylbenzene	103-65-1	3900	5.2 U	5.8 U	6.2 U	370 U
o-Xylene	95-47-6	260	5.2 U	5.8 U	6.2 U	370 U
p-Isopropyltoluene	99-87-6		5.2 U	5.8 U	6.2 U	370 U
sec-Butylbenzene	135-98-8	11000	5.2 U	5.8 U	6.2 U	370 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP2-30 MIP2 05/24/2011	80230-MIP2B-40 MIP2B 05/26/2011	80230-MIP2C-35 MIP2C 06/16/2011	80230-MIP2C-35ME MIP2C 06/16/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Styrene	100-42-5			5.2 U	5.8 U	6.2 U	370 U
tert-Butylbenzene	98-06-6	5900		5.2 U	5.8 U	6.2 U	370 U
Tetrachloroethene	127-18-4	1300		53	5.8 U	840 E	1900 J
Toluene	108-88-3	700		5.2 U	5.8 U	6.2 U	370 U
Total Xylenes	1330-20-7	260		5.2 U	5.8 U	6.2 U	370 U
trans-1,2-Dichloroethene	156-60-5	190		5.2 U	5.8 U	6.2 U	370 U
trans-1,3-Dichloropropene	10061-02-6			5.2 U	5.8 U	6.2 UJ	370 UJ
Trichloroethene	79-01-6	470		5.2 U	5.8 U	7	370 U
Trichlorofluoromethane	75-69-4			5.2 U	5.8 U	6.2 U	370 U
Vinyl Acetate	108-05-4			5.2 U	5.8 U	6.2 UJ	370 UJ
Vinyl Chloride	75-01-4	20		5.2 U	5.8 U	6.2 U	370 U
<b>Semi-Volatile Organic Compounds (µg/kg)</b>							
1,1'-Biphenyl	92-52-4			350 U	390 U	320 U	
2,4,5-Trichlorophenol	95-95-4			710 U	790 U	660 U	
2,4,6-Trichlorophenol	88-06-2			350 U	390 U	320 U	
2,4-Dichlorophenol	120-83-2			350 U	390 U	320 U	
2,4-Dimethylphenol	105-67-9			350 U	390 U	320 UJ	
2,4-Dinitrophenol	51-28-5			710 UJ	790 UJ	660 UJ	
2,4-Dinitrotoluene	121-14-2			350 U	390 U	320 U	
2,6-Dinitrotoluene	606-20-2			350 U	390 U	320 U	
2-Chloronaphthalene	91-58-7			350 U	390 U	320 U	
2-Chlorophenol	95-57-8			350 U	390 U	320 U	
2-Methylnaphthalene	91-57-6			350 UJ	390 UJ	320 U	
2-Methylphenol	95-48-7	330		350 U	390 U	320 U	
2-Nitroaniline	88-74-4			710 U	790 U	660 U	
2-Nitrophenol	88-75-5			350 U	390 U	320 U	
3,3'-Dichlorobenzidine	91-94-1			350 U	390 U	320 U	
3-Nitroaniline	99-09-2			710 U	790 U	660 U	
4,6-Dinitro-2-Methylphenol	534-52-1			710 U	790 U	660 U	
4-Bromophenyl-Phenylether	101-55-3			350 U	390 U	320 U	
4-Chloro-3-Methylphenol	59-50-7			350 U	390 U	320 U	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP2-30	80230-MIP2B-40	80230-MIP2C-35	80230-MIP2C-35ME
			Sample Location	MIP2 05/24/2011	MIP2B 05/26/2011	MIP2C 06/16/2011	MIP2C 06/16/2011
4-Chloroaniline	106-47-8			350 U	390 U	320 U	
4-Chlorophenyl-Phenylether	7005-72-3			350 U	390 U	320 U	
4-Methylphenol	106-44-5	330		350 U	390 U	320 U	
4-Nitroaniline	100-01-6			710 U	790 U	660 U	
4-Nitrophenol	100-02-7			710 U	790 U	660 U	
Acenaphthene	83-32-9	20000		350 U	390 U	320 U	
Acenaphthylene	208-96-8	100000		350 U	390 U	320 U	
Acetophenone	98-86-2			350 U	390 U	320 U	
Anthracene	120-12-7	100000		350 U	390 U	320 U	
Atrazine	1912-24-9			350 U	390 U	320 U	
Benzaldehyde	100-52-7			350 U	390 U	320 U	
Benzo(a)Anthracene	56-55-3	1000		350 U	390 U	320 U	
Benzo(a)Pyrene	50-32-8	1000		350 U	390 U	320 U	
Benzo(b)Fluoranthene	205-99-2	1000		350 U	390 U	320 U	
Benzo(g,h,i)Perylene	191-24-2	100000		350 U	390 U	320 U	
Benzo(k)Fluoranthene	207-08-9	800		350 U	390 U	320 U	
bis(2-Chloroethoxy)Methane	111-91-1			350 U	390 U	320 U	
bis(2-Chloroethyl) Ether	111-44-4			350 U	390 U	320 U	
bis(2-Ethylhexyl)Phthalate	117-81-7			160 J	390 U	48 J	
bis-Chloroisopropyl Ether	108-60-1			350 U	390 U	320 U	
Butylbenzylphthalate	85-68-7			350 U	390 U	320 U	
Caprolactam	105-60-2			350 U	390 U	320 U	
Carbazole	86-74-8			350 U	390 U	320 U	
Chrysene	218-01-9	1000		350 U	390 U	320 U	
Dibenzo(a,h)Anthracene	53-70-3	330		350 U	390 U	320 U	
Dibenzofuran	132-64-9	7000		350 U	390 U	320 U	
Diethylphthalate	84-66-2			350 U	390 U	320 U	
Dimethylphthalate	131-11-3			350 U	390 U	320 U	
di-n-Butylphthalate	84-74-2			350 U	390 U	320 U	
di-n-Octylphthalate	117-84-0			350 U	390 U	320 U	
Fluoranthene	206-44-0	100000		350 U	390 U	320 U	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Sample ID Sample Location Sampling Date	80230-MIP2-30	80230-MIP2B-40	80230-MIP2C-35	80230-MIP2C-35ME
			MIP2 05/24/2011	MIP2B 05/26/2011	MIP2C 06/16/2011	MIP2C 06/16/2011
Fluorene	86-73-7	30000	350 U	390 U	320 U	
Hexachlorobenzene	118-74-1	330	350 U	390 U	320 U	
Hexachlorobutadiene	87-68-3		350 U	390 U	320 U	
Hexachlorocyclopentadiene	77-47-4		350 U	390 U	320 U	
Hexachloroethane	67-72-1		350 U	390 U	320 U	
Indeno(1,2,3-cd)Pyrene	193-39-5	500	350 U	390 U	320 U	
Isophorone	78-59-1		350 U	390 U	320 U	
Naphthalene	91-20-3	12000	350 U	390 U	320 U	
Nitrobenzene	98-95-3		350 U	390 UJ	320 U	
n-Nitroso-di-n-Propylamine	621-64-7		350 U	390 U	320 U	
n-Nitrosodiphenylamine	86-30-6		350 U	390 U	320 U	
Pentachlorophenol	87-86-5	800	710 U	790 U	660 U	
Phenanthrene	85-01-8	100000	350 U	390 U	320 U	
Phenol	108-95-2	330	350 U	390 U	320 U	
Pyrene	129-00-0	100000	350 U	390 U	320 U	
<b>Pesticides (µg/kg)</b>						
4,4'-DDD	72-54-8	3.3		3.9 U	4 U	
4,4'-DDE	72-55-9	3.3		3.9 U	4 U	
4,4'-DDT	50-29-3	3.3		3.9 U	4 U	
Aldrin	309-00-2	5		2 U	2.1 U	
Alpha-BHC	319-84-6	20		2 U	2.1 U	
Alpha-Chlordane	5103-71-9	94		2 U	2.1 U	
Beta-BHC	319-85-7	36		2 U	2.1 U	
Delta-BHC	319-86-8	40		2 U	2.1 U	
Dieldrin	60-57-1	5		3.9 U	4 U	
Endosulfan I	959-98-8	2400		2 U	2.1 U	
Endosulfan II	33213-65-9	2400		3.9 U	4 U	
Endosulfan Sulfate	1031-07-8	2400		3.9 U	4 U	
Endrin	72-20-8	14		3.9 U	4 U	
Endrin Aldehyde	7421-93-4			3.9 U	4 U	
Endrin Ketone	53494-70-5			3.9 U	4 U	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP2-30 MIP2 05/24/2011	80230-MIP2B-40 MIP2B 05/26/2011	80230-MIP2C-35 MIP2C 06/16/2011	80230-MIP2C-35ME MIP2C 06/16/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Gamma-BHC (Lindane)	58-89-9	100			2 U	2.1 U	
Gamma-Chlordane	5103-74-2				2 U	2.1 U	
Heptachlor	76-44-8	42			2 U	2.1 U	
Heptachlor Epoxide	1024-57-3				2 U	2.1 U	
Methoxychlor	72-43-5				20 U	21 U	
Toxaphene	8001-35-2				200 U	210 U	
<b>Polychlorinated Biphenyls (µg/kg)</b>							
Aroclor 1016	12674-11-2	100			39 U	41 U	
Aroclor 1221	11104-28-2	100			39 U	41 U	
Aroclor 1232	11141-16-5	100			39 U	41 U	
Aroclor 1242	53469-21-9	100			39 U	41 U	
Aroclor 1248	12672-29-6	100			39 U	41 U	
Aroclor 1254	11097-69-1	100			39 U	41 U	
Aroclor 1260	11096-82-5	100			39 U	41 U	
<b>Inorganics (mg/kg)</b>							
Aluminum	7429-90-5				3260	4530	
Antimony	7440-36-0				1 BNJ	0.67 U	
Arsenic	7440-38-2	13			1.7 *J	1.5	
Barium	7440-39-3	350			59.1 *J	34.9	
Beryllium	7440-41-7	7.2			0.25 B	0.44	
Cadmium	7440-43-9	2.5			0.071 B	0.17 U	
Calcium	7440-70-2				905	2000	
Chromium	7440-47-3	30			11.3	11.1	
Cobalt	7440-48-4				5.6 *E	5.1	
Copper	7440-50-8	50			10.9 *J	188	
Iron	7439-89-6				11700 *J	11400	
Lead	7439-92-1	63			2.8	4.8	
Magnesium	7439-95-4				1680	1900	
Manganese	7439-96-5	1600			267 *J	205	
Mercury	7439-97-6	0.18			0.047 U	0.043 U	
Nickel	7440-02-0	30			7.6	8.5	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP2-30 MIP2 05/24/2011	80230-MIP2B-40 MIP2B 05/26/2011	80230-MIP2C-35 MIP2C 06/16/2011	80230-MIP2C-35ME MIP2C 06/16/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Potassium	7440-09-7			399	621		
Selenium	7782-49-2	3.9		1.6	U	1	U
Silver	7440-22-4	2		1.6	U	1	U
Sodium	7440-23-5			61.5		72.9	
Thallium	7440-28-0			1.2		0.67	U
Vanadium	7440-62-2			22.8	*J	18.2	
Zinc	7440-66-6	109		13.8		19.7	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID 80230-MIP3-30	MIP3 05/25/2011	80230-MIP3-30-DUP	MIP3 05/25/2011	80230-MIP3B-45	MIP3B 05/26/2011	80230-MIP4B-45	MIP4B 06/17/2011
Chemical Name	CAS#	Part 375 Unrestricted								
<b>Volatile Organic Compounds (µg/kg)</b>										
1,1,1,2-Tetrachloroethane	630-20-6		5.3 U		5.4 U		5.4 U		5.7 U	
1,1,1-Trichloroethane	71-55-6	680	5.3 U		5.4 U		5.4 U		5.7 U	
1,1,2,2-Tetrachloroethane	79-34-5		5.3 U		5.4 U		5.4 U		5.7 U	
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5.3 U		5.4 U		5.4 U		5.7 U	
1,1,2-Trichloroethane	79-00-5		5.3 U		5.4 U		5.4 U		5.7 U	
1,1-Dichloroethane	75-34-3	270	5.3 U		5.4 U		5.4 U		5.7 U	
1,1-Dichloroethene	75-35-4	330	5.3 U		5.4 U		5.4 U		5.7 U	
1,1-Dichloropropene	563-58-6		5.3 U		5.4 U		5.4 U		5.7 UJ	
1,2,3-Trichlorobenzene	87-61-6		5.3 U		5.4 U		5.4 U		5.7 UJ	
1,2,3-Trichloropropane	96-18-4		5.3 U		5.4 U		5.4 U		5.7 U	
1,2,4-Trichlorobenzene	120-82-1		5.3 U		5.4 U		5.4 U		5.7 UJ	
1,2,4-Trimethylbenzene	95-63-6	3600	5.3 U		5.4 U		5.4 U		5.7 U	
1,2-Dibromo-3-Chloropropane	96-12-8		5.3 U		5.4 U		5.4 U		5.7 U	
1,2-Dibromoethane (EDB)	106-93-4		5.3 U		5.4 U		5.4 U		5.7 U	
1,2-Dichlorobenzene	95-50-1	1100	5.3 U		5.4 U		5.4 U		5.7 U	
1,2-Dichloroethane	107-06-2	20	5.3 U		5.4 U		5.4 U		5.7 U	
1,2-Dichloropropane	78-87-5		5.3 U		5.4 U		5.4 U		2.8 J	
1,3,5-Trimethylbenzene	108-67-8	8400	5.3 U		5.4 U		5.4 U		5.7 U	
1,3-Dichlorobenzene	541-73-1	2400	5.3 U		5.4 U		5.4 U		5.7 U	
1,3-Dichloropropane	142-28-9		5.3 U		5.4 U		5.4 U		5.7 U	
1,4-Dichlorobenzene	106-46-7	1800	5.3 U		5.4 U		5.4 U		5.7 U	
2,2-Dichloropropane	594-20-7		5.3 U		5.4 U		5.4 U		5.7 UJ	
2-Butanone (MEK)	78-93-3	120	5.3 R		5.4 R		5.4 R		5.7 R	
2-Chlorotoluene	95-49-8		5.3 U		5.4 U		5.4 U		5.7 U	
2-Hexanone	591-78-6		5.3 U		5.4 U		5.4 U		5.7 UJ	
4-Chlorotoluene	106-43-4		5.3 U		5.4 U		5.4 U		5.7 U	
4-Methyl-2-Pentanone (MIBK)	108-10-1		5.3 U		5.4 U		5.4 U		5.7 U	
Acetone	67-64-1	50	5.3 R		3.4 R		3.1 R		5.7 R	
Benzene	71-43-2	60	5.3 U		5.4 U		5.4 U		5.7 U	
Bromobenzene	108-86-1		5.3 U		5.4 U		5.4 U		5.7 U	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Sample ID Sample Location Sampling Date	80230-MIP3-30	80230-MIP3-30-DUP	80230-MIP3B-45	80230-MIP4B-45
			MIP3 05/25/2011	MIP3 05/25/2011	MIP3B 05/26/2011	MIP4B 06/17/2011
Bromochloromethane	74-97-5	Part 375 Unrestricted	5.3 U	5.4 U	5.4 U	5.7 UJ
Bromodichloromethane	75-27-4		5.3 U	5.4 U	5.4 U	5.7 U
Bromoform	75-25-2		5.3 U	5.4 U	5.4 U	5.7 UJ
Bromomethane	74-83-9		5.3 U	5.4 U	5.4 U	5.7 U
Carbon Disulfide	75-15-0		5.3 U	5.4 U	5.4 U	5.7 U
Carbon Tetrachloride	56-23-5	760	5.3 U	5.4 U	5.4 U	5.7 U
Chlorobenzene	108-90-7	1100	5.3 U	5.4 U	5.4 U	5.7 U
Chloroethane	75-00-3		5.3 U	5.4 U	5.4 U	5.7 U
Chloroform	67-66-3	370	5.3 U	5.4 U	5.4 U	5.7 U
Chloromethane	74-87-3		5.3 U	5.4 U	5.4 U	5.7 U
cis-1,2-Dichloroethene	156-59-2	250	5.4	2.3 J	5.4 U	5.7 U
cis-1,3-Dichloropropene	10061-01-5		5.3 U	5.4 U	5.4 U	5.7 UJ
Cyclohexane	110-82-7		5.3 U	5.4 U	5.4 U	5.7 U
Dibromochloromethane	124-48-1		5.3 U	5.4 U	5.4 U	5.7 U
Dibromomethane	74-95-3		5.3 U	5.4 U	5.4 U	5.7 U
Dichlorodifluoromethane	75-71-8		5.3 U	5.4 U	5.4 U	5.7 U
Ethylbenzene	100-41-4	1000	5.3 U	5.4 U	5.4 U	5.7 U
Hexachlorobutadiene	87-68-3		5.3 U	5.4 U	5.4 U	5.7 UJ
Isopropylbenzene	98-82-8		5.3 U	5.4 U	5.4 U	5.7 U
m,p-Xylene	179601-23-1	260	5.3 U	5.4 U	5.4 U	5.7 U
Methyl Acetate	79-20-9		5.3 U	5.4 U	5.4 U	5.7 UJ
Methyl Iodide	74-88-4		5.3 U	5.4 U	5.4 U	5.7 U
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	930	5.3 U	5.4 U	5.4 U	5.7 U
Methylcyclohexane	108-87-2		5.3 U	5.4 U	5.4 U	5.7 U
Methylene Chloride	75-09-2	50	5.3 U	5.4 U	1.7 J	1.9 J
Naphthalene	91-20-3	12000	5.3 U	5.4 U	5.4 U	5.7 UJ
n-Butylbenzene	104-51-8	12000	5.3 U	5.4 U	5.4 U	5.7 UJ
n-Propylbenzene	103-65-1	3900	5.3 U	5.4 U	5.4 U	5.7 UJ
o-Xylene	95-47-6	260	5.3 U	5.4 U	5.4 U	5.7 U
p-Isopropyltoluene	99-87-6		5.3 U	5.4 U	5.4 U	5.7 U
sec-Butylbenzene	135-98-8	11000	5.3 U	5.4 U	5.4 U	5.7 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP3-30 MIP3 05/25/2011	80230-MIP3-30-DUP MIP3 05/25/2011	80230-MIP3B-45 MIP3B 05/26/2011	80230-MIP4B-45 MIP4B 06/17/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Styrene	100-42-5			5.3 U	5.4 U	5.4 U	5.7 U
tert-Butylbenzene	98-06-6	5900		5.3 U	5.4 U	5.4 U	5.7 U
Tetrachloroethene	127-18-4	1300		6.4	2.2 J	5.4 U	5.7 UJ
Toluene	108-88-3	700		5.3 U	5.4 U	5.4 U	5.7 U
Total Xylenes	1330-20-7	260		5.3 U	5.4 U	5.4 U	5.7 U
trans-1,2-Dichloroethene	156-60-5	190		5.3 U	5.4 U	5.4 U	5.7 U
trans-1,3-Dichloropropene	10061-02-6			5.3 U	5.4 U	5.4 U	5.7 UJ
Trichloroethene	79-01-6	470		1.9 J	5.4 U	5.4 U	5.7 U
Trichlorofluoromethane	75-69-4			5.3 U	5.4 U	5.4 U	5.7 U
Vinyl Acetate	108-05-4			5.3 U	5.4 U	5.4 U	5.7 UJ
Vinyl Chloride	75-01-4	20		5.3 U	5.4 U	5.4 U	5.7 U
<b>Semi-Volatile Organic Compounds (µg/kg)</b>							
1,1'-Biphenyl	92-52-4			360 U	360 U	380 U	330 U
2,4,5-Trichlorophenol	95-95-4			740 U	730 U	770 U	660 U
2,4,6-Trichlorophenol	88-06-2			360 U	360 U	380 U	330 U
2,4-Dichlorophenol	120-83-2			360 U	360 U	380 U	330 U
2,4-Dimethylphenol	105-67-9			360 U	360 U	380 U	330 UJ
2,4-Dinitrophenol	51-28-5			740 UJ	730 UJ	770 UJ	660 UJ
2,4-Dinitrotoluene	121-14-2			360 U	360 U	380 U	330 U
2,6-Dinitrotoluene	606-20-2			360 U	360 U	380 U	330 U
2-Chloronaphthalene	91-58-7			360 U	360 U	380 U	330 U
2-Chlorophenol	95-57-8			360 U	360 U	380 U	330 U
2-Methylnaphthalene	91-57-6			360 UJ	360 UJ	380 UJ	330 U
2-Methylphenol	95-48-7	330		360 U	360 U	380 U	330 U
2-Nitroaniline	88-74-4			740 U	730 U	770 U	660 U
2-Nitrophenol	88-75-5			360 U	360 U	380 U	330 U
3,3'-Dichlorobenzidine	91-94-1			360 U	360 U	380 U	330 U
3-Nitroaniline	99-09-2			740 U	730 U	770 U	660 U
4,6-Dinitro-2-Methylphenol	534-52-1			740 U	730 U	770 U	660 U
4-Bromophenyl-Phenylether	101-55-3			360 U	360 U	380 U	330 U
4-Chloro-3-Methylphenol	59-50-7			360 U	360 U	380 U	330 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP3-30	80230-MIP3-30-DUP	80230-MIP3B-45	80230-MIP4B-45
			Sample Location	MIP3 05/25/2011	MIP3 05/25/2011	MIP3B 05/26/2011	MIP4B 06/17/2011
4-Chloroaniline	106-47-8			360 U	360 U	380 U	330 U
4-Chlorophenyl-Phenylether	7005-72-3			360 U	360 U	380 U	330 U
4-Methylphenol	106-44-5	330		360 U	360 U	380 U	330 U
4-Nitroaniline	100-01-6			740 U	730 U	770 U	660 U
4-Nitrophenol	100-02-7			740 U	730 U	770 U	660 U
Acenaphthene	83-32-9	20000		360 U	360 U	380 U	330 U
Acenaphthylene	208-96-8	100000		360 U	360 U	380 U	330 U
Acetophenone	98-86-2			360 U	360 U	380 U	330 U
Anthracene	120-12-7	100000		360 U	360 U	380 U	330 U
Atrazine	1912-24-9			360 U	360 U	380 U	330 U
Benzaldehyde	100-52-7			360 U	360 U	380 U	330 U
Benzo(a)Anthracene	56-55-3	1000		360 U	360 U	380 U	330 U
Benzo(a)Pyrene	50-32-8	1000		360 U	360 U	380 U	330 U
Benzo(b)Fluoranthene	205-99-2	1000		360 U	360 U	380 U	330 U
Benzo(g,h,i)Perylene	191-24-2	100000		360 U	360 U	380 U	330 U
Benzo(k)Fluoranthene	207-08-9	800		360 U	360 U	380 U	330 U
bis(2-Chloroethoxy)Methane	111-91-1			360 U	360 U	380 U	330 U
bis(2-Chloroethyl) Ether	111-44-4			360 U	360 U	380 U	330 U
bis(2-Ethylhexyl)Phthalate	117-81-7			360 U	360 U	380 U	330 UJ
bis-Chloroisopropyl Ether	108-60-1			360 U	360 U	380 U	330 U
Butylbenzylphthalate	85-68-7			360 U	360 U	380 U	330 U
Caprolactam	105-60-2			360 U	360 U	380 U	330 U
Carbazole	86-74-8			360 U	360 U	380 U	330 U
Chrysene	218-01-9	1000		360 U	360 U	380 U	330 U
Dibenzo(a,h)Anthracene	53-70-3	330		360 U	360 U	380 U	330 U
Dibenzofuran	132-64-9	7000		360 U	360 U	380 U	330 U
Diethylphthalate	84-66-2			360 U	360 U	380 U	330 U
Dimethylphthalate	131-11-3			360 U	360 U	380 U	330 U
di-n-Butylphthalate	84-74-2			360 U	360 U	380 U	330 U
di-n-Octylphthalate	117-84-0			360 U	360 U	380 U	330 U
Fluoranthene	206-44-0	100000		360 U	360 U	380 U	330 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP3-30 MIP3 05/25/2011	80230-MIP3-30-DUP MIP3 05/25/2011	80230-MIP3B-45 MIP3B 05/26/2011	80230-MIP4B-45 MIP4B 06/17/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Fluorene	86-73-7	30000	360 U	360 U	380 U	330 U	
Hexachlorobenzene	118-74-1	330	360 U	360 U	380 U	330 U	
Hexachlorobutadiene	87-68-3		360 U	360 U	380 U	330 U	
Hexachlorocyclopentadiene	77-47-4		360 U	360 U	380 U	330 U	
Hexachloroethane	67-72-1		360 U	360 U	380 U	330 U	
Indeno(1,2,3-cd)Pyrene	193-39-5	500	360 U	360 U	380 U	330 U	
Isophorone	78-59-1		360 U	360 U	380 U	330 U	
Naphthalene	91-20-3	12000	360 U	360 U	380 U	330 U	
Nitrobenzene	98-95-3		360 UJ	360 UJ	380 UJ	330 U	
n-Nitroso-di-n-Propylamine	621-64-7		360 U	360 U	380 U	330 U	
n-Nitrosodiphenylamine	86-30-6		360 U	360 U	380 U	330 U	
Pentachlorophenol	87-86-5	800	740 U	730 U	770 U	660 U	
Phenanthrene	85-01-8	100000	360 U	360 U	380 U	330 U	
Phenol	108-95-2	330	360 U	360 U	380 U	330 U	
Pyrene	129-00-0	100000	360 U	360 U	380 U	330 U	
<b>Pesticides (µg/kg)</b>							
4,4'-DDD	72-54-8	3.3			3.8 U		
4,4'-DDE	72-55-9	3.3			3.8 U		
4,4'-DDT	50-29-3	3.3			3.8 U		
Aldrin	309-00-2	5			2 U		
Alpha-BHC	319-84-6	20			2 UJ		
Alpha-Chlordane	5103-71-9	94			2 U		
Beta-BHC	319-85-7	36			2 UJ		
Delta-BHC	319-86-8	40			2 U		
Dieldrin	60-57-1	5			3.8 U		
Endosulfan I	959-98-8	2400			2 U		
Endosulfan II	33213-65-9	2400			3.8 U		
Endosulfan Sulfate	1031-07-8	2400			3.8 U		
Endrin	72-20-8	14			3.8 U		
Endrin Aldehyde	7421-93-4				3.8 U		
Endrin Ketone	53494-70-5				3.8 U		

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP3-30 MIP3 05/25/2011	80230-MIP3-30-DUP MIP3 05/25/2011	80230-MIP3B-45 MIP3B 05/26/2011	80230-MIP4B-45 MIP4B 06/17/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Gamma-BHC (Lindane)	58-89-9	100				2 U	
Gamma-Chlordane	5103-74-2					2 U	
Heptachlor	76-44-8	42				2 U	
Heptachlor Epoxide	1024-57-3					2 U	
Methoxychlor	72-43-5					20 U	
Toxaphene	8001-35-2					200 U	
<b>Polychlorinated Biphenyls (µg/kg)</b>							
Aroclor 1016	12674-11-2	100				38 U	
Aroclor 1221	11104-28-2	100				38 U	
Aroclor 1232	11141-16-5	100				38 U	
Aroclor 1242	53469-21-9	100				38 U	
Aroclor 1248	12672-29-6	100				38 U	
Aroclor 1254	11097-69-1	100				38 U	
Aroclor 1260	11096-82-5	100				38 U	
<b>Inorganics (mg/kg)</b>							
Aluminum	7429-90-5					3910	
Antimony	7440-36-0					0.52 BNJ	
Arsenic	7440-38-2	13				3.6 *J	
Barium	7440-39-3	350				60.1 *J	
Beryllium	7440-41-7	7.2				0.29	
Cadmium	7440-43-9	2.5				0.11 B	
Calcium	7440-70-2					926	
Chromium	7440-47-3	30				11	
Cobalt	7440-48-4					4.9 *E	
Copper	7440-50-8	50				10.9 *J	
Iron	7439-89-6					12200 *J	
Lead	7439-92-1	63				2.7	
Magnesium	7439-95-4					1860	
Manganese	7439-96-5	1600				375 *J	
Mercury	7439-97-6	0.18					
Nickel	7440-02-0	30				7.7	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP3-30	80230-MIP3-30-DUP	80230-MIP3B-45	80230-MIP4B-45
			Sample Location	MIP3	MIP3	MIP3B	MIP4B
			Sampling Date	05/25/2011	05/25/2011	05/26/2011	06/17/2011
Potassium	7440-09-7					450	
Selenium	7782-49-2	3.9				1.4	U
Silver	7440-22-4	2				1.4	U
Sodium	7440-23-5					60.2	
Thallium	7440-28-0					1.1	
Vanadium	7440-62-2					19.9	*J
Zinc	7440-66-6	109				17.5	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP4C-41 MIP4C 06/17/2011	80230-MIP5-11 MIP5 05/24/2011	80230-MIP5-30 MIP5 05/24/2011	80230-MIP5B-22 MIP5B 05/25/2011
Chemical Name	CAS#	Part 375 Unrestricted					
<b>Volatile Organic Compounds (µg/kg)</b>							
1,1,1,2-Tetrachloroethane	630-20-6		5.6 U	5.5 U	4.6 U	5.8 U	
1,1,1-Trichloroethane	71-55-6	680	5.6 U	5.5 U	4.6 U	5.8 U	
1,1,2,2-Tetrachloroethane	79-34-5		5.6 U	5.5 U	4.6 U	5.8 U	
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5.6 U	5.5 U	4.6 U	5.8 U	
1,1,2-Trichloroethane	79-00-5		5.6 U	5.5 U	4.6 U	5.8 U	
1,1-Dichloroethane	75-34-3	270	5.6 U	5.5 U	4.6 U	5.8 U	
1,1-Dichloroethene	75-35-4	330	5.6 U	5.5 U	4.6 U	5.8 U	
1,1-Dichloropropene	563-58-6		5.6 UJ	5.5 U	4.6 U	5.8 U	
1,2,3-Trichlorobenzene	87-61-6		5.6 UJ	5.5 U	4.6 U	5.8 U	
1,2,3-Trichloropropane	96-18-4		5.6 U	5.5 U	4.6 U	5.8 U	
1,2,4-Trichlorobenzene	120-82-1		5.6 UJ	5.5 U	4.6 U	5.8 U	
1,2,4-Trimethylbenzene	95-63-6	3600	5.6 U	5.5 U	4.6 U	5.8 U	
1,2-Dibromo-3-Chloropropane	96-12-8		5.6 U	5.5 UJ	4.6 UJ	5.8 U	
1,2-Dibromoethane (EDB)	106-93-4		5.6 U	5.5 U	4.6 U	5.8 U	
1,2-Dichlorobenzene	95-50-1	1100	5.6 U	5.5 U	4.6 U	5.8 U	
1,2-Dichloroethane	107-06-2	20	5.6 U	5.5 U	4.6 U	5.8 U	
1,2-Dichloropropane	78-87-5		2.9 J	5.5 U	4.6 U	5.8 U	
1,3,5-Trimethylbenzene	108-67-8	8400	5.6 U	5.5 U	4.6 U	5.8 U	
1,3-Dichlorobenzene	541-73-1	2400	5.6 U	5.5 U	4.6 U	5.8 U	
1,3-Dichloropropane	142-28-9		5.6 U	5.5 U	4.6 U	5.8 U	
1,4-Dichlorobenzene	106-46-7	1800	5.6 U	5.5 U	4.6 U	5.8 U	
2,2-Dichloropropane	594-20-7		5.6 UJ	5.5 U	4.6 U	5.8 U	
2-Butanone (MEK)	78-93-3	120	5.6 R	5.5 R	4.6 R	5.8 R	
2-Chlorotoluene	95-49-8		5.6 U	5.5 U	4.6 U	5.8 U	
2-Hexanone	591-78-6		5.6 UJ	5.5 U	4.6 U	5.8 U	
4-Chlorotoluene	106-43-4		5.6 U	5.5 U	4.6 U	5.8 U	
4-Methyl-2-Pentanone (MIBK)	108-10-1		5.6 U	5.5 U	4.6 U	5.8 U	
Acetone	67-64-1	50	5.6 R	5.5 R	4.6 R	3.7 R	
Benzene	71-43-2	60	5.6 U	5.5 U	4.6 U	5.8 U	
Bromobenzene	108-86-1		5.6 U	5.5 U	4.6 U	5.8 U	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP4C-41	80230-MIP5-11	80230-MIP5-30	80230-MIP5B-22
			Sample Location	MIP4C	MIP5	MIP5	MIP5B
			Sampling Date	06/17/2011	05/24/2011	05/24/2011	05/25/2011
Bromochloromethane	74-97-5			5.6 UJ	5.5 U	4.6 U	5.8 U
Bromodichloromethane	75-27-4			5.6 U	5.5 U	4.6 U	5.8 U
Bromoform	75-25-2			5.6 UJ	5.5 U	4.6 U	5.8 U
Bromomethane	74-83-9			5.6 U	5.5 U	4.6 U	5.8 U
Carbon Disulfide	75-15-0			5.6 U	5.5 U	4.6 U	5.8 U
Carbon Tetrachloride	56-23-5	760		5.6 U	5.5 U	4.6 U	5.8 U
Chlorobenzene	108-90-7	1100		5.6 U	5.5 U	4.6 U	5.8 U
Chloroethane	75-00-3			5.6 U	5.5 U	4.6 U	5.8 U
Chloroform	67-66-3	370		5.6 U	5.5 U	4.6 U	5.8 U
Chloromethane	74-87-3			5.6 U	5.5 U	4.6 U	5.8 U
cis-1,2-Dichloroethene	156-59-2	250		5.6 U	5.5 U	4.6 U	5.8 U
cis-1,3-Dichloropropene	10061-01-5			5.6 UJ	5.5 U	4.6 U	5.8 U
Cyclohexane	110-82-7			5.6 U	5.5 U	4.6 U	5.8 U
Dibromochloromethane	124-48-1			5.6 U	5.5 U	4.6 U	5.8 U
Dibromomethane	74-95-3			5.6 U	5.5 U	4.6 U	5.8 U
Dichlorodifluoromethane	75-71-8			5.6 U	5.5 U	4.6 U	5.8 U
Ethylbenzene	100-41-4	1000		5.6 U	5.5 U	4.6 U	5.8 U
Hexachlorobutadiene	87-68-3			5.6 UJ	5.5 U	4.6 U	5.8 U
Isopropylbenzene	98-82-8			5.6 U	5.5 U	4.6 U	5.8 U
m,p-Xylene	179601-23-1	260		5.6 U	5.5 U	4.6 U	5.8 U
Methyl Acetate	79-20-9			5.6 UJ	5.5 U	4.6 U	5.8 U
Methyl Iodide	74-88-4			5.6 U	5.5 U	4.6 U	5.8 U
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	930		5.6 U	5.5 U	4.6 U	5.8 U
Methylcyclohexane	108-87-2			5.6 U	5.5 U	4.6 U	5.8 U
Methylene Chloride	75-09-2	50		5.6 U	2.6 J	1.9 J	5.8 U
Naphthalene	91-20-3	12000		5.6 UJ	5.5 UJ	4.6 UJ	5.8 U
n-Butylbenzene	104-51-8	12000		5.6 UJ	5.5 U	4.6 U	5.8 U
n-Propylbenzene	103-65-1	3900		5.6 UJ	5.5 U	4.6 U	5.8 U
o-Xylene	95-47-6	260		5.6 U	5.5 U	4.6 U	5.8 U
p-Isopropyltoluene	99-87-6			5.6 U	5.5 U	4.6 U	5.8 U
sec-Butylbenzene	135-98-8	11000		5.6 U	5.5 U	4.6 U	5.8 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP4C-41 MIP4C 06/17/2011	80230-MIP5-11 MIP5 05/24/2011	80230-MIP5-30 MIP5 05/24/2011	80230-MIP5B-22 MIP5B 05/25/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Styrene	100-42-5			5.6 U	5.5 U	4.6 U	5.8 U
tert-Butylbenzene	98-06-6	5900		5.6 U	5.5 U	4.6 U	5.8 U
Tetrachloroethene	127-18-4	1300		47 JB	6	18	16
Toluene	108-88-3	700		5.6 U	5.5 U	4.6 U	5.8 U
Total Xylenes	1330-20-7	260		5.6 U	5.5 U	4.6 U	5.8 U
trans-1,2-Dichloroethene	156-60-5	190		5.6 U	5.5 U	4.6 U	5.8 U
trans-1,3-Dichloropropene	10061-02-6			5.6 UJ	5.5 U	4.6 U	5.8 U
Trichloroethene	79-01-6	470		5.6 U	5.5 U	4.6 U	5.8 U
Trichlorofluoromethane	75-69-4			5.6 U	5.5 U	4.6 U	5.8 U
Vinyl Acetate	108-05-4			5.6 UJ	5.5 U	4.6 U	5.8 U
Vinyl Chloride	75-01-4	20		5.6 U	5.5 U	4.6 U	5.8 U
<b>Semi-Volatile Organic Compounds (µg/kg)</b>							
1,1'-Biphenyl	92-52-4			330 U	380 U	350 U	380 U
2,4,5-Trichlorophenol	95-95-4			660 U	770 U	720 U	770 U
2,4,6-Trichlorophenol	88-06-2			330 U	380 U	350 U	380 U
2,4-Dichlorophenol	120-83-2			330 U	380 U	350 U	380 U
2,4-Dimethylphenol	105-67-9			330 UJ	380 U	350 U	380 U
2,4-Dinitrophenol	51-28-5			660 UJ	770 UJ	720 UJ	770 UJ
2,4-Dinitrotoluene	121-14-2			330 U	380 U	350 U	380 U
2,6-Dinitrotoluene	606-20-2			330 U	380 U	350 U	380 U
2-Chloronaphthalene	91-58-7			330 U	380 U	350 U	380 U
2-Chlorophenol	95-57-8			330 U	380 U	350 U	380 U
2-Methylnaphthalene	91-57-6			330 U	380 UJ	350 UJ	380 UJ
2-Methylphenol	95-48-7	330		330 U	380 U	350 U	380 U
2-Nitroaniline	88-74-4			660 U	770 U	720 U	770 U
2-Nitrophenol	88-75-5			330 U	380 U	350 U	380 U
3,3'-Dichlorobenzidine	91-94-1			330 U	380 U	350 U	380 U
3-Nitroaniline	99-09-2			660 U	770 U	720 U	770 U
4,6-Dinitro-2-Methylphenol	534-52-1			660 U	770 U	720 U	770 U
4-Bromophenyl-Phenylether	101-55-3			330 U	380 U	350 U	380 U
4-Chloro-3-Methylphenol	59-50-7			330 U	380 U	350 U	380 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP4C-41	80230-MIP5-11	80230-MIP5-30	80230-MIP5B-22
			Sample Location	MIP4C	MIP5	MIP5B	MIP5B
		Sampling Date	06/17/2011	05/24/2011	05/24/2011	05/25/2011	05/25/2011
4-Chloroaniline	106-47-8			330 U	380 U	350 U	380 U
4-Chlorophenyl-Phenylether	7005-72-3			330 U	380 U	350 U	380 U
4-Methylphenol	106-44-5	330		330 U	380 U	350 U	380 U
4-Nitroaniline	100-01-6			660 U	770 U	720 U	770 U
4-Nitrophenol	100-02-7			660 U	770 U	720 U	770 U
Acenaphthene	83-32-9	20000		330 U	380 U	350 U	380 U
Acenaphthylene	208-96-8	100000		330 U	380 U	350 U	380 U
Acetophenone	98-86-2			330 U	380 U	350 U	380 U
Anthracene	120-12-7	100000		330 U	380 U	350 U	380 U
Atrazine	1912-24-9			330 U	380 U	350 U	380 U
Benzaldehyde	100-52-7			330 U	380 U	350 U	380 U
Benzo(a)Anthracene	56-55-3	1000		330 U	380 U	350 U	380 U
Benzo(a)Pyrene	50-32-8	1000		330 U	380 U	350 U	380 U
Benzo(b)Fluoranthene	205-99-2	1000		330 U	380 U	350 U	380 U
Benzo(g,h,i)Perylene	191-24-2	100000		330 U	380 U	350 U	380 U
Benzo(k)Fluoranthene	207-08-9	800		330 U	380 U	350 U	380 U
bis(2-Chloroethoxy)Methane	111-91-1			330 U	380 U	350 U	380 U
bis(2-Chloroethyl) Ether	111-44-4			330 U	380 U	350 U	380 U
bis(2-Ethylhexyl)Phthalate	117-81-7			330 UJ	160 J	350 U	380 U
bis-Chloroisopropyl Ether	108-60-1			330 U	380 U	350 U	380 U
Butylbenzylphthalate	85-68-7			330 U	380 U	350 U	380 U
Caprolactam	105-60-2			330 U	380 U	350 U	380 U
Carbazole	86-74-8			330 U	380 U	350 U	380 U
Chrysene	218-01-9	1000		330 U	380 U	350 U	380 U
Dibenzo(a,h)Anthracene	53-70-3	330		330 U	380 U	350 U	380 U
Dibenzofuran	132-64-9	7000		330 U	380 U	350 U	380 U
Diethylphthalate	84-66-2			330 U	380 U	350 U	380 U
Dimethylphthalate	131-11-3			330 U	380 U	350 U	380 U
di-n-Butylphthalate	84-74-2			330 U	380 U	350 U	380 U
di-n-Octylphthalate	117-84-0			330 U	380 U	350 U	380 U
Fluoranthene	206-44-0	100000		330 U	380 U	350 U	380 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP4C-41	80230-MIP5-11	80230-MIP5-30	80230-MIP5B-22
			Sample Location	MIP4C	MIP5	MIP5B	
		Sampling Date		06/17/2011	05/24/2011	05/24/2011	05/25/2011
Fluorene	86-73-7	30000		330 U	380 U	350 U	380 U
Hexachlorobenzene	118-74-1	330		330 U	380 U	350 U	380 U
Hexachlorobutadiene	87-68-3			330 U	380 U	350 U	380 U
Hexachlorocyclopentadiene	77-47-4			330 U	380 U	350 U	380 U
Hexachloroethane	67-72-1			330 U	380 U	350 U	380 U
Indeno(1,2,3-cd)Pyrene	193-39-5	500		330 U	380 U	350 U	380 U
Isophorone	78-59-1			330 U	380 U	350 U	380 U
Naphthalene	91-20-3	12000		330 U	380 U	350 U	380 U
Nitrobenzene	98-95-3			330 U	380 U	350 U	380 UJ
n-Nitroso-di-n-Propylamine	621-64-7			330 U	380 U	350 U	380 U
n-Nitrosodiphenylamine	86-30-6			330 U	380 U	350 U	380 U
Pentachlorophenol	87-86-5	800		660 U	770 U	720 U	770 U
Phenanthrene	85-01-8	100000		330 U	380 U	350 U	380 U
Phenol	108-95-2	330		330 U	380 U	350 U	380 U
Pyrene	129-00-0	100000		330 U	380 U	350 U	380 U
<b>Pesticides (µg/kg)</b>							
4,4'-DDD	72-54-8	3.3					3.8 U
4,4'-DDE	72-55-9	3.3					3.8 U
4,4'-DDT	50-29-3	3.3					3.8 U
Aldrin	309-00-2	5					2 U
Alpha-BHC	319-84-6	20					2 U
Alpha-Chlordane	5103-71-9	94					2 U
Beta-BHC	319-85-7	36					2 U
Delta-BHC	319-86-8	40					2 U
Dieldrin	60-57-1	5					3.8 U
Endosulfan I	959-98-8	2400					2 U
Endosulfan II	33213-65-9	2400					3.8 U
Endosulfan Sulfate	1031-07-8	2400					3.8 U
Endrin	72-20-8	14					3.8 U
Endrin Aldehyde	7421-93-4						3.8 U
Endrin Ketone	53494-70-5						3.8 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP4C-41 MIP4C 06/17/2011	80230-MIP5-11 MIP5 05/24/2011	80230-MIP5-30 MIP5 05/24/2011	80230-MIP5B-22 MIP5B 05/25/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Gamma-BHC (Lindane)	58-89-9	100					2 U
Gamma-Chlordane	5103-74-2						2 U
Heptachlor	76-44-8	42					2 U
Heptachlor Epoxide	1024-57-3						2 U
Methoxychlor	72-43-5						20 U
Toxaphene	8001-35-2						200 U
<b>Polychlorinated Biphenyls (µg/kg)</b>							
Aroclor 1016	12674-11-2	100					38 U
Aroclor 1221	11104-28-2	100					38 U
Aroclor 1232	11141-16-5	100					38 U
Aroclor 1242	53469-21-9	100					38 U
Aroclor 1248	12672-29-6	100					38 U
Aroclor 1254	11097-69-1	100					38 U
Aroclor 1260	11096-82-5	100					38 U
<b>Inorganics (mg/kg)</b>							
Aluminum	7429-90-5						5060
Antimony	7440-36-0						0.34 BNJ
Arsenic	7440-38-2	13					2.8 *J
Barium	7440-39-3	350					45.5
Beryllium	7440-41-7	7.2					0.47
Cadmium	7440-43-9	2.5					0.17
Calcium	7440-70-2						1130
Chromium	7440-47-3	30					34 *J
Cobalt	7440-48-4						11.8 *J
Copper	7440-50-8	50					10.8 *J
Iron	7439-89-6						15200 *J
Lead	7439-92-1	63					8
Magnesium	7439-95-4						2890 *J
Manganese	7439-96-5	1600					348
Mercury	7439-97-6	0.18					0.042 U
Nickel	7440-02-0	30					77.9

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP4C-41	80230-MIP5-11	80230-MIP5-30	80230-MIP5B-22
			Sample Location	MIP4C	MIP5	MIP5B	
			Sampling Date	06/17/2011	05/24/2011	05/24/2011	05/25/2011
Potassium	7440-09-7						953 *J
Selenium	7782-49-2	3.9					0.89 U
Silver	7440-22-4	2					0.89 U
Sodium	7440-23-5						85.6 EJ
Thallium	7440-28-0						1.1
Vanadium	7440-62-2						17.8 *J
Zinc	7440-66-6	109					27.2 *J

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP5C-36.5 MIP5C 06/24/2011	80230-MIP5E-16 MIP5E 06/16/2011	80230-MIP6-10 MIP6 05/23/2011	80230-MIP6-30 MIP6 05/23/2011
Chemical Name	CAS#	Part 375 Unrestricted					
<b>Volatile Organic Compounds (µg/kg)</b>							
1,1,1,2-Tetrachloroethane	630-20-6		5.5 U	5.5 U	5.3 U	5 U	
1,1,1-Trichloroethane	71-55-6	680	5.5 U	5.5 U	5.3 U	5 U	
1,1,2,2-Tetrachloroethane	79-34-5		5.5 U	5.5 U	5.3 U	5 U	
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5.5 U	5.5 U	5.3 U	5 U	
1,1,2-Trichloroethane	79-00-5		5.5 U	5.5 U	5.3 U	5 U	
1,1-Dichloroethane	75-34-3	270	5.5 U	5.5 U	5.3 U	5 U	
1,1-Dichloroethene	75-35-4	330	5.5 U	5.5 U	5.3 U	5 U	
1,1-Dichloropropene	563-58-6		5.5 U	5.5 U	5.3 U	5 U	
1,2,3-Trichlorobenzene	87-61-6		5.5 U	5.5 UJ	5.3 U	5 U	
1,2,3-Trichloropropane	96-18-4		5.5 U	5.5 U	5.3 U	5 U	
1,2,4-Trichlorobenzene	120-82-1		5.5 U	5.5 UJ	5.3 U	5 U	
1,2,4-Trimethylbenzene	95-63-6	3600	5.5 U	5.5 U	5.3 U	5 U	
1,2-Dibromo-3-Chloropropane	96-12-8		5.5 U	5.5 U	5.3 UJ	5 U	
1,2-Dibromoethane (EDB)	106-93-4		5.5 U	5.5 U	5.3 U	5 U	
1,2-Dichlorobenzene	95-50-1	1100	5.5 U	5.5 U	5.3 U	5 U	
1,2-Dichloroethane	107-06-2	20	5.5 U	5.5 U	5.3 U	5 U	
1,2-Dichloropropane	78-87-5		5.5 U	2.5 J	5.3 U	5 U	
1,3,5-Trimethylbenzene	108-67-8	8400	5.5 U	5.5 U	5.3 U	5 U	
1,3-Dichlorobenzene	541-73-1	2400	5.5 U	5.5 U	5.3 U	5 U	
1,3-Dichloropropane	142-28-9		5.5 U	5.5 U	5.3 U	5 U	
1,4-Dichlorobenzene	106-46-7	1800	5.5 U	5.5 U	5.3 U	5 U	
2,2-Dichloropropane	594-20-7		5.5 U	5.5 UJ	5.3 U	5 U	
2-Butanone (MEK)	78-93-3	120	5.5 R	5.5 R	5.3 R	5 R	
2-Chlorotoluene	95-49-8		5.5 U	5.5 U	5.3 U	5 U	
2-Hexanone	591-78-6		5.5 U	5.5 UJ	5.3 U	5 U	
4-Chlorotoluene	106-43-4		5.5 U	5.5 U	5.3 U	5 U	
4-Methyl-2-Pentanone (MIBK)	108-10-1		5.5 U	5.5 U	5.3 U	5 U	
Acetone	67-64-1	50	5.5 R	5.5 R	5.3 R	5 R	
Benzene	71-43-2	60	5.5 U	5.5 U	5.3 U	5 U	
Bromobenzene	108-86-1		5.5 U	5.5 U	5.3 U	5 U	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP5C-36.5	80230-MIP5E-16	80230-MIP6-10	80230-MIP6-30
			Sample Location	MIP5C	MIP5E	MIP6	MIP6
		Sampling Date	06/24/2011	06/16/2011	05/23/2011	05/23/2011	05/23/2011
Bromochloromethane	74-97-5			5.5 U	5.5 UJ	5.3 U	5 U
Bromodichloromethane	75-27-4			5.5 U	5.5 U	5.3 U	5 U
Bromoform	75-25-2			5.5 U	5.5 UJ	5.3 U	5 U
Bromomethane	74-83-9			5.5 U	5.5 U	5.3 U	5 U
Carbon Disulfide	75-15-0			5.5 U	5.5 U	5.3 U	5 U
Carbon Tetrachloride	56-23-5	760		5.5 U	5.5 U	5.3 U	5 U
Chlorobenzene	108-90-7	1100		5.5 U	5.5 U	5.3 U	5 U
Chloroethane	75-00-3			5.5 U	5.5 U	5.3 U	5 U
Chloroform	67-66-3	370		5.5 U	5.5 U	5.3 U	5 U
Chloromethane	74-87-3			5.5 U	5.5 U	5.3 U	5 U
cis-1,2-Dichloroethene	156-59-2	250		5.5 U	5.5 U	5.3 U	5 U
cis-1,3-Dichloropropene	10061-01-5			5.5 U	5.5 UJ	5.3 U	5 U
Cyclohexane	110-82-7			5.5 U	5.5 U	5.3 U	5 U
Dibromochloromethane	124-48-1			5.5 U	5.5 U	5.3 U	5 U
Dibromomethane	74-95-3			5.5 U	5.5 U	5.3 U	5 U
Dichlorodifluoromethane	75-71-8			5.5 U	5.5 U	5.3 U	5 U
Ethylbenzene	100-41-4	1000		5.5 U	5.5 U	5.3 U	5 U
Hexachlorobutadiene	87-68-3			5.5 U	5.5 UJ	5.3 U	5 U
Isopropylbenzene	98-82-8			5.5 U	5.5 U	5.3 U	5 U
m,p-Xylene	179601-23-1	260		5.5 U	5.5 U	5.3 U	5 U
Methyl Acetate	79-20-9			5.5 U	5.5 UJ	5.3 U	5 U
Methyl Iodide	74-88-4			5.5 U	5.5 U	5.3 U	5 U
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	930		5.5 U	5.5 U	5.3 U	5 U
Methylcyclohexane	108-87-2			5.5 U	5.5 U	5.3 U	5 U
Methylene Chloride	75-09-2	50		5.5 U	5.5 U	2 J	1.9 J
Naphthalene	91-20-3	12000		5.5 U	5.5 UJ	5.3 UJ	5 UJ
n-Butylbenzene	104-51-8	12000		5.5 U	5.5 UJ	5.3 U	5 U
n-Propylbenzene	103-65-1	3900		5.5 U	5.5 U	5.3 U	5 U
o-Xylene	95-47-6	260		5.5 U	5.5 U	5.3 U	5 U
p-Isopropyltoluene	99-87-6			5.5 U	5.5 U	5.3 U	5 U
sec-Butylbenzene	135-98-8	11000		5.5 U	5.5 U	5.3 U	5 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP5C-36.5	80230-MIP5E-16	80230-MIP6-10	80230-MIP6-30
			Sample Location	MIP5C	MIP5E	MIP6	MIP6
			Sampling Date	06/24/2011	06/16/2011	05/23/2011	05/23/2011
Styrene	100-42-5			5.5 U	5.5 U	5.3 U	5 U
tert-Butylbenzene	98-06-6	5900		5.5 U	5.5 U	5.3 U	5 U
Tetrachloroethene	127-18-4	1300		5.5 U	4.7 J	3.1 J	4 J
Toluene	108-88-3	700		5.5 U	5.5 U	5.3 U	5 U
Total Xylenes	1330-20-7	260		5.5 U	5.5 U	5.3 U	5 U
trans-1,2-Dichloroethene	156-60-5	190		5.5 U	5.5 U	5.3 U	5 U
trans-1,3-Dichloropropene	10061-02-6			5.5 U	5.5 UJ	5.3 U	5 U
Trichloroethene	79-01-6	470		5.5 U	5.5 U	5.3 U	5 U
Trichlorofluoromethane	75-69-4			5.5 U	5.5 U	5.3 U	5 U
Vinyl Acetate	108-05-4			5.5 U	5.5 UJ	5.3 U	5 U
Vinyl Chloride	75-01-4	20		5.5 U	5.5 U	5.3 U	5 U
<b>Semi-Volatile Organic Compounds (µg/kg)</b>							
1,1'-Biphenyl	92-52-4			360 U	320 U	370 U	380 U
2,4,5-Trichlorophenol	95-95-4			730 U	660 U	760 U	760 U
2,4,6-Trichlorophenol	88-06-2			360 U	320 U	370 U	380 U
2,4-Dichlorophenol	120-83-2			360 U	320 U	370 U	380 U
2,4-Dimethylphenol	105-67-9			360 U	320 UJ	370 U	380 U
2,4-Dinitrophenol	51-28-5			730 U	660 UJ	760 UJ	760 UJ
2,4-Dinitrotoluene	121-14-2			360 U	320 U	370 U	380 U
2,6-Dinitrotoluene	606-20-2			360 U	320 U	370 U	380 U
2-Chloronaphthalene	91-58-7			360 U	320 U	370 U	380 U
2-Chlorophenol	95-57-8			360 U	320 U	370 U	380 U
2-Methylnaphthalene	91-57-6			360 U	320 U	370 UJ	380 UJ
2-Methylphenol	95-48-7	330		360 U	320 U	370 U	380 U
2-Nitroaniline	88-74-4			730 U	660 U	760 U	760 U
2-Nitrophenol	88-75-5			360 U	320 U	370 U	380 U
3,3'-Dichlorobenzidine	91-94-1			360 U	320 U	370 U	380 U
3-Nitroaniline	99-09-2			730 U	660 U	760 U	760 U
4,6-Dinitro-2-Methylphenol	534-52-1			730 U	660 U	760 U	760 U
4-Bromophenyl-Phenylether	101-55-3			360 U	320 U	370 U	380 U
4-Chloro-3-Methylphenol	59-50-7			360 U	320 U	370 U	380 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP5C-36.5	80230-MIP5E-16	80230-MIP6-10	80230-MIP6-30
			Sample Location	MIP5C	MIP5E	MIP6	MIP6
		Sampling Date		06/24/2011	06/16/2011	05/23/2011	05/23/2011
4-Chloroaniline	106-47-8			360 U	320 U	370 U	380 U
4-Chlorophenyl-Phenylether	7005-72-3			360 U	320 U	370 U	380 U
4-Methylphenol	106-44-5	330		360 U	320 U	370 U	380 U
4-Nitroaniline	100-01-6			730 UJ	660 U	760 U	760 U
4-Nitrophenol	100-02-7			730 UJ	660 U	760 U	760 U
Acenaphthene	83-32-9	20000		360 U	320 U	370 U	380 U
Acenaphthylene	208-96-8	100000		360 U	320 U	370 U	380 U
Acetophenone	98-86-2			360 U	320 U	370 U	380 U
Anthracene	120-12-7	100000		360 U	320 U	370 U	380 U
Atrazine	1912-24-9			360 UJ	320 U	370 U	380 U
Benzaldehyde	100-52-7			360 U	320 U	370 U	380 U
Benzo(a)Anthracene	56-55-3	1000		360 U	320 U	370 U	380 U
Benzo(a)Pyrene	50-32-8	1000		360 U	320 U	370 U	380 U
Benzo(b)Fluoranthene	205-99-2	1000		360 U	320 U	370 U	380 U
Benzo(g,h,i)Perylene	191-24-2	100000		360 U	320 U	370 U	380 U
Benzo(k)Fluoranthene	207-08-9	800		360 U	320 U	370 U	380 U
bis(2-Chloroethoxy)Methane	111-91-1			360 U	320 U	370 U	380 U
bis(2-Chloroethyl) Ether	111-44-4			360 U	320 U	370 U	380 U
bis(2-Ethylhexyl)Phthalate	117-81-7			360 U	68 J	50 U	54 J
bis-Chloroisopropyl Ether	108-60-1			360 UJ	320 U	370 U	380 U
Butylbenzylphthalate	85-68-7			360 U	320 U	370 U	380 U
Caprolactam	105-60-2			360 U	320 U	370 U	380 U
Carbazole	86-74-8			360 U	320 U	370 U	380 U
Chrysene	218-01-9	1000		360 U	320 U	370 U	380 U
Dibenzo(a,h)Anthracene	53-70-3	330		360 U	320 U	370 U	380 U
Dibenzofuran	132-64-9	7000		360 U	320 U	370 U	380 U
Diethylphthalate	84-66-2			360 U	320 U	370 U	380 U
Dimethylphthalate	131-11-3			360 U	320 U	370 U	380 U
di-n-Butylphthalate	84-74-2			360 U	320 U	370 U	380 U
di-n-Octylphthalate	117-84-0			360 U	320 U	370 U	380 U
Fluoranthene	206-44-0	100000		360 U	320 U	370 U	380 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP5C-36.5	80230-MIP5E-16	80230-MIP6-10	80230-MIP6-30
			Sample Location	MIP5C 06/24/2011	MIP5E 06/16/2011	MIP6 05/23/2011	MIP6 05/23/2011
Fluorene	86-73-7	30000		360 U	320 U	370 U	380 U
Hexachlorobenzene	118-74-1	330		360 U	320 U	370 U	380 U
Hexachlorobutadiene	87-68-3			360 U	320 U	370 U	380 U
Hexachlorocyclopentadiene	77-47-4			360 U	320 U	370 U	380 U
Hexachloroethane	67-72-1			360 U	320 U	370 U	380 U
Indeno(1,2,3-cd)Pyrene	193-39-5	500		360 UJ	320 U	370 U	380 U
Isophorone	78-59-1			360 U	320 U	370 U	380 U
Naphthalene	91-20-3	12000		360 U	320 U	370 U	380 U
Nitrobenzene	98-95-3			360 U	320 U	370 UJ	380 UJ
n-Nitroso-di-n-Propylamine	621-64-7			360 UJ	320 U	370 U	380 U
n-Nitrosodiphenylamine	86-30-6			360 U	320 U	370 U	380 U
Pentachlorophenol	87-86-5	800		730 U	660 U	760 U	760 U
Phenanthrene	85-01-8	100000		360 U	320 U	370 U	380 U
Phenol	108-95-2	330		360 U	320 U	370 U	380 U
Pyrene	129-00-0	100000		360 U	320 U	370 U	380 U
<b>Pesticides (µg/kg)</b>							
4,4'-DDD	72-54-8	3.3			3.8 U		
4,4'-DDE	72-55-9	3.3			3.8 U		
4,4'-DDT	50-29-3	3.3			3.8 U		
Aldrin	309-00-2	5			1.9 U		
Alpha-BHC	319-84-6	20			1.9 U		
Alpha-Chlordane	5103-71-9	94			1.9 U		
Beta-BHC	319-85-7	36			1.9 U		
Delta-BHC	319-86-8	40			1.9 U		
Dieldrin	60-57-1	5			3.8 U		
Endosulfan I	959-98-8	2400			1.9 U		
Endosulfan II	33213-65-9	2400			3.8 U		
Endosulfan Sulfate	1031-07-8	2400			3.8 U		
Endrin	72-20-8	14			3.8 U		
Endrin Aldehyde	7421-93-4				3.8 U		
Endrin Ketone	53494-70-5				3.8 U		

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP5C-36.5 MIP5C 06/24/2011	80230-MIP5E-16 MIP5E 06/16/2011	80230-MIP6-10 MIP6 05/23/2011	80230-MIP6-30 MIP6 05/23/2011
Chemical Name	CAS#	Part 375 Unrestricted					
Gamma-BHC (Lindane)	58-89-9	100			1.9 U		
Gamma-Chlordane	5103-74-2				1.9 U		
Heptachlor	76-44-8	42			1.9 U		
Heptachlor Epoxide	1024-57-3				1.9 U		
Methoxychlor	72-43-5				19 U		
Toxaphene	8001-35-2				190 U		
<b>Polychlorinated Biphenyls (µg/kg)</b>							
Aroclor 1016	12674-11-2	100			38 U		
Aroclor 1221	11104-28-2	100			38 U		
Aroclor 1232	11141-16-5	100			38 U		
Aroclor 1242	53469-21-9	100			38 U		
Aroclor 1248	12672-29-6	100			38 U		
Aroclor 1254	11097-69-1	100			38 U		
Aroclor 1260	11096-82-5	100			38 U		
<b>Inorganics (mg/kg)</b>							
Aluminum	7429-90-5				7990		
Antimony	7440-36-0				1.1 U		
Arsenic	7440-38-2	13			3.7		
Barium	7440-39-3	350			62.6		
Beryllium	7440-41-7	7.2			0.9		
Cadmium	7440-43-9	2.5			0.27 U		
Calcium	7440-70-2				2970		
Chromium	7440-47-3	30			18.3		
Cobalt	7440-48-4				8.4		
Copper	7440-50-8	50			10.8		
Iron	7439-89-6				18900		
Lead	7439-92-1	63			12.4		
Magnesium	7439-95-4				4980		
Manganese	7439-96-5	1600			652		
Mercury	7439-97-6	0.18			0.039 U		
Nickel	7440-02-0	30			19.4		

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP5C-36.5	80230-MIP5E-16	80230-MIP6-10	80230-MIP6-30
			Sample Location	MIP5C	MIP5E	MIP6	MIP6
			Sampling Date	06/24/2011	06/16/2011	05/23/2011	05/23/2011
Potassium	7440-09-7				1850		
Selenium	7782-49-2	3.9			1.6	U	
Silver	7440-22-4	2			1.6	U	
Sodium	7440-23-5				158		
Thallium	7440-28-0				1.1	U	
Vanadium	7440-62-2				24.7		
Zinc	7440-66-6	109			53		

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP6B-42.5 MIP6B 06/24/2011	80230-MIP6B-42.5-DU MIP6B 06/24/2011	80230-MIP7-38 MIP7 05/23/2011	80230-MIP7-38-DUP MIP7 05/23/2011	80230-MW12S- MW12S 06/24/2011
Chemical Name	CAS#	Part 375 Unrestricted						
<b>Volatile Organic Compounds (µg/kg)</b>								
1,1,1,2-Tetrachloroethane	630-20-6		5 U	4.8 U	5.5 U	5.4 U		5
1,1,1-Trichloroethane	71-55-6	680	5 U	4.8 U	5.5 U	5.4 U		5
1,1,2,2-Tetrachloroethane	79-34-5		5 U	4.8 U	5.5 U	5.4 U		5
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5 U	4.8 U	5.5 U	5.4 U		5
1,1,2-Trichloroethane	79-00-5		5 U	4.8 U	5.5 U	5.4 U		5
1,1-Dichloroethane	75-34-3	270	5 U	4.8 U	5.5 U	5.4 U		5
1,1-Dichloroethene	75-35-4	330	5 U	4.8 U	5.5 U	5.4 U		5
1,1-Dichloropropene	563-58-6		5 U	4.8 U	5.5 U	5.4 U		5
1,2,3-Trichlorobenzene	87-61-6		5 U	4.8 U	5.5 U	5.4 U		5
1,2,3-Trichloropropane	96-18-4		5 U	4.8 U	5.5 U	5.4 U		5
1,2,4-Trichlorobenzene	120-82-1		5 U	4.8 U	5.5 U	5.4 U		5
1,2,4-Trimethylbenzene	95-63-6	3600	5 U	4.8 U	5.5 U	5.4 U		1.3
1,2-Dibromo-3-Chloropropane	96-12-8		5 U	4.8 U	5.5 U	5.4 U		5
1,2-Dibromoethane (EDB)	106-93-4		5 U	4.8 U	5.5 U	5.4 U		5
1,2-Dichlorobenzene	95-50-1	1100	5 U	4.8 U	5.5 U	5.4 U		5
1,2-Dichloroethane	107-06-2	20	5 U	4.8 U	5.5 U	5.4 U		5
1,2-Dichloropropane	78-87-5		5 U	4.8 U	5.5 U	5.4 U		5
1,3,5-Trimethylbenzene	108-67-8	8400	5 U	4.8 U	5.5 U	5.4 U		5
1,3-Dichlorobenzene	541-73-1	2400	5 U	4.8 U	5.5 U	5.4 U		5
1,3-Dichloropropane	142-28-9		5 U	4.8 U	5.5 U	5.4 U		5
1,4-Dichlorobenzene	106-46-7	1800	5 U	4.8 U	5.5 U	5.4 U		5
2,2-Dichloropropane	594-20-7		5 U	4.8 U	5.5 U	5.4 U		5
2-Butanone (MEK)	78-93-3	120	5 R	4.8 R	5.5 R	5.4 R		5
2-Chlorotoluene	95-49-8		5 U	4.8 U	5.5 U	5.4 U		5
2-Hexanone	591-78-6		5 U	4.8 U	5.5 U	5.4 U		5
4-Chlortoluene	106-43-4		5 U	4.8 U	5.5 U	5.4 U		5
4-Methyl-2-Pentanone (MIBK)	108-10-1		5 U	4.8 U	5.5 U	5.4 U		5
Acetone	67-64-1	50	5 R	4.8 R	5.5 R	5.4 R		5
Benzene	71-43-2	60	5 U	4.8 U	5.5 U	5.4 U		5
Bromobenzene	108-86-1		5 U	4.8 U	5.5 U	5.4 U		5

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP6B-42.5	80230-MIP6B-42.5-DU	80230-MIP7-38	80230-MIP7-38-DUP	80230-MW12S-
			Sample Location	MIP6B	MIP6B	MIP7	MIP7	MW12S
		Sampling Date		06/24/2011	06/24/2011	05/23/2011	05/23/2011	06/24/2011
Bromochloromethane	74-97-5			5 U	4.8 U	5.5 U	5.4 U	5
Bromodichloromethane	75-27-4			5 U	4.8 U	5.5 U	5.4 U	5
Bromoform	75-25-2			5 U	4.8 U	5.5 U	5.4 U	5
Bromomethane	74-83-9			5 U	4.8 U	5.5 U	5.4 U	5
Carbon Disulfide	75-15-0			5 U	4.8 U	5.5 U	5.4 U	5
Carbon Tetrachloride	56-23-5	760		5 U	4.8 U	5.5 U	5.4 U	5
Chlorobenzene	108-90-7	1100		5 U	4.8 U	5.5 U	5.4 U	5
Chloroethane	75-00-3			5 U	4.8 U	5.5 U	5.4 U	5
Chloroform	67-66-3	370		5 U	4.8 U	5.5 U	5.4 U	5
Chloromethane	74-87-3			5 U	4.8 U	5.5 U	5.4 U	5
cis-1,2-Dichloroethene	156-59-2	250		5 U	4.8 U	5.5 U	5.4 U	2.3
cis-1,3-Dichloropropene	10061-01-5			5 U	4.8 U	5.5 U	5.4 U	5
Cyclohexane	110-82-7			5 U	4.8 U	5.5 U	5.4 U	5
Dibromochloromethane	124-48-1			5 U	4.8 U	5.5 U	5.4 U	5
Dibromomethane	74-95-3			5 U	4.8 U	5.5 U	5.4 U	5
Dichlorodifluoromethane	75-71-8			5 U	4.8 U	5.5 U	5.4 U	5
Ethylbenzene	100-41-4	1000		5 U	4.8 U	5.5 U	5.4 U	5
Hexachlorobutadiene	87-68-3			5 U	4.8 UJ	5.5 U	5.4 U	5
Isopropylbenzene	98-82-8			5 U	4.8 U	5.5 U	5.4 U	5
m,p-Xylene	179601-23-1	260		5 U	4.8 U	5.5 U	5.4 U	5
Methyl Acetate	79-20-9			5 U	4.8 U	5.5 U	5.4 U	5
Methyl Iodide	74-88-4			5 U	4.8 U	5.5 U	5.4 U	5
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	930		5 U	4.8 U	5.5 U	5.4 U	5
Methylcyclohexane	108-87-2			5 U	4.8 U	5.5 U	5.4 U	5
Methylene Chloride	75-09-2	50		5 U	4.8 U	1.7 J	2.1 J	5
Naphthalene	91-20-3	12000		5 U	1.3 J	5.5 UJ	5.4 UJ	2.1
n-Butylbenzene	104-51-8	12000		5 U	4.8 UJ	5.5 U	5.4 U	5
n-Propylbenzene	103-65-1	3900		5 U	4.8 U	5.5 U	5.4 U	5
o-Xylene	95-47-6	260		5 U	4.8 U	5.5 U	5.4 U	5
p-Isopropyltoluene	99-87-6			5 U	4.8 U	5.5 U	5.4 U	5
sec-Butylbenzene	135-98-8	11000		5 U	4.8 U	5.5 U	5.4 U	5

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID Sample Location Sampling Date	80230-MIP6B-42.5 MIP6B 06/24/2011	80230-MIP6B-42.5-DU MIP6B 06/24/2011	80230-MIP7-38 MIP7 05/23/2011	80230-MIP7-38-DUP MIP7 05/23/2011	80230-MW12S- MW12S 06/24/2011
Chemical Name	CAS#	Part 375 Unrestricted						
Styrene	100-42-5			5 U	4.8 U	5.5 U	5.4 U	5
tert-Butylbenzene	98-06-6	5900		5 U	4.8 U	5.5 U	5.4 U	5
Tetrachloroethene	127-18-4	1300		5 U	1.6 J	5.5 U	5.4 U	880
Toluene	108-88-3	700		5 U	4.8 U	5.5 U	5.4 U	5
Total Xylenes	1330-20-7	260		5 U	4.8 U	5.5 U	5.4 U	5
trans-1,2-Dichloroethene	156-60-5	190		5 U	4.8 U	5.5 U	5.4 U	5
trans-1,3-Dichloropropene	10061-02-6			5 U	4.8 U	5.5 U	5.4 U	5
Trichloroethene	79-01-6	470		5 U	4.8 U	5.5 U	5.4 U	5
Trichlorofluoromethane	75-69-4			5 U	4.8 U	5.5 U	5.4 U	5
Vinyl Acetate	108-05-4			5 U	4.8 U	5.5 U	5.4 U	5
Vinyl Chloride	75-01-4	20		5 U	4.8 U	5.5 U	5.4 U	5
<b>Semi-Volatile Organic Compounds (µg/kg)</b>								
1,1'-Biphenyl	92-52-4					360 U	360 U	
2,4,5-Trichlorophenol	95-95-4					730 U	740 U	
2,4,6-Trichlorophenol	88-06-2					360 U	360 U	
2,4-Dichlorophenol	120-83-2					360 U	360 U	
2,4-Dimethylphenol	105-67-9					360 U	360 U	
2,4-Dinitrophenol	51-28-5					730 UJ	740 UJ	
2,4-Dinitrotoluene	121-14-2					360 U	360 U	
2,6-Dinitrotoluene	606-20-2					360 U	360 U	
2-Chloronaphthalene	91-58-7					360 U	360 U	
2-Chlorophenol	95-57-8					360 U	360 U	
2-Methylnaphthalene	91-57-6					360 UJ	360 UJ	
2-Methylphenol	95-48-7	330				360 U	360 U	
2-Nitroaniline	88-74-4					730 U	740 U	
2-Nitrophenol	88-75-5					360 U	360 U	
3,3'-Dichlorobenzidine	91-94-1					360 U	360 U	
3-Nitroaniline	99-09-2					730 U	740 U	
4,6-Dinitro-2-Methylphenol	534-52-1					730 U	740 U	
4-Bromophenyl-Phenylether	101-55-3					360 U	360 U	
4-Chloro-3-Methylphenol	59-50-7					360 U	360 U	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP6B-42.5	80230-MIP6B-42.5-DU	80230-MIP7-38	80230-MIP7-38-DUP	80230-MW12S-
			Sample Location	MIP6B	MIP6B	MIP7	MIP7	MW12S
		Sampling Date	06/24/2011	06/24/2011	06/24/2011	05/23/2011	05/23/2011	06/24/2011
4-Chloroaniline	106-47-8					360 U	360 U	
4-Chlorophenyl-Phenylether	7005-72-3					360 U	360 U	
4-Methylphenol	106-44-5	330				360 U	360 U	
4-Nitroaniline	100-01-6					730 U	740 U	
4-Nitrophenol	100-02-7					730 U	740 U	
Acenaphthene	83-32-9	20000				360 U	360 U	
Acenaphthylene	208-96-8	100000				360 U	360 U	
Acetophenone	98-86-2					360 U	360 U	
Anthracene	120-12-7	100000				360 U	360 U	
Atrazine	1912-24-9					360 U	360 U	
Benzaldehyde	100-52-7					360 U	360 U	
Benzo(a)Anthracene	56-55-3	1000				48 J	150 J	
Benzo(a)Pyrene	50-32-8	1000				59 J	180 J	
Benzo(b)Fluoranthene	205-99-2	1000				100 J	280 J	
Benzo(g,h,i)Perylene	191-24-2	100000				61 J	160 J	
Benzo(k)Fluoranthene	207-08-9	800				360 U	120 J	
bis(2-Chloroethoxy)Methane	111-91-1					360 U	360 U	
bis(2-Chloroethyl) Ether	111-44-4					360 U	360 U	
bis(2-Ethylhexyl)Phthalate	117-81-7					74 J	69 J	
bis-Chloroisopropyl Ether	108-60-1					360 U	360 U	
Butylbenzylphthalate	85-68-7					360 U	360 U	
Caprolactam	105-60-2					360 U	360 U	
Carbazole	86-74-8					360 U	360 U	
Chrysene	218-01-9	1000				100 J	260 J	
Dibenzo(a,h)Anthracene	53-70-3	330				360 U	38 J	
Dibenzofuran	132-64-9	7000				360 U	360 U	
Diethylphthalate	84-66-2					360 U	360 U	
Dimethylphthalate	131-11-3					360 U	360 U	
di-n-Butylphthalate	84-74-2					360 U	360 U	
di-n-Octylphthalate	117-84-0					360 U	360 U	
Fluoranthene	206-44-0	100000				130 J	350 J	

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Sample ID Sample Location Sampling Date	80230-MIP6B-42.5	80230-MIP6B-42.5-DU	80230-MIP7-38	80230-MIP7-38-DUP	80230-MW12S-
			MIP6B 06/24/2011	MIP6B 06/24/2011	MIP7 05/23/2011	MIP7 05/23/2011	MW12S 06/24/2011
Fluorene	86-73-7	Part 375 Unrestricted			360 U	360 U	
Hexachlorobenzene	118-74-1	30000			360 U	360 U	
Hexachlorobutadiene	87-68-3	330			360 U	360 U	
Hexachlorocyclopentadiene	77-47-4				360 U	360 U	
Hexachloroethane	67-72-1				360 U	360 U	
Indeno(1,2,3-cd)Pyrene	193-39-5	500			46 J	120 J	
Isophorone	78-59-1				360 U	360 U	
Naphthalene	91-20-3	12000			360 U	360 U	
Nitrobenzene	98-95-3				360 UJ	360 UJ	
n-Nitroso-di-n-Propylamine	621-64-7				360 U	360 U	
n-Nitrosodiphenylamine	86-30-6				360 U	360 U	
Pentachlorophenol	87-86-5	800			730 U	740 U	
Phenanthrene	85-01-8	100000			37 J	83 J	
Phenol	108-95-2	330			360 U	360 U	
Pyrene	129-00-0	100000			95 J	250 J	
<b>Pesticides (µg/kg)</b>							
4,4'-DDD	72-54-8	3.3					
4,4'-DDE	72-55-9	3.3					
4,4'-DDT	50-29-3	3.3					
Aldrin	309-00-2	5					
Alpha-BHC	319-84-6	20					
Alpha-Chlordane	5103-71-9	94					
Beta-BHC	319-85-7	36					
Delta-BHC	319-86-8	40					
Dieldrin	60-57-1	5					
Endosulfan I	959-98-8	2400					
Endosulfan II	33213-65-9	2400					
Endosulfan Sulfate	1031-07-8	2400					
Endrin	72-20-8	14					
Endrin Aldehyde	7421-93-4						
Endrin Ketone	53494-70-5						

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID	80230-MIP6B-42.5	80230-MIP6B-42.5-DU	80230-MIP7-38	80230-MIP7-38-DUP	80230-MW12S-
Chemical Name	CAS#	Sample Location	Sampling Date	MIP6B 06/24/2011	MIP6B 06/24/2011	MIP7 05/23/2011	MIP7 05/23/2011	MW12S 06/24/2011
Gamma-BHC (Lindane)	58-89-9	100						
Gamma-Chlordane	5103-74-2							
Heptachlor	76-44-8	42						
Heptachlor Epoxide	1024-57-3							
Methoxychlor	72-43-5							
Toxaphene	8001-35-2							
<b>Polychlorinated Biphenyls (µg/kg)</b>								
Aroclor 1016	12674-11-2	100						
Aroclor 1221	11104-28-2	100						
Aroclor 1232	11141-16-5	100						
Aroclor 1242	53469-21-9	100						
Aroclor 1248	12672-29-6	100						
Aroclor 1254	11097-69-1	100						
Aroclor 1260	11096-82-5	100						
<b>Inorganics (mg/kg)</b>								
Aluminum	7429-90-5							
Antimony	7440-36-0							
Arsenic	7440-38-2	13						
Barium	7440-39-3	350						
Beryllium	7440-41-7	7.2						
Cadmium	7440-43-9	2.5						
Calcium	7440-70-2							
Chromium	7440-47-3	30						
Cobalt	7440-48-4							
Copper	7440-50-8	50						
Iron	7439-89-6							
Lead	7439-92-1	63						
Magnesium	7439-95-4							
Manganese	7439-96-5	1600						
Mercury	7439-97-6	0.18						
Nickel	7440-02-0	30						

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	80230-MIP6B-42.5	80230-MIP6B-42.5-DU	80230-MIP7-38	80230-MIP7-38-DUP	80230-MW12S-
			Sample Location	MIP6B	MIP6B	MIP7	MIP7	MW12S
		Sampling Date	06/24/2011	06/24/2011	06/24/2011	05/23/2011	05/23/2011	06/24/2011
Potassium	7440-09-7							
Selenium	7782-49-2	3.9						
Silver	7440-22-4	2						
Sodium	7440-23-5							
Thallium	7440-28-0							
Vanadium	7440-62-2							
Zinc	7440-66-6	109						

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID	35	80230-MW12S-35ME	PM-SB-1A-A-1112011	PM-SB-1A-B-1112011	PM-SB-1B-A-1112011
Sample Location				MW12S	PM-SB-1A-A	PM-SB-1A-B	PM-SB-1B-A	
Sampling Date				06/24/2011	11/01/2011	11/01/2011	11/02/2011	
Chemical Name	CAS#	Part 375 Unrestricted						
<b>Volatile Organic Compounds (µg/kg)</b>								
1,1,1,2-Tetrachloroethane	630-20-6		U	310	U	5.8	U	5.9
1,1,1-Trichloroethane	71-55-6	680	U	310	U	5.8	U	5.9
1,1,2,2-Tetrachloroethane	79-34-5		U	310	U	5.8	U	5.9
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		U	310	U	5.8	U	5.9
1,1,2-Trichloroethane	79-00-5		U	310	U	5.8	U	5.9
1,1-Dichloroethane	75-34-3	270	U	310	U	5.8	U	5.9
1,1-Dichloroethene	75-35-4	330	U	310	U	5.8	U	5.9
1,1-Dichloropropene	563-58-6		U	310	U	5.8	U	5.9
1,2,3-Trichlorobenzene	87-61-6		U	310	U	5.8	U	5.9
1,2,3-Trichloropropane	96-18-4		U	310	U	5.8	U	5.9
1,2,4-Trichlorobenzene	120-82-1		U	310	U	5.8	UJ	5.9
1,2,4-Trimethylbenzene	95-63-6	3600	J	310	U	5.8	U	5.9
1,2-Dibromo-3-Chloropropane	96-12-8		U	310	U	5.8	UJ	5.9
1,2-Dibromoethane (EDB)	106-93-4		U	310	U	5.8	U	5.9
1,2-Dichlorobenzene	95-50-1	1100	U	310	U	5.8	UJ	5.9
1,2-Dichloroethane	107-06-2	20	U	310	U	5.8	U	5.9
1,2-Dichloropropane	78-87-5		U	310	U	5.8	U	5.9
1,3,5-Trimethylbenzene	108-67-8	8400	U	310	U	5.8	U	5.9
1,3-Dichlorobenzene	541-73-1	2400	U	310	U	5.8	UJ	5.9
1,3-Dichloropropane	142-28-9		U	310	U	5.8	U	5.9
1,4-Dichlorobenzene	106-46-7	1800	U	310	U	5.8	UJ	5.9
2,2-Dichloropropane	594-20-7		U	310	U	5.8	U	5.9
2-Butanone (MEK)	78-93-3	120	R	310	R	5.8	R	5.9
2-Chlorotoluene	95-49-8		U	310	U	5.8	U	5.9
2-Hexanone	591-78-6		U	310	UJ	5.8	UJ	5.9
4-Chlorotoluene	106-43-4		U	310	U	5.8	U	5.9
4-Methyl-2-Pentanone (MIBK)	108-10-1		U	310	U	5.8	U	5.9
Acetone	67-64-1	50	R	310	UJ	17	J	5.9
Benzene	71-43-2	60	U	310	U	5.8	U	5.9
Bromobenzene	108-86-1		U	310	U	5.8	U	5.9

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	35	80230-MW12S-35ME	PM-SB-1A-A-1112011	PM-SB-1A-B-1112011	PM-SB-1B-A-1122011
			Sample Location	MW12S	PM-SB-1A-A 06/24/2011	PM-SB-1A-B 11/01/2011	PM-SB-1B-A 11/02/2011	
Sampling Date								
Bromochloromethane	74-97-5		U	310	U	5.8	U	5.3
Bromodichloromethane	75-27-4		U	310	U	5.8	U	5.3
Bromoform	75-25-2		U	310	U	5.8	U	5.3
Bromomethane	74-83-9		U	310	U	5.8	U	5.3
Carbon Disulfide	75-15-0		U	310	U	5.8	U	5.3
Carbon Tetrachloride	56-23-5	760	U	310	U	5.8	U	5.3
Chlorobenzene	108-90-7	1100	U	310	U	5.8	U	5.3
Chloroethane	75-00-3		U	310	U	5.8	U	5.3
Chloroform	67-66-3	370	U	310	U	5.8	U	5.3
Chloromethane	74-87-3		U	310	U	5.8	U	5.3
cis-1,2-Dichloroethene	156-59-2	250	J	310	U	5.8	U	3.2
cis-1,3-Dichloropropene	10061-01-5		U	310	U	5.8	U	5.3
Cyclohexane	110-82-7		U	310	U	5.8	U	5.3
Dibromochloromethane	124-48-1		U	310	U	5.8	U	5.3
Dibromomethane	74-95-3		U	310	U	5.8	U	5.3
Dichlorodifluoromethane	75-71-8		U	310	U	5.8	U	5.3
Ethylbenzene	100-41-4	1000	U	310	U	5.8	U	5.3
Hexachlorobutadiene	87-68-3		UJ	310	U	5.8	U	5.3
Isopropylbenzene	98-82-8		U	310	U	5.8	U	5.3
m,p-Xylene	179601-23-1	260	U	310	U	5.8	U	5.3
Methyl Acetate	79-20-9		U	310	U	5.8	U	5.3
Methyl Iodide	74-88-4		U	310	U	5.8	UJ	5.3
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	930	U	310	U	5.8	U	5.3
Methylcyclohexane	108-87-2		U	310	U	5.8	U	5.3
Methylene Chloride	75-09-2	50	U	310	U	5.8	U	5.3
Naphthalene	91-20-3	12000	J	310	U	5.8	U	5.3
n-Butylbenzene	104-51-8	12000	UJ	310	U	5.8	U	5.3
n-Propylbenzene	103-65-1	3900	U	310	U	5.8	U	5.3
o-Xylene	95-47-6	260	U	310	U	5.8	U	5.3
p-Isopropyltoluene	99-87-6		U	310	U	5.8	U	5.3
sec-Butylbenzene	135-98-8	11000	U	310	U	5.8	U	5.3

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	35	80230-MW12S-35ME	PM-SB-1A-A-1112011	PM-SB-1A-B-1112011	PM-SB-1B-A-1122011
			Sample Location	MW12S	PM-SB-1A-A	PM-SB-1A-B	PM-SB-1B-A	
		Sampling Date		06/24/2011	11/01/2011	11/01/2011	11/02/2011	
Styrene	100-42-5		U	310	U	5.8	U	5.3
tert-Butylbenzene	98-06-6	5900	U	310	U	5.8	U	5.3
Tetrachloroethene	127-18-4	1300	E	1000		5.8	J	2.7
Toluene	108-88-3	700	U	310	U	7.1	U	5.3
Total Xylenes	1330-20-7	260	U	310	U	5.8	U	5.3
trans-1,2-Dichloroethene	156-60-5	190	U	310	U	5.8	U	5.3
trans-1,3-Dichloropropene	10061-02-6		U	310	U	5.8	U	5.3
Trichloroethene	79-01-6	470	U	310	U	5.8	U	3.2
Trichlorofluoromethane	75-69-4		U	310	U	5.8	U	5.3
Vinyl Acetate	108-05-4		U	310	U	5.8	U	5.3
Vinyl Chloride	75-01-4	20	U	310	U	5.8	U	5.3
Vinyl Chloride	75-01-4	20	U	310	U	5.8	U	5.9
<b>Semi-Volatile Organic Compounds (µg/kg)</b>								
1,1'-Biphenyl	92-52-4							
2,4,5-Trichlorophenol	95-95-4							
2,4,6-Trichlorophenol	88-06-2							
2,4-Dichlorophenol	120-83-2							
2,4-Dimethylphenol	105-67-9							
2,4-Dinitrophenol	51-28-5							
2,4-Dinitrotoluene	121-14-2							
2,6-Dinitrotoluene	606-20-2							
2-Chloronaphthalene	91-58-7							
2-Chlorophenol	95-57-8							
2-Methylnaphthalene	91-57-6							
2-Methylphenol	95-48-7	330						
2-Nitroaniline	88-74-4							
2-Nitrophenol	88-75-5							
3,3'-Dichlorobenzidine	91-94-1							
3-Nitroaniline	99-09-2							
4,6-Dinitro-2-Methylphenol	534-52-1							
4-Bromophenyl-Phenylether	101-55-3							
4-Chloro-3-Methylphenol	59-50-7							

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	35	80230-MW12S-35ME	PM-SB-1A-A-1112011	PM-SB-1A-B-1112011	PM-SB-1B-A-1122011
			Sample Location	MW12S	PM-SB-1A-A 06/24/2011	PM-SB-1A-B 11/01/2011	PM-SB-1B-A 11/02/2011	
Sampling Date								
4-Chloroaniline	106-47-8							
4-Chlorophenyl-Phenylether	7005-72-3							
4-Methylphenol	106-44-5	330						
4-Nitroaniline	100-01-6							
4-Nitrophenol	100-02-7							
Acenaphthene	83-32-9	20000						
Acenaphthylene	208-96-8	100000						
Acetophenone	98-86-2							
Anthracene	120-12-7	100000						
Atrazine	1912-24-9							
Benzaldehyde	100-52-7							
Benzo(a)Anthracene	56-55-3	1000						
Benzo(a)Pyrene	50-32-8	1000						
Benzo(b)Fluoranthene	205-99-2	1000						
Benzo(g,h,i)Perylene	191-24-2	100000						
Benzo(k)Fluoranthene	207-08-9	800						
bis(2-Chloroethoxy)Methane	111-91-1							
bis(2-Chloroethyl) Ether	111-44-4							
bis(2-Ethylhexyl)Phthalate	117-81-7							
bis-Chloroisopropyl Ether	108-60-1							
Butylbenzylphthalate	85-68-7							
Caprolactam	105-60-2							
Carbazole	86-74-8							
Chrysene	218-01-9	1000						
Dibenzo(a,h)Anthracene	53-70-3	330						
Dibenzofuran	132-64-9	7000						
Diethylphthalate	84-66-2							
Dimethylphthalate	131-11-3							
di-n-Butylphthalate	84-74-2							
di-n-Octylphthalate	117-84-0							
Fluoranthene	206-44-0	100000						

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	35	80230-MW12S-35ME	PM-SB-1A-A-1112011	PM-SB-1A-B-1112011	PM-SB-1B-A-1122011
			Sample Location	MW12S	PM-SB-1A-A 06/24/2011	PM-SB-1A-B 11/01/2011	PM-SB-1B-A 11/02/2011	
Fluorene	86-73-7	30000						
Hexachlorobenzene	118-74-1	330						
Hexachlorobutadiene	87-68-3							
Hexachlorocyclopentadiene	77-47-4							
Hexachloroethane	67-72-1							
Indeno(1,2,3-cd)Pyrene	193-39-5	500						
Isophorone	78-59-1							
Naphthalene	91-20-3	12000						
Nitrobenzene	98-95-3							
n-Nitroso-di-n-Propylamine	621-64-7							
n-Nitrosodiphenylamine	86-30-6							
Pentachlorophenol	87-86-5	800						
Phenanthrene	85-01-8	100000						
Phenol	108-95-2	330						
Pyrene	129-00-0	100000						
<b>Pesticides (µg/kg)</b>								
4,4'-DDD	72-54-8	3.3						
4,4'-DDE	72-55-9	3.3						
4,4'-DDT	50-29-3	3.3						
Aldrin	309-00-2	5						
Alpha-BHC	319-84-6	20						
Alpha-Chlordane	5103-71-9	94						
Beta-BHC	319-85-7	36						
Delta-BHC	319-86-8	40						
Dieldrin	60-57-1	5						
Endosulfan I	959-98-8	2400						
Endosulfan II	33213-65-9	2400						
Endosulfan Sulfate	1031-07-8	2400						
Endrin	72-20-8	14						
Endrin Aldehyde	7421-93-4							
Endrin Ketone	53494-70-5							

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID	35	80230-MW12S-35ME	PM-SB-1A-A-1112011	PM-SB-1A-B-1112011	PM-SB-1B-A-1122011
Chemical Name	CAS#	Part 375 Unrestricted	Sample Location	MW12S	PM-SB-1A-A	11/01/2011	PM-SB-1A-B	PM-SB-1B-A
Gamma-BHC (Lindane)	58-89-9	100						
Gamma-Chlordane	5103-74-2							
Heptachlor	76-44-8	42						
Heptachlor Epoxide	1024-57-3							
Methoxychlor	72-43-5							
Toxaphene	8001-35-2							
<b>Polychlorinated Biphenyls (µg/kg)</b>								
Aroclor 1016	12674-11-2	100						
Aroclor 1221	11104-28-2	100						
Aroclor 1232	11141-16-5	100						
Aroclor 1242	53469-21-9	100						
Aroclor 1248	12672-29-6	100						
Aroclor 1254	11097-69-1	100						
Aroclor 1260	11096-82-5	100						
<b>Inorganics (mg/kg)</b>								
Aluminum	7429-90-5							
Antimony	7440-36-0							
Arsenic	7440-38-2	13						
Barium	7440-39-3	350						
Beryllium	7440-41-7	7.2						
Cadmium	7440-43-9	2.5						
Calcium	7440-70-2							
Chromium	7440-47-3	30						
Cobalt	7440-48-4							
Copper	7440-50-8	50						
Iron	7439-89-6							
Lead	7439-92-1	63						
Magnesium	7439-95-4							
Manganese	7439-96-5	1600						
Mercury	7439-97-6	0.18						
Nickel	7440-02-0	30						

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	35	80230-MW12S-35ME	PM-SB-1A-A-1112011	PM-SB-1A-B-1112011	PM-SB-1B-A-1122011
			Sample Location	MW12S	PM-SB-1A-A 06/24/2011	11/01/2011	PM-SB-1A-B 11/01/2011	PM-SB-1B-A 11/02/2011
Potassium	7440-09-7							
Selenium	7782-49-2	3.9						
Silver	7440-22-4	2						
Sodium	7440-23-5							
Thallium	7440-28-0							
Vanadium	7440-62-2							
Zinc	7440-66-6	109						

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

		Sample ID	PM-SB-1B-B-1122011	PM-SB-2-A-1112011	PM-SB-2-B-1112011	PM-SB-3-A-1112011
Chemical Name	CAS#	Sample Location Sampling Date	PM-SB-1B-B 11/02/2011	PM-SB-2-A 11/01/2011	PM-SB-2-B 11/01/2011	PM-SB-3-A 11/01/2011
<b>Volatile Organic Compounds (µg/kg)</b>						
1,1,1,2-Tetrachloroethane	630-20-6		5.5 U	5.9 U	6 U	5.9 U
1,1,1-Trichloroethane	71-55-6	680	5.5 U	5.9 U	6 U	5.9 U
1,1,2,2-Tetrachloroethane	79-34-5		5.5 U	5.9 U	6 U	5.9 U
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5.5 U	5.9 U	6 U	5.9 U
1,1,2-Trichloroethane	79-00-5		5.5 U	5.9 U	6 U	5.9 U
1,1-Dichloroethane	75-34-3	270	5.5 U	5.9 U	6 U	5.9 U
1,1-Dichloroethene	75-35-4	330	5.5 U	5.9 U	6 U	5.9 U
1,1-Dichloropropene	563-58-6		5.5 U	5.9 U	6 U	5.9 U
1,2,3-Trichlorobenzene	87-61-6		5.5 U	5.9 U	6 U	5.9 U
1,2,3-Trichloropropane	96-18-4		5.5 U	5.9 U	6 U	5.9 U
1,2,4-Trichlorobenzene	120-82-1		5.5 U	5.9 UJ	6 UJ	5.9 UJ
1,2,4-Trimethylbenzene	95-63-6	3600	5.5 U	5.9 U	6 U	5.9 U
1,2-Dibromo-3-Chloropropane	96-12-8		5.5 U	5.9 UJ	6 UJ	5.9 UJ
1,2-Dibromoethane (EDB)	106-93-4		5.5 U	5.9 U	6 U	5.9 U
1,2-Dichlorobenzene	95-50-1	1100	5.5 U	5.9 UJ	6 UJ	5.9 UJ
1,2-Dichloroethane	107-06-2	20	5.5 U	5.9 U	6 U	5.9 U
1,2-Dichloropropane	78-87-5		5.5 U	5.9 U	6 U	5.9 U
1,3,5-Trimethylbenzene	108-67-8	8400	5.5 U	5.9 U	6 U	5.9 U
1,3-Dichlorobenzene	541-73-1	2400	5.5 U	5.9 UJ	6 UJ	5.9 UJ
1,3-Dichloropropane	142-28-9		5.5 U	5.9 U	6 U	5.9 U
1,4-Dichlorobenzene	106-46-7	1800	5.5 U	5.9 UJ	6 UJ	5.9 UJ
2,2-Dichloropropane	594-20-7		5.5 U	5.9 U	6 U	5.9 U
2-Butanone (MEK)	78-93-3	120	5.5 R	5.9 R	6 R	5.9 R
2-Chlorotoluene	95-49-8		5.5 U	5.9 U	6 U	5.9 U
2-Hexanone	591-78-6		5.5 U	5.9 UJ	6 UJ	5.9 UJ
4-Chlorotoluene	106-43-4		5.5 U	5.9 U	6 U	5.9 U
4-Methyl-2-Pentanone (MIBK)	108-10-1		5.5 U	5.9 U	6 U	5.9 U
Acetone	67-64-1	50	5.5 R	5.9 R	6 R	5.9 R
Benzene	71-43-2	60	5.5 U	5.9 U	6 U	5.9 U
Bromobenzene	108-86-1		5.5 U	5.9 U	6 U	5.9 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-1B-B-1122011	PM-SB-2-A-1112011	PM-SB-2-B-1112011	PM-SB-3-A-1112011
			Sample Location	PM-SB-1B-B 11/02/2011	PM-SB-2-A 11/01/2011	PM-SB-2-B 11/01/2011	PM-SB-3-A 11/01/2011
Bromochloromethane	74-97-5			5.5 U	5.9 U	6 U	5.9 U
Bromodichloromethane	75-27-4			5.5 U	5.9 U	6 U	5.9 U
Bromoform	75-25-2			5.5 U	5.9 U	6 U	5.9 U
Bromomethane	74-83-9			5.5 UJ	5.9 U	6 U	5.9 U
Carbon Disulfide	75-15-0			5.5 U	5.9 U	6 U	5.9 U
Carbon Tetrachloride	56-23-5	760		5.5 UJ	5.9 UJ	6 UJ	5.9 UJ
Chlorobenzene	108-90-7	1100		5.5 U	5.9 U	6 U	5.9 U
Chloroethane	75-00-3			5.5 U	5.9 U	6 U	5.9 U
Chloroform	67-66-3	370		5.5 U	5.9 U	6 U	5.9 U
Chloromethane	74-87-3			5.5 UJ	5.9 U	6 U	5.9 U
cis-1,2-Dichloroethene	156-59-2	250		5.5 U	3.5 J	14	5.9 U
cis-1,3-Dichloropropene	10061-01-5			5.5 U	5.9 U	6 U	5.9 U
Cyclohexane	110-82-7			5.5 U	5.9 U	6 U	5.9 U
Dibromochloromethane	124-48-1			5.5 U	5.9 U	6 U	5.9 U
Dibromomethane	74-95-3			5.5 U	5.9 U	6 U	5.9 U
Dichlorodifluoromethane	75-71-8			5.5 U	5.9 U	6 U	5.9 U
Ethylbenzene	100-41-4	1000		5.5 U	5.9 U	6 U	5.9 U
Hexachlorobutadiene	87-68-3			5.5 U	5.9 U	6 U	5.9 U
Isopropylbenzene	98-82-8			5.5 UJ	5.9 U	6 U	5.9 U
m,p-Xylene	179601-23-1	260		5.5 U	5.9 U	6 U	5.9 U
Methyl Acetate	79-20-9			5.5 U	5.9 U	6 U	5.9 U
Methyl Iodide	74-88-4			5.5 U	5.9 UJ	6 UJ	5.9 UJ
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	930		5.5 U	5.9 U	6 U	5.9 U
Methylcyclohexane	108-87-2			5.5 U	5.9 U	6 U	5.9 U
Methylene Chloride	75-09-2	50		2.9 J	5.9 U	2 J	5.9 U
Naphthalene	91-20-3	12000		5.5 UJ	5.9 UJ	6 UJ	5.9 UJ
n-Butylbenzene	104-51-8	12000		5.5 U	5.9 U	6 U	5.9 U
n-Propylbenzene	103-65-1	3900		5.5 U	5.9 U	6 U	5.9 U
o-Xylene	95-47-6	260		5.5 U	5.9 U	6 U	5.9 U
p-Isopropyltoluene	99-87-6			5.5 U	5.9 U	6 U	5.9 U
sec-Butylbenzene	135-98-8	11000		5.5 U	5.9 U	6 U	5.9 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-1B-B-1122011	PM-SB-2-A-1112011	PM-SB-2-B-1112011	PM-SB-3-A-1112011
			Sample Location	PM-SB-1B-B 11/02/2011	PM-SB-2-A 11/01/2011	PM-SB-2-B 11/01/2011	PM-SB-3-A 11/01/2011
Styrene	100-42-5			5.5 U	5.9 U	6 U	5.9 U
tert-Butylbenzene	98-06-6	5900		5.5 U	5.9 U	6 U	5.9 U
Tetrachloroethene	127-18-4	1300		5.5 U	5.9 J	9.2 J	22 J
Toluene	108-88-3	700		5.5 U	5.9 U	6 U	5.9 U
Total Xylenes	1330-20-7	260		5.5 U	5.9 U	6 U	5.9 U
trans-1,2-Dichloroethene	156-60-5	190		5.5 U	5.9 U	6 U	5.9 U
trans-1,3-Dichloropropene	10061-02-6			5.5 U	5.9 U	6 U	5.9 U
Trichloroethene	79-01-6	470		5.5 U	5.9 U	1.2 J	5.9 U
Trichlorofluoromethane	75-69-4			5.5 U	5.9 U	6 U	5.9 U
Vinyl Acetate	108-05-4			5.5 U	5.9 U	6 U	5.9 U
Vinyl Chloride	75-01-4	20		5.5 U	5.9 U	6 U	5.9 U
<b>Semi-Volatile Organic Compounds (µg/kg)</b>							
1,1'-Biphenyl	92-52-4						
2,4,5-Trichlorophenol	95-95-4						
2,4,6-Trichlorophenol	88-06-2						
2,4-Dichlorophenol	120-83-2						
2,4-Dimethylphenol	105-67-9						
2,4-Dinitrophenol	51-28-5						
2,4-Dinitrotoluene	121-14-2						
2,6-Dinitrotoluene	606-20-2						
2-Chloronaphthalene	91-58-7						
2-Chlorophenol	95-57-8						
2-Methylnaphthalene	91-57-6						
2-Methylphenol	95-48-7	330					
2-Nitroaniline	88-74-4						
2-Nitrophenol	88-75-5						
3,3'-Dichlorobenzidine	91-94-1						
3-Nitroaniline	99-09-2						
4,6-Dinitro-2-Methylphenol	534-52-1						
4-Bromophenyl-Phenylether	101-55-3						
4-Chloro-3-Methylphenol	59-50-7						

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-1B-B-1122011	PM-SB-2-A-1112011	PM-SB-2-B-1112011	PM-SB-3-A-1112011
			Sample Location	PM-SB-1B-B 11/02/2011	PM-SB-2-A 11/01/2011	PM-SB-2-B 11/01/2011	PM-SB-3-A 11/01/2011
4-Chloroaniline	106-47-8						
4-Chlorophenyl-Phenylether	7005-72-3						
4-Methylphenol	106-44-5	330					
4-Nitroaniline	100-01-6						
4-Nitrophenol	100-02-7						
Acenaphthene	83-32-9	20000					
Acenaphthylene	208-96-8	100000					
Acetophenone	98-86-2						
Anthracene	120-12-7	100000					
Atrazine	1912-24-9						
Benzaldehyde	100-52-7						
Benzo(a)Anthracene	56-55-3	1000					
Benzo(a)Pyrene	50-32-8	1000					
Benzo(b)Fluoranthene	205-99-2	1000					
Benzo(g,h,i)Perylene	191-24-2	100000					
Benzo(k)Fluoranthene	207-08-9	800					
bis(2-Chloroethoxy)Methane	111-91-1						
bis(2-Chloroethyl) Ether	111-44-4						
bis(2-Ethylhexyl)Phthalate	117-81-7						
bis-Chloroisopropyl Ether	108-60-1						
Butylbenzylphthalate	85-68-7						
Caprolactam	105-60-2						
Carbazole	86-74-8						
Chrysene	218-01-9	1000					
Dibenzo(a,h)Anthracene	53-70-3	330					
Dibenzofuran	132-64-9	7000					
Diethylphthalate	84-66-2						
Dimethylphthalate	131-11-3						
di-n-Butylphthalate	84-74-2						
di-n-Octylphthalate	117-84-0						
Fluoranthene	206-44-0	100000					

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Sample ID Sample Location Sampling Date	PM-SB-1B-B-1122011	PM-SB-2-A-1112011	PM-SB-2-B-1112011	PM-SB-3-A-1112011
			PM-SB-1B-B 11/02/2011	PM-SB-2-A 11/01/2011	PM-SB-2-B 11/01/2011	PM-SB-3-A 11/01/2011
Fluorene	86-73-7	Part 375 Unrestricted				
Hexachlorobenzene	118-74-1	30000				
Hexachlorobutadiene	87-68-3					
Hexachlorocyclopentadiene	77-47-4					
Hexachloroethane	67-72-1					
Indeno(1,2,3-cd)Pyrene	193-39-5	500				
Isophorone	78-59-1					
Naphthalene	91-20-3	12000				
Nitrobenzene	98-95-3					
n-Nitroso-di-n-Propylamine	621-64-7					
n-Nitrosodiphenylamine	86-30-6					
Pentachlorophenol	87-86-5	800				
Phenanthrene	85-01-8	100000				
Phenol	108-95-2	330				
Pyrene	129-00-0	100000				
<b>Pesticides (µg/kg)</b>						
4,4'-DDD	72-54-8	3.3				
4,4'-DDE	72-55-9	3.3				
4,4'-DDT	50-29-3	3.3				
Aldrin	309-00-2	5				
Alpha-BHC	319-84-6	20				
Alpha-Chlordane	5103-71-9	94				
Beta-BHC	319-85-7	36				
Delta-BHC	319-86-8	40				
Dieldrin	60-57-1	5				
Endosulfan I	959-98-8	2400				
Endosulfan II	33213-65-9	2400				
Endosulfan Sulfate	1031-07-8	2400				
Endrin	72-20-8	14				
Endrin Aldehyde	7421-93-4					
Endrin Ketone	53494-70-5					

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID	PM-SB-1B-B-1122011	PM-SB-2-A-1112011	PM-SB-3-A-1112011
Chemical Name	CAS#	Sample Location Sampling Date	PM-SB-1B-B 11/02/2011	PM-SB-2-A 11/01/2011	PM-SB-2-B 11/01/2011	PM-SB-3-A 11/01/2011
Gamma-BHC (Lindane)	58-89-9	100				
Gamma-Chlordane	5103-74-2					
Heptachlor	76-44-8	42				
Heptachlor Epoxide	1024-57-3					
Methoxychlor	72-43-5					
Toxaphene	8001-35-2					
<b>Polychlorinated Biphenyls (µg/kg)</b>						
Aroclor 1016	12674-11-2	100				
Aroclor 1221	11104-28-2	100				
Aroclor 1232	11141-16-5	100				
Aroclor 1242	53469-21-9	100				
Aroclor 1248	12672-29-6	100				
Aroclor 1254	11097-69-1	100				
Aroclor 1260	11096-82-5	100				
<b>Inorganics (mg/kg)</b>						
Aluminum	7429-90-5					
Antimony	7440-36-0					
Arsenic	7440-38-2	13				
Barium	7440-39-3	350				
Beryllium	7440-41-7	7.2				
Cadmium	7440-43-9	2.5				
Calcium	7440-70-2					
Chromium	7440-47-3	30				
Cobalt	7440-48-4					
Copper	7440-50-8	50				
Iron	7439-89-6					
Lead	7439-92-1	63				
Magnesium	7439-95-4					
Manganese	7439-96-5	1600				
Mercury	7439-97-6	0.18				
Nickel	7440-02-0	30				

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-1B-B-1122011	PM-SB-2-A-1112011	PM-SB-2-B-1112011	PM-SB-3-A-1112011
			Sample Location	PM-SB-1B-B 11/02/2011	PM-SB-2-A 11/01/2011	PM-SB-2-B 11/01/2011	PM-SB-3-A 11/01/2011
Potassium	7440-09-7						
Selenium	7782-49-2	3.9					
Silver	7440-22-4	2					
Sodium	7440-23-5						
Thallium	7440-28-0						
Vanadium	7440-62-2						
Zinc	7440-66-6	109					

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

		Sample ID Sample Location Sampling Date	PM-SB-4-A-1112011 PM-SB-4-A 11/01/2011	PM-SB-5-A-1112011 PM-SB-5-A 11/01/2011	PM-SB-5-B-1112011 PM-SB-5-B 11/01/2011	PM-SB-6-A-1112011 PM-SB-6-A 11/01/2011
Chemical Name	CAS#	Part 375 Unrestricted				
<b>Volatile Organic Compounds (µg/kg)</b>						
1,1,1,2-Tetrachloroethane	630-20-6		5.9 U	6.8 U	5.4 U	6.7 U
1,1,1-Trichloroethane	71-55-6	680	5.9 U	6.8 U	5.4 U	6.7 U
1,1,2,2-Tetrachloroethane	79-34-5		5.9 U	6.8 U	5.4 U	6.7 U
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5.9 U	6.8 U	5.4 U	6.7 U
1,1,2-Trichloroethane	79-00-5		5.9 U	6.8 U	5.4 U	6.7 U
1,1-Dichloroethane	75-34-3	270	5.9 U	6.8 U	5.4 U	6.7 U
1,1-Dichloroethene	75-35-4	330	5.9 U	6.8 U	5.4 U	6.7 U
1,1-Dichloropropene	563-58-6		5.9 U	6.8 U	5.4 U	6.7 U
1,2,3-Trichlorobenzene	87-61-6		5.9 U	6.8 U	5.4 U	6.7 U
1,2,3-Trichloropropane	96-18-4		5.9 U	6.8 U	5.4 U	6.7 U
1,2,4-Trichlorobenzene	120-82-1		5.9 UJ	6.8 UJ	5.4 UJ	6.7 UJ
1,2,4-Trimethylbenzene	95-63-6	3600	1.8 J	6.8 U	5.4 U	6.7 U
1,2-Dibromo-3-Chloropropane	96-12-8		5.9 UJ	6.8 UJ	5.4 UJ	6.7 UJ
1,2-Dibromoethane (EDB)	106-93-4		5.9 U	6.8 U	5.4 U	6.7 U
1,2-Dichlorobenzene	95-50-1	1100	5.9 UJ	6.8 UJ	5.4 UJ	6.7 UJ
1,2-Dichloroethane	107-06-2	20	5.9 U	6.8 U	5.4 U	6.7 U
1,2-Dichloropropane	78-87-5		5.9 U	6.8 U	5.4 U	6.7 U
1,3,5-Trimethylbenzene	108-67-8	8400	5.9 U	6.8 U	5.4 U	6.7 U
1,3-Dichlorobenzene	541-73-1	2400	5.9 UJ	6.8 UJ	5.4 UJ	6.7 UJ
1,3-Dichloropropane	142-28-9		5.9 U	6.8 U	5.4 U	6.7 U
1,4-Dichlorobenzene	106-46-7	1800	5.9 UJ	6.8 UJ	5.4 UJ	6.7 UJ
2,2-Dichloropropane	594-20-7		5.9 U	6.8 U	5.4 U	6.7 U
2-Butanone (MEK)	78-93-3	120	5.9 R	6.8 R	5.4 R	6.7 R
2-Chlorotoluene	95-49-8		5.9 U	6.8 U	5.4 U	6.7 U
2-Hexanone	591-78-6		5.9 UJ	6.8 UJ	5.4 UJ	6.7 UJ
4-Chlorotoluene	106-43-4		5.9 U	6.8 U	5.4 U	6.7 U
4-Methyl-2-Pentanone (MIBK)	108-10-1		5.9 U	6.8 U	5.4 U	6.7 U
Acetone	67-64-1	50	14 J	6.8 R	5.4 R	6.7 R
Benzene	71-43-2	60	5.9 U	6.8 U	5.4 U	6.7 U
Bromobenzene	108-86-1		5.9 U	6.8 U	5.4 U	6.7 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-4-A-1112011	PM-SB-5-A-1112011	PM-SB-5-B-1112011	PM-SB-6-A-1112011
			Sample Location	PM-SB-4-A 11/01/2011	PM-SB-5-A 11/01/2011	PM-SB-5-B 11/01/2011	PM-SB-6-A 11/01/2011
Bromochloromethane	74-97-5			5.9 U	6.8 U	5.4 U	6.7 U
Bromodichloromethane	75-27-4			5.9 U	6.8 U	5.4 U	6.7 U
Bromoform	75-25-2			5.9 U	6.8 U	5.4 U	6.7 U
Bromomethane	74-83-9			5.9 U	6.8 U	5.4 U	6.7 U
Carbon Disulfide	75-15-0			5.9 U	6.8 U	5.4 U	6.7 U
Carbon Tetrachloride	56-23-5	760		5.9 UJ	6.8 U	5.4 UJ	6.7 UJ
Chlorobenzene	108-90-7	1100		5.9 U	6.8 U	5.4 U	6.7 U
Chloroethane	75-00-3			5.9 U	6.8 U	5.4 U	6.7 U
Chloroform	67-66-3	370		5.9 U	6.8 U	5.4 U	6.7 U
Chloromethane	74-87-3			5.9 U	6.8 U	5.4 U	6.7 U
cis-1,2-Dichloroethene	156-59-2	250		5.9 U	12	5.4 U	12
cis-1,3-Dichloropropene	10061-01-5			5.9 U	6.8 U	5.4 U	6.7 U
Cyclohexane	110-82-7			5.9 U	6.8 U	5.4 U	6.7 U
Dibromochloromethane	124-48-1			5.9 U	6.8 U	5.4 U	6.7 U
Dibromomethane	74-95-3			5.9 U	6.8 U	5.4 U	6.7 U
Dichlorodifluoromethane	75-71-8			5.9 U	6.8 U	5.4 U	6.7 U
Ethylbenzene	100-41-4	1000		5.9 U	6.8 U	5.4 U	6.7 U
Hexachlorobutadiene	87-68-3			5.9 U	6.8 U	5.4 U	6.7 U
Isopropylbenzene	98-82-8			5.9 U	6.8 U	5.4 U	6.7 U
m,p-Xylene	179601-23-1	260		5.9 U	6.8 U	5.4 U	6.7 U
Methyl Acetate	79-20-9			5.9 U	6.8 U	5.4 U	6.7 U
Methyl Iodide	74-88-4			5.9 UJ	6.8 U	5.4 UJ	6.7 UJ
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	930		5.9 U	6.8 U	5.4 U	6.7 U
Methylcyclohexane	108-87-2			5.9 U	6.8 UJ	5.4 U	6.7 U
Methylene Chloride	75-09-2	50		5 J	6.8 U	3.2 J	3.8 J
Naphthalene	91-20-3	12000		5.8 J	6.8 U	5.4 UJ	6.7 UJ
n-Butylbenzene	104-51-8	12000		5.9 U	6.8 U	5.4 U	6.7 U
n-Propylbenzene	103-65-1	3900		5.9 U	6.8 U	5.4 U	6.7 U
o-Xylene	95-47-6	260		5.9 U	6.8 U	5.4 U	6.7 U
p-Isopropyltoluene	99-87-6			5.9 U	6.8 U	5.4 U	6.7 U
sec-Butylbenzene	135-98-8	11000		5.9 U	6.8 U	5.4 U	6.7 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-4-A-1112011	PM-SB-5-A-1112011	PM-SB-5-B-1112011	PM-SB-6-A-1112011
			Sample Location	PM-SB-4-A 11/01/2011	PM-SB-5-A 11/01/2011	PM-SB-5-B 11/01/2011	PM-SB-6-A 11/01/2011
Styrene	100-42-5			5.9 U	6.8 U	5.4 U	6.7 U
tert-Butylbenzene	98-06-6	5900		5.9 U	6.8 U	5.4 U	6.7 U
Tetrachloroethene	127-18-4	1300		16 J	110 J	4.5 J	26 J
Toluene	108-88-3	700		3.8 J	2.3 J	1.9 J	6.7 U
Total Xylenes	1330-20-7	260		5.9 U	6.8 U	5.4 U	6.7 U
trans-1,2-Dichloroethene	156-60-5	190		5.9 U	6.8 U	5.4 U	6.7 U
trans-1,3-Dichloropropene	10061-02-6			5.9 U	6.8 U	5.4 U	6.7 U
Trichloroethene	79-01-6	470		5.9 U	16 J	5.4 U	6.6 J
Trichlorofluoromethane	75-69-4			5.9 U	6.8 U	5.4 U	6.7 U
Vinyl Acetate	108-05-4			5.9 U	6.8 U	5.4 U	6.7 U
Vinyl Chloride	75-01-4	20		5.9 U	6.8 U	5.4 U	6.7 U
<b>Semi-Volatile Organic Compounds (µg/kg)</b>							
1,1'-Biphenyl	92-52-4						
2,4,5-Trichlorophenol	95-95-4						
2,4,6-Trichlorophenol	88-06-2						
2,4-Dichlorophenol	120-83-2						
2,4-Dimethylphenol	105-67-9						
2,4-Dinitrophenol	51-28-5						
2,4-Dinitrotoluene	121-14-2						
2,6-Dinitrotoluene	606-20-2						
2-Chloronaphthalene	91-58-7						
2-Chlorophenol	95-57-8						
2-Methylnaphthalene	91-57-6						
2-Methylphenol	95-48-7	330					
2-Nitroaniline	88-74-4						
2-Nitrophenol	88-75-5						
3,3'-Dichlorobenzidine	91-94-1						
3-Nitroaniline	99-09-2						
4,6-Dinitro-2-Methylphenol	534-52-1						
4-Bromophenyl-Phenylether	101-55-3						
4-Chloro-3-Methylphenol	59-50-7						

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-4-A-1112011	PM-SB-5-A-1112011	PM-SB-5-B-1112011	PM-SB-6-A-1112011
			Sample Location	PM-SB-4-A 11/01/2011	PM-SB-5-A 11/01/2011	PM-SB-5-B 11/01/2011	PM-SB-6-A 11/01/2011
4-Chloroaniline	106-47-8						
4-Chlorophenyl-Phenylether	7005-72-3						
4-Methylphenol	106-44-5	330					
4-Nitroaniline	100-01-6						
4-Nitrophenol	100-02-7						
Acenaphthene	83-32-9	20000					
Acenaphthylene	208-96-8	100000					
Acetophenone	98-86-2						
Anthracene	120-12-7	100000					
Atrazine	1912-24-9						
Benzaldehyde	100-52-7						
Benzo(a)Anthracene	56-55-3	1000					
Benzo(a)Pyrene	50-32-8	1000					
Benzo(b)Fluoranthene	205-99-2	1000					
Benzo(g,h,i)Perylene	191-24-2	100000					
Benzo(k)Fluoranthene	207-08-9	800					
bis(2-Chloroethoxy)Methane	111-91-1						
bis(2-Chloroethyl) Ether	111-44-4						
bis(2-Ethylhexyl)Phthalate	117-81-7						
bis-Chloroisopropyl Ether	108-60-1						
Butylbenzylphthalate	85-68-7						
Caprolactam	105-60-2						
Carbazole	86-74-8						
Chrysene	218-01-9	1000					
Dibenzo(a,h)Anthracene	53-70-3	330					
Dibenzofuran	132-64-9	7000					
Diethylphthalate	84-66-2						
Dimethylphthalate	131-11-3						
di-n-Butylphthalate	84-74-2						
di-n-Octylphthalate	117-84-0						
Fluoranthene	206-44-0	100000					

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Sample ID Sample Location Sampling Date	PM-SB-4-A-1112011	PM-SB-5-A-1112011	PM-SB-5-B-1112011	PM-SB-6-A-1112011
			PM-SB-4-A 11/01/2011	PM-SB-5-A 11/01/2011	PM-SB-5-B 11/01/2011	PM-SB-6-A 11/01/2011
Fluorene	86-73-7	Part 375 Unrestricted				
Hexachlorobenzene	118-74-1	30000				
Hexachlorobutadiene	87-68-3					
Hexachlorocyclopentadiene	77-47-4					
Hexachloroethane	67-72-1					
Indeno(1,2,3-cd)Pyrene	193-39-5	500				
Isophorone	78-59-1					
Naphthalene	91-20-3	12000				
Nitrobenzene	98-95-3					
n-Nitroso-di-n-Propylamine	621-64-7					
n-Nitrosodiphenylamine	86-30-6					
Pentachlorophenol	87-86-5	800				
Phenanthrene	85-01-8	100000				
Phenol	108-95-2	330				
Pyrene	129-00-0	100000				
<b>Pesticides (µg/kg)</b>						
4,4'-DDD	72-54-8	3.3				
4,4'-DDE	72-55-9	3.3				
4,4'-DDT	50-29-3	3.3				
Aldrin	309-00-2	5				
Alpha-BHC	319-84-6	20				
Alpha-Chlordane	5103-71-9	94				
Beta-BHC	319-85-7	36				
Delta-BHC	319-86-8	40				
Dieldrin	60-57-1	5				
Endosulfan I	959-98-8	2400				
Endosulfan II	33213-65-9	2400				
Endosulfan Sulfate	1031-07-8	2400				
Endrin	72-20-8	14				
Endrin Aldehyde	7421-93-4					
Endrin Ketone	53494-70-5					

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

		Sample ID	PM-SB-4-A-1112011	PM-SB-5-A-1112011	PM-SB-5-B-1112011	PM-SB-6-A-1112011
Chemical Name	CAS#	Sample Location Sampling Date	PM-SB-4-A 11/01/2011	PM-SB-5-A 11/01/2011	PM-SB-5-B 11/01/2011	PM-SB-6-A 11/01/2011
Gamma-BHC (Lindane)	58-89-9	100				
Gamma-Chlordane	5103-74-2					
Heptachlor	76-44-8	42				
Heptachlor Epoxide	1024-57-3					
Methoxychlor	72-43-5					
Toxaphene	8001-35-2					
<b>Polychlorinated Biphenyls (µg/kg)</b>						
Aroclor 1016	12674-11-2	100				
Aroclor 1221	11104-28-2	100				
Aroclor 1232	11141-16-5	100				
Aroclor 1242	53469-21-9	100				
Aroclor 1248	12672-29-6	100				
Aroclor 1254	11097-69-1	100				
Aroclor 1260	11096-82-5	100				
<b>Inorganics (mg/kg)</b>						
Aluminum	7429-90-5					
Antimony	7440-36-0					
Arsenic	7440-38-2	13				
Barium	7440-39-3	350				
Beryllium	7440-41-7	7.2				
Cadmium	7440-43-9	2.5				
Calcium	7440-70-2					
Chromium	7440-47-3	30				
Cobalt	7440-48-4					
Copper	7440-50-8	50				
Iron	7439-89-6					
Lead	7439-92-1	63				
Magnesium	7439-95-4					
Manganese	7439-96-5	1600				
Mercury	7439-97-6	0.18				
Nickel	7440-02-0	30				

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-4-A-1112011	PM-SB-5-A-1112011	PM-SB-5-B-1112011	PM-SB-6-A-1112011
			Sample Location	PM-SB-4-A 11/01/2011	PM-SB-5-A 11/01/2011	PM-SB-5-B 11/01/2011	PM-SB-6-A 11/01/2011
Potassium	7440-09-7						
Selenium	7782-49-2	3.9					
Silver	7440-22-4	2					
Sodium	7440-23-5						
Thallium	7440-28-0						
Vanadium	7440-62-2						
Zinc	7440-66-6	109					

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

		Sample ID	PM-SB-6-B-1112011	PM-SB-7-B-1122011	PM-SB-99-110211
Chemical Name	CAS#	Sample Location Sampling Date	PM-SB-6-B 11/01/2011	PM-SB-7-B 11/02/2011	PM-SB-99 11/02/2011
<b>Volatile Organic Compounds (µg/kg)</b>					
1,1,1,2-Tetrachloroethane	630-20-6		5.7 U	5.2 U	6 U
1,1,1-Trichloroethane	71-55-6	680	5.7 U	5.2 U	6 U
1,1,2,2-Tetrachloroethane	79-34-5		5.7 U	5.2 U	6 U
1,1,2-Trichloro-1,2,2-Trifluoroethane	76-13-1		5.7 U	5.2 U	6 U
1,1,2-Trichloroethane	79-00-5		5.7 U	5.2 U	6 U
1,1-Dichloroethane	75-34-3	270	5.7 U	5.2 U	6 U
1,1-Dichloroethene	75-35-4	330	5.7 U	5.2 U	6 U
1,1-Dichloropropene	563-58-6		5.7 U	5.2 U	6 U
1,2,3-Trichlorobenzene	87-61-6		5.7 U	5.2 U	6 U
1,2,3-Trichloropropane	96-18-4		5.7 U	5.2 U	6 U
1,2,4-Trichlorobenzene	120-82-1		5.7 UJ	5.2 UJ	6 U
1,2,4-Trimethylbenzene	95-63-6	3600	5.7 U	5.2 U	6 U
1,2-Dibromo-3-Chloropropane	96-12-8		5.7 UJ	5.2 U	6 U
1,2-Dibromoethane (EDB)	106-93-4		5.7 U	5.2 U	6 U
1,2-Dichlorobenzene	95-50-1	1100	5.7 UJ	5.2 U	6 U
1,2-Dichloroethane	107-06-2	20	5.7 U	5.2 U	6 U
1,2-Dichloropropane	78-87-5		5.7 U	5.2 U	6 U
1,3,5-Trimethylbenzene	108-67-8	8400	5.7 U	5.2 U	6 U
1,3-Dichlorobenzene	541-73-1	2400	5.7 UJ	5.2 U	6 U
1,3-Dichloropropane	142-28-9		5.7 U	5.2 U	6 U
1,4-Dichlorobenzene	106-46-7	1800	5.7 UJ	5.2 U	6 U
2,2-Dichloropropane	594-20-7		5.7 U	5.2 U	6 U
2-Butanone (MEK)	78-93-3	120	5.7 R	5.2 R	6 R
2-Chlorotoluene	95-49-8		5.7 U	5.2 U	6 U
2-Hexanone	591-78-6		5.7 UJ	5.2 U	6 U
4-Chlorotoluene	106-43-4		5.7 U	5.2 U	6 U
4-Methyl-2-Pentanone (MIBK)	108-10-1		5.7 U	5.2 U	6 U
Acetone	67-64-1	50	5.7 R	5.2 R	9 R
Benzene	71-43-2	60	5.7 U	5.2 U	6 U
Bromobenzene	108-86-1		5.7 U	5.2 U	6 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-6-B-1112011	PM-SB-7-B-1122011	PM-SB-99-110211
			Sample Location	PM-SB-6-B 11/01/2011	PM-SB-7-B 11/02/2011	PM-SB-99 11/02/2011
Bromochloromethane	74-97-5			5.7 U	5.2 U	6 U
Bromodichloromethane	75-27-4			5.7 U	5.2 U	6 U
Bromoform	75-25-2			5.7 U	5.2 U	6 U
Bromomethane	74-83-9			5.7 U	5.2 UJ	6 U
Carbon Disulfide	75-15-0			5.7 U	5.2 U	6 U
Carbon Tetrachloride	56-23-5	760		5.7 UJ	5.2 UJ	6 U
Chlorobenzene	108-90-7	1100		5.7 U	5.2 U	6 U
Chloroethane	75-00-3			5.7 U	5.2 U	6 U
Chloroform	67-66-3	370		5.7 U	5.2 U	6 U
Chloromethane	74-87-3			5.7 U	5.2 UJ	6 U
cis-1,2-Dichloroethene	156-59-2	250		5.7 U	5.2 U	6 U
cis-1,3-Dichloropropene	10061-01-5			5.7 U	5.2 UJ	6 U
Cyclohexane	110-82-7			5.7 U	5.2 U	6 U
Dibromochloromethane	124-48-1			5.7 U	5.2 U	6 U
Dibromomethane	74-95-3			5.7 U	5.2 U	6 U
Dichlorodifluoromethane	75-71-8			5.7 U	5.2 UJ	6 U
Ethylbenzene	100-41-4	1000		5.7 U	5.2 U	6 U
Hexachlorobutadiene	87-68-3			5.7 U	5.2 U	6 U
Isopropylbenzene	98-82-8			5.7 U	5.2 UJ	6 U
m,p-Xylene	179601-23-1	260		5.7 U	5.2 U	6 U
Methyl Acetate	79-20-9			5.7 U	5.2 U	6 U
Methyl Iodide	74-88-4			5.7 UJ	5.2 U	6 UJ
Methyl Tert-Butyl Ether (MTBE)	1634-04-4	930		5.7 U	5.2 U	6 U
Methylcyclohexane	108-87-2			5.7 U	5.2 UJ	6 U
Methylene Chloride	75-09-2	50		3 J	2.9 J	6 U
Naphthalene	91-20-3	12000		5.7 UJ	5.2 UJ	6 U
n-Butylbenzene	104-51-8	12000		5.7 U	5.2 U	6 U
n-Propylbenzene	103-65-1	3900		5.7 U	5.2 U	6 U
o-Xylene	95-47-6	260		5.7 U	5.2 U	6 U
p-Isopropyltoluene	99-87-6			5.7 U	5.2 U	6 U
sec-Butylbenzene	135-98-8	11000		5.7 U	5.2 U	6 U

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-6-B-1112011	PM-SB-7-B-1122011	PM-SB-99-110211
			Sample Location	PM-SB-6-B 11/01/2011	PM-SB-7-B 11/02/2011	PM-SB-99 11/02/2011
Styrene	100-42-5			5.7 U	5.2 U	6 U
tert-Butylbenzene	98-06-6	5900		5.7 U	5.2 U	6 U
Tetrachloroethene	127-18-4	1300		5.7 U	5.2 U	2.3 J
Toluene	108-88-3	700		5.7 U	5.2 U	5.6 U
Total Xylenes	1330-20-7	260		5.7 U	5.2 U	6 U
trans-1,2-Dichloroethene	156-60-5	190		5.7 U	5.2 U	6 U
trans-1,3-Dichloropropene	10061-02-6			5.7 U	5.2 U	6 U
Trichloroethene	79-01-6	470		5.7 U	5.2 U	6 U
Trichlorofluoromethane	75-69-4			5.7 U	5.2 U	6 U
Vinyl Acetate	108-05-4			5.7 U	5.2 U	6 U
Vinyl Chloride	75-01-4	20		5.7 U	5.2 U	6 U
<b>Semi-Volatile Organic Compounds (µg/kg)</b>						
1,1'-Biphenyl	92-52-4					
2,4,5-Trichlorophenol	95-95-4					
2,4,6-Trichlorophenol	88-06-2					
2,4-Dichlorophenol	120-83-2					
2,4-Dimethylphenol	105-67-9					
2,4-Dinitrophenol	51-28-5					
2,4-Dinitrotoluene	121-14-2					
2,6-Dinitrotoluene	606-20-2					
2-Chloronaphthalene	91-58-7					
2-Chlorophenol	95-57-8					
2-Methylnaphthalene	91-57-6					
2-Methylphenol	95-48-7	330				
2-Nitroaniline	88-74-4					
2-Nitrophenol	88-75-5					
3,3'-Dichlorobenzidine	91-94-1					
3-Nitroaniline	99-09-2					
4,6-Dinitro-2-Methylphenol	534-52-1					
4-Bromophenyl-Phenylether	101-55-3					
4-Chloro-3-Methylphenol	59-50-7					

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Sample ID Sample Location Sampling Date	PM-SB-6-B-1112011	PM-SB-7-B-1122011	PM-SB-99-110211
			PM-SB-6-B 11/01/2011	PM-SB-7-B 11/02/2011	PM-SB-99 11/02/2011
4-Chloroaniline	106-47-8				
4-Chlorophenyl-Phenylether	7005-72-3				
4-Methylphenol	106-44-5	330			
4-Nitroaniline	100-01-6				
4-Nitrophenol	100-02-7				
Acenaphthene	83-32-9	20000			
Acenaphthylene	208-96-8	100000			
Acetophenone	98-86-2				
Anthracene	120-12-7	100000			
Atrazine	1912-24-9				
Benzaldehyde	100-52-7				
Benzo(a)Anthracene	56-55-3	1000			
Benzo(a)Pyrene	50-32-8	1000			
Benzo(b)Fluoranthene	205-99-2	1000			
Benzo(g,h,i)Perylene	191-24-2	100000			
Benzo(k)Fluoranthene	207-08-9	800			
bis(2-Chloroethoxy)Methane	111-91-1				
bis(2-Chloroethyl) Ether	111-44-4				
bis(2-Ethylhexyl)Phthalate	117-81-7				
bis-Chloroisopropyl Ether	108-60-1				
Butylbenzylphthalate	85-68-7				
Caprolactam	105-60-2				
Carbazole	86-74-8				
Chrysene	218-01-9	1000			
Dibenzo(a,h)Anthracene	53-70-3	330			
Dibenzofuran	132-64-9	7000			
Diethylphthalate	84-66-2				
Dimethylphthalate	131-11-3				
di-n-Butylphthalate	84-74-2				
di-n-Octylphthalate	117-84-0				
Fluoranthene	206-44-0	100000			

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Sample ID Sample Location Sampling Date	PM-SB-6-B-1112011	PM-SB-7-B-1122011	PM-SB-99-110211
			PM-SB-6-B 11/01/2011	PM-SB-7-B 11/02/2011	PM-SB-99 11/02/2011
Fluorene	86-73-7	Part 375 Unrestricted			
Hexachlorobenzene	118-74-1	30000			
Hexachlorobutadiene	87-68-3				
Hexachlorocyclopentadiene	77-47-4				
Hexachloroethane	67-72-1				
Indeno(1,2,3-cd)Pyrene	193-39-5	500			
Isophorone	78-59-1				
Naphthalene	91-20-3	12000			
Nitrobenzene	98-95-3				
n-Nitroso-di-n-Propylamine	621-64-7				
n-Nitrosodiphenylamine	86-30-6				
Pentachlorophenol	87-86-5	800			
Phenanthrene	85-01-8	100000			
Phenol	108-95-2	330			
Pyrene	129-00-0	100000			
<b>Pesticides (µg/kg)</b>					
4,4'-DDD	72-54-8	3.3			
4,4'-DDE	72-55-9	3.3			
4,4'-DDT	50-29-3	3.3			
Aldrin	309-00-2	5			
Alpha-BHC	319-84-6	20			
Alpha-Chlordane	5103-71-9	94			
Beta-BHC	319-85-7	36			
Delta-BHC	319-86-8	40			
Dieldrin	60-57-1	5			
Endosulfan I	959-98-8	2400			
Endosulfan II	33213-65-9	2400			
Endosulfan Sulfate	1031-07-8	2400			
Endrin	72-20-8	14			
Endrin Aldehyde	7421-93-4				
Endrin Ketone	53494-70-5				

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

			Sample ID	PM-SB-6-B-1112011	PM-SB-7-B-1122011	PM-SB-99-110211
Chemical Name	CAS#	Sample Location	PM-SB-6-B Sampling Date 11/01/2011	PM-SB-7-B 11/02/2011	PM-SB-99 11/02/2011	
Gamma-BHC (Lindane)	58-89-9	100				
Gamma-Chlordane	5103-74-2					
Heptachlor	76-44-8	42				
Heptachlor Epoxide	1024-57-3					
Methoxychlor	72-43-5					
Toxaphene	8001-35-2					
<b>Polychlorinated Biphenyls (µg/kg)</b>						
Aroclor 1016	12674-11-2	100				
Aroclor 1221	11104-28-2	100				
Aroclor 1232	11141-16-5	100				
Aroclor 1242	53469-21-9	100				
Aroclor 1248	12672-29-6	100				
Aroclor 1254	11097-69-1	100				
Aroclor 1260	11096-82-5	100				
<b>Inorganics (mg/kg)</b>						
Aluminum	7429-90-5					
Antimony	7440-36-0					
Arsenic	7440-38-2	13				
Barium	7440-39-3	350				
Beryllium	7440-41-7	7.2				
Cadmium	7440-43-9	2.5				
Calcium	7440-70-2					
Chromium	7440-47-3	30				
Cobalt	7440-48-4					
Copper	7440-50-8	50				
Iron	7439-89-6					
Lead	7439-92-1	63				
Magnesium	7439-95-4					
Manganese	7439-96-5	1600				
Mercury	7439-97-6	0.18				
Nickel	7440-02-0	30				

**Appendix C-2**  
**Soil Sample Results**  
**Former Paul Miller Dry Cleaners Site**  
**Port Richmond, Richmond County, New York**

Chemical Name	CAS#	Part 375 Unrestricted	Sample ID	PM-SB-6-B-1112011	PM-SB-7-B-1122011	PM-SB-99-110211
			Sample Location	PM-SB-6-B 11/01/2011	PM-SB-7-B 11/02/2011	PM-SB-99 11/02/2011
Potassium	7440-09-7					
Selenium	7782-49-2	3.9				
Silver	7440-22-4	2				
Sodium	7440-23-5					
Thallium	7440-28-0					
Vanadium	7440-62-2					
Zinc	7440-66-6	109				

## Appendix D

### Data Usability Reports

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Total Metals Analyses**

**Samples Collected: May 23<sup>rd</sup> & 24<sup>th</sup>, 2011**

**Samples Received: May 25, 2011**

**Laboratory: Spectrum**

**Sample Delivery Group: K0909**

**Laboratory Reference Numbers:**

Lab ID	Client ID	Matrix	Date Sampled
K0909-05	80230-GW-MIP6-33.5	Aqueous	23-May-11
K0909-07	80230-GW-MIP6-15	Aqueous	23-May-11
K0909-08	80230-FB-110523	Aqueous	23-May-11
K0909-09	80230-FB-AQ-110523	Aqueous	23-May-11
K0909-10	80230-GW-MIP5-28	Aqueous	24-May-11
K0909-10 MS	80230-GW-MIP5-28 MS	Aqueous	24-May-11
K0909-10 MSD	80230-GW-MIP5-28 MSD	Aqueous	24-May-11
K0909-12	80230-FB-AQ-110524	Aqueous	24-May-11

Water samples were validated for inorganic analyses by the US EPA Region II data validation SOP (HW-2, Revision 13). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - Holding Times
- \* - Calibration Verification
  - CRDL Standard
  - \* - Laboratory Control Sample
  - Serial Dilutions
- \* - Calibration Blanks
  - Field Blank
  - \* - Preparation Blanks
  - \* - Matrix Spike
  - \* - Matrix Duplicate
  - \* - ICP Interference Check Sample
  - \* - Detection Limit Results
  - \* - Linear Range
  - \* - Sample Results

\* - Indicates that all criteria were met for this parameter.

**Data Validation Summary**

A CRDL standard was not analyzed with this sample delivery group.

The minor problem with the serial dilution should be noted.

No other problems were found that would affect the use of the data.

## Holding Times

All samples were analyzed within the required holding times.

## CRDL Standards

A CRDL standard was not analyzed with this sample delivery group.

## Initial and Continuing Calibrations

No problems were detected with any of the calibrations associated with this sample delivery group.

## Preparation Blank

No compounds were detected in the one preparation blank associated with the digestions of these samples at concentrations above the CRDL. Several analytes were found in the preparation blank at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table.

## Calibration Blanks

Several analytes were found in the continuing calibration blanks at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table and do not affect the end use of the data.

## Field Blank

Several analytes were found in the continuing field blanks at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table and do not affect the end use of the data.

## ICP Interference Check Sample

No problems were detected with the reported ICP Interference Check Sample recoveries.

## Matrix Spike Recovery

Sample K0909-10 / 80230-GW-MIP5-28 was used as the matrix spike. All recoveries were within the 75% - 125% quality control limits.

## Duplicate Analysis

Sample K0909-10 / 80230-GW-MIP5-28 was used as the matrix duplicate. All RPDs that could be accurately calculated were less than 20%.

### **Laboratory Control Sample**

No problems were detected with the recoveries of the LCS standards.

### **Serial Dilutions**

Sample K0909-10 / 80230-GW-MIP5-28 was used as the serial dilution. All percent differences that could be accurately calculated were less than 10% with the exception of barium (11%).

The barium data were flagged with the "J" qualifier and are estimated values.

### **Instrument Detection Limit**

No problems were found with the instrument detection limits.

### **ICP Linear Ranges**

No problems were detected with the linear ranges.

### **Sample Results**

No problems were detected with any of these samples.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Water Pesticide Analyses**

**Samples Collected: May 23<sup>rd</sup> & 24<sup>th</sup>, 2011**

**Samples Received: May 25, 2011**

**Laboratory: Spectrum**

**Sample Delivery Group: K0909**

**Laboratory Reference Numbers:**

Lab ID	Client ID	Matrix	Date Sampled
K0909-08	80230-FB-110523	Aqueous	23-May-11

Soil and water samples were validated for analyses of pesticides by the US EPA Region II data validation SOP (HW-44, Revision 1). Data were reviewed for usability according to the following criteria:

- Data Completeness
- \* - Holding Times
- \* - Laboratory Blanks
- \* - Field Blanks
- \* - Surrogate Recoveries
- \* - Surrogate Retention Times
  - Matrix Spike / Matrix Spike Duplicate
  - \* - Surrogate Spike Recovery
  - \* - Laboratory Control Sample
- \* - Calibrations
- \* - Method Blanks
  - Florisil Cartridge Check
  - GPC Calibration
- \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

Form IX for a florisil cleanup was not included in the data package.

No significant problems were detected with any of the data.

**Holding Times**

All extractions and analyses were performed within the required holding times.

**Surrogate Recoveries**

All surrogate recoveries were within the required limits.

### **Matrix Spike**

A matrix spike was not analyzed with this sample delivery group.

### **Laboratory Control Samples**

All of the laboratory control sample recoveries were within the required limits.

### **Initial Calibrations**

No problems were detected with the initial calibrations associated with the analyses of the samples. All % RSDs were less than 20%.

### **Continuing Calibrations**

All %Ds in the continuing calibrations directly with the sample were less than 20%.

### **Florisil Cartridge Check**

Form IX for a florisil cleanup was not included in the data package.

### **GPC Calibration**

A GPC cleanup was not performed on these samples.

### **Method Blanks**

No problems were detected with any of the method blanks.

### **Calibration Blanks**

No problems were detected with the calibration blanks associated with this sample delivery group.

### **Field Blank**

No compounds were detected in the field blank.

### **Sample Results**

No problems were detected with the sample data.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Water PCB Analyses**

**Samples Collected: May 23<sup>rd</sup> & 24<sup>th</sup>, 2011**

**Samples Received: May 25, 2011**

**Laboratory: Spectrum**

**Sample Delivery Group: K0909**

**Laboratory Reference Numbers:**

Lab ID	Client ID	Matrix	Date Sampled
K0909-08	80230-FB-110523	Aqueous	23-May-11

One water sample was validated for analyses of PCBs by the US EPA Region II data validation SOP (HW-44, Revision 1). Data were reviewed for usability according to the following criteria:

- Data Completeness
- \* - Holding Times
- \* - Laboratory Blanks
- \* - Field Blanks
- \* - Surrogate Recoveries
- \* - Surrogate Retention Times
- Matrix Spike / Matrix Spike Duplicate
- \* - System Monitoring Spike Recovery
- \* - Laboratory Control Sample
- \* - Calibrations
- \* - Method Blanks
- GPC Calibration
- \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

Only compounds noted as target compounds by the laboratory's data system were reported in the raw data.

No other significant problems were detected with any of the data.

**Holding Times**

All extractions and analyses were performed within the required holding times.

**Surrogate Recoveries**

The recovery of the DCB surrogate on the primary column (40%) was at the 40% quality control limit. The data were not required to be qualified.

### **Matrix Spike**

A matrix spike was not analyzed with this sample delivery group.

### **Laboratory Control Samples**

All of the laboratory control sample recoveries were within the required limits.

### **Initial Calibrations**

No problems were detected with the initial calibrations associated with the analyses of the samples. All %RSDs were less than 20%.

### **Continuing Calibrations**

All %Ds were less than 15%.

### **Florisil Cartridge Check**

Form IX for a florisil cleanup was not included in the data package.

### **GPC Calibration**

A GPC cleanup was not performed on these samples.

### **Method Blanks**

No problems were detected with any of the method blanks.

### **Calibration Blanks**

No problems were detected with the calibration blanks associated with this sample delivery group.

### **Field Blank**

No compounds were detected in the field blank.

### **Sample Results**

No problems were detected with the sample data.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Semivolatile Organic Analyses**

**Samples Collected: May 23<sup>rd</sup> & 24<sup>th</sup>, 2011**

**Samples Received: May 25, 2011**

**Laboratory: Spectrum**

**Sample Delivery Group: K0909**

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0909-01	80230-MIP6-30	Soil	23-May-11
K0909-02	80230-MIP7-38	Soil	23-May-11
K0909-03	80230-DUP1-50	Soil	23-May-11
K0909-04	80230-MIP6-10	Soil	23-May-11
K0909-05	80230-GW-MIP6-33.5	Aqueous	23-May-11
K0909-07	80230-GW-MIP6-15	Aqueous	23-May-11
K0909-08	80230-FB-110523	Aqueous	23-May-11
K0909-09	80230-FB-AQ-110523	Aqueous	23-May-11
K0909-10	80230-GW-MIP5-28	Aqueous	24-May-11
K0909-10 MS	80230-GW-MIP5-28 MS	Aqueous	24-May-11
K0909-10 MSD	80230-GW-MIP5-28 MSD	Aqueous	24-May-11
K0909-12	80230-FB-AQ-110524	Aqueous	24-May-11

Soil and water samples were validated for analyses of semivolatile organics by the US EPA Region II data validation SOP (HW-22, Revision 3). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
- Calibrations
- \* - Laboratory Blanks
- \* - Field Blank
- Laboratory Control Sample
- Surrogate Compound Recoveries
- \* - Internal Standard Recoveries
- Matrix Spike / Matrix Spike Duplicate
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The minor problems with the laboratory control samples, matrix spike and calibrations should be noted. These are discussed in detail below.

## Holding Times

All samples were extracted and analyzed within the contractual and technical times required by the US EPA Region II protocols.

Continuous extraction of water samples for semivolatile analysis was started within 7 days of the date of collection and soil samples 14 days.

Extracts were analyzed within 40 days of the date of extraction.

## Tunes

No problems were detected with any of the tunes associated with the samples of this delivery group.

## Surrogate Recoveries

All surrogate recoveries were within the required limits with the following exceptions:

	A S1	A S2	B S3	B S4	A S5	B S6
MB-59737						127%
LCS-59737						135%
K0909-05	80230-GW-MIP6-33.5					127%
K0909-07	80230-GW-MIP6-15					129%
K0909-08	80230-FB-110523					131%
K0909-09	80230-FB-AQ-110523					126%
K0909-10 MS	80230-GW-MIP5-28 MS					128%
K0909-12	80230-FB-AQ-110524					

There should not be a problem with surrogate recoveries in a blank and laboratory control sample.

The data were not required to be qualified since the NYS DEC ASP requirements allow one surrogate in each fraction to be outside of the limits (as long as it is greater than 10%).

## Laboratory Control Samples

All of the recoveries of the LCS related to the water samples were within the required limits.

All of the recoveries of the LCS related to the soil samples were within the required limits with the exception of 2-methylnaphthalene (121%) which was above the 125% quality control limit.

This compound was not detected in any of the samples and the data were not affected by the high recovery.

## Matrix Spike / Matrix Spike Duplicate

Sample K0909-10 /80230-GW-MIP5-28 from this sample delivery group was used as the matrix spike and matrix spike duplicate. All recoveries and RPDs which could be

accurately calculated were within the required limits with the exceptions of the caprolactam matrix spike recoveries (18% & 16%) and the RPD of 2,4-dinitrophenol (63%).

The data for these two compounds were flagged with the "J" qualifier. It is possible that low concentrations of these compounds may have been overlooked.

### **Calibrations**

The %RSDs of all compounds in the initial calibration associated with the analysis of all of the samples were less than 15% with the exceptions of 2,4-dinitrophenol (26%) and benzaldehyde (18%).

None of these compounds were detected in any of the samples and the data were not required to be qualified for the high %RSD.

The %Ds of all compounds in the continuing calibration associated with the analyses of samples -05, -07, -08, -09, -10 & -12 were less than 20% with the exceptions of 2-methylnaphthalene (34%) and 2,4-dinitrophenol (30%).

The %Ds of all compounds in the continuing calibration associated with the analysis of samples -01, -02, -03 & -04 were less than 20% with the exceptions of nitrobenzene (22%), 2-methylnaphthalene (32%) and 2,4-dinitrophenol (30%).

The data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Method Blanks**

No target compounds were detected in the method blanks.

### **Field Blank**

No target compounds were detected in the field blank.

### **Internal Standard Areas and Retention Times**

All internal standard recoveries and retention times were within the required limits.

### **Sample Results**

No problems were found with the results of any of the samples of this delivery group.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Total Metals Analyses**

**Samples Collected:** May 24, 2011

**Samples Received:** May 26, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K0918

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0918-01	80230-GW-MIP5-15	Aqueous	24-May-11
K0918-03	80230-DUP1-GW	Aqueous	24-May-11
K0918-06	80230-FB-110524	Aqueous	24-May-11

Water samples were validated for inorganic analyses by the US EPA Region II data validation SOP (HW-2, Revision 13). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - Holding Times
- \* - Calibration Verification
  - CRDL Standard
- \* - Laboratory Control Sample
  - Serial Dilutions
  - Calibration Blanks
  - Field Blank
- \* - Preparation Blanks
  - Matrix Spike
  - Matrix Duplicate
- \* - ICP Interference Check Sample
- \* - Detection Limit Results
- \* - Linear Range
- \* - Sample Results

\* - Indicates that all criteria were met for this parameter.

**Data Validation Summary**

A CRDL standard was not analyzed with this sample delivery group.

No other problems were found that would affect the use of the data.

## **Holding Times**

All samples were analyzed within the required holding times.

## **CRDL Standards**

A CRDL standard was not analyzed with this sample delivery group.

## **Initial and Continuing Calibrations**

No problems were detected with any of the calibrations associated with this sample delivery group.

## **Preparation Blank**

No compounds were detected in the one preparation blank associated with the digestions of these samples at concentrations above the CRDL. Several analytes were found in the preparation blank at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table.

## **Calibration Blanks**

Several analytes were found in the continuing calibration blanks at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table and do not affect the end use of the data.

## **Field Blank**

Low concentrations of sodium (271 mg/l) were detected in the field blank. The sodium concentrations in the samples were too high to be affected by the field blank.

## **ICP Interference Check Sample**

No problems were detected with the reported ICP Interference Check Sample recoveries.

## **Matrix Spike Recovery**

A matrix spike was not analyzed with this sample delivery group.

## **Duplicate Analysis**

A matrix duplicate was not analyzed with this sample delivery group.

## **Laboratory Control Sample**

No problems were detected with the recoveries of the LCS standards.

### **Serial Dilutions**

A serial dilution was not analyzed with this sample delivery group.

### **Instrument Detection Limit**

No problems were found with the instrument detection limits.

### **ICP Linear Ranges**

No problems were detected with the linear ranges.

### **Sample Results**

No problems were detected with any of these samples.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Water Pesticide Analyses**

**Samples Collected: May 24, 2011**

**Samples Received: May 26, 2011**

**Laboratory: Spectrum**

**Sample Delivery Group: K0918**

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0918-06	80230-FB-110524	Aqueous	24-May-11

One water sample was validated for analyses of pesticides by the US EPA Region II data validation SOP (HW-44, Revision 1). Data were reviewed for usability according to the following criteria:

- Data Completeness
- \* - Holding Times
- \* - Laboratory Blanks
- \* - Field Blanks
- Surrogate Recoveries
- \* - Surrogate Retention Times
- Matrix Spike / Matrix Spike Duplicate
- \* - Surrogate Spike Recovery
- \* - Laboratory Control Sample
- \* - Calibrations
- \* - Method Blanks
- Florisil Cartridge Check
- GPC Calibration
- \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

Form IX for a florisil cleanup was not included in the data package.

No significant problems were detected with any of the data.

**Holding Times**

All extractions and analyses were performed within the required holding times.

**Surrogate Recoveries**

The laboratory's case narrative states:

80230-FB-110524 (K0918-06C), recovery is below criteria for Decachlorobiphenyl on rear column at 25% with criteria of (30-135) and Decachlorobiphenyl on front column at 28% with criteria of (30-135).

The data were not qualified since only a field blank sample was submitted.

### **Matrix Spike**

A matrix spike was not analyzed with this sample delivery group.

### **Laboratory Control Samples**

All of the laboratory control sample recoveries were within the required limits.

### **Initial Calibrations**

No problems were detected with the initial calibrations associated with the analyses of the samples. All % RSDs were less than 20%.

### **Continuing Calibrations**

All %Ds in the continuing calibrations directly with the sample were less than 20%.

### **Florisil Cartridge Check**

Form IX for a florisil cleanup was not included in the data package.

### **GPC Calibration**

A GPC cleanup was not performed on these samples.

### **Method Blanks**

No problems were detected with any of the method blanks.

### **Calibration Blanks**

No problems were detected with the calibration blanks associated with this sample delivery group.

### **Field Blank**

No compounds were detected in the field blank.

### **Sample Results**

No problems were detected with the sample data.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Water PCB Analyses**

**Samples Collected:** May 24, 2011

**Samples Received:** May 26, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K0918

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0918-06	80230-FB-110524	Aqueous	24-May-11

One water sample was validated for analyses of PCBs by the US EPA Region II data validation SOP (HW-44, Revision 1). Data were reviewed for usability according to the following criteria:

- Data Completeness
- \* - Holding Times
- \* - Laboratory Blanks
- \* - Field Blanks
- \* - Surrogate Recoveries
- \* - Surrogate Retention Times
- Matrix Spike / Matrix Spike Duplicate
- \* - System Monitoring Spike Recovery
- \* - Laboratory Control Sample
- \* - Calibrations
- \* - Method Blanks
- GPC Calibration
- \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

Only compounds noted as target compounds by the laboratory's data system were reported in the raw data.

No other significant problems were detected with any of the data.

**Holding Times**

All extractions and analyses were performed within the required holding times.

**Surrogate Recoveries**

The laboratory's case narrative states:

*80230-FB-110524 (K0918-06C), recovery is below criteria for Decachlorobiphenyl on rear column at 28% with criteria of (40-135) and Decachlorobiphenyl on front column at 28% with criteria of (40-135).*

The data were not qualified since only a field blank sample was submitted.

### **Matrix Spike**

A matrix spike was not analyzed with this sample delivery group.

### **Laboratory Control Samples**

All of the laboratory control sample recoveries were within the required limits.

### **Initial Calibrations**

No problems were detected with the initial calibrations associated with the analyses of the samples. All %RSDs were less than 20%.

### **Continuing Calibrations**

All %Ds were less than 15%.

### **Florisil Cartridge Check**

Form IX for a florisil cleanup was not included in the data package.

### **GPC Calibration**

A GPC cleanup was not performed on these samples.

### **Method Blanks**

No problems were detected with any of the method blanks.

### **Calibration Blanks**

No problems were detected with the calibration blanks associated with this sample delivery group.

### **Field Blank**

No compounds were detected in the field blank.

### **Sample Results**

No problems were detected with the sample data.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Semivolatile Organic Analyses**

**Samples Collected:** May 24, 2011

**Samples Received:** May 26, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K0918

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0918-01	80230-GW-MIP5-15	Aqueous	24-May-11
K0918-03	80230-DUP1-GW	Aqueous	24-May-11
K0918-04	80230-MIP5-11	Soil	24-May-11
K0918-05	80230-MIP5-30	Soil	24May-11
K0918-06	80230-FB-110524	Aqueous	24-May-11
K0918-07	80230-MIP1-16.5	Soil	24-May-11
K0918-08	80230-MIP2-30	Soil	24-May-11

Soil and water samples were validated for analyses of semivolatile organics by the US EPA Region II data validation SOP (HW-22, Revision 3). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
- \* - Laboratory Blanks
- \* - Field Blank
  - Laboratory Control Sample
  - Surrogate Compound Recoveries
- \* - Internal Standard Recoveries
  - Matrix Spike / Matrix Spike Duplicate
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The minor problems with the laboratory control samples and calibrations should be noted. These are discussed in detail below.

## Holding Times

All samples were extracted and analyzed within the contractual and technical times required by the US EPA Region II protocols.

Continuous extraction of water samples for semivolatile analysis was started within 7 days of the date of collection and soil samples 14 days.

Extracts were analyzed within 40 days of the date of extraction.

## Tunes

No problems were detected with any of the tunes associated with the samples of this delivery group.

## Surrogate Recoveries

All surrogate recoveries were within the required limits with the following exceptions:

	A S1	A S2	B S3	B S4	A S5	B S6
MB-59437						127%
LCS-59437						135%
MB-59433						132%
LCS-59433						126%
LCSD-59433						132%

There should not be a problem with surrogate recoveries in method blanks and laboratory control samples.

## Laboratory Control Samples

All of the recoveries of the LCS related to the water samples were within the required limits with the exception of caprolactam (18%).

The data for this compound were flagged with the "J" qualifier and are estimated values.

All of the recoveries of the LCS related to the soil samples were within the required limits with the exception of 2-methylnaphthalene (125%) and caprolactam (18%).

The data for caprolactam were flagged with the "J" qualifier and are estimated values.

2-Methylnaphthalene was not detected in any of the samples and the data were not affected by the high recovery.

## Matrix Spike / Matrix Spike Duplicate

A matrix spike and matrix spike duplicate were not analyzed with this sample delivery group.

## **Calibrations**

The %RSDs of all compounds in the initial calibration associated with the analysis of all of the samples were less than 15% with the exceptions of 2,4-dinitrophenol (26%) and benzaldehyde (18%).

None of these compounds were detected in any of the samples and the data were not required to be qualified for the high %RSD.

The %Ds of all compounds in the continuing calibration associated with the analysis of all the samples were less than 20% with the exceptions of 2-methylnaphthalene (34%) and 2,4-dinitrophenol (30%)

The data for these compounds were flagged with the "J" qualifier and are estimated values.

## **Method Blanks**

No target compounds were detected in the method blanks.

## **Field Blank**

No target compounds were detected in the field blank.

## **Internal Standard Areas and Retention Times**

All internal standard recoveries and retention times were within the required limits.

## **Sample Results**

No problems were found with the results of any of the samples of this delivery group.

**SUMMARY OF THE ANALYTICAL DATA USABILITY**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Volatile Organic Analyses – Method 8260C**

**Samples Collected:** May 24, 2011

**Samples Received:** May 26, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K0918

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0918-01	80230-GW-MIP5-15	Aqueous	24-May-11
K0918-02	TRIP BLANK	Aqueous	24-May-11
K0918-03	80230-DUP1-GW	Aqueous	24-May-11
K0918-04	80230-MIP5-11	Soil	24-May-11
K0918-05	80230-MIP5-30	Soil	24May-11
K0918-06	80230-FB-110524	Aqueous	24-May-11
K0918-07	80230-MIP1-16.5	Soil	24-May-11
K0918-08	80230-MIP2-30	Soil	24-May-11

Soil and water samples were validated for analyses of volatile organics by the US EPA Region II data validation SOP (HW-24, Revision 2). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
  - Laboratory Blanks
- \* - Trip Blanks
- \* - Field Blanks
- \* - System Monitoring Compound Recoveries
- \* - Internal Standard Recoveries
  - Matrix Spike / Matrix Spike Duplicate
  - Laboratory Control Samples
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The problems with the calibrations and laboratory control samples should be noted. These are described in detail below.

No other significant problems were found with this sample delivery group which would affect the usability of the data.

## **Holding Times**

The samples of this delivery group were validated against the Region II technical holding time requirements:

All of the samples were analyzed within 14 days of collection.

## **Tunes**

No problems were detected with the tunes associated with the samples of this delivery group.

## **System Monitoring Compound Recoveries**

All of the sample surrogate recoveries were within the required limits.

## **Calibrations**

### **Initial Calibrations**

All relative percent differences in the 5/05 initial calibration associated with the analyses of samples -02, -06, -01, -01DL and -03 were less than 20% with the exception of naphthalene (23%). .

The RRF of 2-butanone (0.038) was less than the 0.050 quality control limit in the above calibration.

All relative percent differences in the 5/31 initial calibration associated with the analyses of samples -04, -05, -07 and 08 were less than 20% with the exceptions of methylene chloride (21%), sec-butylbenzene (22%), 4-isopropyltoluene (21%), naphthalene (28%) and methylcyclohexane (24%). The data for these compounds were flagged with the "J" qualifier and are estimated values.

The RRFs of acetone (0.024) and 2-butanone (0.039) were less than the 0.050 quality control limit in the above calibration. This compound was not detected in any of the samples and the data were flagged with the "R" qualifier and technically rejected.

Detected data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Continuing Calibrations**

All of the percent differences in the continuing calibration associated with the analysis of samples -02, -06 and -01 were less than 20% with the exceptions of dichlorodifluoromethane (31%) and acetone (34%).

The relative response factors of acetone (0.036) and 2-butanone (0.037) in this continuing calibration were less than 0.050.

All of the percent differences in the continuing calibration associated with the analysis of samples -03 and -01DL were less than 20% with the exceptions of

dichlorodifluoromethane (28%), 1,2,4-trichlorobenzene (21%), hexachlorobutadiene (23%), 1,2,3-trichlorobenzene (21%), and methyl acetate (23%).

The relative response factors of 2-butanone (0.044) in this continuing calibration were less than 0.050.

All of the percent differences in the continuing calibration associated with the analysis of sample -04, -05, -07, and -08 were less than 20% with the exceptions of 1,2-dibromo-3-chloropropane (21%) and naphthalene (24%). The data for these compounds were flagged with the "J" qualifier and are estimated values.

The relative response factors of acetone (0.020) and 2-butanone (0.037) in this continuing calibration were less than 0.050.

The compounds with percent differences greater than 20% were flagged with the "J" qualifier and are estimated values.

Acetone and 2-butanone were not detected in any of the samples. The data for these compounds were flagged with the "R" qualifier in the associated samples and are technically rejected.

### **Matrix Spike and Matrix Spike Duplicate**

A matrix spike and matrix spike duplicate were not analyzed.

### **Laboratory Control Sample**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

All of the laboratory control sample recoveries associated with soil samples -01, -02 and -06 were within the required limits with the exception of dichlorodifluoromethane (63%) and acetone (69%).

The data for these compounds were flagged with the "J" qualifier and are estimated values.

All of the other LCS recoveries were within the required limits.

### **Method Blanks**

Low concentrations of 1,2,4-trichlorobenzene (1.3J ug/kg), 1,2,3-trichlorobenzene (1.4J ug/kg) and naphthalene (2.8J ug/kg) were detected in the method blank associated with samples -04, -05, -07 and -08.

None of these compounds were detected in any of these samples and the blank contamination does not affect the usability of the data.

No target compounds were detected in the other method blanks.

### **Trip Blank**

No target compounds were detected in the trip blank.

### **Field Blank**

No target compounds were detected in the field blank.

### **Internal Standard Areas and Retention Times**

The recoveries and retention times of all internal standards were within the required 50% - 100% quality control limits.

### **Sample Results**

No problems were detected with the samples.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Total Metals Analyses**

**Samples Collected:** May 25, 2011

**Samples Received:** May 31, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K0940

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0940-02	80230-MIP1B-14	Soil	25-May-11
K0940-03	80230-FB-110525	Aqueous	25-May-11
K0940-04	80230-MIP5B-22	Soil	25-May-11
K0940-04 MS	80230-MIP5B-22 MS	Soil	25-May-11
K0940-04 MSD	80230-MIP5B-22 MSD	Soil	25-May-11

Soil and water samples were validated for inorganic analyses by the US EPA Region II data validation SOP (HW-2, Revision 13). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - Holding Times
- \* - Calibration Verification
- CRDL Standard
- \* - Laboratory Control Sample
  - Serial Dilutions
- \* - Calibration Blanks
  - Field Blank
- \* - Preparation Blanks
  - Matrix Spike
  - Matrix Duplicate
- \* - ICP Interference Check Sample
- \* - Detection Limit Results
- \* - Linear Range
- \* - Sample Results

\* - Indicates that all criteria were met for this parameter.

**Data Validation Summary**

A CRDL standard was not analyzed with this sample delivery group.

The problems with the matrix spike, matrix duplicate and serial dilution should be noted. These are described in detail below.

## Holding Times

All samples were analyzed within the required holding times.

## CRDL Standards

A CRDL standard was not analyzed with this sample delivery group.

## Initial and Continuing Calibrations

No problems were detected with any of the calibrations associated with this sample delivery group.

## Preparation Blank

No compounds were detected in the one preparation blank associated with the digestions of these samples at concentrations above the CRDL. Several analytes were found in the preparation blank at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table.

## Calibration Blanks

Several analytes were found in the continuing calibration blanks at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table and do not affect the end use of the data.

## Field Blank

A low concentration of calcium (172 ug/l) was detected in the field blank. The concentrations of calcium in the samples were too high to be affected by the blank contamination.

## ICP Interference Check Sample

No problems were detected with the reported ICP Interference Check Sample recoveries.

## Matrix Spike Recovery

Sample K0940-04 / 80230-MIP5B-22 was used as the matrix spike. All recoveries were within the 75% - 125% quality control limits with the exception of antimony (59%).

The antimony data in the soil samples were flagged with the "J" qualifier and are estimated values.

## Duplicate Analysis

Sample K0940-04 / 80230-MIP5B-22 was used as the matrix duplicate. All RPDs that could be accurately calculated were less than 20% with the following exceptions:

<b>Analyte</b>	<b>RPD</b>
Arsenic.	23%
Chromium	32%
Cobalt	27%
Copper	27%
Iron	21%
Magnesium	25%
Potassium	24%
Vanadium	22%
Zinc	29%

The data for these analytes were flagged with the "J' qualifier and are estimated values.

### **Laboratory Control Sample**

No problems were detected with the recoveries of the LCS standards.

### **Serial Dilutions**

Sample K0940-04 / 80230-MIP5B-22 was used as the serial dilution. All percent differences that could be accurately calculated were less than 10% with the exception of sodium (16%).

The sodium data were flagged with the "J' qualifier and are estimated values.

### **Instrument Detection Limit**

No problems were found with the instrument detection limits.

### **ICP Linear Ranges**

No problems were detected with the linear ranges.

### **Sample Results**

No problems were detected with any of these samples.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Pesticide Analyses**

**Samples Collected: May 25, 2011**

**Samples Received: May 31, 2011**

**Laboratory: Spectrum**

**Sample Delivery Group: K0940**

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0940-02	80230-MIP1B-14	Soil	25-May-11
K0940-03	80230-FB-110525	Aqueous	25-May-11
K0940-04	80230-MIP5B-22	Soil	25-May-11
K0940-04 MS	80230-MIP5B-22 MS	Soil	25-May-11
K0940-04 MSD	80230-MIP5B-22 MSD	Soil	25-May-11

Soil and water samples were validated for analyses of pesticides by the US EPA Region II data validation SOP (HW-44, Revision 1). Data were reviewed for usability according to the following criteria:

- Data Completeness
- \* - Holding Times
- \* - Laboratory Blanks
- \* - Field Blanks
- Surrogate Recoveries
- \* - Surrogate Retention Times
- \* - Matrix Spike / Matrix Spike Duplicate
- \* - Surrogate Spike Recovery
- \* - Laboratory Control Sample
- \* - Calibrations
- \* - Method Blanks
- Florisil Cartridge Check
- GPC Calibration
- \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

Form IX for a florisil cleanup was not included in the data package.

No significant problems were detected with any of the data.

### **Holding Times**

All extractions and analyses were performed within the required holding times.

### **Surrogate Recoveries**

All surrogate recoveries were within the required limits.

### **Matrix Spike**

Sample K0940-04 / 80230-MIP5B-22 was used as the matrix spike and matrix spike duplicate. All recoveries and RPDs were within the required limits.

### **Laboratory Control Samples**

The laboratory's case narrative states:

*LCS-59481 in batch 59481, recovery is above criteria for gamma-Chlordane on rear column at 278% with criteria of (60-125).*

*LCS-59497 in batch 59497, recovery is above criteria for alpha-Chlordane on front column at 120% with criteria of (65-120).*

These two compounds were not detected in any of the samples and the high recoveries do not affect the use of the data.

All of the other laboratory control sample recoveries were within the required limits.

### **Initial Calibrations**

No problems were detected with the initial calibrations associated with the analyses of the samples. All % RSDs were less than 20%.

### **Continuing Calibrations**

All %Ds in the continuing calibrations directly with the sample were less than 20%.

### **Florisil Cartridge Check**

Form IX for a florisil cleanup was not included in the data package.

### **GPC Calibration**

A GPC cleanup was not performed on these samples.

### **Method Blanks**

No problems were detected with any of the method blanks.

### **Calibration Blanks**

No problems were detected with the calibration blanks associated with this sample delivery group.

### **Field Blank**

No compounds were detected in the field blank.

### **Sample Results**

No problems were detected with the sample data.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water PCB Analyses**

**Samples Collected:** May 25, 2011

**Samples Received:** May 31, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K0940

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0940-02	80230-MIP1B-14	Soil	25-May-11
K0940-03	80230-FB-110525	Aqueous	25-May-11
K0940-04	80230-MIP5B-22	Soil	25-May-11
K0940-04 MS	80230-MIP5B-22 MS	Soil	25-May-11
K0940-04 MSD	80230-MIP5B-22 MSD	Soil	25-May-11

Soil and water sample were validated for analyses of PCBs by the US EPA Region II data validation SOP (HW-44, Revision 1). Data were reviewed for usability according to the following criteria:

- Data Completeness
- \* - Holding Times
- \* - Laboratory Blanks
- \* - Field Blanks
- \* - Surrogate Recoveries
- \* - Surrogate Retention Times
- \* - Matrix Spike / Matrix Spike Duplicate
- \* - System Monitoring Spike Recovery
- \* - Laboratory Control Sample
- \* - Calibrations
- \* - Method Blanks
- GPC Calibration
- \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

Only compounds noted as target compounds by the laboratory's data system were reported in the raw data.

No other significant problems were detected with any of the data.

## Holding Times

All extractions and analyses were performed within the required holding times.

## Surrogate Recoveries

The laboratory's case narrative states:

*80230-FB-110525 (K0940-03C), recovery is below criteria for Decachlorobiphenyl on front column at 37% with criteria of (40-135).*

The data were not qualified since the NYS DEC ASP protocol allow for one surrogate to be outside the quality control limits.

## Matrix Spike

Sample K0940-04 / 80230-MIP5B-22 was used as the matrix spike and matrix spike duplicate. All recoveries and RPDs were within the required limits.

## Laboratory Control Samples

All of the laboratory control sample recoveries were within the required limits.

## Initial Calibrations

No problems were detected with the initial calibrations associated with the analyses of the samples. All %RSDs were less than 20%.

## Continuing Calibrations

Several of the percent differences were greater than 15% (21%).

The data were not qualified since no compounds were detected in any of the samples and the areas of the standards were increasing.

## Florisil Cartridge Check

Form IX for a florisil cleanup was not included in the data package.

## GPC Calibration

A GPC cleanup was not performed on these samples.

## Method Blanks

No problems were detected with any of the method blanks.

## Calibration Blanks

No problems were detected with the calibration blanks associated with this sample delivery group.

### **Field Blank**

No compounds were detected in the field blank.

### **Sample Results**

No problems were detected with the sample data.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Semivolatile Organic Analyses**

**Samples Collected:** May 25, 2011

**Samples Received:** May 31, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K0940

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0940-01	80230-MIP3-30	Soil	25-May-11
K0940-02	80230-MIP1B-14	Soil	25-May-11
K0940-03	80230-FB-110525	Aqueous	25-May-11
K0940-04	80230-MIP5B-22	Soil	25-May-11
K0940-04 MS	80230-MIP5B-22 MS	Soil	25-May-11
K0940-04 MSD	80230-MIP5B-22 MSD	Soil	25-May-11
K0940-05	80230-DUP1-SO	Soil	25-May-11

Soil and water samples were validated for analyses of semivolatile organics by the US EPA Region II data validation SOP (HW-22, Revision 3). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
  - Laboratory Blanks
  - Field Blank
  - Laboratory Control Sample
  - Surrogate Compound Recoveries
- \* - Internal Standard Recoveries
- \* - Matrix Spike / Matrix Spike Duplicate
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The minor problems with the laboratory control samples and calibrations should be noted. These are discussed in detail below.

## Holding Times

All samples were extracted and analyzed within the contractual and technical times required by the US EPA Region II protocols.

Continuous extraction of water samples for semivolatile analysis was started within 7 days of the date of collection and soil samples 14 days.

Extracts were analyzed within 40 days of the date of extraction.

## Tunes

No problems were detected with any of the tunes associated with the samples of this delivery group.

## Surrogate Recoveries

All surrogate recoveries were within the required limits with the following exceptions:

	A S1	A S2	B S3	B S4	A S5	B S6
MB-59483						136%
LCS-59483						139%
LCSD-59483						138%
K0940-03		80230-FB-110525				128%
MB-59499						127%

There should not be a problem with surrogate recoveries in method blanks, field blanks and laboratory control samples.

The data were not required to be qualified since the NYS DEC ASP requirements allow one surrogate in each fraction to be outside of the limits (as long as it is greater than 10%).

## Laboratory Control Samples

All of the recoveries of the LCS related to the field blank were within the required limits with the exceptions of 2-methylnaphthalene (117%) and caprolactam (18%).

2-Methylnaphthalene was not detected in the sample and the data were not affected by the high recovery.

The data for caprolactam were flagged with the "J" qualifier and are estimated values.

All of the recoveries of the LCS related to the soil samples were within the required limits with the exception of 2-methylnaphthalene (108%).

2-Methylnaphthalene was not detected in any of the samples and the data were not affected by the high recovery.

### **Matrix Spike / Matrix Spike Duplicate**

Sample K0940-04 / 80230-MIP5B-22 was used as the matrix spike and matrix spike duplicate.

All recoveries and RPDs were within the required limits.

### **Calibrations**

The %RSDs of all compounds in the initial calibration associated with the analysis of all of the samples were less than 15% with the exceptions of 2,4-dinitrophenol (26%) and benzaldehyde (18%).

None of these compounds were detected in any of the samples and the data were not required to be qualified for the high %RSD.

The %Ds of all compounds in the continuing calibration associated with the analysis of the field blank were less than 20% with the exceptions of nitrobenzene (21%), 2-methylnaphthalene (32%) and 2,4-dinitrophenol (35%).

The %Ds of all compounds in the continuing calibration associated with the analysis of the soil samples were less than 20% with the exceptions of nitrobenzene (27%), 2-methylnaphthalene (33%) and 2,4-dinitrophenol (43%)

The data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Method Blanks**

A low concentration of bis(2-ethylhexyl)phthalate (13 ug/l) was detected in the field blank associated with the analysis of the field blank.

No target compounds were detected in the soil method blank.

### **Field Blank**

A low concentration of bis(2-ethylhexyl)phthalate (7.5 ug/l) was detected in the field blank.

Only low concentrations of this compound, less than the CRDL were detected in these samples. All of the data for this compound were flagged with the "U" qualifier during the data validation.

### **Internal Standard Areas and Retention Times**

All internal standard recoveries and retention times were within the required limits.

### **Sample Results**

No problems were found with the results of any of the samples of this delivery group.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Total Metals Analyses**

**Samples Collected: May 26, 2011**

**Samples Received: May 31, 2011**

**Laboratory: Spectrum**

**Sample Delivery Group: K0941**

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0941-01	80230-FB-110526	Aqueous	26-May-11
K0941-02	80230-MIP3B-45	Soil	26-May-11
K0941-02 MS	80230-MIP3B-45 MS	Soil	26-May-11
K0941-02 MSD	80230-MIP3B-45 MSD	Soil	26-May-11
K0941-03	80230-MIP2B-40	Soil	26-May-11
K0941-04	80230-MIP1D-37.5	Soil	26-May-11

Soil and water samples were validated for inorganic analyses by the US EPA Region II data validation SOP (HW-2, Revision 13). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - Holding Times
- \* - Calibration Verification
  - CRDL Standard
- \* - Laboratory Control Sample
  - Serial Dilutions
  - Calibration Blanks
  - Field Blank
- \* - Preparation Blanks
  - Matrix Spike
  - Matrix Duplicate
- \* - ICP Interference Check Sample
- \* - Detection Limit Results
- \* - Linear Range
- \* - Sample Results

\* - Indicates that all criteria were met for this parameter.

**Data Validation Summary**

A CRDL standard was not analyzed with this sample delivery group.

The problems with the matrix spike, matrix duplicate and serial dilution should be noted. These are described in detail below.

## **Holding Times**

All samples were analyzed within the required holding times.

## **CRDL Standards**

A CRDL standard was not analyzed with this sample delivery group.

## **Initial and Continuing Calibrations**

No problems were detected with any of the calibrations associated with this sample delivery group.

## **Preparation Blank**

No compounds were detected in the one preparation blank associated with the digestions of these samples at concentrations above the CRDL. Several analytes were found in the preparation blank at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table.

## **Calibration Blanks**

Several analytes were found in the continuing calibration blanks at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table and do not affect the end use of the data.

## **Field Blank**

Low concentrations of calcium (267 ug/l) and zinc (25.8 ug/l) were detected in the field blank.

The concentrations of calcium and zinc in the samples were too high to be affected by the blank contamination.

## **ICP Interference Check Sample**

No problems were detected with the reported ICP Interference Check Sample recoveries.

## **Matrix Spike Recovery**

Sample K0941-02 / 80230-MIP3B-45 was used as the matrix spike. All recoveries were within the 75% - 125% quality control limits with the exception of antimony (70%).

The antimony data in the soil samples were flagged with the "J" qualifier and are estimated values.

## Duplicate Analysis

Sample K0941-02 / 80230-MIP3B-45 was used as the matrix duplicate. All RPDs that could be accurately calculated were less than 20% with the following exceptions:

Analyte	RPD
Arsenic.	58%
Barium	22%
Copper	32%
Iron	20%
Manganese	22%
Vanadium	25%

The data for these analytes were flagged with the "J" qualifier and are estimated values.

## Laboratory Control Sample

No problems were detected with the recoveries of the LCS standards.

## Serial Dilutions

Sample K0941-02 / 80230-MIP3B-45 was used as the serial dilution. All percent differences that could be accurately calculated were less than 10% with the exception of cobalt (12%).

The cobalt data were flagged with the "J" qualifier and are estimated values.

## Instrument Detection Limit

No problems were found with the instrument detection limits.

## ICP Linear Ranges

No problems were detected with the linear ranges.

## Sample Results

No problems were detected with any of these samples.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Pesticide Analyses**

**Samples Collected: May 26, 2011**

**Samples Received: May 31, 2011**

**Laboratory: Spectrum**

**Sample Delivery Group: K0941**

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0941-01	80230-FB-110526	Aqueous	26-May-11
K0941-02	80230-MIP3B-45	Soil	26-May-11
K0941-02 MS	80230-MIP3B-45 MS	Soil	26-May-11
K0941-02 MSD	80230-MIP3B-45 MSD	Soil	26-May-11
K0941-03	80230-MIP2B-40	Soil	26-May-11
K0941-04	80230-MIP1D-37.5	Soil	26-May-11

Soil and water samples were validated for analyses of pesticides by the US EPA Region II data validation SOP (HW-44, Revision 1). Data were reviewed for usability according to the following criteria:

- Data Completeness
- \* - Holding Times
- \* - Laboratory Blanks
- \* - Field Blanks
- Surrogate Recoveries
- \* - Surrogate Retention Times
- Matrix Spike / Matrix Spike Duplicate
- Laboratory Control Sample
- Calibrations
- \* - Method Blanks
- Florisil Cartridge Check
- GPC Calibration
- \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The minor problems with the continuing calibrations should be noted.

Form IX for a florisil cleanup was not included in the data package.

No other significant problems were detected with any of the data.

## Holding Times

All extractions and analyses were performed within the required holding times.

## Surrogate Recoveries

The laboratory's case narrative states:

*80230-FB-110526 (K0941-01C), recovery is below criteria for Decachlorobiphenyl on rear column at 29% with criteria of (30-135).*

The data were not qualified since the NYS DEC ASP protocol allow for one surrogate to be outside the quality control limits.

## Matrix Spike

Sample K0941-02 / 80230-MIP3B-45 was used as the matrix spike and matrix spike duplicate.

The laboratory's case narrative states:

*80230-MIP3B-45 (K0941-02AMSD), recovery is above criteria for alpha-BHC on rear column at 127% with criteria of (60-125), beta-BHC on rear column at 130% with criteria of (60-125) and beta-BHC on front column at 131% with criteria of (60-125).*

*Replicate RPDs were within the QC limits.*

The data were not affected since no compounds were detected in any of the samples.

## Laboratory Control Samples

The laboratory's case narrative states:

*LCS-59481 in batch 59481, recovery is above criteria for gamma-Chlordane on rear column at 278% with criteria of (60-125).*

*LCS-59497 in batch 59497, is above criteria for alpha-Chlordane on front column at 120% with criteria of (65-120).*

These two compounds were not detected in any of the samples and the high recoveries do not affect the use of the data.

All of the other laboratory control sample recoveries were within the required limits.

## Initial Calibrations

No problems were detected with the initial calibrations associated with the analyses of the samples. All % RSDs were less than 20%.

### **Continuing Calibrations**

The percent differences of alpha-BHC (30%) and delta-BHC (30%) the PEMVQ continuing calibration standard associated with the analyses of sample K0941-02 / 80230-MIP3B-45 were above the 20% quality control limit.

The data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Florisil Cartridge Check**

Form IX for a florisil cleanup was not included in the data package.

### **GPC Calibration**

A GPC cleanup was not performed on these samples.

### **Method Blanks**

No problems were detected with any of the method blanks.

### **Calibration Blanks**

No problems were detected with the calibration blanks associated with this sample delivery group.

### **Field Blank**

No compounds were detected in the field blank.

### **Sample Results**

No problems were detected with the sample data.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water PCB Analyses**

**Samples Collected:** May 26, 2011

**Samples Received:** May 31, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K0941

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0941-01	80230-FB-110526	Aqueous	26-May-11
K0941-02	80230-MIP3B-45	Soil	26-May-11
K0941-02 MS	80230-MIP3B-45 MS	Soil	26-May-11
K0941-02 MSD	80230-MIP3B-45 MSD	Soil	26-May-11
K0941-03	80230-MIP2B-40	Soil	26-May-11
K0941-04	80230-MIP1D-37.5	Soil	26-May-11

Soil and water samples were validated for analyses of PCBs by the US EPA Region II data validation SOP (HW-44, Revision 1). Data were reviewed for usability according to the following criteria:

- Data Completeness
- \* - Holding Times
- \* - Laboratory Blanks
- \* - Field Blanks
  - Surrogate Recoveries
  - \* - Surrogate Retention Times
  - \* - Matrix Spike / Matrix Spike Duplicate
  - \* - System Monitoring Spike Recovery
  - \* - Laboratory Control Sample
  - \* - Calibrations
  - \* - Method Blanks
    - GPC Calibration
  - \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

Only compounds noted as target compounds by the laboratory's data system were reported in the raw data.

No other significant problems were detected with any of the data.

## Holding Times

All extractions and analyses were performed within the required holding times.

## Surrogate Recoveries

The laboratory's case narrative states:

*80230-FB-110526 (K0941-01C), recovery is below criteria for Decachlorobiphenyl on rear column at 35% with criteria of (40-135) and Decachlorobiphenyl on front column at 32% with criteria of (40-135).*

The data were not qualified since only the field blank was affected.

There should not be a problem with surrogate recoveries in a field blank.

## Matrix Spike

Sample K0941-02 / 80230-MIP3B-45 was used as the matrix spike and matrix spike duplicate. All recoveries and RPDs were within the required limits.

## Laboratory Control Samples

All recoveries were within the required limits.

## Initial Calibrations

No problems were detected with the initial calibrations associated with the analyses of the samples. All %RSDs were less than 20%.

## Continuing Calibrations

Several of the percent differences were greater than 15% (21%).

The data were not qualified since no compounds were detected in any of the samples and the areas of the standards were increasing.

## Florisil Cartridge Check

Form IX for a florisil cleanup was not included in the data package.

## GPC Calibration

A GPC cleanup was not performed on these samples.

## Method Blanks

No problems were detected with any of the method blanks.

## Calibration Blanks

No problems were detected with the calibration blanks associated with this sample delivery group.

### **Field Blank**

No compounds were detected in the field blank.

### **Sample Results**

No problems were detected with the sample data.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Semivolatile Organic Analyses**

**Samples Collected:** May 26, 2011

**Samples Received:** May 31, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K0941

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0941-01	80230-FB-110526	Aqueous	26-May-11
K0941-02	80230-MIP3B-45	Soil	26-May-11
K0941-02 MS	80230-MIP3B-45 MS	Soil	26-May-11
K0941-02 MSD	80230-MIP3B-45 MSD	Soil	26-May-11
K0941-03	80230-MIP2B-40	Soil	26-May-11
K0941-04	80230-MIP1D-37.5	Soil	26-May-11
K0941-05	80230-MIP1C-26	Soil	26-May-11

Soil and water samples were validated for analyses of semivolatile organics by the US EPA Region II data validation SOP (HW-22, Revision 3). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
  - Laboratory Blanks
  - Field Blank
  - Laboratory Control Sample
  - Surrogate Compound Recoveries
- \* - Internal Standard Recoveries
- \* - Matrix Spike / Matrix Spike Duplicate
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The minor problems with the laboratory control samples and calibrations should be noted. These are discussed in detail below.

## Holding Times

All samples were extracted and analyzed within the contractual and technical times required by the US EPA Region II protocols.

Continuous extraction of water samples for semivolatile analysis was started within 7 days of the date of collection and soil samples 14 days.

Extracts were analyzed within 40 days of the date of extraction.

## Tunes

No problems were detected with any of the tunes associated with the samples of this delivery group.

## Surrogate Recoveries

All surrogate recoveries were within the required limits with the following exceptions:

	A S1	A S2	B S3	B S4	A S5	B S6
MB-59483						136%
LCS-59483						139%
LCSD-59483						138%
K0941-01		80230-FB-110526				131%
MB-59499						127%

There should not be a problem with surrogate recoveries in method blanks, field blanks and laboratory control samples.

The data were not required to be qualified since the NYS DEC ASP requirements allow one surrogate in each fraction to be outside of the limits (as long as it is greater than 10%).

## Laboratory Control Samples

All of the recoveries of the LCS related to the field blank were within the required limits with the exceptions of 2-methylnaphthalene (117%) and caprolactam (18%).

2-Methylnaphthalene was not detected in the sample and the data were not affected by the high recovery.

The data for caprolactam were flagged with the "J" qualifier and are estimated values.

All of the recoveries of the LCS related to the soil samples were within the required limits with the exception of 2-methylnaphthalene (108%).

2-Methylnaphthalene was not detected in any of the samples and the data were not affected by the high recovery.

### **Matrix Spike / Matrix Spike Duplicate**

Sample K0941-02 / 80230-MIP3B-45 was used as the matrix spike and matrix spike duplicate.

All recoveries and RPDs were within the required limits.

### **Calibrations**

The %RSDs of all compounds in the initial calibration associated with the analysis of all of the samples were less than 15% with the exceptions of 2,4-dinitrophenol (26%) and benzaldehyde (18%).

None of these compounds were detected in any of the samples and the data were not required to be qualified for the high %RSD.

The %Ds of all compounds in the continuing calibration associated with the analysis of the field blank were less than 20% with the exceptions of nitrobenzene (21%), 2-methylnaphthalene (32%) and 2,4-dinitrophenol (35%).

The %Ds of all compounds in the continuing calibration associated with the analysis of the soil samples were less than 20% with the exceptions of nitrobenzene (27%), 2-methylnaphthalene (33%) and 2,4-dinitrophenol (43%)

The data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Method Blanks**

A low concentration of bis(2-ethylhexyl)phthalate (13 ug/l) was detected in the method blank associated with the analysis of the field blank.

No target compounds were detected in the soil method blank.

### **Field Blank**

A low concentration of bis(2-ethylhexyl)phthalate (5.9 ug/l) was detected in the field blank.

Only low concentrations of this compound, less than the CRDL were detected in these samples. All of the data for this compound were flagged with the "U" qualifier during the data validation.

### **Internal Standard Areas and Retention Times**

All internal standard recoveries and retention times were within the required limits.

### **Sample Results**

No problems were found with the results of any of the samples of this delivery group.

**SUMMARY OF THE ANALYTICAL DATA USABILITY**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Volatile Organic Analyses – Method 8260C**

**Samples Collected:** May 26, 2011

**Samples Received:** May 31, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K0941

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K0941-01	80230-FB-110526	Aqueous	26-May-11
K0941-02	80230-MIP3B-45	Soil	26-May-11
K0941-02 MS	80230-MIP3B-45 MS	Soil	26-May-11
K0941-02 MSD	80230-MIP3B-45 MSD	Soil	26-May-11
K0941-03	80230-MIP2B-40	Soil	26-May-11
K0941-04	80230-MIP1D-37.5	Soil	26-May-11
K0941-05	80230-MIP1C-26	Soil	26-May-11

Soil and water samples were validated for analyses of volatile organics by the US EPA Region II data validation SOP (HW-24, Revision 2). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
  - Laboratory Blanks
  - Trip Blanks
- \* - Field Blanks
- \* - System Monitoring Compound Recoveries
- \* - Internal Standard Recoveries
  - Matrix Spike / Matrix Spike Duplicate
  - Laboratory Control Samples
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The problems with the calibrations should be noted. These are described in detail below.

No other significant problems were found with this sample delivery group which would affect the usability of the data.

## **Holding Times**

The samples of this delivery group were validated against the Region II technical holding time requirements:

Soil and preserved aqueous samples were analyzed within 14 days of collection.

## **Tunes**

No problems were detected with the tunes associated with the samples of this delivery group.

## **System Monitoring Compound Recoveries**

All of the sample surrogate recoveries were within the required limits.

## **Calibrations**

### **Initial Calibrations**

All relative percent differences in the 6/03 initial calibration associated with the analyses of samples, -02, -04, and -05 were less than 20% with the exceptions of acetone (43%), trichloroethene (24%) and hexachlorobutadiene (22%).

The RRFs of acetone (0.043) and 2-butanone (0.027) were less than the 0.050 quality control limit in the above calibration.

All relative percent differences in the 6/2 initial calibration associated with the analyses of sample -01 were less than 20% with the exceptions of chloroethane (31%) and bromoform (29%).

The RRFs of acetone (0.023) and 2-butanone (0.026) were less than the 0.050 quality control limit in the above calibration.

Detected data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Continuing Calibrations**

All of the percent differences in the continuing calibration associated with the analysis of samples -02, -04, and -05 were less than 20% with the exception of 2-butanone (23%).

The relative response factors of acetone (0.044) and 2-butanone (0.033) in this continuing calibration were less than 0.050.

All of the percent differences in the continuing calibration associated with the analysis of sample -03 were less than 20%.

The relative response factors of acetone (0.038) and 2-butanone (0.031) in this continuing calibration were less than 0.050.

All of the percent differences in the continuing calibration associated with the analysis of sample -01 were less than 20% with the exceptions of acetone (21%), tetrachloroethene (21%).

The relative response factors of acetone (0.029) and 2-butanone (0.028) in this continuing calibration were less than 0.050.

The compounds with percent differences greater than 20% were flagged with the "J" qualifier and are estimated values.

Acetone and 2-butanone were not detected in any of the samples. The data for these compounds were flagged with the "R" qualifier and are technically rejected.

### **Matrix Spike and Matrix Spike Duplicate**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

Soil sample K0941-02 / 80230-MIP3B-45 was used as the matrix spike and matrix spike duplicate. All recoveries and RPDs were within the required limits with the exceptions of the acetone recoveries in the matrix spike (54%) and matrix spike duplicate (57%):

The data for acetone were technically rejected due to a low relative response factor.

### **Laboratory Control Sample**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

All of the laboratory control sample recoveries associated with the field blank were within the required limits with the exception of dichlorodifluoromethane (66%)

The data for this compound was flagged with the "J" qualifier and are estimated values.

All of the other LCS recoveries were within the required limits.

### **Method Blanks**

A low concentration of acetone (2.6J ug/kg) was detected in the method blank associated with soil samples -02, -04 and -05.

Only low concentrations of acetone, less than 10X the concentration in the method blank were detected in the samples. The data for acetone were previously technically rejected due to the low relative response factor.

A low concentration of hexachlorocyclopentadiene (1.2J ug/l) was detected in the method blank associated with the field blank.

This was not detected in the field blank and the blank contamination did not affect the use of the data.

No compounds were detected in the method blank associated with sample -03.

**Trip Blank**

A trip blank was not analyzed with this sample delivery group.

**Field Blank**

No target compounds were detected in the field blank.

**Internal Standard Areas and Retention Times**

The recoveries and retention times of all internal standards were within the required 50% - 100% quality control limits.

**Sample Results**

No problems were detected with the samples.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Total Metals & Cyanide Analyses**

**Samples Collected:** June 16<sup>th</sup> & 17<sup>th</sup>, 2011

**Samples Received:** June 22, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K1072

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K1072-01	80230-FB-110617	Aqueous	17-Jun-11
K1072-05	80230-MIP2C-35	Soil	16-Jun-11
K1072-06	80230-MIP5E-16	Soil	16-Jun-11

Soil and water samples were validated for inorganic analyses by the US EPA Region II data validation SOP (HW-2, Revision 13). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - Holding Times
- \* - Calibration Verification
  - CRDL Standard
  - \* - Laboratory Control Sample
    - Serial Dilutions
  - \* - Calibration Blanks
    - Field Blanks
  - \* - Preparation Blanks
    - Matrix Spike
    - Matrix Duplicate
  - \* - ICP Interference Check Sample
  - \* - Detection Limit Results
  - \* - Linear Range
  - \* - Sample Results

\* - Indicates that all criteria were met for this parameter.

**Data Validation Summary**

No problems were found that would affect the use of the data.

## **Holding Times**

All samples were analyzed within the required holding times.

## **CRDL Standards**

A CRDL standard was not analyzed with this sample delivery group.

## **Initial and Continuing Calibrations**

No problems were detected with any of the calibrations associated with this sample delivery group.

## **Preparation Blank**

No compounds were detected in the one preparation blank associated with the digestions of these samples at concentrations above the CRDL. Several analytes were found in the preparation blank at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table.

## **Calibration Blanks**

Several analytes were found in the continuing calibration blanks at concentrations between the CRDL and instrument detection limit. These very low concentrations are not required to be noted in the data validation summary table and do not affect the end use of the data.

## **Field Blank**

Low concentrations of several analytes were detected in the field blank. The concentrations of these in the sample were more than 10X the concentration in the field blank.

## **ICP Interference Check Sample**

No problems were detected with the reported ICP Interference Check Sample recoveries.

## **Matrix Spike Recovery**

A matrix spike was not analyzed for this sample delivery group.

## **Duplicate Analysis**

A matrix duplicate was not analyzed for this sample delivery group.

## **Laboratory Control Sample**

No problems were detected with the recoveries of the LCS standards.

### **Serial Dilutions**

A serial dilution was not analyzed for this sample delivery group.

### **Instrument Detection Limit**

No problems were found with the instrument detection limits.

### **ICP Linear Ranges**

No problems were detected with the linear ranges.

### **Sample Results**

No problems were detected with any of these samples.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil Pesticide Analyses**

**Samples Collected:** June 16<sup>th</sup> & 17<sup>th</sup>, 2011

**Samples Received:** June 22, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K1072

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K1072-05	80230-MIP2C-35	Soil	16-Jun-11
K1072-06	80230-MIP5E-16	Soil	16-Jun-11

Soil samples were validated for analyses of pesticides by the US EPA Region II data validation SOP (HW-44, Revision 1). Data were reviewed for usability according to the following criteria:

- Data Completeness
- \* - Holding Times
- \* - Laboratory Blanks
- \* - Field Blanks
- \* - Surrogate Recoveries
- \* - Surrogate Retention Times
- Matrix Spike / Matrix Spike Duplicate
- \* - Surrogate Spike Recovery
- \* - Laboratory Control Sample
  - Calibrations
- \* - Method Blanks
  - Florisil Cartridge Check
  - GPC Calibration
- \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

Form IX for a florisil cleanup was not included in the data package.

No significant problems were detected with any of the data.

**Holding Times**

All extractions and analyses were performed within the required holding times.

**Surrogate Recoveries**

All surrogate recoveries were within the required limits.

### **Matrix Spike**

A matrix spike was not analyzed with this sample delivery group.

### **Laboratory Control Samples**

All of the laboratory control sample recoveries were within the required limits.

### **Initial Calibrations**

No problems were detected with the initial calibrations associated with the analyses of the samples. All % RSDs were less than 20%.

### **Continuing Calibrations**

Several percent differences in the continuing calibrations were greater than 20%. (as high as 28%) In all cases, the calibration factors in the continuing calibration had increased over the mean in the initial calibration. None of the affected compounds were detected in the samples and the high percent differences did not affect the use of the data.

All other %Ds in the continuing calibrations directly with the sample were less than 20%.

### **Florisil Cartridge Check**

Form IX for a florisil cleanup was not included in the data package.

### **GPC Calibration**

A GPC cleanup was not performed on these samples.

### **Method Blanks**

No problems were detected with any of the method blanks.

### **Calibration Blanks**

No problems were detected with the calibration blanks associated with this sample delivery group.

### **Field Blank**

No compounds were detected in the field blank.

### **Sample Results**

No problems were detected with the sample data.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil PCB Analyses**

**Samples Collected:** June 16<sup>th</sup> & 17<sup>th</sup>, 2011

**Samples Received:** June 22, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K1072

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K1072-05	80230-MIP2C-35	Soil	16-Jun-11
K1072-06	80230-MIP5E-16	Soil	16-Jun-11

Soil samples were validated for analyses of PCBs by the US EPA Region II data validation SOP (HW-44, Revision 1). Data were reviewed for usability according to the following criteria:

- Data Completeness
- \* - Holding Times
- \* - Laboratory Blanks
- \* - Field Blanks
- \* - Surrogate Recoveries
- \* - Surrogate Retention Times
  - Matrix Spike / Matrix Spike Duplicate
- \* - System Monitoring Spike Recovery
- \* - Laboratory Control Sample
- \* - Calibrations
- \* - Method Blanks
  - GPC Calibration
- \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

Only compounds noted as target compounds by the laboratory's data system were reported in the raw data.

No other significant problems were detected with any of the data.

**Holding Times**

All extractions and analyses were performed within the required holding times.

**Surrogate Recoveries**

All surrogate recoveries were within the required limits.

### **Matrix Spike**

A matrix spike was not analyzed with this sample delivery group.

### **Laboratory Control Samples**

All of the laboratory control sample recoveries were within the required limits.

### **Initial Calibrations**

No problems were detected with the initial calibrations associated with the analyses of the samples. All %RSDs were less than 20%.

### **Continuing Calibrations**

All %Ds were less than 15%.

### **Florisil Cartridge Check**

Form IX for a florisil cleanup was not included in the data package.

### **GPC Calibration**

A GPC cleanup was not performed on these samples.

### **Method Blanks**

No problems were detected with any of the method blanks.

### **Calibration Blanks**

No problems were detected with the calibration blanks associated with this sample delivery group.

### **Field Blank**

No compounds were detected in the field blank.

### **Sample Results**

No problems were detected with the sample data.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Semivolatile Organic Analyses**

**Samples Collected:** June 16<sup>th</sup> & 17<sup>th</sup>, 2011

**Samples Received:** June 22, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K1072

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K1072-01	80230-FB-110617	Aqueous	17-Jun-11
K1072-02	80230-MIP4C-41	Soil	17-Jun-11
K1072-03	80230-MIP4B-45	Soil	17-Jun-11
K1072-03 MS	80230-MIP4B-45 MS	Soil	17-Jun-11
K1072-03 MSD	80230-MIP4B-45 MSD	Soil	17-Jun-11
K1072-04	80230-FB-110616	Aqueous	16-Jun-11
K1072-05	80230-MIP2C-35	Soil	16-Jun-11
K1072-06	80230-MIP5E-16	Soil	16-Jun-11

Soil and water samples were validated for analyses of semivolatile organics by the US EPA Region II data validation SOP (HW-22, Revision 3). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
- \* - Laboratory Blanks
  - Field Blank
  - Laboratory Control Sample
  - Surrogate Compound Recoveries
- \* - Internal Standard Recoveries
- \* - Matrix Spike / Matrix Spike Duplicate
- \* - Compound Identification
  - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

Percent moisture data for the soil samples were not included on the FORM I's. These were calculated from the percent moisture report (Page 567) during the data validation.

The minor problems with the matrix spike, laboratory control samples and calibrations should be noted. These are discussed in detail below.

### **Holding Times**

All samples were extracted and analyzed within the contractual and technical times required by the US EPA Region II protocols.

Continuous extraction of water samples for semivolatile analysis was started within 7 days of the date of collection and soil samples 14 days.

Extracts were analyzed within 40 days of the date of extraction.

### **Tunes**

No problems were detected with any of the tunes associated with the samples of this delivery group.

### **Surrogate Recoveries**

All surrogate recoveries were within the required limits with the following exceptions:

80230-FB-110617 (K1072-01B), recovery is above criteria for 2,4,6-Tribromophenol at 126% with criteria of (40-125).

80230-FB-110616 (K1072-04B), recovery is above criteria for 2,4,6-Tribromophenol at 126% with criteria of (40-125).

LCS-59922, recovery is above criteria for 2,4,6-Tribromophenol at 134% with criteria of (40-125).

LCSD-59922, recovery is above criteria for 2,4,6-Tribromophenol at 136% with criteria of (40-125).

MB-59922, recovery is above criteria for 2,4,6-Tribromophenol at 127% with criteria of (40-125).

There should not be a problem with surrogate recoveries in method blanks, field blanks and laboratory control samples.

The data were not required to be qualified since the NYS DEC ASP requirements allow one surrogate in each fraction to be outside of the limits (as long as it is greater than 10%).

### **Laboratory Control Samples**

All of the recoveries of the LCS related to the field blanks were within the required limits with the exceptions of 3,3'-dichlorobenzidine (115%) and caprolactam (14%).

3,3'-Dichlorobenzidine was not detected in either field blank and the data were not affected by the high recovery.

The data for caprolactam were flagged with the "J" qualifier and are estimated values.

All of the recoveries of the LCS related to the soil samples were within the required limits.

### **Matrix Spike / Matrix Spike Duplicate**

Sample K1072-03 / 80230-MIP4B-45 was used as the matrix spike and matrix spike duplicate.

All recoveries and RPDs were within the required limits with the exceptions of the RPDs for 4-chloroanaline (42%), pentachlorophenol (43%), 3,3'-Dichlorobenzidine (41%) and benzaldehyde (128%).

None of these compounds were detected in the soil samples and the high RPDs did not affect the use of the data.

### **Calibrations**

The %RSDs of all compounds in the initial calibration associated with the analysis of all of the samples were less than 15% with the exceptions of 2,4-dinitrophenol (26%) and benzaldehyde (18%).

None of these compounds were detected in any of the samples and the data were not required to be qualified for the high %RSD.

The %Ds of all compounds in the continuing calibration associated with the analysis of the field blanks were less than 20% with the exceptions of 2,4-dimethylphenol (24%) and 2,4-dinitrophenol (29%).

The %Ds of all compounds in the continuing calibration associated with the analysis of the soil samples were less than 20% with the exceptions of 2,4-dimethylphenol (23%), 2,4-dinitrophenol (28%) and bis (2-ethylhexyl)phthalate (21%)..

The data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Method Blanks**

No target compounds were detected in the method blanks.

### **Field Blank**

A low concentration of bis(2-ethylhexyl)phthalate (1.6 ug/l) was detected in the 6/17 field blank.

Only low concentrations of this compound, less than the CRDL were detected in these samples. All of the data for this compound were flagged with the "U" qualifier during the data validation.

No target compounds were detected in the 6/16 field blank.

### **Internal Standard Areas and Retention Times**

All internal standard recoveries and retention times were within the required limits.

## **Sample Results**

Percent moisture data for the soil samples were not included on the FORM I's. These were calculated from the percent moisture report (Page 567) during the data validation.

No other problems were found with the results of any of the samples of this delivery group.

**SUMMARY OF THE ANALYTICAL DATA USABILITY**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Volatile Organic Analyses – Method 8260C**

**Samples Collected:** June 16<sup>th</sup> & 17<sup>th</sup>, 2011

**Samples Received:** June 22, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K1072

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K1072-01	80230-FB-110617	Aqueous	17-Jun-11
K1072-02	80230-MIP4C-41	Soil	17-Jun-11
K1072-03	80230-MIP4B-45	Soil	17-Jun-11
K1072-03 MS	80230-MIP4B-45 MS	Soil	17-Jun-11
K1072-03 MSD	80230-MIP4B-45 MSD	Soil	17-Jun-11
K1072-04	80230-FB-110616	Aqueous	16-Jun-11
K1072-05	80230-MIP2C-35	Soil	16-Jun-11
K1072-05 DL	80230-MIP2C-35 DL	Soil	16-Jun-11
K1072-06	80230-MIP5E-16	Soil	16-Jun-11

Soil and water samples were validated for analyses of volatile organics by the US EPA Region II data validation SOP (HW-24, Revision 2). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
  - Laboratory Blanks
  - Trip Blanks
  - Field Blanks
- \* - System Monitoring Compound Recoveries
- \* - Internal Standard Recoveries
  - Matrix Spike / Matrix Spike Duplicate
  - Laboratory Control Samples
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The problems with the calibrations, laboratory control samples and blanks should be noted. These are described in detail below.

No other significant problems were found with this sample delivery group which would affect the usability of the data.

## Holding Times

The samples of this delivery group were validated against the Region II technical holding time requirements:

Soil and preserved aqueous samples were analyzed within 14 days of collection.

## Tunes

No problems were detected with the tunes associated with the samples of this delivery group.

## System Monitoring Compound Recoveries

All of the sample surrogate recoveries were within the required limits.

## Calibrations

### Initial Calibrations

All relative percent differences in the 6/10 initial calibration associated with the medium level analysis of sample K1072-05 / 80230-MIP2C-35 were less than 20% with the exceptions of acetone (23%), hexachlorobutadiene (28%) and 1,2,3-trichlorobenzene (25%).

The RRFs of acetone (0.022) and 2-butanone (0.022) were less than the 0.050 quality control limit in the above calibration.

All relative percent differences in the 6/22 initial calibration associated with the analyses of samples K1072-05 / 80230-MIP2C-35, K1072-06 / 80230-MIP5E-16, K1072-02 / 80230-MIP4C-41 and K1072-03 / 80230-MIP4B-45 were less than 20% with the exceptions of acetone (23%) and 1,2-dibromo-3-chloropropane (23%).

The RRFs of acetone (0.036) and 2-butanone (0.027) were less than the 0.050 quality control limit in the above calibration.

All relative percent differences in the 6/21 initial calibration associated with the analyses of the two field blanks were less than 20% with the exceptions of iodomethane (49%) and styrene (23%).

The RRFs of acetone (0.030) and 2-butanone (0.044) were less than the 0.050 quality control limit in the above calibration.

Detected data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Continuing Calibrations**

All of the percent differences in the 6/24 continuing calibration associated with the analysis of K1072-05 / 80230-MIP2C-35 and K1072-06 / 80230-MIP5E-16 were less than 20% with the exception of n-butylbenzene (22%).

The relative response factors of acetone (0.034) and 2-butanone (0.024) in this continuing calibration were less than 0.050.

All of the percent differences in the continuing calibration associated with the medium level analysis of sample K1072-05 / 80230-MIP2C-35 were less than 20% with the exceptions of dichlorodifluoromethane (27%), 1,2-dibromo-3-chloronethane (29%), 1,2,4-trichlorobenzene (26%), hexachlorobutadiene (31%), 1,2,3-trichlorobenzene (34%) and naphthalene (34%).

The relative response factors of acetone (0.019) and 2-butanone (0.019) in this continuing calibration were less than 0.050.

All of the percent differences in the 6/29 continuing calibration associated with the analysis of samples K1072-02 / 80230-MIP4C-41 and K1072-03 / 80230-MIP4B-45 were less than 20% with the exceptions of acetone (22%), 1,1-dichloropropane (26%), tetrachloroethene (35%), n-propylbenzene (25%) and hexachlorobutadiene (28%).

The relative response factors of acetone (0.032) and 2-butanone (0.028) in this continuing calibration were less than 0.050.

All of the percent differences in the 6/23 continuing calibration associated with the analyses of the two field blanks were less than 20% with the exceptions of dichlorodifluoromethane (22%), 1,1-dichloroethane (21%), acetone (22%) and tetrachloroethene (21%).

The relative response factors of acetone (0.036) and 2-butanone (0.044) in this continuing calibration were less than 0.050.

The compounds with percent differences greater than 20% were flagged with the "J" qualifier and are estimated values.

When acetone and 2-butanone were not detected in a sample, the data for these compounds were flagged with the "R" qualifier and are technically rejected.

When acetone and / or 2-butanone were detected in a sample, the data were flagged with the "J" qualifier and are estimated values.

### **Matrix Spike and Matrix Spike Duplicate**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

Soil sample K1072-03 / 80230-MIP4B-45 was used as the matrix spike and matrix spike duplicate. All recoveries and RPDs were within the required limits with the following exceptions

<b>Compound</b>	<b>MS</b>	<b>MSD</b>
1,2,3-Trichlorobenzene	54%	50%
1,2,4-Trichlorobenzene	53%	52%
2,2-Dichloropropane	55%	57%
2-Butanone	65%	
2-Hexanone	63%	59%
Acetone	67%	54%
Bromoform	55%	60%
cis-1,3-Dichloropropene	66%	
Dibromochloromethane	66%	67%
Hexachlorobutadiene	68%	
Methyl acetate		62%
n-Butylbenzene		69%
Naphthalene	64%	63%
Tetrachloroethene	173%	177%
trans-1,3-Dichloropropene	64%	65%
Vinyl acetate	40%	41%

The soil data for compounds with low recoveries were flagged with the "J" qualifier and are estimated values.

Tetrachloroethene was detected in all of the soil samples and the data were flagged with the "J" qualifier. The soil tetrachloroethene concentrations are estimated values.

### **Laboratory Control Sample**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

All of the laboratory control sample recoveries associated with the field blanks were within the required limits with the exception of dichlorodifluoromethane (63%)

The data for this compound were flagged with the "J" qualifier and are estimated values.

All of the laboratory control sample recoveries associated with samples K1072-05 / 80230-MIP2C-35 and K1072-06 / 80230-MIP5E-16 were within the required limits with the exception of hexachlorobutadiene (136%)

The compound was not detected in either of the two samples and the high recovery did not affect the use of the data.

All of the laboratory control sample recoveries associated with the medium level analysis of sample K1072-05 / 80230-MIP2C-3 were within the required limits with the exception of dichlorodifluoromethane (67%)

The data for this compound was flagged with the "J" qualifier and are estimated values.

All of the other LCS recoveries were within the required limits.

### **Method Blanks**

A low concentration of tetrachloroethene (3.3J ug/kg) was detected in the method blank associated with soil samples K1072-02 / 80230-MIP4C-41 and K1072-03 / 80230-MIP4B-45.

The concentration of tetrachloroethene in sample K1072-02 / 80230-MIP4C-41 (40 ug/l) was too high to be affected by the low level method blank contamination.

Only a low concentration of the tetrachloroethene (4 ug/l) was detected in sample K1072-03 / 80230-MIP4B-45.

The data for tetrachloroethene in this sample was flagged with the "U" qualifier and reported at the CRDL.

### **Trip Blank**

A trip blank was not analyzed with this sample delivery group.

### **Field Blank**

Acetone (6.8 ug/l) and methylene chloride (3.1 ug/) were detected in the 6/17 field blank K1072-04 / 80230-FB-110616.

Acetone (3.4 ug/l) was detected in the 6/16 field blank K1072-01 / 80230-FB-110617.

Low concentrations of these compounds, less than the CRDL were found in several samples. These were flagged with the "U" qualifier and reported at the CRDL/

### **Internal Standard Areas and Retention Times**

The recoveries and retention times of all internal standards were within the required 50% - 100% quality control limits.

### **Sample Results**

No problems were detected with the samples.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Semivolatile Organic Analyses**

**Samples Collected:** June 24, 2011

**Samples Received:** June 30, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K1157

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>
K1157-01	80230-FB-110624	Aqueous
K1157-02	80230-MIP5C-36.5	Soil

Soil and water samples were validated for analyses of semivolatile organics by the US EPA Region II data validation SOP (HW-22, Revision 3). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
- \* - Laboratory Blanks
- \* - Field Blank
  - Laboratory Control Sample
- \* - Surrogate Compound Recoveries
- \* - Internal Standard Recoveries
  - Matrix Spike / Matrix Spike Duplicate
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The minor problems with the laboratory control samples and calibrations should be noted. These are discussed in detail below.

## Holding Times

All samples were extracted and analyzed within the contractual and technical times required by the US EPA Region II protocols.

Continuous extraction of water samples for semivolatile analysis was started within 7 days of the date of collection and soil samples 14 days.

Extracts were analyzed within 40 days of the date of extraction.

## Tunes

No problems were detected with any of the tunes associated with the samples of this delivery group.

## Surrogate Recoveries

All surrogate recoveries were within the required limits.

## Laboratory Control Samples

All of the recoveries of the LCS related to the field blank were within the required limits with the following exceptions:

Compound	MS % Rec.	MS % Rec.
2,4-Dinitrophenol	168%	182%
2,4-Dinitrotoluene	135%	138%
2,6-Dinitrotoluene	123%	125%
2-Nitroaniline		117%
2-Nitrophenol	123%	126%
4,6-Dinitro-2-methylphenol	149%	155%

None of these compounds were detected in the sample and the data were not affected by the high recoveries.

All of the recoveries of the LCS related to the soil sample were within the required limits.

## Matrix Spike / Matrix Spike Duplicate

A matrix spike and matrix spike duplicate were not analyzed with this sample delivery group.

## Calibrations

The %RSDs of all compounds in the initial calibration associated with the analysis of sample K1157-02 / 80230-MIP5C-36.5 were less than 15% with the exceptions of 2,4-dinitrophenol (26%) and benzaldehyde (18%).

None of these compounds were detected in the sample and the data were not required to be qualified for the high %RSD.

Several compounds had %RSDs greater than 15% in the initial calibration associated with the analysis of the field blank.

No compounds were detected in the field blank and the high %RSDs do not affect the use of the data.

The %Ds of all compounds in the continuing calibration associated with the analysis of the soil sample were less than 20% with the following exceptions:

<b>Compound</b>	<b>%D</b>
2,2'-oxybis(1-Chloropropane)	22%
4-Nitroaniline	22%
4-Nitrophenol	28%
Atrazine	21%
Indeno(1,2,3-cd)pyrene	29%
N-Nitroso-di-n-propylamine	21%

The data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Method Blanks**

No target compounds were detected in the method blanks.

### **Field Blank**

No compounds were detected in the field blank.

### **Internal Standard Areas and Retention Times**

All internal standard recoveries and retention times were within the required limits.

### **Sample Results**

No problems were found with the results of any of the samples of this delivery group.

**SUMMARY OF THE ANALYTICAL DATA USABILITY**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Volatile Organic Analyses – Method 8260C**

**Samples Collected:** June 24, 2011

**Samples Received:** June 30, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K1157

**Laboratory Reference Numbers:**

Lab ID	Client ID	Matrix
K1157-01	80230-FB-110624	Aqueous
K1157-02	80230-MIP5C-36.5	Soil
K1157-03	80230-MIP6B-42.5	Soil
K1157-04	80230-MW12S-35	Soil
K1157-04 DL	80230-MW12S-35 DL	Soil
K1157-05	80230-DUP3-50	Soil

Soil and water samples were validated for analyses of volatile organics by the US EPA Region II data validation SOP (HW-24, Revision 2). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
  - Laboratory Blanks
  - Trip Blanks
  - Field Blanks
- \* - System Monitoring Compound Recoveries
- \* - Internal Standard Recoveries
  - Matrix Spike / Matrix Spike Duplicate
  - Laboratory Control Samples
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The problems with the calibrations, laboratory control samples and blanks should be noted. These are described in detail below.

No other significant problems were found with this sample delivery group which would affect the usability of the data.

## **Holding Times**

The samples of this delivery group were validated against the Region II technical holding time requirements:

Soil and preserved aqueous samples were analyzed within 14 days of collection.

## **Tunes**

No problems were detected with the tunes associated with the samples of this delivery group.

## **System Monitoring Compound Recoveries**

All of the sample surrogate recoveries were within the required limits.

## **Calibrations**

### **Initial Calibrations**

All relative percent differences in the 6/22 initial calibration were less than 20% with the exception of 1,2-dibromo-3-chloropropane (23%). This initial calibration was associated with the analyses of the following samples:

K1157-02	80230-MIP5C-36.5
K1157-03	80230-MIP6B-42.5
K1157-04	80230-MW12S-35
K1157-05	80230-DUP3-50

The RRFs of acetone (0.036) and 2-butanone (0.027) were less than the 0.050 quality control limit in the above calibration.

All relative percent differences in the 7/06 initial calibration associated with the analyses of samples K1157-01 / 80230-FB-110624 and K1157-04 DL / 80230-MW12S-35 DL were less than 20%.

The RRFs of acetone (0.035) and 2-butanone (0.038) were less than the 0.050 quality control limit in the above calibration.

Detected data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Continuing Calibrations**

All of the percent differences in the 6/30 continuing calibration associated with the analysis samples K1157-04 / 80230-MW12S-35 and K1157-05 / 80230-DUP3-50 were less than 20% with the exceptions of n-butylbenzene (22%) and hexachlorobutadiene (34%).

The relative response factors of acetone (0.034) and 2-butanone (0.026) in this continuing calibration were less than 0.050.

All of the percent differences in the 6/30 continuing calibration associated with the analysis samples K1157-02 / 80230-MIP5C-36.5 and K1157-03 / 80230-MIP6B-42.5 were less than 20%.

The relative response factors of acetone (0.035) and 2-butanone (0.026) in this continuing calibration were less than 0.050.

All of the percent differences in the continuing calibration associated with the level analysis of samples K1157-01 / 80230-FB-110624 and K1157-04 DL / 80230-MW12S-35 DL were less than 20% with the exceptions of acetone (44%), 2-butanone (29%) and 2-hexanone (31%)

The relative response factor of 2-butanone (0.049) in this continuing calibration was less than 0.050.

The compounds with percent differences greater than 20% were flagged with the "J" qualifier and are estimated values.

When acetone and or 2-butanone, with RRFs less than 0.050, were not detected in a sample, the data for these compounds were flagged with the "R" qualifier and are technically rejected.

When acetone and / or 2-butanone, with RRFs less than 0.050, were detected in a sample, the data were flagged with the "J" qualifier and are estimated values.

### **Matrix Spike and Matrix Spike Duplicate**

A matrix spike was not analyzed with this sample delivery group.

### **Laboratory Control Sample**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

All of the laboratory control sample recoveries associated with the medium level analysis of sample K1157-04 / 80230-MW12S-35 were within the required limits with the exception of acetone (137%).

The data for this compound were flagged with the "J" qualifier and are estimated values.

All of the other LCS recoveries were within the required limits.

### **Method Blanks**

Low concentrations of carbon disulfide (0.67J ug/kg) and naphthalene (0.60 ug/kg) was detected in the method blank associated with the analyses of the field blank.

Neither of these compounds were detected in the sample and the blank contamination did not affect the use of the data.

A low concentration of naphthalene (56J ug/kg) was detected in the method blank associated with the medium level analysis of sample K1157-04 / 80230-MW12S-35.

This compound was not detected in the sample and the blank contamination did not affect the use of the data.

### **Trip Blank**

A trip blank was not analyzed with this sample delivery group.

### **Field Blank**

Acetone (2.68 ug/l) and methylene chloride (1.9 ug/l) were detected in the field blank.

Neither of these compounds were detected in the samples and the data were not affected by the blank contamination.

### **Internal Standard Areas and Retention Times**

The recoveries and retention times of all internal standards were within the required 50% - 100% quality control limits.

### **Sample Results**

No problems were detected with the samples.

**SUMMARY OF THE ANALYTICAL DATA USABILITY**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Methane, Ethane and Ethylene Analyses**

**Samples Collected: March 5<sup>th</sup> through 8<sup>th</sup>, 2012**

**Samples Received: March 9<sup>th</sup> through 9<sup>th</sup>, 2012**

**Laboratory: Spectrum**

**Sample Delivery Group: L0441**

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Date Sampled</b>
L0441-02	80230-MW09D-030512	3/5/2012
L0441-02MS	80230-MW09D-030512MS	3/5/2012
L0441-02MSD	80230-MW09D-030512MSD	3/5/2012
L0441-03	80230-MW09S-030512	3/5/2012
L0441-04	80230-FB-030512	3/5/2012
L0441-05	80230-MW10D-030512	3/5/2012
L0441-06	80230-MW10S-030512	3/5/2012
L0441-08	80230-MW16S-030612	3/6/2012
L0441-09	80230-MW01-030612	3/6/2012
L0441-10	80230-FB-030612	3/6/2012
L0441-11	80230-MW04-030612	3/6/2012
L0441-12	80230-MW02-030612	3/6/2012
L0441-14	80230-MW15D-030712	3/7/2012
L0441-15	80230-MW13S-030712	3/7/2012
L0441-16	80230-MW13D-030712	3/7/2012
L0441-17	80230-FB-030712	3/7/2012
L0441-18	80230-MW08S-030712	3/7/2012
L0441-19	80230-MW03S-030712	3/7/2012
L0441-21	80230-MW11D-030812	3/8/2012
L0441-22	80230-MW14S-030812	3/8/2012
L0441-23	80230-MW11S-030812	3/8/2012
L0441-24	80230-FB-030812	3/8/2012
L0441-25	80230-MW12S-030812	3/8/2012
L0441-26	80230-MW112S-030812	3/8/2012

Water samples were received for analyses of ethane, ethene and methane by US EPA Region II protocols. A complete analytical validation was performed based upon the following parameters:

- \* - Data Completeness
- \* - Holding Times
- Field Blanks
- \* - Matrix Spike / Matrix Spike Duplicate
- \* - Laboratory Control Sample Recovery
- \* - Method Blank
- \* - Calibrations
- \* - Compound Identification

\* - Indicates that all criteria were met for this parameter.

## DATA USABILITY SUMMARY

The methane and ethane contamination in some of the field blanks should be noted. These are described in detail below.

No other problems were found that would affect that would affect the use of the data.

### Holding Times

All samples were analyzed within 14 days of sample collection..

### System Monitoring Compound Recoveries

Surrogates were not used for this analysis.

### Calibrations

All of the %RSDs in the initial calibration were less than 20%.

The laboratory noted in a supplemental email that the initial calibration standards were 5 ug/l, 2,000 ug/l, 4,000 ug/l and 10,000 ug/l. The concentrations were not included in the raw data.

All of the percent differences in the continuing calibrations were less than 20%.

### Matrix Spike / Matrix Spike Duplicate

Sample L0441-02 / 80230-MW09D-030512 was used as the matrix spike and matrix spike duplicate. All recoveries and RPDs were within the required limits.

### Laboratory Control Sample

The recoveries of the LCS standard were within the 75% - 125% quality assurance limits.

### Field Blanks

Methane (2.8 ug/l) and ethane (120 ug/l) were detected in the L0441-10 / 80230-FB-030612 field blank.

Methane (2.2 ug/l) was detected in the L0441-17 / 80230-FB-030712 field blank.

Methane (1.8 ug/l) was detected in the L0441-24 / 80230-FB-030812 field blank.

Concentrations of these compounds in the associated samples less than five times the concentration in the corresponding blanks were flagged with the "U" qualifier.

### Method Blanks

No compounds were detected in any of the method blanks.

## **Sample Results**

No problems were found with the reported results of any of the samples of this delivery group.

**SUMMARY OF THE ANALYTICAL DATA USABILITY**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Water Volatile Organic Analyses – Method 8260C**

**Samples Collected: March 5<sup>th</sup> through 8<sup>th</sup>, 2012**

**Samples Received: March 9<sup>th</sup> through 9<sup>th</sup>, 2012**

**Laboratory: Spectrum**

**Sample Delivery Group: L0441**

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Date Sampled</b>
L0441-01	80230-TB-030512	3/5/2012
L0441-02	80230-MW09D-030512	3/5/2012
L0441-02MS	80230-MW09D-030512MS	3/5/2012
L0441-02MSD	80230-MW09D-030512MSD	3/5/2012
L0441-03	80230-MW09S-030512	3/5/2012
L0441-04	80230-FB-030512	3/5/2012
L0441-05	80230-MW10D-030512	3/5/2012
L0441-06	80230-MW10S-030512	3/5/2012
L0441-07	80230-TB-030612	3/6/2012
L0441-08	80230-MW16S-030612	3/6/2012
L0441-08DL	80230-MW16S-030612DL	3/6/2012
L0441-09	80230-MW01-030612	3/6/2012
L0441-09DL	80230-MW01-030612DL	3/6/2012
L0441-10	80230-FB-030612	3/6/2012
L0441-11	80230-MW04-030612	3/6/2012
L0441-12	80230-MW02-030612	3/6/2012
L0441-13	80230-TB-030712	3/7/2012
L0441-14	80230-MW15D-030712	3/7/2012
L0441-15	80230-MW13S-030712	3/7/2012
L0441-15DL	80230-MW13S-030712DL	3/7/2012
L0441-16	80230-MW13D-030712	3/7/2012
L0441-17	80230-FB-030712	3/7/2012
L0441-18	80230-MW08S-030712	3/7/2012
L0441-18DL	80230-MW08S-030712DL	3/7/2012
L0441-19	80230-MW03S-030712	3/7/2012
L0441-20	80230-TB-030812	3/8/2012
L0441-21	80230-MW11D-030812	3/8/2012
L0441-22	80230-MW14S-030812	3/8/2012
L0441-22DL	80230-MW14S-030812DL	3/8/2012
L0441-23	80230-MW11S-030812	3/8/2012
L0441-23DL	80230-MW11S-030812DL	3/8/2012
L0441-24	80230-FB-030812	3/8/2012
L0441-25	80230-MW12S-030812	3/8/2012
L0441-25DL	80230-MW12S-030812DL	3/8/2012
L0441-26	80230-MW112S-030812	3/8/2012
L0441-26DL	80230-MW112S-030812DL	3/8/2012

Water samples were validated for analyses of volatile organics by the US EPA Region II data validation SOP (HW-24, Revision 2). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
- Calibrations
- \* - Laboratory Blanks
- \* - Trip Blank
- Field Blank
- \* - System Monitoring Compound Recoveries
- \* - Internal Standard Recoveries
- Matrix Spike / Matrix Spike Duplicate
- Laboratory Control Samples
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

## **DATA VALIDATION SUMMARY**

The problems with the calibrations, matrix spikes, laboratory control samples and blanks should be noted. These are described in detail below.

No other significant problems were found with this sample delivery group which would affect the usability of the data.

### **Holding Times**

The samples of this delivery group were validated against the Region II technical holding time requirements:

Soil and preserved aqueous samples were analyzed within 14 days of collection.

### **Tunes**

No problems were detected with the tunes associated with the samples of this delivery group.

### **System Monitoring Compound Recoveries**

All of the sample surrogate recoveries were within the required limits.

### **Calibrations**

#### **Initial Calibrations**

All relative percent differences in the 3/12 initial calibration (Instrument V1) were less than 20.

The RRFs of acetone (0.042) and 2-butanone (0.040) were less than the 0.050 quality control limit in the above calibration.

This calibration was associated with the analyses of the following samples:

L0441-08DL	80230-MW16S-030612DL
L0441-09DL	80230-MW01-030612DL
L0441-10	80230-FB-030612
L0441-11	80230-MW04-030612
L0441-15DL	80230-MW13S-030712DL
L0441-16	80230-MW13D-030712
L0441-17	80230-FB-030712
L0441-18DL	80230-MW08S-030712DL
L0441-19	80230-MW03S-030712
L0441-20	80230-TB-030812
L0441-21	80230-MW11D-030812
L0441-22	80230-MW14S-030812
L0441-23	80230-MW11S-030812
L0441-25	80230-MW12S-030812
L0441-26	80230-MW112S-030812

All relative percent differences in the 3/16 initial calibration (Instrument V1) were less than 20% with the exceptions of acetone (24%), methylene chloride (21%) and naphthalene (30%).

The RRF of 2-butanone (0.038) was less than the 0.050 quality control limit in the above calibration.

This initial calibration was associated with the analyses of the following samples:

L0441-22DL	80230-MW14S-030812DL
L0441-23DL	80230-MW11S-030812DL
L0441-24	80230-FB-030812
L0441-25DL	80230-MW12S-030812DL
L0441-26DL	80230-MW112S-030812DL

All relative percent differences in the 3/7 initial calibration (Instrument V6) were less than 20% with the exceptions of bromomethane (36%), iodomethane (21%), 1,2-dichlorobenzene (21%), hexachlorobutadiene (23%), 1,2,3-trichlorobenzene (23%) and naphthalene (28%).

The RRFs of acetone (0.032) and 2-butanone (0.030) were less than the 0.050 quality control limit in the above calibration.

This initial calibration was associated with the analyses of the following samples:

L0441-01	80230-TB-030512
L0441-07	80230-TB-030612

All relative percent differences in the 3/7 initial calibration (Instrument V10) were less than 20% with the exceptions of bromomethane (36%) and iodomethane (54%).

The RRFs of acetone (0.048) and 2-butanone (0.043) were less than the 0.050 quality control limit in the above calibration.

This initial calibration was associated with the analyses of the following samples:

L0441-02	80230-MW09D-030512
L0441-03	80230-MW09S-030512
L0441-04	80230-FB-030512
L0441-05	80230-MW10D-030512
L0441-06	80230-MW10S-030512
L0441-08	80230-MW16S-030612
L0441-09	80230-MW01-030612
L0441-12	80230-MW02-030612
L0441-13	80230-TB-030712
L0441-14	80230-MW15D-030712
L0441-15	80230-MW13S-030712
L0441-18	80230-MW08S-030712

Detected data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Continuing Calibrations**

All relative percent differences in the 3/13 continuing calibration were less than 20%.

The RRFs of acetone (0.036) and 2-butanone (0.036) were less than the 0.050 quality control limit in the above calibration.

This continuing calibration was associated with the analyses of the following samples:

L0441-08DL	80230-MW16S-030612DL
L0441-09DL	80230-MW01-030612DL
L0441-10	80230-FB-030612
L0441-11	80230-MW04-030612
L0441-15DL	80230-MW13S-030712DL
L0441-16	80230-MW13D-030712
L0441-17	80230-FB-030712
L0441-18DL	80230-MW08S-030712DL
L0441-19	80230-MW03S-030712

All percent differences in the 3/14 continuing calibration were less than 20% with the exceptions of dichlorodifluoromethane (24%), vinyl chloride (21%), chloroethane (22%), acetone (43%), 2-butanone (32%), 2,2-dichloropropane (25%), 2-hexanone (29%) and naphthalene (22%).

The RRFs of acetone (0.024) and 2-butanone (0.027) were less than the 0.050 quality control limit in the above calibration.

This continuing calibration was associated with the analyses of the following samples:

L0441-20	80230-TB-030812
L0441-21	80230-MW11D-030812
L0441-22	80230-MW14S-030812
L0441-23	80230-MW11S-030812
L0441-25	80230-MW12S-030812
L0441-26	80230-MW112S-030812

All percent differences in the 3/16 continuing calibration were less than 20% with the exceptions of 2,2-dichloropropane (26%), 1,1,1-trichloroethane (26%), carbon tetrachloride (28%) and 1,2,3-trichloropropane (24%).

The RRF of 2-butanone (0.043) were less than the 0.050 quality control limit in the above calibration.

This continuing calibration was associated with the analyses of the following samples:

L0441-22DL	80230-MW14S-030812DL
L0441-23DL	80230-MW11S-030812DL
L0441-24	80230-FB-030812
L0441-25DL	80230-MW12S-030812DL
L0441-26DL	80230-MW112S-030812DL

All percent differences in the 3/7 continuing calibration were less than 20%.

The RRFs of acetone (0.039) and 2-butanone (0.034) were less than the 0.050 quality control limit in the above calibration.

This continuing calibration was associated with the analyses of the following samples:

L0441-01	80230-TB-030512
L0441-07	80230-TB-030612

All percent differences in the 3/9 continuing calibration were less than 20% with the exceptions of iodomethane (70%), 1,2-dibromo-3-chloropropane (21%), 1,2,3-trichlorobenzene (24%) and naphthalene (24%).

The RRFs of acetone (0.045) and 2-butanone (0.041) were less than the 0.050 quality control limit in the above calibration.

This continuing calibration was associated with the analyses of the following samples:

L0441-02	80230-MW09D-030512
L0441-03	80230-MW09S-030512
L0441-04	80230-FB-030512
L0441-05	80230-MW10D-030512
L0441-06	80230-MW10S-030512

L0441-08	80230-MW16S-030612
L0441-09	80230-MW01-030612
L0441-12	80230-MW02-030612
L0441-13	80230-TB-030712
L0441-14	80230-MW15D-030712
L0441-15	80230-MW13S-030712
L0441-18	80230-MW08S-030712

When acetone and / or 2-butanone with a low relative response factor were detected in a sample, they were flagged with the "J" qualifier.

When these compounds were not detected in a sample they were flagged with the "R" qualifier and are technically rejected.

The compounds with percent differences greater than 20% were flagged with the "J" qualifier and are estimated values.

### **Matrix Spike and Matrix Spike Duplicate**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

Soil sample L0441-02 / 80230-MW09D-030512 was used as the matrix spike and matrix spike duplicate. All recoveries and RPDs were within the required limits with the following exceptions:

<b>Compound</b>	<b>MS % Rec</b>	<b>MSD % Rec</b>	<b>RPD</b>
1,1,2-Trichloro-1,2,2-trifluoroethane	69%65%		
2,2-Dichloropropane	66%62%		
2-Butanone	65%		
2-Hexanone	61%66%		
Acetone	44%46%		
Chloroethane	62%60%		
Chloromethane	60%63%		
Cyclohexane	66%65%		
Dichlorodifluoromethane	63%65%		
Methyl acetate	67%64%		
Methylcyclohexane	68%63%		
Naphthalene	59%		
Vinyl chloride	64%64%		

The data for compounds with low recoveries were flagged with the "J" qualifier and are estimated values.

An RPD of 30% was used for the purposes of the validation.

### **Laboratory Control Sample**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

All of the laboratory control sample recoveries in LCS-65104 / LCSD-65104, were within the required limits with the exception of acetone (57%). These laboratory control samples were associated with the following samples:

Lab ID	Client ID
L0441-20	80230-TB-030812
L0441-21	80230-MW11D-030812
L0441-22	80230-MW14S-030812
L0441-23	80230-MW11S-030812
L0441-25	80230-MW12S-030812
L0441-26	80230-MW112S-030812

The data for these compounds were flagged with the "J" qualifier and are estimated values.

All of the laboratory control sample recoveries in LCS-65160 were within the required limits with the exceptions of iodomethane (131%), 2,2-dichloropropane (152%), 1,1,1-trichloroethane (144%) and carbon tetrachloride (155%). This laboratory control sample was associated with the following samples:

Lab ID	Client ID
L0441-22DL	80230-MW14S-030812DL
L0441-23DL	80230-MW11S-030812DL
L0441-24	80230-FB-030812
L0441-25DL	80230-MW12S-030812DL
L0441-26DL	80230-MW112S-030812DL

None of these compounds were detected in any of these samples and the data were not affected by the high recoveries. The data were not required to be qualified.

All of the other LCS recoveries were within the required limits.

### **Method Blanks**

No compounds were detected in any of the method blanks.

### **Trip Blank**

No compounds were detected in the trip blanks.

### **Field Blank**

Low concentrations of methylene chloride were detected in most of the field blanks.

This compound was not detected in any of the samples and the field blank contamination does not affect the use of the data.

A low concentration of trichloroethene was detected in field blank 80230-FB-030612.

Trichloroethene was detected in samples 80230-MW04-030612 (L0441-11) and 80230-MW02-030612(L0441-12) at concentrations of 2.5 ug/l. The

trichloroethene data in these samples were flagged with the "U" qualifier and reported at the CRDL.

The trichloroethene concentrations in the other associated samples were too high to be affected by the field blank contamination.

A low concentration of tetrachloroethene (4.5 J ug/l) was detected in field blank 80230-FB-030812.

The trichloroethene concentrations in the associated samples were too high to be affected by the field blank contamination.

### **Internal Standard Areas and Retention Times**

The recoveries and retention times of all internal standards were within the required 50% - 100% quality control limits.

### **Sample Results**

No problems were detected with the samples.

**SUMMARY OF THE ANALYTICAL DATA VALIDATION**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Wet Chemistry Analyses**

**Samples Collected:** March 5<sup>th</sup> through 8<sup>th</sup>, 2012

**Samples Received:** March 9<sup>th</sup> through 9<sup>th</sup>, 2012

**Laboratory:** Spectrum

**Sample Delivery Group:** L0441

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Date Sampled</b>
L0441-02	80230-MW09D-030512	3/5/2012
L0441-02MS	80230-MW09D-030512MS	3/5/2012
L0441-02MSD	80230-MW09D-030512MD	3/5/2012
L0441-03	80230-MW09S-030512	3/5/2012
L0441-04	80230-FB-030512	3/5/2012
L0441-05	80230-MW10D-030512	3/5/2012
L0441-06	80230-MW10S-030512	3/5/2012
L0441-08	80230-MW16S-030612	3/6/2012
L0441-09	80230-MW01-030612	3/6/2012
L0441-10	80230-FB-030612	3/6/2012
L0441-11	80230-MW04-030612	3/6/2012
L0441-12	80230-MW02-030612	3/6/2012
L0441-14	80230-MW15D-030712	3/7/2012
L0441-15	80230-MW13S-030712	3/7/2012
L0441-16	80230-MW13D-030712	3/7/2012
L0441-17	80230-FB-030712	3/7/2012
L0441-18	80230-MW08S-030712	3/7/2012
L0441-19	80230-MW03S-030712	3/7/2012
L0441-21	80230-MW11D-030812	3/8/2012
L0441-22	80230-MW14S-030812	3/8/2012
L0441-23	80230-MW11S-030812	3/8/2012
L0441-24	80230-FB-030812	3/8/2012
L0441-25	80230-MW12S-030812	3/8/2012
L0441-26	80230-MW112S-030812	3/8/2012

Water samples were received for analyses of the wet chemistry analyte list by NYS DEC ASP protocols. A complete analytical validation was performed based upon the following parameters:

- \* - Chloride
- \* - Ammonia
- \* - Nitrate / Nitrite
- \* - Sulfide
- \* - Sulfate
- \* - Total Alkalinity
- \* - Total Dissolved Solids
- \* - Total Suspended Solids
- \* - Total Kjeldahl Nitrogen
- \* - Total Organic Carbon
- \* - Total Hardness

\* - Indicates that all criteria were met for this parameter.

## DATA VALIDATION SUMMARY

The problems with the field blanks and matrix spikes should be noted. These are described in detail below.

### Holding Times

All samples were analyzed within the required holding times.

### Initial and Continuing Calibrations

No problems were detected with any of the calibrations associated with this sample delivery group.

### Preparation Blank

No analytes were detected in any of the preparation blanks.

### Calibration Blanks

No analytes were detected in any of the calibration blanks.

### Field Blank

A low concentration of nitrate 0.032J mg/l was detected in field blank L0441-04 / 80230-FB-030512.

Concentrations of nitrate less than 10X the concentration in the field blank in associated samples were flagged with the "U" qualifier and reported at the CRDL.

A low concentration of nitrate 0.019J mg/l was detected in field blank L0441-17 / 80230-FB-030712

Concentrations of nitrate less than 10X the concentration in the field blank were flagged with the "U" qualifier and reported at the CRDL.

A low concentration of chloride (0.50J mg/l) was detected in field blank L0441-10 / 80230-FB-030612.

All of the chloride concentrations in the samples were too high to be affected by the field blank contamination.

### Matrix Spike and Matrix Spike Duplicate Recovery

Sample L0441-02 / 80230-MW09D-030512 was used for the designated matrix spike for the sample delivery group.

All recoveries were within the 75% - 125% quality control limits used for the validation with the exceptions of ammonia and TKN which were not recovered in either the matrix spike or matrix spike duplicate (0%).

The data for ammonia and TKN were flagged with the "R" qualifier and technically rejected.

### **Duplicate Analysis**

All matrix duplicate RPDs were less than 20%.

### **Laboratory Control Sample**

No problems were detected with the recoveries of the LCS standards.

### **Linear Ranges**

No problems were detected with the linear ranges. The reported concentrations of all samples in this delivery group were within their linear range for each analyte.

### **Sample Results**

No other problems were detected with any of the samples.

**SUMMARY OF THE ANALYTICAL DATA USABILITY**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Volatile Organic Analyses – Method 8260C**

**Samples Collected:** November 1, 2011

**Samples Received:** November 4, 2011

**Laboratory:** Spectrum

**Sample Delivery Group:** K2296

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>
K2296-01	PM-TB-1112011	Aqueous
K2296-02 PM-	FB-SO-1112011	Aqueous
K2296-03 PM-	SB-1a-A-1112011	Soil
K2296-04 PM-	SB-1a-B-1112011	Soil
K2296-05 PM-	SB-2-A-1112011	Soil
K2296-06 PM-	SB-2-B-1112011	Soil
K2296-07 PM-	SB-3-A-1112011	Soil
K2296-08 PM-	SB-4-A-1112011	Soil
K2296-09 PM-	SB-5-A-1112011	Soil
K2296-10 PM-	SB-5-B-1112011	Soil
K2296-11 PM-	SB-6-A-1112011	Soil
K2296-12 PM-	SB-6-B-1112011	Soil

Soil and water samples were validated for analyses of volatile organics by the US EPA Region II data validation SOP (HW-24, Revision 2). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
  - Laboratory Blanks
- \* - Trip Blank
- \* - Field Blank
- \* - System Monitoring Compound Recoveries
- \* - Internal Standard Recoveries
  - Matrix Spike / Matrix Spike Duplicate
  - Laboratory Control Samples
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

**DATA VALIDATION SUMMARY**

The problems with the calibrations, matrix spikes, laboratory control samples and method blanks should be noted. These are described in detail below.

No other significant problems were found with this sample delivery group that would affect the usability of the data.

### **Holding Times**

The samples of this delivery group were validated against the Region II technical holding time requirements:

Soil and preserved aqueous samples were analyzed within 14 days of collection.

### **Tunes**

No problems were detected with the tunes associated with the samples of this delivery group.

### **System Monitoring Compound Recoveries**

All of the sample surrogate recoveries were within the required limits.

### **Calibrations**

#### **Initial Calibrations**

All relative percent differences in the 11/10 initial calibration associated with the analyses of the field and trip blanks were less than 20% with the exceptions of hexachlorobutadiene (21%) and 1,2,3-trichlorobenzene (23%).

The RRFs of acetone (0.036) and 2-butanone (0.028) were less than the 0.050 quality control limit in the above calibration.

All relative percent differences in the 11/10 initial calibration were less than 20% with the exceptions of acetone (35%), iodomethane (33%) and hexachlorobutadiene (22%). This initial calibration was associated with the analyses of the following samples:

K2296-04	PM-SB-1a-B-1112011
K2296-05	PM-SB-2-A-1112011
K2296-06	PM-SB-2-B-1112011
K2296-07	PM-SB-3-A-1112011
K2296-08	PM-SB-4-A-1112011
K2296-10	PM-SB-5-B-1112011
K2296-11	PM-SB-6-A-1112011
K2296-12	PM-SB-6-B-1112011

The RRFs of acetone (0.032) and 2-butanone (0.034) were less than the 0.050 quality control limit in the above calibration.

All relative percent differences in the 11/14 initial calibration were less than 20% with the exceptions of acetone (31%), toluene (39%) and naphthalene (23%). This initial calibration was associated with the analyses of samples K2296-09 / PM-SB-5-A-1112011 and K2296-03 / PM-SB-1a-A-1112011.

The RRFs of acetone (0.032) and 2-butanone (0.032) were less than the 0.050 quality control limit in the above calibration.

Detected data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Continuing Calibrations**

All relative percent differences in the 11/11 continuing calibration associated with the analyses of the field and trip blanks were less than 20% with the exceptions of acetone (47%), tetrachloroethene (35%), 2-hexanone (22%), hexachlorobutadiene (27%), 1,2,3-trichlorobenzene (22%), and naphthalene (21%).

The RRFs of acetone (0.036) and 2-butanone (0.023) were less than the 0.050 quality control limit in the above calibration.

All percent differences in the 11/12 continuing calibration were less than 20% with the exceptions of iodomethane (33%), carbon tetrachloride (24%) and naphthalene (22%). This continuing calibration was associated with the analyses of the following samples:

K2296-04	PM-SB-1a-B-1112011
K2296-05	PM-SB-2-A-1112011
K2296-06	PM-SB-2-B-1112011
K2296-07	PM-SB-3-A-1112011
K2296-08	PM-SB-4-A-1112011
K2296-10	PM-SB-5-B-1112011
K2296-11	PM-SB-6-A-1112011
K2296-12	PM-SB-6-B-1112011

The RRFs of acetone (0.032) and 2-butanone (0.033) were less than the 0.050 quality control limit in the above calibration.

All percent differences in the 11/14 continuing calibration were less than 20% with the exception of methylcyclohexane (21%). This continuing calibration was associated with the analyses of sample K2296-09 / PM-SB-5-A-1112011.

The RRFs of acetone (0.034) and 2-butanone (0.035) were less than the 0.050 quality control limit in the above calibration.

All percent differences in the 11/15 continuing calibration were less than 20% with the exception of iodomethane (48%). This continuing calibration was associated with the analyses of sample K2296-03 / PM-SB-1a-A-1112011.

The RRFs of acetone (0.027) and 2-butanone (0.035) were less than the 0.050 quality control limit in the above calibration.

When acetone and/ or 2-butanone were detected in a sample, they were flagged with the "J" qualifier.

When these compounds were not detected in a sample they were flagged with the "R" qualifier and are technically rejected.

The compounds with percent differences greater than 20% were flagged with the "J" qualifier and are estimated values.

### **Matrix Spike and Matrix Spike Duplicate**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

Soil sample K2296-09 / PM-SB-5-A-1112011 was used as the matrix spike and matrix spike duplicate. All recoveries and RPDs were within the required limits with the following exceptions:

<b>Compound</b>	<b>MS</b>	<b>MSD</b>	<b>%RSD</b>
1,2,4-Trichlorobenzene	44%	61%	31%
1,2-Dibromo-3-chloropropane	67%		
1,2-Dichlorobenzene	67%		
1,3-Dichlorobenzene	68%		
1,4-Dichlorobenzene	62%		
2-Hexanone	63%		
Tetrachloroethene	141%	368%	89%
Trichloroethene		131%	

The data for compounds with low recoveries were flagged with the "J" qualifier and are estimated values.

Data for compounds with high recoveries were only qualified (J) when they were detected in a sample. High recoveries for undetected compounds do not affect the usability of the data.

### **Laboratory Control Sample**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

All of the laboratory control sample recoveries in LCS-62940, associated with the field and trip blanks were within the required limits with the exceptions of acetone (42%), tetrachloroethene (67%) and 2-hexanone (69%).

The data for this compound was flagged with the "J" qualifier and are estimated values.

All of the laboratory control sample recoveries in LCS-62994/ LCDD-62994, were within the required limits with the exception of bromomethane (140%). These LCSs were associated with the analyses of the following samples:

K2296-04	PM-SB-1a-B-1112011
K2296-05	PM-SB-2-A-1112011
K2296-06	PM-SB-2-B-1112011
K2296-07	PM-SB-3-A-1112011

K2296-08	PM-SB-4-A-1112011
K2296-10	PM-SB-5-B-1112011
K2296-11	PM-SB-6-A-1112011
K2296-12	PM-SB-6-B-1112011

Bromomethane was not detected in any of these samples and the data were not affected by the high recoveries.

All of the laboratory control sample recoveries in LCS-63069 / LCDD-63069, were within the required limits with the exceptions of chloromethane in the LCS (132%) and iodomethane in the LCS and LCSD (45% & 48%). These LCSs were associated with the analyses of sample K2296-03 / PM-SB-1a-A-1112011:

Chloromethane was not detected in the sample and the data were not affected by the high recoveries.

The data for iodomethane were flagged with the "J" qualifier and are estimated values.

All of the other LCS recoveries were within the required limits.

### **Method Blanks**

Low concentrations of toluene (4.5J ug/l) and naphthalene (1.0J ug/kg) were detected in the method blank associated with soil sample K2296-03 / PM-SB-1a-A-1112011

Only a low concentrations of toluene, less than 2X the concentration in the method blank were detected in the sample. The data for toluene were reported at the CRDL with the "U" qualifier.

### **Trip Blank**

No compounds were detected in the trip blank.

### **Field Blank**

No target compounds were detected in the field blank.

### **Internal Standard Areas and Retention Times**

The recoveries and retention times of all internal standards were within the required 50% - 100% quality control limits.

### **Sample Results**

No problems were detected with the samples.

**SUMMARY OF THE ANALYTICAL DATA USABILITY**  
**DEC-Former Paul Miller Site**  
**D004437-23**

**Soil and Water Volatile Organic Analyses – Method 8260C**

**Samples Collected: November 2<sup>nd</sup> & 3<sup>rd</sup>, 2011**

**Samples Received: November 4, 2011**

**Laboratory: Spectrum**

**Sample Delivery Group: K2297**

**Laboratory Reference Numbers:**

<b>Lab ID</b>	<b>Client ID</b>	<b>Matrix</b>	<b>Date Sampled</b>
K2297-01	PM-GWS-7-1122011	Aqueous	11/02/2011
K2297-01 MS	PM-GWS-7-1122011 MS	Aqueous	11/02/2011
K2297-01 MSD	PM-GWS-7-1122011 MSD	Aqueous	11/02/2011
K2297-02	PM-GWS-1b-1122011	Aqueous	11/02/2011
K2297-03	PM-GWS-2-1122011	Aqueous	11/02/2011
K2297-04	PM-FB-SO-1122011	Aqueous	11/02/2011
K2297-05	PM-FB-dAQ-1122011	Aqueous	11/02/2011
K2297-06	PM-TB-1122011	Aqueous	11/02/2011
K2297-07	PM-SB-7-B-1122011	Soil	11/02/2011
K2297-08	PM-SB-1b-B-1122011	Soil	11/02/2011
K2297-09	PM-SB-1b-A-1122011	Soil	11/02/2011
K2297-10	PM-SB-99-110211	Soil	11/02/2011
K2297-11	PM-GWS-5-110311	Aqueous	11/03/2011
K2297-12	PM-GWS-6-110311	Aqueous	11/03/2011
K2297-13	PM-TB-110311	Aqueous	11/03/2011
K2297-14	PM-GWS-1a-110311	Aqueous	11/03/2011
K2297-15	PM-GWS-99-110311	Aqueous	11/03/2011

Soil and water samples were validated for analyses of volatile organics by the US EPA Region II data validation SOP (HW-24, Revision 2). Data were reviewed for usability according to the following criteria:

- \* - Data Completeness
- \* - GC/MS Tuning
- \* - Holding Times
  - Calibrations
  - Laboratory Blanks
- \* - Trip Blank
  - Field Blank
- \* - System Monitoring Compound Recoveries
- \* - Internal Standard Recoveries
  - Matrix Spike / Matrix Spike Duplicate
  - Laboratory Control Samples
- \* - Compound Identification
- \* - Compound Quantitation

\* - Indicates that all criteria were met for this parameter.

## DATA VALIDATION SUMMARY

The problems with the calibrations, matrix spikes, laboratory control samples and blanks should be noted. These are described in detail below.

No other significant problems were found with this sample delivery group which would affect the usability of the data.

### Holding Times

The samples of this delivery group were validated against the Region II technical holding time requirements:

Soil and preserved aqueous samples were analyzed within 14 days of collection.

### Tunes

No problems were detected with the tunes associated with the samples of this delivery group.

### System Monitoring Compound Recoveries

The recovery of the Dibromofluoromethane system monitoring compound in sample K2297-01 MS / PM-GWS-7-1122011 MS (116%) was just above the 115% quality control limit.

The recovery of all of the system monitoring samples were within the required limits in the analysis of the original sample. The slightly high recovery in the matrix spike did not affect the use of the data.

All of the other sample surrogate recoveries were within the required limits.

### Calibrations

#### Initial Calibrations

All relative percent differences in the 11/10 initial calibration (Instrument V1) were less than 20% with the exceptions of hexachlorobutadiene (21%) and 1,2,3-trichlorobenzene (23%). This calibration was associated with the analyses of the following samples:

K2297-01	PM-GWS-7-1122011
K2297-02	PM-GWS-1b-1122011
K2297-03	PM-GWS-2-1122011
K2297-04	PM-FB-SO-1122011
K2297-05	PM-FB-dAQ-1122011
K2297-06	PM-TB-1122011
K2297-11	PM-GWS-5-110311
K2297-12	PM-GWS-6-110311
K2297-13	PM-TB-110311
K2297-14	PM-GWS-la-110311
K2297-15	PM-GWS-99-110311

The RRFs of acetone (0.036) and 2-butanone (0.028) were less than the 0.050 quality control limit in the above calibration.

All relative percent differences in the 11/10 initial calibration (Instrument V10) were less than 20% with the exceptions of acetone (35%), iodomethane (33%) and hexachlorobutadiene (22%). This initial calibration was associated with the analyses of the following samples:

K2297-07	PM-SB-7-B-1122011
K2297-08	PM-SB-1b-B-1122011
K2297-09	PM-SB-1b-A-1122011

The RRFs of acetone (0.032) and 2-butanone (0.034) were less than the 0.050 quality control limit in the above calibration.

All relative percent differences in the 11/14 initial calibration (Instrument V10) were less than 20% with the exceptions of acetone (31%), toluene (39%) and naphthalene (23%). This initial calibration was associated with the analysis of sample K2297-10 / PM-SB-99-110211.

The RRFs of acetone (0.032) and 2-butanone (0.032) were less than the 0.050 quality control limit in the above calibration.

Detected data for these compounds were flagged with the "J" qualifier and are estimated values.

### **Continuing Calibrations**

All relative percent differences in the 11/11 continuing calibration were less than 20% with the exceptions of acetone (47%), tetrachloroethene (35%), 2-hexanone (22%), hexachlorobutadiene (27%), 1,2,3-trichlorobenzene (22%), and naphthalene (21%). This continuing calibration was associated with the analyses of the following samples:

K2297-02	PM-GWS-1b-1122011
K2297-03	PM-GWS-2-1122011
K2297-06	PM-TB-1122011

The RRFs of acetone (0.036) and 2-butanone (0.023) were less than the 0.050 quality control limit in the above calibration.

All percent differences in the 11/12 continuing calibration were less than 20% with the exceptions of chloromethane (22%), bromomethane (42%), iodomethane (24%), carbon tetrachloride (24%) and naphthalene (22%). This continuing calibration was associated with the analyses of the following samples:

K2297-07	PM-SB-7-B-1122011
K2297-08	PM-SB-1b-B-1122011
K2297-09	PM-SB-1b-A-1122011

The RRFs of acetone (0.028) and 2-butanone (0.033) were less than the 0.050 quality control limit in the above calibration.

All percent differences in the 11/14 continuing calibration were less than 20% with the exceptions of acetone (22%), 2-butanone (32%) tetrachloroethene (26%), bromoform (30%) and 1,2,3-trichloropropane (22%). This continuing calibration was associated with the analyses of the following samples:

K2297-01	PM-GWS-7-1122011
K2297-12	PM-GWS-6-110311
K2297-13	PM-TB-110311

The RRFs of acetone (0.044) and 2-butanone (0.037) were less than the 0.050 quality control limit in the above calibration.

All percent differences in the 11/15 continuing calibration were less than 20% with the exceptions of acetone (31%), dibromochloromethane (21%), bromoform (33%), n-butylbenzene, (22%), hexachlorobutadiene (34%), 1,2,3-trichlorobenzene (28%) and naphthalene (27%). This continuing calibration was associated with the analyses of the following samples:

K2297-04	PM-FB-SO-1122011
K2297-05	PM-FB-dAQ-1122011
K2297-11	PM-GWS-5-110311
K2297-14	PM-GWS-la-110311
K2297-15	PM-GWS-99-110311

The RRFs of acetone (0.023) and 2-butanone (0.029) were less than the 0.050 quality control limit in the above calibration.

All percent differences in the 11/14 continuing calibration were less than 20% with the exceptions of acetone (31%), dibromochloromethane (21%), bromoform (33%), n-butylbenzene, (22%), hexachlorobutadiene (34%), 1,2,3-trichlorobenzene (28%) and naphthalene (27%). This continuing calibration was associated with the analyses of the following samples:

K2297-04	PM-FB-SO-1122011
K2297-05	PM-FB-dAQ-1122011
K2297-11	PM-GWS-5-110311
K2297-14	PM-GWS-la-110311
K2297-15	PM-GWS-99-110311

The RRFs of acetone (0.023) and 2-butanone (0.029) were less than the 0.050 quality control limit in the above calibration.

All percent differences in the 11/15 continuing calibration (Instrument V10) were less than 20% with the exception of iodomethane (48%). This continuing calibration was associated with the analysis of sample K2297-10 / PM-SB-99-110211.

The RRFs of acetone (0.027) and 2-butanone (0.035) were less than the 0.050 quality control limit in the above calibration.

When acetone and/ or 2-butanone were detected in a sample, they were flagged with the "J" qualifier.

When these compounds were not detected in a sample they were flagged with the "R" qualifier and are technically rejected.

The compounds with percent differences greater than 20% were flagged with the "J" qualifier and are estimated values.

### **Matrix Spike and Matrix Spike Duplicate**

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

Soil sample K2297-01 / PM-GWS-7-1122011 was used as the matrix spike and matrix spike duplicate. All recoveries and RPDs were within the required limits with the following exceptions:

<b>Compound</b>	<b>MS</b>	<b>MSD</b>	<b>RPD</b>
1,1,1-Trichloroethane			40%
1,1-Dichloroethane			35%
1,1-Dichloroethene			42%
1,2,4-Trichlorobenzene	54%		43%
1,2,4-Trimethylbenzene			43%
1,2-Dichloroethane			42%
2-Butanone		11%	146%
Acetone	145%	15%	162%
Bromomethane		62%	47%
Carbon disulfide			41%
Chloroethane			56%
Chloromethane			32%
cis-1,2-Dichloroethene			35%
cis-1,3-Dichloropropene	51%		43%
Dichlorodifluoromethane		62%	57%
Methyl acetate	508%		127%
Methylcyclohexane	59%		47%
Methylene chloride			35%
Trichlorofluoromethane			47%
Vinyl chloride			55%

The data for compounds with low recoveries were flagged with the "J" qualifier and are estimated values.

Data for compounds with high recoveries were only qualified (J) when they were detected in a sample. High recoveries for undetected compounds do not affect the usability of the data.

An RPD of 30% was used for the purposes of the validation. Compounds with RPDs greater than 30% were only qualified when they were detected in a sample.

### Laboratory Control Sample

Some of the laboratory's in-house quality control limits were as low as 30% and as high as 163%. Quality control limits of 70% - 130% were used for the validation.

All of the laboratory control sample recoveries in LCS-62940, were within the required limits with the exceptions of acetone (42%), tetrachloroethene (67%) and 2-hexanone (69%). This laboratory control samples is associated with the following samples:

K2297-02	PM-GWS-1b-1122011
K2297-03	PM-GWS-2-1122011
K2297-06	PM-TB-1122011

The data for these compounds were flagged with the "J" qualifier and are estimated values.

All of the laboratory control sample recoveries in LCS-63069/ LCSD-63069, were within the required limits with the exceptions of chloromethane (132%) and iodomethane (45% & 48%). These LCSs were associated with the analyses of the sample :PM-SB-99-110211 (K2297-10)

The data for compounds with low recoveries were flagged with the "J" qualifier and are estimated values.

Data for compounds with high recoveries were only qualified (J) when they were detected in a sample. High recoveries for undetected compounds do not affect the usability of the data.

All of the other LCS recoveries were within the required limits.

### Method Blanks

Low concentrations of toluene (4.5J ug/l) and naphthalene (1.0J ug/kg) were detected in the method blank associated with soil sample K2297-10 / PM-SB-99-110211

Only a low concentration of toluene (4 ug/l), less than 2X the concentration in the method blank were detected in the sample. The data for toluene were reported at the CRDL with the "U" qualifier.

No compounds were detected in any of the other method blanks.

### Trip Blank

No compounds were detected in the trip blanks.

### Field Blank

Low concentrations of 2-butanone (3.2ug/l) and acetone (6.9 ug/l) were detected in the soil field blank collected on 11/02/2011.

**Only a low concentration of acetone (7 ug/l) was detected in sample PM-SB-99-110211 (K2297-10).**

**This was reported as 9U ug/kg and flagged with the “R” qualifier due to the low relative response factor.**

Low concentrations of 2-butanone (3 ug/l) and acetone (6.8 ug/l) were detected in the water field blank, K2297-05 / PM-FB-dAQ-1122011 collected on 11/02/2011.

When detected in sample PM-GWS-7-1122011 (K2297-01) the concentrations of these two compounds were too high to be affected by the low level field blank contamination.

### **Internal Standard Areas and Retention Times**

The recoveries and retention times of all internal standards were within the required 50% - 100% quality control limits.

### **Sample Results**

No problems were detected with the samples.

## Appendix E

### Field Logbooks

Paul Miller Site

LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/6/12

SAMPLERS: 2K  
Fenses 1m = ~~3.04~~  
~~10 ft~~ D. Int. ~ 0.54  
WEATHER CONDITIONS: Sunny 30° F  
SAMPLE ID: 80230-MW01-030612 SAMPLE TIME: 0910  
CLP ID:

WELL #: MW-1  
DEPTH OF PUMP INTAKE: 15  ft TIC or ft BGS (circle one)  
SCREENED/OPEN BOREHOLE INTERVAL: 6-16  ft TIC or ft BGS (circle one)  
SAMPLE FLOW RATE: 150 ml/minute

Instrument Type/Model:  
Complete and/or Circle at right

CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	pH (± 0.1 SU)	SPECIFIC CONDUCTIVITY (± 3%)	DISSOLVED OXYGEN (± 10%)	TEMP. °C	REDOX POTENTIAL (± 10 mV)	Instrument: YSI Model # <u>650</u> / Horiba U-22 (circle one)	
										Instrument: YSI model 650	
0813	9.15	200			6.51	4.230	0.99	11.74	-131.6	7999	
0821	10.30	200			6.48	4.187	0.89	11.95	-126	7999	
0826	10.75	200			6.48	4.226	1.25	12.91	-117.3	80.1	
0831	11.30	200			6.48	4.306	1.56	14.83	-99.8	68	
0834	11.55	200			6.48	4.330	1.63	15.57	-91.3	68	
0837	11.59	200			6.49	4.777	1.77	15.71	-109.6	69	
0840	11.75	100			6.49	4.783	1.64	16.10	-110.3	68	
0843	11.65	100			6.51	4.778	1.43	15.61	-110.7	65	
0846	11.65	100			6.50	4.570	1.60	15.09	-107.6	61	
0849	11.65	100			6.50	4.562	1.60	15.36	-106.8	60	

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values: DO = 0.3 - 10 mg/L Redox Potential = -100 - +600 mV  
Spec. Conductivity ( $\mu\text{S}/\text{cm}$ ) = 0.01 - 5,000; up to 10,000 in industrial, -55,000 in high salt content water. Note: 1,000  $\mu\text{S}/\text{cm}$  = 1 mS/cm  
TIC = Top of Inner Casing BGS = Below Ground Surface

67099124EP

Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/6/12

SAMPLERS: EK

WEATHER CONDITIONS: *Sunny* 30°f  
SAMPLE ID: 80230-MW01-030612 S  
CLP ID:

WELL #: mhw-1

**DEPTH OF PUMP INTAKE:** 15  ft TIC or ft BGS (circle one)  
**SCREENED/OPEN BOREHOLE INTERVAL:** 6-16  ft TIC or ft BGS  
**SAMPLE FLOW RATE:** 150 ml/minute

**Instrument Type/Model:  
Complete and/or Circle at right**

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values:  
 $DO = 0.3 - 10 \text{ mg/L}$   
 $Redox \text{ Potential} = -100 - +600 \text{ mV}$   
 $Spec. \text{ Conductivity } (\mu\text{S/cm}) = 0.01 - 5,000$ ; up to 10,000 in industrial, -55,000 in high salt content water.  
 $Turbidity = 0 - >500 \text{ NTUs}$   
Note:  $1,000 \mu\text{S/cm} = 1 \text{ mS/cm}$

TIC = Top of Inner Casing      BGS = Below Ground Surface

Paul Miller Site

LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/6/12

SAMPLERS: ✓ ✓

Ferrus Iron = 10.2 Oktan = 0.92 mlf  
= 9.2 mlf WELL #: M-107

SAMPLERS: 2 R  
WEATHER CONDITION  
SAMPLE ID: 80230-1  
CLP ID: N/A

SUNNY 42° F

SAMPLERS: 2 R  
WEATHER CONDITIONS: Sun/NW 42° F  
SAMPLE ID: #0230-MW02-030612  
SAMPLE TIME: 1200

## DEPTH OF PUMP INTAKE

SCREENED OPEN BOREH

**DEPT OF PUMP INTAKE:** 10 ft TIC or ft BGS (circle one) **ft TIC** **circle**

### **Instrument Type/Model:**

### **Instrument Type/Model:**

SAMPLE TIME: 12:00

SAMPLE FLOW RATE: 250

### DEPTH OF PUMP INTAKE:

**DEPTH OF PUMP INTAKE:** 10 ft TIC or ft BGS (circle one)

(YSI Model# 650 / Horiba U-22 (circle one)  
Other (specify) \_\_\_\_\_

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values:  
 $\text{DO} = 0.3 - 10 \text{ mg/L}$   
 $\text{Redox Potential} = -100 - +600 \text{ mV}$   
 $\text{Spec. Conductivity } (\mu\text{s/cm}) = 0.01 - 5,000; \text{ up to } 10,000 \text{ in industrial, } -55,000 \text{ in high salt content water.}$   
 $\text{Turbidity} = 0 - >500 \text{ NTUs}$   
 $\text{Note: } 1,000 \mu\text{s/cm} = 1 \text{ mS/cm}$

TIC = Top of Inner Casing  
BGS = Below Ground Surface

Paul Miller Site

**LOW FLOW GROUNDWATER SAMPLING PURGE RECORD**

DATE: 3/7/12      FLOW RATE: 10% 0.6 min = 2.5 GPM      WELL #: MW-03  
SAMPLERS: EK  
WEATHER CONDITIONS: sunny 45°f  
80-23°  
SAMPLE ID: MW-035-030712      SAMPLE TIME: 1150      SAMPLE FLOW RATE: 225 ml/minute  
SCREENED OPEN BOREHOLE INTERVAL: 2.5-6 m (11 ft) TIC or ft BGS (circle one)  
DEPTH OF PUMP INTAKE: 1 / TIC or ft BGS (circle one)  
CLIP ID: (circle one)

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values:  
 $\text{DO} = 0.3 - 10 \text{ mg/L}$   
 $\text{Redox Potential} = -100 - +600 \text{ mV}$   
 $\text{Spec. Conductivity } (\mu\text{S/cm}) = 0.01 - 5,000; \text{ up to } 10,000 \text{ in industrial}$   
 $\text{Turbidity} = 0 - >500 \text{ NTUs}$   
 $\text{Note: } 1,000 \mu\text{S/cm} = 1 \text{ mS/cm}$

## Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/6/12      FORM *bore = 18% Dilution*      WELL #: 17W-04  
 SAMPLERS: F. Robin      DILUTION: 0.51 m3/l      DEPTH OF PUMP INTAKE: 9 ft TIC or ft BGS (circle one)

WEATHER CONDITIONS: Clear 35°

SAMPLE ID: 80230-13004-03612      CLP ID:

Instrument Type/Model:  
Complete and/or Circle at right

YSI Model # 500 / Horiba U-22 (circle one)  
Other (specify) \_\_\_\_\_

CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	pH	SPECIFIC CONDUCTIVITY (± 3%)	DISSOLVED OXYGEN (± 10%)	TEMP. °C	REDOX POTENTIAL (± 10 mV)	TURBIDITY NTUs	Instrument: Cassette 2020
24-Hour	gallons / liters (circle one)	ft TIC / ft BGS (circle one)	Units: SU	ft TIC / ft BGS (circle one)	SU	S/cm, mS/cm <sup>2</sup> or μS/cm (circle one)	mg/L (not %)	Units: °C	mV		
1/00	3.85	150		7.79	7.41	1.410	-1.61	11.60	-28.6	53.2	
1/15	4.52	150		7.98	1.434	1.10	13.03	-42.2	77.7		
1/20	4.70	150		7.34	1.486	1.04	13.55	-37.5	74.6		
1/21	4.77	150		7.22	1.610	1.03	13.74	-52.6	54.8		
1/30	5.02	160		7.15	1.681	1.04	14.31	-55.2	48.1		
1/35	5.13	150		7.11	1.729	1.08	14.51	-58.3	44.4		
1/40	5.18	150		7.05	1.829	1.02	13.90	-63.3	38.0		
1/45	5.25	150		7.06	1.890	1.13	14.63	-61.9	38.5		
1/50	5.32	150		7.02	1.896	1.18	14.75	-59.9	34.4		
1/55	4.5	150	7.00	1.902	1.17	14.89	-60.7	30.5			

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values:

DO = 0.3 - 10 mg/L      Redox Potential = -100 - +600 mV  
 Spec. Conductivity (μS/cm) = 0.01 - 5,000; up to 10,000 in industrial, ~55,000 in high salt content water. Note: 1,000 μS/cm = 1 mS/cm

TIC = Top of Inner Casing

BGS = Below Ground Surface

Turbidity = 0 - >500 NTUs

Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/7/2

Torous Iraa = 0.05 ms/J

SAMPLERS: F. Robin

WEATHER CONDITIONS: Clear 5°

WELL #: 705 - 85

DEPTH OF PUMP INTAKE: 30 ft TIC or ft BGS (circle one)  
 SCREENED/OPEN BOREHOLE INTERVAL: 25 - 35 ft TIC or ft BGS (circle one)

SAMPLE ID: 80236 - 705BGS - 030712  
CLP ID:Instrument Type/Model:  
Complete and/or Circle at right

XSI Model # 600 X-L / Horiba U-22 (circle one)  
 Other (specify)

Instrument:  
 2420

XS1 Model # 600 X-L / Horiba U-22 (circle one)  
 Other (specify)

XS1 Model # 600 X-L / Horiba U-22 (circle one)  
 Other (specify)

CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	pH (± 0.1 SU)	SPECIFIC CONDUCTIVITY (± 3%)	DISSOLVED OXYGEN (± 10%)	TEMP. °C (± 10%)	REDOX POTENTIAL (± 10 mV)	Units: mV	Instrument: 2420	
											S/cm <sup>-1</sup> ft BGS (circle one)	S/cm <sup>-1</sup> ft BGS (circle one)
10:50	6.97	250	8.06	0.546	6.30	15.69	185.4	21000				
11:05	8.19	300	7.09	0.541	4.53	17.40	148.0	20000				
11:10	8.17	300	7.07	0.558	4.72	17.57	154.2	2008				
11:15	8.17	300	7.04	0.570	4.80	17.57	161.4	208				
11:20	8.17	300	7.03	0.569	4.81	17.72	154.8	153				
11:25	8.21	300	7.03	0.568	4.81	17.76	155.2	103				
11:30	8.18	300	7.01	0.565	4.80	17.91	162.1	76.0				
11:35	8.21	300	7.01	0.564	4.77	17.99	157.3	54.1				

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values: DO = 0.3 - 10 mg/L  
TIC = Top of Inner Casing

Redox Potential = -100 - +600 mV  
Spec. Conductivity (µS/cm) = 0.01 - 5,000; up to 10,000 in industrial, ~55,000 in high salt content water. Note: 1,000 µS/cm = 1 mS/cm

Turbidity = 0 - >500 NTUs  
BGS = Below Ground Surface

Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/5/12

Flow from = 0.00 ml/s

SAMPLERS: EK

WEATHER CONDITIONS: Sunny, Windy 40°f

SAMPLE ID: 80230-MW095-030512 SAMPLE TIME: 1055

CLP ID: N/A

WELL #: MW-95

DEPTH OF PUMP INTAKE: 30 ft TIC or ft BGS (circle one)

SCREENED OPEN BOREHOLE INTERVAL: 25-35 ft TIC or ft BGS (circle one)

SAMPLE FLOW RATE: 250 ml/minute

Instrument Type/Model:

Complete and/or Circle at right

YSI Model # 630 / Horiba U-22 (circle one)  
Other (specify)

CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	pH (± 0.1 SU)	SPECIFIC CONDUCTIVITY (± 3%)	DISSOLVED OXYGEN (± 10%)	TEMP. (± 10%)	REDOX POTENTIAL (± 10 mV)	TURBIDITY (± 10%)	Instrument: 630 /	
											NTUs	
0958	4.36'	250	250	6.30	1.063	2.98	14.44	15.47	2.55			
1020	5.15	250	250	6.78	0.845	0.47	16.04	-2.7	281			
1025	5.16	250	250	6.77	0.967	0.56	15.93	-21.1	1257			
1030	5.16	250	250	6.80	0.838	0.46	16.20	-20.8	286			
1035	5.16	250	250	6.80	0.826	0.44	16.13	-25.2	280			
1040	4	5.16	250	6.81	0.820	0.42	16.40	-28.1	216			
1043	5.16	250	250	6.82	0.807	0.42	16.64	-28.6	209			
1046	5.16	250	250	6.82	0.805	0.42	16.67	-29.1	147			
1049	5.5	5.16	250	6.82	0.804	0.42	16.61	-29.4	139			
1055	Sample Time											

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values: TIC = Top of Inner Casing Spec. Conductivity ( $\mu\text{S}/\text{cm}$ ) = 0.01 - 5,000; up to 10,000 in industrial, ~55,000 in high salt content water. Note: 1,000  $\mu\text{S}/\text{cm}$  = 1 mS/cm

Redox Potential = -100 - +600 mV  
Spec. Conductivity ( $\mu\text{S}/\text{cm}$ ) = 0.01 - 5,000; up to 10,000 in industrial, ~55,000 in high salt content water. Note: 1,000  $\mu\text{S}/\text{cm}$  = 1 mS/cm

BGS = Below Ground Surface



Paul Miller Site

**LOW FLOW GROUNDWATER SAMPLING PURGE RECORD**

DATE: 3/5/12

SAMPLERS: *J. Robins* Sensors  $1_{\text{top}} = 1.63 \text{ mg/L}$

WEATHER CONDITIONS:

SAMPLE ID: #80230 - new DS - 030512 CLP ID:

WELL #: 1700-10 S

DEPTH OF PUMP INTAKE: 28'  ft TIC or ft BGS (circle one)  
SCREENED/OPEN BOREHOLE INTERVAL: 22'-32'  ft TIC or ft BGS (circle one)

SAMPLE TIME: /400 SAMPLE FLOW RATE: ml/minute

Instrument Type/Model:  
Complete and/or Circle at right

CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	pH (± 0.1 SU)	SPECIFIC CONDUCTIVITY (± 3%)	DISSOLVED OXYGEN (± 10%)	TEMP. (± 10%)	REDOX POTENTIAL (± 10 mV)	TURBIDITY (± 10%)	Instrument: <input checked="" type="checkbox"/> 2000	
											YSI Model # <u>600XL</u> Other (specify)	/ Horiba U-22 (circle one)
24-Hour gallons/liters (circle one)	ft TIC / ft BGS (circle one)	ft TIC / ft BGS (circle one)	ml/min	ft TIC / ft BGS (circle one)	SU	SI/cm, <u>mS/cm</u> <input checked="" type="checkbox"/> or µS/cm (circle one)	mg/L (not %)	°C	Units: mV	NTUs		
12:50	4.74	250		6.82	1.392	1.36	14.58	3.3				72000
13:05	5.687.80	200		6.67	1.462	1.36	15.51	-4.8				232
13:16	9.988.75	200		6.66	1.463	1.30	15.65	-4.9				239
13:17	9.55	200		6.65	1.409	1.27	16.56	-8.4				185
13:20	10.20	200		6.66	1.407	1.32	16.05	-11.3				210
13:25	10.92	200		6.66	1.408	1.28	15.92	-14.7				177
13:30	11.93	200		6.65	1.405	1.46	16.49	-18.3				111
13:35	12.90	200		6.65	1.408	1.49	17.27	-22.2				217
13:40	14.03	200		6.66	1.409	1.50	17.69	-22.5				179
13:45	15.65	200		6.65	1.411	1.47	17.71	-29.9				161
13:50	16.15	200		6.64	1.409	1.50	17.89	-31.8				182

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values: DO = 0.3 - 10 mg/L  
Spec. Conductivity ( $\mu\text{S}/\text{cm}$ ) = 0.01 - 5,000; up to 10,000 in industrial, ~55,000 in high salt content water. Note: 1,000  $\mu\text{S}/\text{cm}$  = 1 mS/cm

Redox Potential = -100 - +600 mV  
Turbidity = 0 - >500 NTUs

TIC = Top of Inner Casing BGS = Below Ground Surface

**Paul Miller Site**

DATE: 3/5/12      Sampling Depth: 0000 ft  
 SAMPLERS: EK

**LOW FLOW GROUNDWATER SAMPLING PURGE RECORD**

WEATHER CONDITIONS: Sunny, Windy 41°F  
 SAMPLE ID: 02230-mw10P - 03651/2  
 CLP ID:

WELL #: MW-10D

DEPTH OF PUMP INTAKE: 65 ft TIC or ft BGS (circle one)  
 SCREENED/OPEN BOREHOLE INTERVAL: 65 ± 70 ft TIC or ft BGS (circle one)  
 SAMPLE TIME: 1345  
 SAMPLE FLOW RATE: 250 ml/minute

Instrument Type/Model:  
 Complete and/or Circle at right

CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	YSI Model # 650 / Horiba U-22		(circle one)	Instrument: LaMotte	
					PH (± 0.1 SU)	SPECIFIC CONDUCTIVITY (± 3%) S/cm, mS/cm or µS/cm (circle one)	DISSOLVED OXYGEN (± 10%) mg/L (not %)	TEMP. REDOX POTENTIAL (± 10%) mV	
1255	5.44	325	325	7.25	0.739	1.66	14.77	54.8	191
1310	5.44	325	325	7.26	0.748	0.82	15.27	43.5	75
1315	5.44	325	325	7.26	0.755	0.81	15.07	43.3	66
1320	5.44	325	325	7.26	0.754	0.74	15.12	44.1	56
1325	5.44	325	325	7.26	0.767	0.69	15.09	40.9	45
1330	5.44	325	325	7.27	0.768	0.70	15.22	43.1	27.5
1333	5.44	325	325	7.27	0.770	0.68	15.25	44.3	23.7
1336	5.44	326	326	7.26	0.771	0.67	15.27	45.2	16.2
1339	7	5.44	325	7.26	0.773	0.67	15.33	46.0	14.3
1345	Sampling TIME								

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values: DO = 0.3 - 10 mg/l. Redox Potential = -100 - +600 mV Spec. Conductivity (µS/cm) = 0.01 - 5,000, up to 10,000 in industrial, ~55,000 in high salt content water. Note: 1,000 µS/cm = 1 mS/cm  
 TIC = Top of Inner Casing      BGS = Below Ground Surface  
 Turbidity = 0 - >500 NTUs

## Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/8/12

Fenses Iron = 0.06 mg/l  
WELL #: NW. 115

SAMPLERS: E/K

WEATHER CONDITIONS: Partly Sunny 45°

Temp 23°

DO

SAMPLE ID: NW/115-0308/2  
CLP ID:

DEPTH OF PUMP INTAKE: 27 ft TIC or ft BGS (circle one)

SCREENED OPEN BOREHOLE INTERVAL: 27 - 32 ft TIC or ft BGS (circle one)

SAMPLE TIME: 0925  
SAMPLE FLOW RATE: 250 ml/minuteInstrument Type/Model:  
Complete and/or Circle at right

CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	SPECIFIC CONDUCTIVITY (± 3%)		DISSOLVED OXYGEN (± 10%)	TEMP. °C	REDOX POTENTIAL (± 10 mV)	TURBIDITY NTUs	Instrument: Lamotte 2020
					YSI Model #	Other (specify)					
0754	6.05	200		6.21	1.363	3.73	14.44	83.8	>999		
0833	6.90	200		6.20	1.462	0.90	16.70	58.7	>999		
0836	5	200		6.20	1.459	0.88	16.78	63.3	>999		
0839	6.90	200		6.20	1.453	0.95	16.90	67.8	>999		
0842	6.90	200		6.20	1.448	0.99	16.76	74.1	>999		
0845	60.70	Wait For Turbidity To Come Down (cleared out flow thru cell)		6.19	1.456	0.94	16.91	80.3	103		
0905	6.90	200		6.19	1.455	0.91	17.10	80.5	74		
0910	6.90	200		6.19	1.455	0.93	17.18	82.2	60		
0915	6.90	200		6.19	1.455	0.93	17.13	83.5	51		
0920	4 Gallons	6.90	200	6.19	1.456	0.94	17.13	83.5	51		

0925 Sample Time

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values:

DO = 0.3 - 10 mg/L

Redox Potential = -100 - +600 mV

Spec. Conductivity ( $\mu\text{S}/\text{cm}$ ) = 0.01 - 5,000; up to 10,000 in industrial, ~55,000 in high salt content water.Note: 1,000  $\mu\text{S}/\text{cm} = 1 \text{ mS}/\text{cm}$ 

TIC = Top of Inner Casing

BGS = Below Ground Surface

Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/3/12

$$f_{\text{error}}(x) = 0.67 \sin(x)$$

SAMPLERS: E/K

WELL #: MW-11D

SAMPLERS: EK  
WEATHER CONDITIONS: Partly sunny 45°  
SAMPLE ID: MW1D-030617  
SAMPLE TIME: 0900  
SAMPLE FLOW RATE: 250 ml/minute  
SCREENED/OPEN BOREHOLE INTERVAL: 60'-70' TIC or ft BGS (circle one)  
DEPTH OF PUMP INTAKE: 65' ft TIC or ft BGS (circle one)  
(circle one)

## Instrument Type/Model:

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values:  
 $\text{DO} = 0.3 - 10 \text{ mg/L}$   
 $\text{Redox Potential} = -100 - +600 \text{ mV}$   
 $\text{Spec. Conductivity } (\mu\text{s/cm}) = 0.01 - 5,000; \text{ up to } 10,000 \text{ in industrial, } \sim 55,000 \text{ in high salt content water. Note: } 1,000 \mu\text{s/cm} = 1 \text{ mS/cm}$   
 $\text{Turbidity} = 0 - >500 \text{ NTUs}$

Redox Potential = -100 - +600 mV

### Spec. Conductivity

BGS = Below-Ground Surface

Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/8/12 SAMPLERS: FR/CK WEATHER CONDITIONS: Partly cloudy with  
SAMPLE ID: 80230 - MW 1125-036812 CLP ID: 80230 - MW 1125-036812 (Duplicate)  
WELL #: MW-12S DEPTH OF PUMP INTAKE: 30' SCREENED/OPEN BOREHOLE INTERVAL:  
SAMPLE TIME: 11:15 SAMPLE FLOW RATE: 250 ml/minute  
(ft TiC or ft BGS (circle one)) (ft TiC or ft BGS (circle one))

Instrument Type/Model: Complete and/or Circle at right		PSI Model# <u>600 x L</u>		/ Horiba U-22 (circle one)		Instrument: <u>Siemens</u> <u>2020</u>				
CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	pH (± 0.1 SU)	SPECIFIC CONDUCTIVITY (± 3%)	DISSOLVED OXYGEN (± 10%)	TEMP. (± 10%)	REDOX POTENTIAL (± 10 mV)	TURBIDITY (± 10%)
24-Hour	gallons/liters	ft TIC/ ft BGS	Units: <u>m/min</u>	ft TIC/ ft BGS (circle one)	SU	S/cm, mS/cm <sup>2</sup> , or µS/cm (circle one)	mg/L (not %)	Units: °C	mV	NTUs
1025	6.12	375		6.81	0.650	1.81	1665	159.2	70000	
1045	6.30	400		6.87	0.674	1.83	17.45	83.8	147	
1050	6.30	400		6.74	0.676	1.80	17.60	80.7	75.2	
1055	6.30	400		6.82	0.680	1.81	17.46	81.6	53.6	
1100	6.30	400		6.81	0.686	1.75	17.39	82.7	29.8	
1105	6.30	400		6.79	0.688	1.82	17.41	83.1	26.6	
1110	4.5	6.30	400	6.78	0.692	1.81	17.39	85.4	21.9	

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values:  
 $DO = 0.3 - 10 \text{ mg/L}$   
 $\text{Spec. Conductivity } (\mu\text{S/cm}) = 0.01 - 5,000$ ; up to 10,000 in industrial, ~55,000 in high salt content water. Note: 1,000  $\mu\text{S/cm} = 1 \text{ mS/cm}$

Redox Potential = -100 - +600 mV  
 Turbidity = 0 - >500 NTUs

Paul Miller Site

**LOW FLOW GROUNDWATER SAMPLING PURGE RECORD**

DATE: 3/1/12

Ferm Iron = 0.53 mS/cm

WELL #: MW-135

SAMPLERS: EK

WEATHER CONDITIONS: Sunny 35°F  
8623°

SAMPLE ID: MW/135-030712  
CLP ID:

DEPTH OF PUMP INTAKE: 30 ft TIC or ft BGS (circle one)  
SCREENED/OPEN BOREHOLE INTERVAL: 25 - 35 ft TIC or ft BGS (circle one)

SAMPLE TIME: 0920

SAMPLE FLOW RATE: 250 ml/minute

Instrument Type/Model:  
Complete and/or Circle at right

CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	YSI Model # <u>650</u> / Horiba U-22 (circle one)			Instrument: <i>Comfit</i>
					pH (± 0.1 SU)	SPECIFIC CONDUCTIVITY (± 3%)	DISSOLVED OXYGEN (± 10%)	
0815	8.53	ft BGS (circle one)	ml/min	ft TIC (circle one)	6.58	1,343	4.98	16.09 - 34.8 17
0840	8.65	400		ft TIC (circle one)	6.47	1,397	3.58	16.39 - 54.9 7
0845	8.65	400		ft TIC (circle one)	6.45	1,413	3.33	16.42 - 49.2 5
0850	8.65	400		ft TIC (circle one)	6.44	1,416	3.37	16.51 - 44.3 5
0855	8.65	400		ft TIC (circle one)	6.43	1,426	2.54	16.44 - 42.5 6
0900	8.65	400		ft TIC (circle one)	6.43	1,430	2.10	16.67 - 41.0 4
0905	8.65	400		ft TIC (circle one)	6.43	1,433	2.15	16.68 - 39.5 5
0910	6.5	8.65	400	ft TIC (circle one)	6.42	1,430	2.17	16.72 - 37.5 5
0920 Sample Time								

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values: DO = 0.3 - 10 mg/L  
Redox Potential = -100 - +600 mV  
Spec. Conductivity (µS/cm) = 0.01 - 5,000; up to 10,000 in industrial, ~55,000 in high salt content water. Note: 1,000 µS/cm = 1 mS/cm

Turbidity = 0 - >500 NTUs  
Turbidity = Top of Inner Casing Below Ground Surface

Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

$$\text{Ferric Iron} = 0.01 \text{ mg/l}$$

DATE: 3/2/12  
SAMPLERS: CK

WELL #: MW-13D

SAMPLERS: *ck*

DEPTH OF PUMP INTAKE: 65 ft TIC or ft BGS (circle one)

WEATHER CONDITIONS: Sunny 35°  
80-23°  
SAMPLE ID: MW-13D-030712  
GLP ID:

SAMPLE ID: MW-13D-030712  
CLP ID:

SAMPLE TIME: 07/0

107

**SCREENED OPEN BOREHOLE INTERVAL:** 60 - 70  
**ft TIC**  **ft BGS**  
(Circle one)

(circle one)

**Instrument Type/Model:**  
Complete and/or Circle all that apply

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values: DO = 0.3 - 10 mg/L Redox Potential = -100 - +600 mV Turbidity = 0 - >500 NTUs  
Spec. Conductivity ( $\mu\text{S}/\text{cm}$ ) = 0.01 - 5,000; up to 10,000 in industrial, ~55,000 in high salt content water. Note: 1,000  $\mu\text{S}/\text{cm}$  = 1  $\text{mS}/\text{cm}$

Turbidity = 0 :  $\geq 500 \text{ NTU}$

Redox Potential = -100 - +600 mV

**Typical values:** DO = 0.3 - 10 mg/L Spec. Conductivity (S)

BGS = Below Ground Surface

TIC = Top of Inner Casing

## Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/8/12

SAMPLERS: F. Roline

Conc/m = 0.65 mg/l

WELL #: MW-145

WEATHER CONDITIONS: Partly cloudy 60°

DEPTH OF PUMP INTAKE: 29'  ft TIC or ft BGS (circle one)SCREENED OPEN BOREHOLE INTERVAL: 24 - 34  ft TIC or ft BGS (circle one)

SAMPLE ID: 86 236-MW/145-030812 CLP ID:

SAMPLE TIME: 0900

SAMPLE FLOW RATE: 100 ml/minute

Instrument Type/Model:  
Complete and/or Circle at right

CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	SI Model: <input checked="" type="checkbox"/> CasxL / Horiba U-22 (circle one)		Instrument: <input checked="" type="checkbox"/> <input type="checkbox"/> <input type="checkbox"/>
					PH (± 0.1 SU)	SPECIFIC CONDUCTIVITY (± 3%)	
24-Hour	Gallons/liter <input checked="" type="checkbox"/> ft BGS (circle one)	ft TIC <input checked="" type="checkbox"/> ft BGS (circle one)	nl/min	SU	S/cm, <input checked="" type="checkbox"/> $\mu\text{S}/\text{cm}$ (circle one)	mg/L (not %)	NTUs
0750	6.78	275		6.31	1.708	1.00	14.84 -67.7 7000
0815	11.38	200		6.62	1.691	2.91	15.58 0.6 7000
0820	12.00	200		6.62	1.690	2.96	15.71 2.5 317
0825	12.40	200		6.62	1.680	2.98	15.88 12.1 355
0830	12.88	200		6.62	1.690	2.89	16.15 9.7 337
0835	13.02	100		6.63	1.693	2.81	16.38 7.4 287
0840	13.00	100		6.62	1.692	2.78	16.53 8.9 241
0845	13.00	100		6.62	1.692	2.76	16.48 11.4 241
0850	13.40	100		6.62	1.688	2.72	16.91 11.5 156
0855	3.5	13.00	100	6.62	1.687	2.71	16.30 19.8 124

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values: DO = 0.3 - 10 mg/l Redox Potential = -100 - +600 mV  
Spec. Conductivity ( $\mu\text{S}/\text{cm}$ ) = 0.01 - 5,000; up to 10,000 in industrial, ~55,000 in high salt content water. Note: 1,000  $\mu\text{S}/\text{cm}$  = 1 mS/cm

TIC = Top of Inner Casing BGS = Below Ground Surface

Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/7/2 FLOW: 0.13 mg/l  
ECD: 1000 GROUNDWATER SAMPLING PURGE RE  
WEH #: MW-150

### SAMPLERS: ← Robin

DEPARTAMENTO

WEATHER CONDITIONS: Clear 40°  
SAMPLE ID: 80236 - MW50-03072 SAMPLE TIME: 07:55  
SCREENED/OPEN BOREHOLE INTERVAL: 60 - 170' ft TIC or ft BGS (circle one)  
SAMPLE FLOW RATE: 250 ml/minute  
DEPTH OF PUMP IN TUBE: 63 ft TIC or ft BGS (circle one)  
(circle one)

SAMPLE ID: 80236 - 37W15D - 030712 SAMPLE TIME: 0855  
CLP ID:

**Instrument Type/Model:  
Complete and/or Circle at right**

Instrument Type/Model: Complete and/or Circle at right		YSI Model # 650XL Other (specify)		Horiba U-22 (circle one)		Instrument: Lumex 20-20				
CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	pH (± 0.1 SU)	SPECIFIC CONDUCTIVITY (± 3%)	DISSOLVED OXYGEN (± 10%)	TEMP. (± 10%)	REDOX POTENTIAL (± 10 mV)	TURBIDITY (± 10%)
24-Hour	(gallons) / liters (circle one)	(ft TTD) / ft BGS (circle one)	Units: ml/min	ft TTD / ft BGS (circle one)	SU	S/cm, <u>mS/cm</u> or <u>µS/cm</u> (circle one)	mg/L (not %)	Units: °C	mV	NTUs
0805	6.76	300			6.84	0.693	2.43	14.30	114.9	>10000
0825	6.77	300			7.09	0.717	2.16	14.31	125.8	>10000
0830	6.77	300			7.11	0.717	2.17	14.19	126.7	>10000
0831	6.77	300			7.12	0.719	2.19	15.28	144.6	279
0840	6.78	300			7.13	0.721	2.13	15.13	142.3	187
0845	6.78	300			7.14	0.724	2.09	15.28	140.9	122
0850	6.78	300			7.14	0.722	2.06	15.32	155.2	90.3

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

**Typical values:**

Redox Potential =  $-100 - +600$  mV

Turbidity = 0 - >500 NTUs  
water. Note: 1,000  $\mu$ S/cm = 1 mS/cm

## Paul Miller Site

## LOW FLOW GROUNDWATER SAMPLING PURGE RECORD

DATE: 3/6/12

Average flow = 0.07 ml/s WELL #: MW-165

SAMPLERS: Frank Elgin

DEPTH OF PUMP INTAKE: 30'

WEATHER CONDITIONS: Clear ~30°

SAMPLE ID: 80230-MW/S - 030612

CLP ID:

SCREENED/OPEN BOREHOLE INTERVAL: 25 - 35 ft TIC or ft BGS (circle one)

SAMPLE TIME: 0915 SAMPLE FLOW RATE: 200 ml/minute

Instrument Type/Model:  
Complete and/or Circle at right

YSI Model # 600 / Horiba U-22 (circle one)

Other (specify)

CURRENT TIME	VOLUME PURGED	DEPTH TO WATER	FLOW RATE	DRAWDOWN (± 0.3 FT)	pH (± 0.1 SU)	SPECIFIC CONDUCTIVITY (± 3%)	DISSOLVED OXYGEN (± 10%)	REDOX POTENTIAL (± 10 mV)	TURBIDITY (± 10%)	Instrument: Lcomat 2020
24-Hour	(gallons/liters) (circle one)	(ft TIC) (circle one)	ft BGS	Units: ft TIC / ft BGS (circle one)	SU	S/cm, <u>mS/cm²</u> or µS/cm (circle one)	mg/L (not %)	Units: mV °C	NTUs	
0810	9.51	200		6.30	2.060	2.012	12.95	216.2	>1000	
0825	10.61	200		6.98	1.961	1.97	13.68	150.9	295	
0830	10.73	200		6.98	1.945	1.98	13.92	149.2	271	
0835	10.74	200		6.92	1.913	1.93	15.10	142.7	215	
0840	10.75	200		6.88	1.886	1.63	14.86	147.7	215	
0845	10.75	200		6.88	1.877	1.62	15.14	142.7	138	
0850	10.77	200		6.86	1.872	1.68	14.82	139.8	163.6	
0855	10.77	200		6.86	1.866	1.62	15.22	132.8	93.4	
0900	10.77	200		6.85	1.859	1.62	15.33	129.9	37.2	
0905	4.5	10.72	200	6.85	1.859	1.63	15.37	129.9	28.6	

Drawdown is not to exceed 0.3 feet. Flow rate should not exceed 500 ml/min during purging or 250 ml/min during sampling. Readings should be taken every three to five minutes. The well is considered stabilized and ready for sampling when the indicator parameters have stabilized for three consecutive readings by the measurements indicated in parenthesis.

Typical values: DO = 0.3 - 10 mg/L Redox Potential = -100 - +600 mV  
Spec. Conductivity ( $\mu\text{S}/\text{cm}$ ) = 0.01 - 5,000, up to 10,000 in industrial, -55,000 in high salt content water. Note: 1,000  $\mu\text{S}/\text{cm}$  = 1 mS/cm

TIC = Top of Inner Casing BGS = Below Ground Surface

Turbidity = 0 - >500 NTUs



**ALL-WEATHER  
ENVIRONMENTAL FIELD BOOK**

Name Seth Kellogg  
CJM-Smith

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Project Former Paul Miller Site  
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NYSDEC : Job # 0897-80230

**This book is printed on "Rite in the Rain" All-Weather Writing Paper - A** unique paper created to shed water and enhance the written image. It is widely used throughout the world for recording critical field data in all kinds of weather. For best results, use a pencil or an all-weather pen.

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Location Staten Island NY

Project / Client Paul Miller / 20480 EC

## Synoptic Water levels

now

3/8/12

Date \_\_\_\_\_

3

0630.	F. Robinson onsite		
Clear	33°		
0640.	Scott Kellogg onsite		
0650.	Ed Kuliksky onsite		
0755.	Pine on site dropping off equipment		
0810-	finished synoptic round of water levels		
MW#	OTW (ft TIC)	PTD (PPM)	Depth to Bottom (ft TIC)
1	8.94	49	16
2	3.41	0.3	12
3	3.60	0.5	13
4	3.56	0	12
8S	7.00	18	35
9S	4.30	0	35
90	4.32	0	70
10S	4.55	2	32
10D	5.90	0.1	70
11S	6.00	603	35
11D	5.95	2.3	70
12S	6.13	2.78	35
13S	8.70	112	35
13D	8.28	2.5	70
14S	6.70	9999	34
15D	6.68	17.5	70
16S	9.34	57	35

Total Mv 3/8/12

4

Location SI, NY

Date 3/5/12

Project / Client Paul Miller

MW-95/90

0920. Cal K. calibrated c/w 3 YSI water quality instruments and 2 PLE 2 LaMotte 2020 turbidity meters. See Cal Sheets for details.

0940. Setting up at MW-95 + MW-90

1050. Sampled MW-090 w/ m/s/mso

Lat. fw Parameters:

pH = 7.09 Sp. cond = 0.809 O<sub>2</sub> = 2.38

T °C = 15.82 Redox = 12.67 turb = 51.0

Sample designation: 80230-MW090-030512

1055. Sampled MW-095

pH = 6.82 Sp. cond = 0.804 O<sub>2</sub> = 0.42

T °C = 16.11 Redox = -28.4 TDS = 139

1145. Ferrass Iron via Hach Acc Vac's:

MW-093 = 0.50 mg/l MW-090 = 0.06 mg/l

Sample suit for all MW's:

VOC, Dissolved gases, TOC, Nitrogen (ammonia and organic), hardness, total sulfides and TSS, TDS, Alkalinity and iron chromo. (low) ms/mso only for VOC + dissolved gases (mEE)

1200. Decommissioning Grunefix pumps: ran 8 min. w/ Alkanox + water, then 5 min tap water rinse then 5 min w/ deion. water.

1215. Collected Field Blank of Grunefix Pump after decommissioning

Frank Ma 3/5/12

MON

3/5/12

Location

SI, NY

Date

3/5/12

Date

3/5/12

Date

3/5/12

5

Project / Client

Paul Miller

MW-103/100

1215. Field Blank

80230-FB-030512

1235. Setting up at MW-105 + 100

1250. Starting to pump MW-25

1345. Sampled MW-100

" 80230-MW100-030512"

Final fw Parameters:

pH = 7.26 Cond = 0.773 O<sub>2</sub> = 0.62

T °C = 15.33 ORP = 46.0 Turb = 14.3

Ferrass Iron = 0.00 mg/l

1400. Sampled MW-105

" 80230-MW105-030512"

Final fw Parameters:

pH = 6.64 Cond = 1.409 O<sub>2</sub> = 1.50

T °C = 17.88 ORP = -31.8 Turb = 187 NTU

Ferrass Iron = 1.63 mg/l

1430. Decommissioning pumps as described earlier.

1500. Leaving the site, Ed taking the car

to Geddes: A/B/E 8764 5236 7140

3/5/12

Frank Ma

6

Location

SI, NY

Tue

Date

3/6/12

Project / Client

Paul Miller / NYIDEC

MW-1 / MW-16S

0630 - K. Robinson on site

Clear 28° light wind

0645 - Ed Kuliksky on site

0740 - Calibrated 2 YSI water quality meters and  
2 Lamott 2020 turbidity meters. See C/I  
Sheets for details

- Setting up at MW-1 and MW-16S

0910 - Sampled MW-02

" 80230 - MW-02 - 030612" same sample unit  
as yesterday for all samples

Final GW Parameters:

pH = 6.85 Cond = 4.996 DO<sub>2</sub> = 1.73

T°C = 15.90 ORP = -96.8 Turb = 54 NTU

Ferrous Iron (10<sup>-3</sup> dilution) = 5.4 mg/l

0915 - Sampled MW-16S

" 80230 - MW16S - 030612"

Final GW Parameters:

pH = 6.85 Cond = 1.859 DO<sub>2</sub> = 1.63

T°C = 15.37 ORP = 129.3 Turb = 28.6 NTU

Ferrous Iron = 0.07 mg/l

1005 - Setting up on MW-02 + MW-04

1015 - Decommissioning pumps as per yesterday's procedure

1030 - D62 Collected field blank of decommissioned

Pump "80230 - F6 - 030612"

JL 3/6/12

Tue

Location

SI, NY

Tues

Date

3/6/12

Project / Client

Paul Miller / NYIDEC

MW-02 / MW-04

1200 - Sampled MW-02

" 80230 - MW02 - 030612 "

Final GW Parameters:

pH = 6.86 Cond = 1.667 DO<sub>2</sub> = 0.79

T°C = 13.40 ORP = -63.5 Turb = 11 NTU

Ferrous Iron (10<sup>-3</sup> dilution) = 0.92 × 10 = 9.2 mg/l

1210 - Sampled MW-04

" 80230 - MW04 - 030612 "

Final GW Parameters:

pH = 7.00 Cond = 1.902 DO<sub>2</sub> = 1.17

T°C = 14.89 ORP = -60.7 turb = 30.5 NTU

Ferrous Iron (10<sup>-3</sup>, Dilution) = 0.51 × 10 = 5.1 mg/l

NOTE: GW Parameters Units:

Specific Conductivity = mS/cm

DO<sub>2</sub> = mg/l

T = °C

Redox Potential (ORP) = mV

Turbidity = NTU

1240 - Decommissioning pumps as before

All purge + decom water going into

55 gal drums.

1255 - Ed will be taking cooler w/ samples  
to Fedex (Edison)

4B # 8764 5236 7/39

JK 3/6/12

8

Location SI, NY

Tues

Date

3/6/12

Project / Client Paul Miller / NYSDEC

1330 - Leaving the site: Ed's bring the sample cooler to Kedex in Edison and getting supplies in the warehouse.

1830 - Finished making up labels and making up bottle sets. Took longer than should have due to problem with labels paper backing hard to separate off the adhesive.

~~3/6/12~~

Location SI, NY

Wed

Date 3/7/12

Project / Client Paul Miller / NYSDEC

MW-15D / MW-13S/D

0630 - F. Robinson onsite

Clear 37°

0645 - E. Kuliksky onsite

0740 - Finished calibrating successfully 3 YSI water quality instruments and 2 Lamotte 2020 turbidity meters. See all sheets for details

0745 - Setting up at MW-15D and MW-13 S/D.

0855 - Sampled MW-15D

" 80230 - MW-15D - 030712 "

Initial GW Parameters:

 $\text{pH} = 7.4$  Cond: 0.722  $\text{DO}_2 = 2.06$  $T^{\circ}\text{C} = 15.32$  ORP: 155.2 turb: 90.3 NTU

Previous Iron: 0.13 mg/l

0910 - Sampled MW-13D

" 80230 - MW-13D - 030712 "

Initial GW Parameters:

 $\text{pH} = 7.25$  Cond: 0.790  $\text{DO}_2 = 1.81$  $T^{\circ}\text{C} = 15.94$  ORP: 80.4 turb: 30 NTU

Previous Iron: 0.01 mg/l

0920 - Sampled MW-13S

" 80230 - MW-13S - 030712 "

End of 3/6/12

10

Location SI, NY

Wet

Date 3/2/12

Project / Client Paul Miller / NYSDEC

MW-3 / MW-85

0920 (cont'd) Final GW Parameters for MW-1, 3, 5:

 $pH = 6.42$  Cond = 1.430  $D_{O_2} = 2.17$  $T^{\circ}C = 16.72$  ORP = -37.5 turb = 5 NTU

Ferrous Iron = 0.53 mg/l

0945 - Decommissioning all 3 pumps as indicated  
on the 1st day.1010 - Collected Field Blank of discarded pumps  
using lab supplied water w/ full suite of bottles  
" 80230 - FB - 030712"

1030 - Setting up at MW-3 and MW-85

1140 - Sampled MW-85

" 80230 - MW 085 - 030712"

Final GW Parameters:

 $pH = 7.01$  Cond = 0.564  $D_{O_2} = 4.77$  $T^{\circ}C = 17.99$  ORP = 157.3 turb = 54.1 NTU

Ferrous Iron = 0.05 mg/l

1150 Sampled MW-3

" 80230 - MW 03 - 030712"

Final GW Parameters:

 $pH = 7.06$  Cond = 1.408  $D_{O_2} = 0.55$  $T^{\circ}C = 13.46$  ORP = -125.2 turb = 7 NTU

Ferrous Iron: 10% dilution = 2.56

$$\frac{25.6}{10} \text{ mg/l}$$

JUL 12 2012

wed

Location SI, NY

wed

Date 3/2/12

Project / Client Paul Miller / NYSDEC

1230 - Finished Decommissioning both Gravel  
pumps as described earlier.  
mw-3 - Pickup up cooler : Ed will take  
to Edison Federal office

AIA # 876453 42

AB# 876452 36 7128

1245 - E.K. leaving the site for Edison Federal  
and getting supplies @ the Warehouse

1300 - Leaving the site

1445 - Received cooler from Kedex  
with 1DW sample bottles1620 - Labeled bottles and makeup COC  
form.

3/2/12

CT-200

15

12

Location SI, NY

Thurs  
Date 3/8/12

Project / Client Paul Miller / NYDEC

MW-145 / MW-115/0

0620 - F. Robinson on site

Partly Cloudy 54° winds fm SW

0630 - Ed K. on site

0730 - Calibrated 3 YSI + 2 Lamo &amp; turbidity instruments successfully. See cal Sheets for details

0735 - Setting up at MW-145 and MW-115 + 110.

0900 Sampled MW-145

" 80230 - MW145-030812"

Final GW Parameters:

pH = 6.62 Cond = 1.687 DO<sub>2</sub> = 2.71

T°C = 16.30 ORP = 19.8 Turb = 124 NTU

Ferrous Iron = 0.65 mg/l

0900 - Sampled MW-110

" 80230 - MW110-030812"

Final GW Parameters:

pH = 7.30 Cond = 0.796 DO<sub>2</sub> = 1.53

T°C = 15.75 ORP = 114.8 Turb = 188 NTU

Ferrous Iron = 0.67 mg/l

0925 - Sampled MW-115

" 80230 - MW115-030812"

Total Iron 368 mg/l

Thurs

3/8/12

Date

Location SI, NY

Project / Client Paul Miller / NYDEC

MW-125 / IDW Sampling

0925 - MW-115 Final GW Parameters

pH = 6.19 Cond = 1.456 DO<sub>2</sub> = 0.94

T°C = 17.13 ORP = 83.5 Turb = 51 NTU

Ferrous Iron = 0.06 mg/l

1000 - Collected Field Blank of deionized Grumman sub. pump using bid supplied water. " 80230 - FB-030812"

1015 - Setting up at MW-125

1110 - Sampled MW-125 + MW-112S (Duplicate Sample)

" 80230 - MW125-030812"

" 80230 - MW112S-030812" (Dip)

Final GW Parameters:

pH = 6.78 Cond = 0.692 DO<sub>2</sub> = 1.81

T°C = 17.39 ORP = 85.1 Turb 21.9 NTU

Ferrous Iron = 0.22

1100 - Sampled IDW - Soil

" 80230 - IDW-Soil"

For PCB's, RCI VOC and Full TCLP

1150 - Sampled IDW - Purge water

" 80230 - IDW - AQ"

SVOC, Pest. Herb. PCB's, RCI, metals and VOC's and for FR.

JR 3/8/12

13

14

Location

S I, NY

Thru

Date 3/8/12

Project / Client Paul Miller / NYSDER

Surveyor.

1130 - Terry Mc Kivin (Bryant Assoc.)

Surveyor.

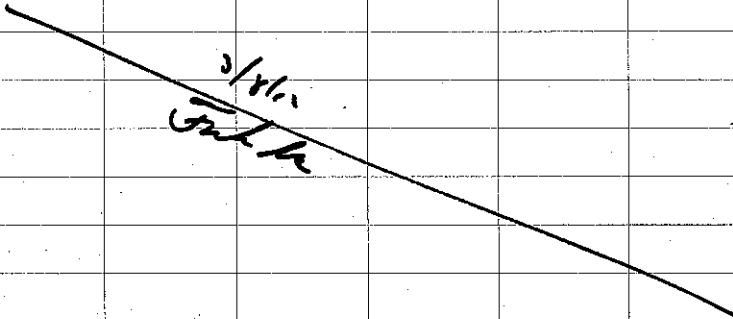
1215 - Ed K. leaving the site to drop off  
both coolers & Tdex in Edison and  
also bring equipment back to Pine.

1240 - Seth Kellogg (com) leaving the site

1330 - Surveyor cannot get his instrument  
to work, wanted to use GPS but  
Seth said no.1335 - Seth told me to help him out  
shutting each well1430 - Finished getting elevations of  
MW's 1, 2, 3 + 4 and other site  
control point. Also did elevations of MW-16S  
to check with elevations (16S done previously)  
Kevin w/ GPS location of MW's.

1445 - Leaving the site

~~3/8/12~~  
~~Leave site~~



Location

S I, NY

Thru

Date

3/8/12

Project / Client

Paul Miller / NYSDER

NOTES

- Purgings and Sampling of the MW's were done with a 2" Grubbs sub. pump using 3/8" ID tubing lined w/ Teflon.

- Pumps were deconned by running them through an alconox/potable water for 5 min, then a potable water rinse for 5 min and finally reagent grade water for 5 minutes.

- The sampling suite for all MW's were: VOC's, Dissolved Gases, TOC, Nitrogen (ammonia + organic), Hardness, total sulfides and TSS, TDS, Alkalinity, and ion Chrom. (low).

- Ferrous Iron was determined by a Hach colorimeter DR/890 using Ferrass Iron AccuVAC Ampules.

- Water Quality Instrument were a YSI 600 XL with a flow cell and LaMotte 2020E turbidity meters.

- IDW Sampling:

Soil sample was collected using a disposable trowel and placing soil into a disposable aluminum pan to make a composite sample

Fresh to 3/8/12

15

Location SI, NY  
 Project / Client Paul Miller/NYSDEC

Thurs  
3/8/92

NOTES

IDW Sampling

Water Sampling (Aqueous) was done by using a disposable beaker to sample each drum and combining them to make a composite sample in an empty 10 liter lab grade reagent water container.

\* GW Parameters Units:

pH = SU Specific Conductivity = mS/cm

Dissolved Oxygen = mg/l T = °C

Redox Potential (ORP) = mV

Turbidity = NTU

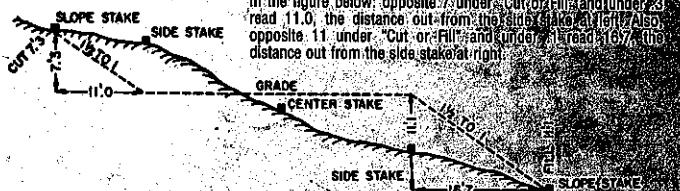
2/2/92

Location \_\_\_\_\_ Date \_\_\_\_\_

Project / Client \_\_\_\_\_

DISTANCES FROM SIDE STAKES FOR CROSS-SECTIONING

Roadway of any Width. Side Slopes:  $\frac{1}{2}$  to 1.  
In the figure below: opposite 7 under Cut or Fill, and under 3 read 11.0, the distance out from the side stake at left. Also, opposite 11 under Cut or Fill, and under 1 read 16.7, the distance out from the side stake at right.



Cut or Fill	0	.1	.2	.3	.4	.5	.6	.7	.8	.9	.0
	Distance out from Side or Shoulder Stake										
0	0.0	0.2	0.3	0.5	0.6	0.8	0.93	1.1	1.2	1.34	1.0
1	1.5	1.7	1.8	2.0	2.1	2.3	2.4	2.6	2.7	2.89	2.1
2	3.0	3.2	3.3	3.5	3.6	3.8	3.9	4.1	4.2	4.4	3.2
3	4.5	4.7	4.8	5.0	5.1	5.3	5.4	5.6	5.7	5.9	4.3
4	6.0	6.2	6.3	6.5	6.6	6.8	6.9	7.1	7.2	7.4	5.4
5	7.5	7.7	7.8	8.0	8.1	8.3	8.4	8.6	8.7	8.9	6.5
6	9.0	9.2	9.3	9.5	9.6	9.8	9.9	10.1	10.2	10.45	7.6
7	10.5	10.7	10.8	11.0	11.1	11.3	11.4	11.6	11.7	11.9	8.7
8	12.0	12.2	12.3	12.5	12.6	12.8	12.9	13.1	13.2	13.4	9.8
9	13.5	13.7	13.8	14.0	14.1	14.3	14.4	14.6	14.7	14.9	10.9
10	15.0	15.2	15.3	15.5	15.6	15.8	15.9	16.1	16.2	16.4	11.0
11	16.5	16.7	16.8	17.0	17.1	17.3	17.4	17.6	17.7	17.9	11.1
12	18.0	18.2	18.3	18.5	18.6	18.8	18.9	19.1	19.2	19.4	12.2
13	19.5	19.7	19.8	20.0	20.1	20.3	20.4	20.6	20.7	20.9	13.3
14	21.0	21.2	21.3	21.5	21.6	21.8	21.9	22.1	22.2	22.4	14.4
15	22.5	22.7	22.8	23.0	23.1	23.3	23.4	23.6	23.7	23.9	15.5
16	24.0	24.2	24.3	24.5	24.6	24.8	24.9	25.1	25.2	25.4	16.6
17	25.5	25.7	25.8	26.0	26.1	26.3	26.4	26.6	26.7	26.9	17.7
18	27.0	27.2	27.3	27.5	27.6	27.8	27.9	28.1	28.2	28.4	18.8
19	28.5	28.7	28.8	29.0	29.1	29.3	29.4	29.6	29.7	29.9	19.9
20	30.0	30.2	30.3	30.5	30.6	30.8	30.9	31.1	31.2	31.4	20.0
21	31.5	31.7	31.8	32.0	32.1	32.3	32.4	32.6	32.7	32.9	21.1
22	33.0	33.2	33.3	33.5	33.6	33.8	33.9	34.1	34.2	34.4	22.2
23	34.5	34.7	34.8	35.0	35.1	35.3	35.4	35.6	35.7	35.9	23.3
24	36.0	36.2	36.3	36.5	36.6	36.8	36.9	37.1	37.2	37.4	24.4
25	37.5	37.7	37.8	38.0	38.1	38.3	38.4	38.6	38.7	38.9	25.5
26	39.0	39.2	39.3	39.5	39.6	39.8	39.9	40.1	40.2	40.4	26.6
27	40.5	40.7	40.8	41.0	41.1	41.3	41.4	41.6	41.7	41.9	27.7
28	42.0	42.2	42.3	42.5	42.6	42.8	42.9	43.1	43.2	43.4	28.8
29	43.5	43.7	43.8	44.0	44.1	44.3	44.4	44.6	44.7	44.9	29.9
30	45.0	45.2	45.3	45.5	45.6	45.8	45.9	46.1	46.2	46.4	30.0
31	46.5	46.7	46.8	47.0	47.1	47.3	47.4	47.6	47.7	47.9	31.1
32	48.0	48.2	48.3	48.5	48.6	48.8	48.9	49.1	49.2	49.4	32.2
33	49.5	49.7	49.8	50.0	50.1	50.3	50.4	50.6	50.7	50.9	33.3
34	51.0	51.2	51.3	51.5	51.6	51.8	51.9	52.1	52.2	52.4	34.4
35	52.5	52.7	52.8	53.0	53.1	53.3	53.4	53.6	53.7	53.9	35.5
36	54.0	54.2	54.3	54.5	54.6	54.8	54.9	55.1	55.2	55.4	36.6
37	55.5	55.7	55.8	56.0	56.1	56.3	56.4	56.6	56.7	56.9	37.7
38	57.0	57.2	57.3	57.5	57.6	57.8	57.9	58.1	58.2	58.4	38.8
39	58.5	58.7	58.8	59.0	59.1	59.3	59.4	59.6	59.7	59.9	39.9
40	60.0	60.2	60.3	60.5	60.6	60.8	60.9	61.1	61.2	61.4	40.0

NYSDEC STANDBY

Paul Miller Drycleaners  
1465 Forest Ave Port Richmond  
Site # 2-43-018 Staten Island  
NY

CDM Project Mgr: Dave Kiel  
516-496-8400

ChemTech Ari 908 789 8900x  
108

Boston Market (Maria, Tom, Sherry)  
718 815 1198

Pine Environmental  
800 301 9663 (Donna)  
PO# 0897-63417

Delta Well  
631 981 2255

The paper in this book is  
made of 50% high grade rag stock with  
a WATER RESISTING surface sizing.

## CONTENTS

R&Chenf 9/4/13

- 0600 RGC arrives on site. 71° Hwy  
 0610 Delta arriving B11 B11, Brian  
 Delta's plan today is to install  
 flush-mount manholes at SP  
 wells and patch existing sidewall  
 No development planned for today.  
 0700 Saw-cutting patch and manhole areas  
 in front of Boston Market on Forest Ave.  
 One man left with drill rig -  
 Bill & Brian remain to patching  
 manholes.  
 0715 Cut patch manholes on side  
  of Boston Market - move to  
 Michael's parking lot location.  
 0755 Fired up at Michael's KFC.  
 Diggers head to hydrant to fill water tank.  
 0825 D Grove arrives / AC leaves  
 1015 DRILLERS POUR CONCRETE  
 AROUND WEEKS  
 1025 DRIVER BEGIN PLACING  
 WEEKS FOR MANHOLE  
 PLACEMENT AND CONCRETE WORK.  
 WEEKS ARE LOCATED ON SIDE  
 OF BOSTON MARKET next to  
 KFC DRIVEWAY. 3 weeks

D Brown

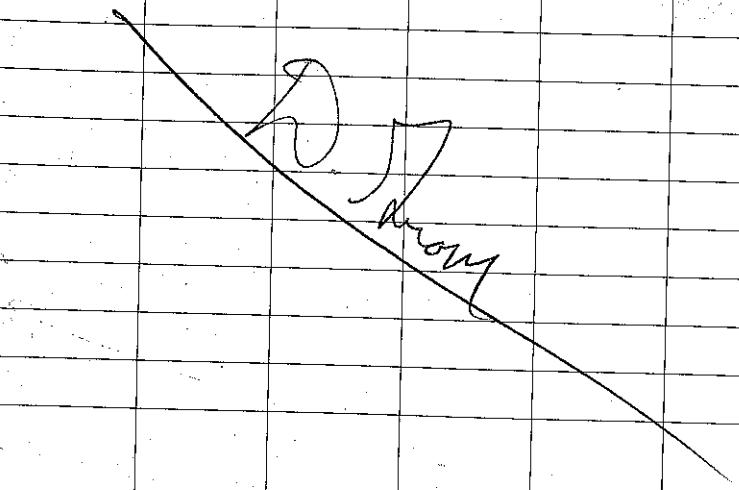
2. P. MILLER WELD INSTALL  
9/14/08 D. GROVE

1115 FINISH WORKS ON 3 WELDS  
IN BETWEEN BOSTON  
MARKET AND KFC. 2 HOLES  
PATCHED w/ HOT PATCH - ONE  
WELL w/ CEMENT.

1120 DRILLERS BEGIN BACK DOWN  
OF EQUIPMENT THAT WAS TO  
GO BACK TO SIFOP. DRUM  
DELIVERY HAS BEEN SCRAPPED  
FOR LATER TODAY for 15  
DRUMS.

1230 DRUM SHIPMENT ARRIVES  
AT C140. TOTAL 15 DRUMS

1250 WELLS SECURED. SITE  
CLEANED AND DEMOBED  
LEAVE SITE w/ DRILLERS



4/15/08 Rec Check

3

0800 Arrive at site, Clear 78°  
Delta drilling equipment - Bill R.  
Monolith completed yesterday include  
wells in KFC lot and well  
on Boston Market lot next to KFC,  
plus to holes abandoned at this location  
were patched with mortar.  
AC (CP4 and P3? location) (11 S location)

0830 Worked on one well and  
abandoned lines in front of KFC  
(16 S location)

0740 Finished 2 16S - add some  
cement to 11 S location

0800 Move to Michael's parking lot

1100 Finished 4 monolith on st. check  
preliminary drums of grout / cement  
are set at wells in Michael's lot  
to allow concrete to set.

0830 plastic road "clay" set  
on well in front of Boston Market -  
prior well will this flow. AC  
too low to just for tape to wrap  
around drums at Michael's.

1145 - Wood over from ground  
drums at Michael's - less sh

Rec 4/5/08

4

09/08/2008

0600 S.Britch arrive on site.  
Delta already on site.  
Weather: ~70°F, slight breeze.  
DPE: Level D.

0618 Begin developing MW9D  
Depth to bottom from unercase:  
69'. DTW: 32.5'. Using a Grumius  
pump + plastic tubing

0630 Starting to pump into a  
55 gal drum.

0635 Stop pumping b/c no  
material

0637 Tested pump in a 5 gal  
bucket of water. No flow, pump  
appears to be jammed. Switching  
pumps

0641 testing pump #2 Grumius  
in 5 gal bucket of water

0642 Pump #2 no flow, switch  
to #3.

0645 testing pump #3 in 5 gal  
bucket of water + works

0653 Start pumping MW9D w/  
Grumius pump #3, plastic  
tubing powered by bobcat battery.

SJB 09/08/2008

5

09/08/2008

0657 Depth to water = 32.5' IS  
incorrect. DTW = 8'. Water has  
slight sheen

0820 DTW pumped 9.1'. Cristina  
called + said we have no  
access for wells MW9D, 9S, 10D,  
10S so can't develop until state  
gets access agreement.  
~85gals / 90min = ~1gal/min

0838 Begin developing MW13D.  
DTW = 12', DT Bottom = 69.6'

0900 Starting to pump MW13D  
w/ Grumius pump

1055 DTW = 12.1" Flushed pump.  
~110gall / ~120min = ~1gal/min

1059 Begin developing MW13S  
DTW = 12.0"

1105 Starting to pump MW13S  
w/ Grumius pump + plastic tube

1116 Lunch break while  
MW13S ~~OSB~~ nos

recovers b/c it isn't yielding  
water (may be due to material  
screen is up)

1146 lunch over. DTW = 20'. Let  
recover a little more

SJB 09/08/2008

6

09/08/2008

1200 DTW = 18'. Still recovering  
 1216 DTW = 16.5' start pumping  
 again

1251 Lost yield (pumped ~2gal)  
 Let recover

1301 DTW = ~19'. Begin  
 pumping for first time today.  
 Let recover overnight +  
 continue tomorrow

1305 finished pumping MW13S  
 lost yield after ~2gal. Will  
 let recover overnight. Cleaning  
 & securing site.

1330 SB + Delta off site

gms dead

SJB  
 09/08/08  
 SJB

09/09/2008 7

0559 SB on site. Weather:  
 ~70°F, breezy. Level PPC:D  
 0615 Delta on site

0619 Begin developing MW15D  
 DTW = 10.5', DTB = 69.5'

0625 Starting to pump MW15D  
 w/ Grumius pump + plastic  
 tubing into drum

0628 Surging MW13S to  
 clean out well so can pump  
 0640 finished surging MW13S  
 for now

0725 Surging MW16S

0737 finished surging MW16S

0758 finished pumping MW15D  
 110gal / ~90min = ~1.2gal/min

DTW = 10.7'

0810 Begin developing MW16S  
 DTW = 13.7'

0813 Starting pumping MW16S  
 with Grumius pump + plastic  
 tubing

0825 No more yield from MW16S  
 Let recover for a while. yielded about  
 5gal

SJB 09/09/2008

8

09/09/2008

- 0857 taking waste sample  
 D00443723-WC-W-080909-01. ~~D00443723-WC-W-080909-01~~  
 from drums 34-36 (wells 13D + 13S)  
 (3) 1L Amber jars - BNA, Pest, Herb TC2P,  
 11L plastic - corrosivity, 1500ml plastic  
 metals, 2 40mL vials - VOC)  
 0900 taking waste sample  
 D00443723-WC-W-080909-02 from  
 drum 37-39 (wells 15D + 16S)  
 (3) 1L Amber jars - BNA, Pest, Herb TC2P,  
 11L plastic - corrosivity, 1500ml plastic  
 metals, 2 40mL vials - VOC)  
 0937 Starting pumping MW16S  
 0946 no more yield, let recover  
 1028 Starting pumping MW16S  
 DTW = 18'  
 1046 Start pumping MW13S  
 1050 Chemtech on site to pick  
 up samples (1 cooler) ~~NE 080909-01~~  
~~080909-01~~ D00443723-WC-W-080909-  
 01 + 02  
 1052 Chemtech (Lawrence) off site  
 1120 pumping done, cleaning +  
 securing site

SJB 09/09/2008

9

09/09/2008

- 1110 talked w/ Maria from  
 Boston Market + told her that  
 we are done + we will get  
 drums off property within the  
 next few wks  
 1130 lunch break  
 1200 lunch over, packing up  
 1230 Site cleaned + secured.  
 Delta + SB off site.  
 40 Drums used  
 (e. Drums left on site empty  
 46 Total

09/09/2008  
 SJB  
 09/09/2008  
 SJB  
 09/09/2008  
 SJB  
 09/09/2008  
 SJB

10

09/29/2008

0745 Stef Brtch arrived on Site.  
 Pat Connolly already here. Weather ~65°F, slight breeze, clear, Sun. Level PPE: D modified: nitrile gloves, boots.

0805 DTW TOC = 5.32' MWQS

0818 Begin developing MWQS w/ whale pump

0822 Pat left site, stopped developing

0827 Calibrated Hach 2100P Turbidimeter w/ 0.1, 20, 100, 800.

0.1 Reading = 20.0

20 " = 100.0

100 " = 800.0

800 " = 000.0

800 test = 800 NTU  
 20 test = 19.6 NTU

0845 Pat back on Site

0850 Begin pumping MWQS

0900 Turbidity reading = E 3 (too high) initially put pump to bottom + pulled up 5 feet now w/L at top of pump before water source (surface)

SJB 09/29/2008

09/29/2008

11

0913 turbidity = E 3, Last flow so lower pump 2'

0921 DTW 24.30', Turbidity: 1000m

0930 Turb = 672 ntu dtw = 27.69'

0940 Turb = 536 dtw 29.11'

0950 Turb = 662 dtw = 29.86'

1000 Turb = 974 dtw = 29.6'

1010 Turb = 540 dtw = below top of pump. Top of pump dtw = 30.23'

1020 = turb = 640 dtw = top of pump

1030 turb = 998

1040 turb = 1000 dtw top of pump

1050 flow stopped, flushed developing. Volume purged = 37gal  
~~2(11.5')<sup>2</sup>(34.5) = 28053 ft<sup>3</sup>~~

11:23 ~~11.5' x 34.5~~ MWQS dtw = 8.50'

dtb = 24.98

11:35 dtw 7.77' begin developing MWQS

11:40 loss of flow dtw = 24'  
 top of pump = 26' pump + tube because disconnected. Resumed pumping at 11:45. Water extremely turbid.

SJB 09/29/2008

12

09/29/2008

- 1148 loss flow. dtw = 27.75  
 1151 pump is having difficulty maintaining flow. Pulling pump to clean out  
 1155 WL = 25.8 after pump cleaned out. Begin pumping again  
 1158 loss of flow. top of pump.  
 25.95'

\*~~11~~ meas. is top of casing  
 1202 pump removed in order to surge MN10S

1215 attempted to surge MN10S for about 15 min but no water coming out. Water is in the well though.

1220 ~~at 13~~ clogged out surge block b/c clogged w/ mud. attempted to surge again.

1223 clogged w/ mud cleared, out dtw = 26.09'. dtb = 30.60' total volume purged = 3.4 gal

1303 MN10S dtw = 6.36'

dtb = 69.48

1345 began developing MW10D

SJB 09/29/2008

09/29/2008

13

- 1355 Turbid = 164 dtw = 9.40  
 1405 Turbid = 29.3 dtw = 9.40  
 1415 Turbid = 12.9 dtw = 9.40  
 1425 Turbid = 6.86 dtw = 9.40  
 1435 Turbid = 5.31 dtw = 9.40  
 1445 Turbid = 4.62 dtw = 9.40  
 flushed developing MW10D  
 Total Volume purged = 25.2  
 1506 Developing MW10S again  
 dtw = 16.5  
 1510 began developing pump  
 1515 loss of flow until at top of pump at 26.8 ft. flushed developing MW10S  
 1615 took waste sample D00443723-NC-N-050929 (31L amber TCLP Post herb, BNA, 1-1L plastic RCRA Char, 1-500mL Metals, 2 40mL vials TCLP VOC)

4.3 drums total, 3 empty on site  
 1700 Pat + Stef leave site

~~5/13~~  
 09/29/2008

## G.W. SAMPLING

10/13/8 D.GROUP

0700 ARRIVE AT SITE W  
V. Eugene off com.

WEATHER: P. CLOUDY 50-60%

WIND: WEST 15.WEST 0 -10 MPH

CREW: D.GROVE COM ENC.

V. Eugene 11 11

PPE: S.T. BOOTS, S. VESTS, S. GLASSES  
GLOVES

## EQUIPMENT

PID	02601	COM
G.W. PROB	11271	PUMP
W.H. METER	31133	COM
W.H. METER	02496	COM
HORIBA U22	06224	PURE
HORIBA U22	10129	PURE

0710 BEGIN PURGE AT MW-95

WELL PID D.T.P. DTW COMMENTS

95	6.0	-	16.81
9M	0.0	-	10.21
10S	0.0	-	8.1841
10D	0.0	-	9.670
8S	1.2	-	11.60
16S	95	-	10.13
MW-1	0.0	-	11.98
MW-14S	303	-	12.94
MW-4	0.0	-	8.03

J. Brown

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## G.W. SAMPLING

10/13/8 17.6 GONE

PID	D.T.P	DTW
15D	1.3	10.95
12S	0.0	10.35
11D	0.0	9.52
11S	4.3	9.90
13D	0.0	12.51
13S	3.2	12.58

CANNOT FIND MW3 ON MW 2

P-35 CANNOT OPEN

P-3D CANNOT OPEN

P-5 OPENED BUT BLOCKED 2' DOWN °OPM

P-2 CANNOT OPEN

P-4D CANNOT FIND

P-4S CANNOT FIND

P-1 CANNOT OPEN

0930 BEGIN PURGE AT MW-95

\* SEE FIELD PARAMETER SHEET FOR  
ALL FIELD READS.

D.T.W = 11.81 D.T.B = 33.75

DIA = 2" LOW FLOW PUMP  
2" SUB. PUMP.

1013 END PURGE AT MW-95

TOTAL PURGE = 14 GAL D.T.W. = 11.84

1016 COLLECT SAMPLE OFF PUMP

1408C FOR VOC'S AND SVOC'S  
(VOC \$200.8 + 10, SVOC \$27.00 + 120)

J. Brown

15

G.W. SAMPLING  
10/18/8 D.GROUP

D.T.W. = 11.84

ALL VOC AND SVOC SAMPLES WILL  
USE MENTIONED LAB METHODS.

0845 BEGIN PURGE AT MW-9D

D.T.W. = 10.21 D.T.B. = 69.0

DIA = 2" LOW FLOW PURGE

PUMP = 2" SUB.

0958 END PURGE AT MW-9D

TOTAL PURGE = 15.4 GAL D.T.W. = 10.21

1000 COLLECT SAMPLE FROM

MW-9D THROUGH PUMP 1105P

FOR VOC'S AND SVOC'S. DTN = 10.21

1100 BEGIN PURGE AT MW-10D

D.T.W. = 9.63 D.T.B. = 69.0

DIA = 2" LOW FLOW PURGE

PUMP = 2" SUB

1143 END PURGE AT MW-10D

TOTAL PURGE = 9.0 GAL DTW = 9.63

1145 COLLECT SAMPLE FROM MW-10D

THROUGH PUMP 1105 FOR VOC'S

AND SVOC'S. COLLECT ONE SAMPLE

CALLED MW-10X FOR VOC'S

AND SVOC'S AT 1150

1150 BEGIN PURGE AT MW-10S

D.T.W. = 8.84 D.T.B. = 35.25

J. M.  
J. Mowry

16

G.W. SAMPLING

10/18/8 D.GROUP

17

DIA = 2" LOW FLOW PURGE

PUMP = 2" SUB.

1237 SEND PURGE AT MW-10S

TOTAL PURGE = 13.5 GAL D.T.W. = 28.12

1240 COLLECT SAMPLE FROM

MW-10S THROUGH PUMP 1405P

FOR VOC'S AND SVOC'S. D.T.W. = 28.12

1245 BEGIN PURGE AT MW-9

D.T.W. = 8.03 D.T.B. = 12.50

DIA = 2" LOW FLOW PURGE

PUMP = PERISTALTIC

1245 WEAR IS DRY. WILL CHECK  
ON RECHARGE IN 1 HOUR.

1326 AS PER C.RAMACCIOTTI

1440 ARRIVES ON SITE. COLLECT

ANOTHER 1 LITER AMBER FOR

SVOC'S FROM MW-9B (MW-93B)

AND MW-9D (MW-90B) AT 1325

AND 1330. CHANGE TOTAL

NUMBER OF SAMPLE BOTTLES

COLLECTED FROM 2 VOC'S AND

1 1000 ML AMBER (AS PER LAB) TO

2 VOC'S AND 2 1000 ML AMBERS.

1437 BEGIN PURGE AT MW-8S

D.T.W. = 11.60 D.T.B. = 35.25

J. M.  
J. Mowry

G.W. SAMPL

10/13/08 D.GROUP

DIA. = 2" LOW FLOW PUMP

PUMP = 2" SUB

1544 END PUMP AT MW-85

TOTAL PURGE = 15.5 GAL D.T.W. 13.21

1545 COLLECT TEL 1 GALLON

SAMPLE FROM PUMP HIGH

VOC'S, SVOC'S, PCB'S, PCP610005,  
MERCH AND CYANIDE.

COLLECT MOL/MED SAMPLE

FOR VOC'S (6 VOMERO VIALS)

AND SVOC (4 1000 ML AMBIENS)

CHECK MW-4 AGAIN. NO WATER

IN WELL. WILL SAMPLE TOMORROW

1ST SHIFT IN MORNING.

PACK SAMPLES, DEMOB, DECON  
EQUIPMENT.

1610 COLLECT F.B. - FB3981013

FOR VOC'S AND SVOC'S.

1645 LEAVE SITE FOR EDISON

W.H.

*D. Stevens*

18

G.W. SAMPL

19

10/14/08 D.GROUP

0715 ARRIVE AT SITE.

1306IN SET UP AT MW-165

WEATHER: SUNNY 60-75°

EQUIPMENT, FIELD CREW, PIPE

same 142 10/13

0730 V. EUGENE ARRIVED AT

STIC.

CHECK MW WITH MOTORS AND

OLW PROBE ON 165

OLW PROBE 13.13

W.H. 31133 13.13

W.H. 02496 13.13

CAL SOLUTIONS FOR CAL CHECKS

DONE ON 10/13, 14/08

100 ppm ISOTOPES

LOT # EXP DATE

HQA 13A 80L

LOT # 64245 7/24/9

1306IN PURGE AT MW-165

D.T.W. = D.T.B. =

DIA = 2" LOW FLOW PUMP

PUMP = 2" SUB

END PURGE AT MW-165

TOTAL PURGE = D.T.W.

0900 COLLECT SAMPLE FROM

*D. Stevens*

B.W. SANDP.

10/14/8 D. Groves

20

MW-165 through pump tubing  
for VOC's AND SVOC's D.T.W.=

BEGIN PURGE AT MW-1

NO FIELD READS TAKEN. WATER  
HAS 84CON. VERY TURBID

WEAL OXY. WILL CHECK ON  
W.L. AND SAMPLE WHEN WELD  
RECHARGES. USED PERISTALTIC

PUMP. TOTAL PURGE =

0945 BEGIN PURGE ON MW-135

D.T.W. = 12.71 D.T.B. = 35.0

DIA. = 2" LOW FLOW PUMP

PUMP = 2" SUB.

1039 END PURGE AT MW-135

TOTAL PURGE = 11.3 GAL D.T.W. = 1284

1040 COLLECT SAMPLE FROM MW-135  
THROUGH PUMP TUBING FOR  
VOC'S AND SVOC'S. D.T.W. = 12.8d

\* MW-135 COLLECT FULL TOP SAMP.

w/ PESTICIDES, PCB'S, METALS, CHANDEL

0950 BEGIN PURGE AT MW-180

D.T.W. = 12.6 D.T.B. = 68.5

DIA. = 2" LOW FLOW PUMP

PUMP = 2" SUB.

1050 END PURGE AT MW-180

D. Groves

B.W. SANDP.

21

10/14/8 D. Groves

TOTAL PURGE = 9.5 GAL D.T.W. = 12.90

1100 COLLECT SAMPLE FROM MW-130

THROUGH PUMP TUBING FOR

VOC'S AND SVOC'S. D.T.W. = 12.90

1135 BEGIN PURGE AT MW-115

D.T.W. = 10.34 D.T.B. = 35.27

DIA. = 2" LOW FLOW PUMP

PUMP = 2" SUB.

1303 END PURGE AT MW-115

TOTAL PURGE = 18.5 GAL D.T.W. = 11.96

1305 COLLECT SAMPLE FROM  
MW-115 THROUGH PUMP ITOSO  
FOR VOC'S AND SVOC'S.

D.T.W. = 11.96

1145 BEGIN PURGE AT MW-110

D.T.W. = 10.23 D.T.B. = 64.37

DIA. = 2" LOW FLOW PUMP

PUMP = 2" SUB.

1308 END PURGE AT MW-110

TOTAL PURGE = 10.5 GAL D.T.W. = 10.40

1310 COLLECT SAMPLE FROM  
MW-110 THROUGH PUMP ITOSO  
FOR FULL TECH SAMPLE (VOC,  
SVOC, PESTICIDES, PCB'S, METALS,

CHANDEL) D.T.W. = 10.40

D. Groves

G.W. STAMP  
10/14/8 D.Grove

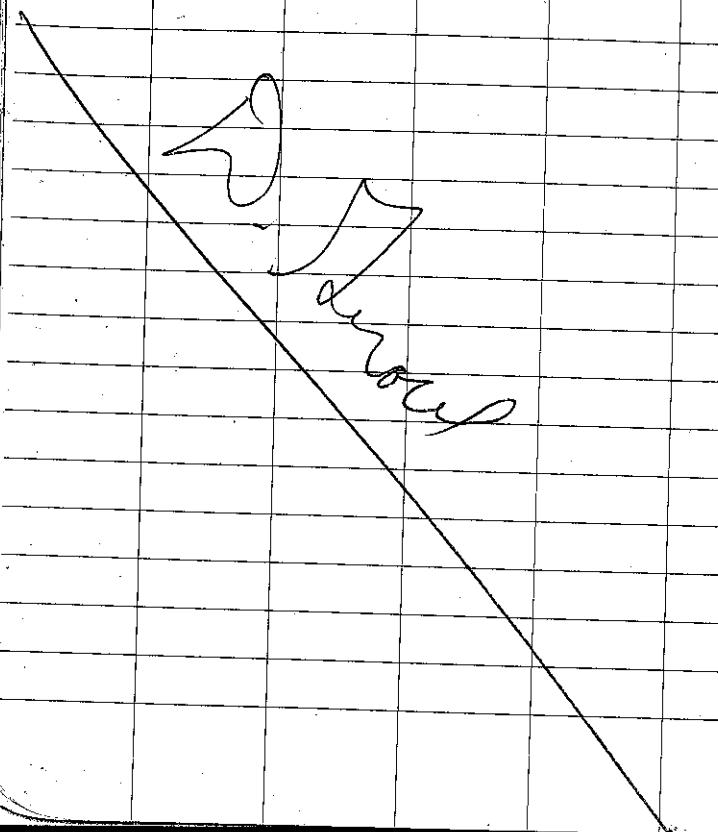
SECURE SITE, DEMOB, DECOOL  
EQUIPMENT.

1415 LEAVE SITE AND RETURN  
TO EDISON W.H. TO PACK, PACK  
SAMPLES FROM 10/13 AND 10/14  
FOR PICK-UP BY LAB.

COC # 075 759

075 764

1600 FIELD BLANK COLLECTED  
FOR VOC'S AND SVOC'S



22

G.W. STAMP.

10/15/8 D.Grove

0730 ARRIVED ON SITE w/  
V. EUGENE at 08M.

BEGIN SET-UP ON MW-125

0818 BEGIN PURGE AT MW-125

D.T.W. = 10.61 D.T.B. = 35.3

PRODUCT DETECTED AT 16.21'

DIA. = 2" FLOW FLOW PURGE

PUMP = 2" SUB

0915 END PURGE AT MW-125

TOTAL PURGE = 8.5 GAL D.T.W. = 10.83

0920 COLLECT SAMPLE FROM

MW-125 THROUGH PUMP TUBING

FOR VOC'S AND SVOC'S, DTW. = 10.83

1010 BEGIN PURGE AT MW-15D

D.T.W. = 11.0 D.T.B. = 30.0

DIA. = 2" FLOW FLOW PURGE

PUMP = 2" SUB

1055 END PURGE AT MW-15D

TOTAL PURGE = 8.0 GAL D.T.W. = 11.03

1100 COLLECT SAMPLE FROM MW-15D

THROUGH PUMP TUBING FOR

VOC'S AND SVOC'S. COLLECT DND

SAMPLE (MW-15X 1105) FOR

VOC'S AND SVOC. DT.W. = 11.03

1115 COLLECT FIELD BLANK

23

J. Knott

G.W. SAMPLING  
10/10/08 D. Gravel

261

(CFD 081015) for VOC's AND  
SUOC'S. 0:

0905 BEGIN PUMP ON MW-145

D.T.W. = 12.94 D.T.B. = 32.6

DIA. = 2" DON KNOW PUMP

PUMP = 2" 8418

1030 WELL DRY. PUMP NOT

BURNING RIGHT DUE TO  
VERY TURBID WATER (SUSPENDED)

WELL ATTEMPT SAMPLE AT

11:30

11:30 MW-145 VERY WINDY  
ACROSSAGE. WATER VERY TURBID.

COLLECT SAMPLE w/ TAINER.

PREP SAMPLE FOR TURN-UP  
FROM W.I. OCT 073769.

SECURE SITE; SECURE DRUMS,  
DEMOB/DECON EQUIPMENT.

1315 LEAVE SITE

40. 7  
loop

25

12/11/2008

0900 CDM(SB) ARRIVED ON SITE

WEATHER: ~37°F, RAINING

0930 ENTERED NORTHFIELD BANK +  
MET W/ JENNIFER. WENT TO  
BASEMENT TO SET UP FOR THE DOH  
SURVEY. THEY HAVE NO AIR DUCT  
WORK + CENTRAL AIR. THE BOILER  
IS IN THE BASEMENT. THE  
BATHROOM + KITCHEN ARE LOCATED  
THERE TOO. WILL MOST LIKELY  
DO BASEMENT SUB-SB IN  
CLOSET. JENNIFER SAID WE ARE  
ALLOWED TO SET UP AMBIENT AIR  
SAMPLE ON MAIN LEVEL BEHIND  
TELLERS.

1000 LEFT BANK.

1030 SURVEYED WHERE TO PUT  
OUTDOOR AIR SAMPLE AT BANK.  
COULD POSSIBLY ATTACH IT TO  
LIGHT POSTS, DRIVE THROUGH  
FENCE THAT SEPARATES BOSTON  
MARKET + BANK.

1100 ENTERING KFC FOR ENVIRON.  
WALK THROUGH. MANAGER NOT  
AVAILABLE. WILL RETURN LATER.

SJB 12/11/2008

12/11/2008

26

1230 entered Boston Market + met w/ Sherry. Walk through + did survey. Have exhausts that vent to roof. All systems are on roof. Use cleaning supplies regularly. Basement was locked + couldn't find key so will have to finish survey on tuesday. Also not sure about driller gettting room. Will call on monday to find out. Will put outdoor sample in garbage ~~in SBP 1108 shed.~~

1300 left Boston Market

1305 entered KFC. Walked through dining room + kitchens. wasn't able to speak w/ anyone so will return tomorrow morning.

1315 left KFC.

1330 leave Site

~~St. James J. Butler  
12/11/2008~~

12/12/2008

27

0930 am re-on site - CDM (SB) Weather: ~40°F, breezy, cloudy. On site to finish DOH Survey w/ KFC + to straighten things out w/ Boston Market.

1000 entered KFC + met w/ Nadine. Walked around again. All systems on roof. Use cleaning products regularly. Have exhausts in kitchen + bathroom. Wants us to put port in kitchen, can't because of cleaning supplies. Will do it in service area most likely.

1050 left KFC.

1100 leave Site

~~St. James J. Butler  
12/12/2008~~

12/16/2008

28

0830 MK + SB (COM) arrive  
on site. Weather: ~39°F,  
Slight breeze, Rain. Level ppe:  
Steel toe boots.

0930 arrive at Northfield  
Bank + Met w/Jer.

0945 MK calibrate pid at  
105 while SB detect metal  
using cable locator. Found  
spot in closet. pid = 0.0 ppm

1009 Setting up tracer test.  
Seal OK - pid = 1.9 ppm.

1018 MK puts tubing down  
hole + connects canister.  
While SB records sample  
id, start time + initial ~~sample~~  
pressure (243018 - SB01, 243018-  
+ canister # + regulator #. +  
initial pid reading + location  
Loc. Sample Id. pid St Time, Init. P.

BankBase. SB01 243018- 1.9 ppm 1026 -28  
" 243018- 1A01 — 1026 -28

Samp. Id.	Can#	reg#
SB01	10299	10517
1A01	10268	10481

SJB 12/16/2008

12/16/2008

29

all also recorded on table  
2 - Sample Location Information  
Summary.

~~1018 SB 1009~~

1025 finished in basement.  
JER sets up indoor air  
canister on first floor by  
tellers. SB records info  
which is also recorded on  
Table 2.

Loc Samp Id PID St Time Init P.  
Bank 243018- FF01 — 1029 -27

Sample Id Can# Reg#  
FF01 10003 10240

1035 MK + SB clean up +  
Set up ambient air sample  
(outside) at the back  
entrance of the bank.

Location Samp Id St. Time Init P.  
Bank outside 243018- 1042 -29

Samp Id Can# Reg#  
1A01 10006 10246

(also recorded on Table 2)

1045 SB + MK leave bank

1055 SB + MK arrive at KFC

SJB 12/16/2008

12/16/2008

30

1100 MK begins drilling hole for Subslab air sample by the garbage bin inside at the Forest Ave entrance.

1110 MK is unable to drill through slab so call Cristina Ramacciotti + she suggests to send someone from office to help. Waiting for Joe Yenchak to arrive.

1237 Joe Yenchak on site. Also Andrew from PW Grosser who is representing the property owner also arrives on site to oversee work.

1240 Drilling by JY begins.

1302 Setting up for tracer test. Seal passed.

1310 take pid reading. Reading = 0.2 ppm. ~~SB 121608~~

1312 Setting up air (SB + MK) canisters for air sampling (SB + MK). Recording info. Info also on Table 2. Indoor air + Subslab.

SJB 12/16/2008

12/16/2008

31

Location	Sample Id	PID	ST Time	Int.P
KFC	243018-SB02	0.2	1325	-30
"	243018-IA02		1325	-30

Sample Id	Can#	Reg#
SB02	10323	10486
IA02	10605	10700

1327 Clean up + leave KFC

1330 Arrive at Boston Market. Talk w/ Sherry + Tom (managers) + get access to basement.

1350 begin drilling (JY) for subslab sample.

1405 Set up for tracer test (MK). Seal passed.

1420 Setting up canisters for Subslab + indoor air plus duplicates for both. Recording info. Info also recorded on Table 2.

Location	Sample Id	PID	ST Time	Int.P
Boston Market Basement	243018-IA03	-	1430	-30
"	243018-IA03D	-	1430	-30
"	243018-SB03	-	1430	-30
"	243018-SB03D	-	1430	-28

SJB 12/16/2008

32 12/16/2008

Sample Id	Can #	Reg#
1A03	10269	10575
1A03D	10025	10181
SB03	10489	10254
SB03D	10258	10509

1437 Setting up outside ambient air sample canister in garbage shed by grease dumpster. Record info. Info also on table 2.

Location Sample ID Sftime In/H/P  
Boston Market 24308-1A02 1440 -30  
Garbage shed

Sample Id	Can #	Reg#
1A02	10445	10214

1447 Clean up equipment.

1453 Enter Boston Market main floor.

1458 JY begins drilling for subslab sample by service area next to refrig/freezer.

1505 MK sets up for tracer test. Seal passed. Pid reading taken. Reading = 166 ppm.

1512 Setting up ~~for SB03~~ Canisters for Subslab + indoor air

SJB 12/16/2008

12/16/2008

33

Sampling. Recording info.

Info also recorded on Table 2  
Location Sample ID Sftime In/H/P  
Boston Market 24308-1A02 166 1517 -30  
main floor SB03 — 1516 -30  
" 24308-1A02 — 1516 -30

Sample ID	Can #	Reg#
SB04	10492	10476
1A04	10313	10551

1526 Cleaning up site, taking pictures of 1A01 + 1A02  
1615 SB, MK, JY + Andrew leave site.

Note: asking customers if they would like earplugs while drilling

12/16/2008

12/16/2008

12/16/2008

12/16/2008

34

12/17/2008

0930 Stefanie Brtch (CDM) on site.  
Weather: ~38°F, light rain,  
Overcast. Level ppe: Steel toe  
boots.

0950 Mel Koberle on site. (CDM)  
1000 enter bank.

1005 SB takes pictures (SB121708)  
pictures of the air canisters  
for documentation. Also SB  
finishes DOH Survey + takes  
pictures of products that  
could effect sample.

1015 MK checking to see if  
canisters' pressure low  
enough.

1020 MK preparing to collect  
samples. Recording info. Also  
recorded on Table 2.

Sample Id Time Collect Final Press.

243018-1A01 1026 -5

243018-FF01 1029 -5

Waiting until SB01 pressure  
go down a bit more.

1040 preparing to collect SB01

Sample Id time collect final P. PID

243018-SB01 1045 -7.5 6.0

SJB 12/17/2008

12/17/2008

35.

1050 MK filling hole in  
slab w/concrete to make  
like original before drilling.  
1100 cleaning up inside  
bank. Also preparing to collect  
1A01.

Sample Id Time Collect Final P.  
243018-1A01 1108 0

Note: pid reading taken for  
sub slab in bank = 6.0 ppm.

1110 leave bank. Break in  
work b/c need to let canisters  
sit for 24 hrs. Waiting until  
that time.

1330 enter KFC. Prepare to  
collect canisters. Recording  
info. Recordings also on Table  
2.

Sample Id Time Collect Final P PID

243018-SB02 1338 -6 0.2

243018-1A02 1338 -6

1343 MK patching up sub slab  
hole. Cleaning up equipment.

1355 leave KFC

1420 enter Boston Market

SJB 12/17/2008



3/9/16 D. Groves

0806 AIR SAMPLING.

0830 ARRIVE AT SITE.

BEGUN EXTERIOR DATA SURVEY  
AT NORTH FIELD BANK BLDG.

WEATHER: P CLOUDY / WINDY

TEMP 40-45°

WIND: NORTH 5-15 MPH

GOT 3406 DIMENSIONS, PICTURES  
(see photo o.d.)

NOTE STORM DRAIN IN BACK  
WEST SIDE OF BLDG.) AND  
LARGE OPEN FENCE IN  
STORM DRAIN IN SOUTH 310P  
AT BLDG.

0910 BANK MANAGER LETS  
ME IN TO PERFORM INTERIOR  
DATA SURVEY. TAKE PICS  
AND DIMENSIONS OF 1ST  
FLOOR. NOTE BATHROOM AND  
ENCLOSING SURFACES. SEE  
VIDEO DATA SETCTI.

0950 GO TO BASEMENT AND  
BEGIN DATA SURVEY. BASEMENT  
CONSISTS OF KITCHEN/ EATING  
AREA, MULTIPLE STORAGE

38

3/9/16 D. Groves

CLOSETS AND UTILITY ROOMS  
AND BATHROOM. ALSO  
DIMENSIONS AND PICS.

ONE LOCKED ROOM CONTAINS  
A OPEN PVC FLOOR PIPE (FOR  
DRAINAGE?) IT MAY BE BURIED  
TO MARKING BUT NO VC ROOM  
CONTAINS WATER METER.  
NOT TWO SCOURP SMALL  
AMOUNT OF CLEANING  
SUPPLIES. NO OBVIOUS CRACKS  
OR OPENINGS IN FLOOR.  
MAKE ARRANGEMENT WITH  
STAFF BRANCH MGR. TO RETURN  
TOMORROW TO PLACE SANDED  
CANISTER.

1005 CONTACT SITFT MGR  
AT 16PC TO SEE IF I  
CAN OBTAIN ACCESS FOR DATA  
SURVEY. HE WILL NOT GIVE  
ME HIS NAME NOR TELON  
ACCESS BUT DOES GIVE M/T  
PHONE NUMBER FOR RCG.  
MGR. (ROY). CONTACT AND  
LEAVE MESSAGE w/ ROY

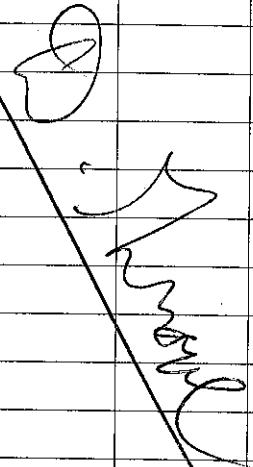
39

D. Groves

319/11 D-GROUP

(40)

CONTACT C-RANCHER AT  
COM. 3 NOC INSTRUCTED ME  
TO WAIT UNTIL 12:00 FOR  
REPLY AND THEN LEAVE SITE.  
1216 ~~PER~~ I RECENT ROY OF  
KFC AND HE ALLOWS ACCESS  
TO LEAVE SITE.



J. MUELLER TO 3 NOC  
319/11 D-GROUP

(41)

0900 ARRIVED AT SITE 319/11 D-GROUP  
10-11 AM 3-4 MPH CLOUDY SKIES  
WEATHER: CLOUDY/WEATHER 40-45°F  
WIND SOUTH EAST 10-20 MPH.  
\*HEAVY RAIN EXPECTED DURING  
SAMPLING.

CONTACT BRANCH MGR AND GAIN  
ACCESS TO BASEMENT TO  
PLACE SAMPLER CANNISTER  
TYPE = 3 M 3530 0 VM  
LOT # 0228-11 FOR  
PCP

BASMENT TEMP = 73° HUMID. = 60%  
SAMPLE ID = PM8AN121  
24 HR GRAB NYSDOH METHOD 311-9  
OPENED 1025 3/11 15:15

319/11 ID = PM8AN1240  
24 HR GRAB NYSDOH METHOD 311-9  
OPENED 1025 3/11 15:15  
SEE ATTACHED FOR PLACEMENT LOCATION  
PLACE OUTSIDE AIR SAMPLER  
UNDER DIRT THRUOUT AREA OF  
DANK APPROX 4' ABOVE GROUND.

D-Group

3/11/11 D. G. GORE

EXAMPLE I-175 P.M. AMB AIR

201 HR 6948 RV 180018 NORWOOD 311-9

SPINCO 1035 S/N 4411

INFORM DOWNT MGR I WILL  
RETURN ON 3/11 IN THE  
MORNING TO PICK UP SAMPLES  
1110 LEAVE 2100

(42)

~~3/11 1035 S/N 4411~~

3/11/11 D. GORE

0945 ARRIVE RT 8176 70

PICCO 4A RV 180018

WEATHER CLOUDY SUNNY 45-50°

WIND NORTHEAST 10-30 MPH

HEAVY RAIN (2-3") FELL DURING  
THE NIGHT AND EARLY MORNING  
HOURS.

1005 SIGNAL AND PACK UP 7  
BASEMENT SAMPLES.

1025 ESCAPE AND PACK UP 147 DOOR  
811 SAMPLER. UNIT IS INCT.

1050 RETURN TO RT AND RETURN  
TO BALLOON WITH UNIT I  
PACK AND JACKET 10 AM 1123

BACK TO 2101  
FEDEX # 0415764720748

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~~3/11 1035 S/N 4411~~

5/4/2011

44

8:00 Alan Eisinger Meets AGS Chris Cull  
on site for Geophysical Survey. Chris  
said the Rain is too heavy for him to  
start work and the spray paint will  
wash off if he goes to mark out  
in these conditions.

8:10 Alan and Chris go for a site walk  
to locate some of the locations  
that will need to be surveyed.

8:35 Dennis Grove Arrives, Rain continues.

8:45 Call Cristina R. and Discuss the weather  
conditions. We have decided to do the  
work tomorrow when dry weather is  
forecasted.

8:50 Walk the site with Dennis, Meet with  
Store / Bank managers and let them know  
that we will be working in the area.

9:30 Finish meeting store owners/rangers  
All personnel off site

*Alan Eisinger* 5/4/2011

5/5/2011

45

Personnel: A. Eisinger C.D.M.  
C.Cul. AGS

Weather: 55 sunny 5-15 mph winds

Scope: Conduct Geophysical Survey of MIP  
Proposed and Proposed Contingency Points  
20 points total.

8:00 Arrive on site meet with Chris Cull  
and go over the scope of the project  
for the day

8:15 Move to location behind KFC and start  
to survey area. Near MIP. (455 Forest Ave)

Materials: Fisher TWG Pipe + Cable locator  
RF RD4000  
GPR + 400 MHz Antenna  
MScope metal detector

8:50 Finish location Behind KFC move to location  
Behind Boston Market off the corner of  
the concrete pad (Dumplin Area) 1465 Forest Ave

9:15 Discovered Buried concrete pad under asphalt  
Moved MIP location closer to the fence  
Finished location. Moved to MIP.  
Concrete pad approximately 6 inches thick and  
burned 6" to 1" under asphalt.

*Alan Eisinger* 5/5/2011

5/5/2011

(46)

9:35 Finish MIP-7 move to MIP6  
and MIP5

9:40 Alan goes around to the local businesses  
and hands out cards. Explains what will be  
going on over the next 2 weeks and  
invites people to contact him should they  
have any questions or concerns

10:00

Finish MIP5 move to MIP6.

MIP-6 Relocated Due to Buried Gas line  
and other buried utilities

10:15

Finish MIP6 move to Northfield Bank  
Near the ATM (far Driverguide)

And Further Back from the location (North)

Near a dumpster there is a location in the  
Road we are simultaneously investigating  
Upon Discussing the location of the  
Stormdrain ~~to~~ <sup>to</sup> Forest Ave

10:30

Christie told me we will need to contact someone  
to find its location underground

It may run under MIP5,6,7

It might be 8ft long and 12 ft wide

11:05

Finish location banking Northfield  
Bank west of the one done previously  
Near a dumpster. Mark out the rest  
of the locations in chalk

5/5/2011

(47)

11:47 Finish MIP2 and location  
By Chalk and Shear drop birds (Behind KFC)  
Move to MIP3

11:55 Finish MIP3 Move to MIP-4 and  
the location further out than MIP-1

12:05 Speaking to a gentler from the D.E.P about  
the culvert in front of the Northfield  
Bank. I was advised to go to their office  
to view plans for this area and be given  
phone # and address. Called Christie R.  
to inquire as to whether this has been done  
already or if I should look into it further.  
There was no answer. I left a detailed message.

1:35 Finished all locations on Boston market  
Side of the road and all locations in the parking  
lots Behind Boston Market move across  
the street to vacant parking lot to  
clear the location in the grass there.

1:48 Location in grass By abandoned parking lot  
(is cleared) move across the street to  
ST Bank and Trust

2:00 Finish location outside ST Bank + trust

2:10 Call Christie about Culvert if needed to relocate  
MIP5,6+7 further in from the street

2:40 Done for the day all personnel off  
site

*Mark H* 5/5/2011

5/9/11

48

Personnel: A. Eischen, C. Ransciotti, Rob Smith  
 Weather: Steve Vosilla  
 Weather: Sunny 60°

Activities: Concrete Saw, 58 gal drum for tailings

Scope: Hand Auger, Generator, Jackhammer

Clear MIP locations. MIP 1-7

9:00 Arrive on site. Zebra + (kittens are already here)

9:15 Health and safety talk

9:25 Start work at MIP-6. and calibrate

PID 98 ppm  $\rightarrow$  100 ppm

0-5" Asphalt Concrete

5-12" Light Red-Brown Fine Sand Some Silt  
trace Gravel moist

24-30" Light Red-Brown fine sand some silt  
fine - med - coarse Sand Some Silt Little  
fine - med Gravel moist

49" Light Red Brown fine - med - coarse Sand  
Some Silt 1/4" fine to med  
gravel

10:20 finish MIP-6 to 5 feet Backfill  
with tailings fill top with cold patch  
and ramp down

10:30 more to MIP-1. Start to clear  
location

5/9/11

49

MIP-6



Asphalt Concrete

6"

fine Sand some Silt trace  
Gravel

24" fine Sand some Silt trace  
Gravel

30"

49" fine Sand some Silt trace  
Gravel

60"

"Take it easy down to 6-7 ft" As per Rob's  
Recommendation

5/9/4

(50)

- 0-6" Asphalt Concrete  
6-12 fine S and NE Brown fm G and fm St<sup>s</sup>  
12-18" Red Brown fm S a. fm rounded/subround G  
18" wet  
24"-30" Gray Brown fm S l. fm G l\$ wet  
24"-30" 42"-45" D-Grey fm S l+G l\$ wet  
54"-58" D-Grey fm S l+G fm G wet  
11:52 Finish MIP-1 5 ft bgs backfill hole  
Patch Asphalt temp dam  
move to MIP-2 Break through  
Asphalt  
12:25 Break for Lunch  
12:50 Continue work on MIP-2  
0"-6" Red Brown fm S a. fm G l\$ t C wet  
24"-30" fm S a. fm G t \$  
54"-60" Grey fm S l \$  
12:50 13:20 finish MIP-2 Back fill, Apply  
Asphalt cold patch and temp  
13:30 move to MIP-3 and start work  
0-6" D-Brown fm S s. fm G moist  
15" large Boulders  
Discontinue location move 5' and start New location

(51)

Asphalt 6"-12"

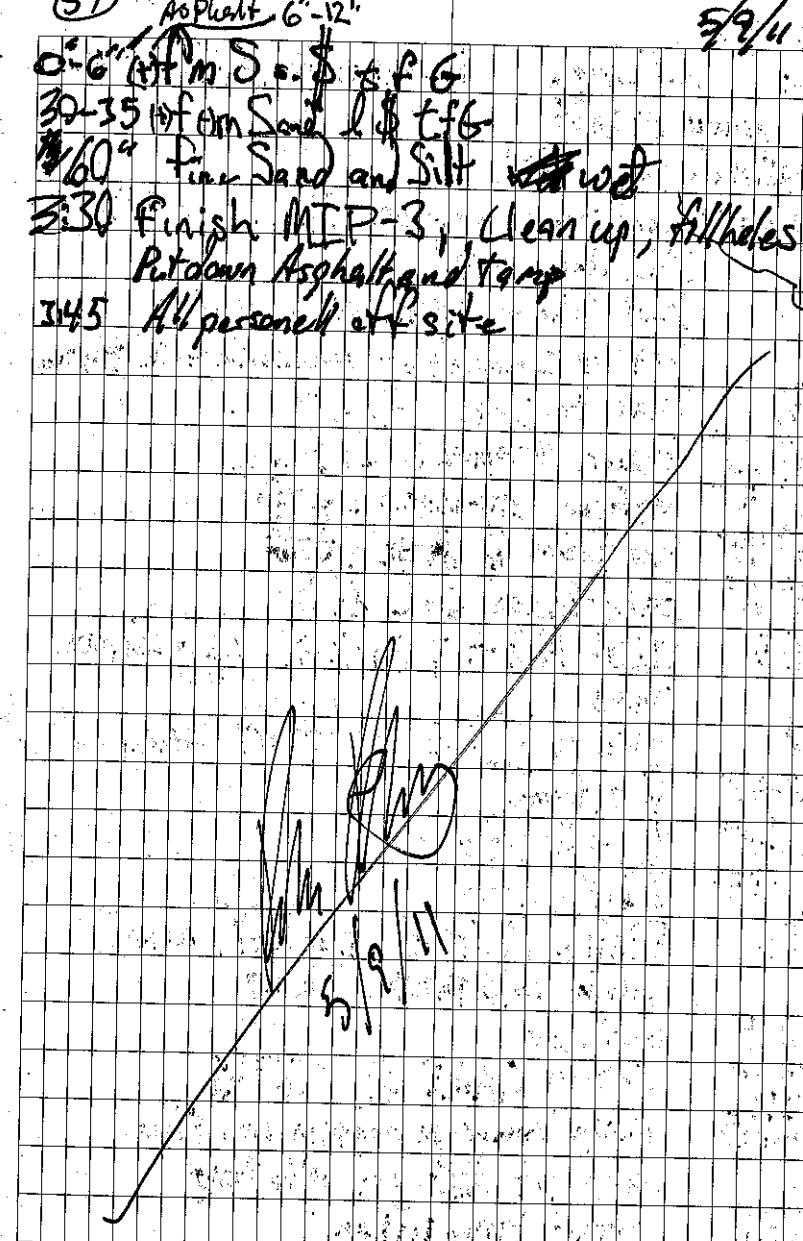
6"-6" (ft) fm S o. \$ t f G  
30-35 ft fm Sand l \$ t f G

54"-60" fine Sand and Silt wet

3:30 finish MIP-3, Clean up, fill holes  
Put down Asphalt and temp

3:45 All personnel off site

5/9/4



5/10/11

Personnel A. Eishberg, C. Rambaschiatti, John Piroozi  
 Quincy Brandt,  
 Weather: Sunny 65°  
 Materials: hand Auger, Geoprobe 7720 DT #GP34,  
 John Deere Gator

8:30

Meet John + C listing at job site,

9:40

Set up communication monitoring + Col. equipment

Start to clear MTP4

24"-26" Brown - (ff) m/s S lf g l \$

36"-40" Red-Brown - (ff) m/s S s lf g

40"-44" Red-Brown \$ ff m/s lf g

55"-60" Grey-Brown \$y l f s

10:25 finish MTP-4, Backfill, Apply cold patch  
Tamp

10:50

Move to MTP-5 and start clearing 06"

25"-30" \$ s from S

40"-45" \$ lf s lf g

50"-55" \$ l f s lf g

11:23 Finish MTP-5 backfill hole,  
Apply cold patch, Tamp

11:45

Go to lunch

12:15 Set up CAMPA at MTP-1

12:45 Move Geoprobe and map into place

and let MTP get up to temp

Start time 1:11 pm

John Piroozi

5/10/11

(52)

(53)

5/10/11

Start log @ Rubber Ring 6" below ground surface  
 his log will report this as 0ft or ground surfaceFID PID Flow Pressure Start/End  
 Off 1600  
 If 2400

Depth may have gotten a little off around 18-19 ft.  
 the Rods needed tightening and were twisted  
 thus tracking the MTP into thinking it was  
 moving through soil, when this was not the case.  
 Not the case we wanted the Rods and looked  
 at the depth and they appear correct. There  
 is a line on the side of the drill rig that's hooked  
 up to the computer that lets it know the depth.

	ECD	FID	PID	Flow	Pressure	Start/End
CDP 100	585	✓	✓	64.7	13.66	1/11/10

10 ft 1600

atmos.

11.5 2400 (57.5) (0.97)

(PID 12.5 zeroed to 10000)

14.55 ECD Up to 2500 76.77 - 8000 ECD

18.48 FID (10000)

↓ ECD Down to 2000

John Piroozi

5/10/11

5/10/11

(54)

4:30 Pull Rod's, Email Logs from MPP  
(CDMMIP1), Decontaminate equipment,  
Put equipment away.

5:15 Fill hole with bentonite chips & ½ bag  
went in, then patch with Asphalt patch  
All personnel off site.

5:50

All personnel off site.

M. Sarnowicz  
5/10/11

5/11/11

(55)

Weather: 65° Mostly Sunny wind 10 mph  
Personnel: A. Edberg, C. Ramascati, John Diamond,  
Quincy Branch

Scope: Continue MIP work.

7:30 Onsite Start to set up Community Air  
Monitoring and calibrate Allegemant

10:05 Finish set up on MIP-2 and start work

On with MIP after equipment cal + warm up

Equipment: Geoprobe 7720DT GP34,  
John Deere Gator with MIP Setup, 2xPID, 2x  
DataRAM 4.

10:50 finish troubleshooting a gas pressure  
issue with the MIP and start to advance  
MIP.

11:10 Notes: MIP-2 Started at 6" Below ground surface  
- Around 18-19 ft bgs first indicates of  
contamination in MIP-2

- ① 28 ft Very High Readings
- ② 38 ft Theys calm down

CR note - K. Sarnowicz onsite at noon. We  
reconnoiter site and progress.

1:30S - Zebra and CR discuss restore  
today. MIP unit needs to be mobilized  
to a storage unit today. With late  
start, MIP-2 will not be finished

5-11-11

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until around 13:30-14:00 today.  
We will finish MIP-2 and do 0  
MIP 3 and 4 tomorrow. Zebra will  
also string up 25 ft of trunk line  
since we are achieving such great  
depths with the MIP. Design of trunk  
line may need to be modified due to  
presence of high concentrations  
from roughly 6 to 39 ft bgs.

14:30 End MIP-2 @ 97.60 ft bgs  
Start to pull rods and Dismantle  
Equipment

15:20 Fill hole with Bentonite Chips  
And continue to clean up around  
the site

15:45 Patch Asphalt with concrete  
Patch and Black Dye

16:00 Leave 2 cones by the wet cement so that  
No one drives into it. End day all  
personell off Site

*M. May* 5/11/11

(57)

5/12/11

Personell: A. Eisberg, John Diamond, Quincy Brandt  
Weather: Sunny 70°

Materials: Geoprobe, John Deere MIP, Comutec  
Monitoring equipment

7:30 on site calibrate equipment test hammer  
8:15 Zebra on site and start to setup

on MIP-3 and warm up equipment

9:15 calibrate MIP

9:36 Start MIP-3

MIP Start point + 6" bgs

9:50 Removal @ 14.15 ft bgs

10:05 Move to New location and Start MIP-3 there  
5' East of original

10:25 Move to another location to avoid bolder

11:06 New location cleared Start MIP  
Work: Soil is 9" bgs. Set up for

11:15 Start MIP-3 at its new location

11:18 Hit obstruction in New location we are  
going to attempt to pull out the MIP  
and put in regular Rod to attempt  
to Break through it

11:40 After 10-12 min of hammering we have  
made no progress, End at 12 ft bgs

12:00 Alarm went off while moving the Geoprobe  
close to the P.I.D

5/12/2011

- 1205 Clear a new and final attempt at MIP-3.  
1230 Location Cleared quick break for lunch  
1255 Zebra Returns from Lunch  
1302 Start MIP-3 @ 3 inches bgs  
1310 We have made it to 9 ft and a portion  
The MPP broke probably a fuse. Reuse MPP  
Complete MIP. Requires push down to  
9ft bgs and continue from there.

Went up 18-19 ft bgs

- 1611 Maxed out Meter 37ft bgs to 39ft bgs  
Stop @ 91.90 ft bgs. problems with  
the membranes - 43-44 end hits until  
51-52 some smaller hits

Summary of today: we hand dug to 2ft, 5ft, and 9ft  
and pushed the MIP to 12ft, 14 $\frac{1}{2}$ ft, and 91.9 ft.

- 1736 Finishing packing up. Apply Asphalt to  
hole, After filling with bentonite, And tamp

- 1755 All personnel off site

Note: One of the datarams did not record data today.

(58)

(59)

5/13/2011

- Personnel: A.Fisberg, T.Diamond, Q.Brandt  
Weather: Cloudy, 60°  
Scope: Complete MIP-4, if time allows either  
Do MIP-5 or clear some of the 4 locations (crushing)  
L as pointed out to me yesterday.  
Materials: John Deere MIP, Commandry or Monitoring  
Equipment

7:35 A. Fisberg on site calibrates and sets up  
Commandry or monitoring Equipment

- 8:05 T. Diamond arrives and starts to set up  
The MIP equipment and perform maintenance

- 8:30 Q. Brandt arrives and starts to set up  
The Geoprobe

- 9:10 Finish maintenance on MIP, and continue to  
Set up.

Note: I have been having trouble with the Alarms  
on the Dust monitor going off this morning.  
I have contacted Pine and made some adjustments,  
they said there is more that we can try. I need to  
call back. As of now I have moved the monitors  
a little further away from the machinery which  
has reportedly been setting them off.

- 9:47 Start MIP-4 @ 2 inches bgs

Slight Rise from 19-20 ft to 16-17 ft bgs

5/13/2011

24 ft had a quick spike

28-29 ft bgs had a high spike  
mixed out meter 38 ft bgs to <sup>approx 52</sup> ?

11:32 We had a small problem with the computer for the MIP. It stopped displaying new data. The last data I saw stopped at 79.1 ft. Then the MIP was advanced approximately 20 ft after that John made a new file and things

+100 ft P4A 1 ft CDM MIP 4A 30' station so continued work from there he said even though we could not see the data it was being recorded.

The meter was mixed out from 38 ft bgs to approximately 48-52 ft bgs after that it went into a pattern of spiking high and falling to a level and sustaining there

12:16 at 103.5 ft bgs membrane may be clogged or the soil is too tight gas flow has reduced to low to continue we will not get accurate readings. We do continue up until we stopped at 103.5 we sustained that rise and fall pattern described earlier.

Start to pull rod out of the hole

Rods are cut, put away more equipment, fill hole with Bentonite →

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5/13/2011

and Patch with asphalt

1:44 Finish Patch and putting away equipment  
Do some paper work with Zebra

2:00 All personnel off Site

2011

5/13

11:30

5/16/2011

Personnel: A. Eisberg, J. Diamond, Q. Brandt

Weather: Light Rain, Cloudy, 60°, wind 5-10 mph

Scope: Continue work on MIP-5 and MIP-6 if time allows for clear MIP-7.

Materials: CAMP equipment, John Deere MIP Geoprobe.

7:40 Arrive on site, Prepare CAMP equipment to be deployed, Calibrate equipment and ready it in its box

8:45 Zebra arrives on site and starts to set up. At this time the CAMP is also set up, cones are deployed in appropriate locations, and Zebra equipment

9:15 downwind PID goes off and sets off alarm. The PID may be picking up exhaust fumes from traffic passing by. Unfortunately this appears to be the only location where the downwind CAMP can be set up. Call Crisfield for recommendations regarding the PID and wet weather/high humidity and its behavior.

Start Set up on MIP-5

9:45 Start MIP work on MIP-5 @ 3 inches below ground surface

9:53 The downwind PID alarm continues to go off in fresh air. Also the fan keeps

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(63)

5/16/2011

stoping. worker may be getting in to the device and making it difficult to draw air through the wet filter. The PID is turned off due to wet weather and not reading concentrations of VOC's in fresh air also when compared to the other PID the readings are off. However they were almost identical 7.3, running after calibration

10:00 Put the MIP up from 14 ftags. The device is malfunctioning and needs to be troubleshooting. I was contacted by Crisfield and was told that the PID may get fuzzy in high humidity and rain. I turn the fan off, dry out and put it back out of for this the PID lasted 15 min before going off.

The PID has been shut down and put into the truck to keep it out of the elements. 10:30 John has told me that there is something lodged in the trunkline and he is unsure of how long it will take to clear it or if he will need a new trunkline. He said one of this length may take days to get in.

10:43 John is going to have dept to get a compressor to try to blow out the particle in the trunkline

OVER →

5/16/2011

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John said this is a trick his Mentor has had some success with in the past-  
11:34 John Returns with compressor

to blow out the lines.

12:37 We have gotten several small  
small chunks out of the trunk line.  
John continues to purge the line  
as we break for lunch.

12:50

Continue to purge the trunk line  
and John starts to get the other lines  
ready to connect them to the MIP

2:30

Contact Cristina. Let her know  
we have been fighting with a clog in  
the trunk line all day and now  
we may need to replace it. We were  
given the option a 75ft line that  
they have already or a new 200ft line.  
The 75ft line could be ready and hooked up  
before work tomorrow. The 200ft line  
would be ordered and express shipped and  
potentially hooked up by 12 noon.

Cristina chose the 75ft line since  
most of the contamination is above 75ft  
bgs.

2:40

Pack up for the day.

3:15

END Day All personnel off site

5/17/2011

(65)

Personnel: A. Fisberg, John Diamond, Q. Bragot, C. Rangwala,  
Weather: 55° Rain Cloudy 5-15 mph wind

Scope: Complete MIP-5 and MIP-6 in time.

Allow 2' clear first 5' of MIP-7.

7:45 Arrive Zebraison site setting up.

8:09 Prepare and calibrate CAMP equipment.

8:43 CAMP + Geoprobe are set up and in position

MIP is still getting ready.

Egyptian Site: CAMP Egypt site (Dormitory, PDD, Radio,  
Weather Resistant enclosure, Typewriter, Battery) Geoprobe,

John Deere Gator powered MIP with 75ft+ truss line.

8:55 PM we continue MIP-5 from 140ft bgs

9-10 ft + bgs not a spike this well in the open hole  
from yesterday John says if it continues to go down  
down like this we may consider it a hit.

14:30 Strong hit, marked out water

Up to 28ft the reading keeps turning out the  
metres and just dipping down into a zone that can  
be read by the meter then it turns out again  
this pattern ends at around 30ft bgs

where the pattern turns into a declining spike  
and the pattern declines with depth.

at 45ft the second tall bar is rather small  
considering speaking nothing around 60ft  
where the levels off through the end at 77.8

10:30 start to pull rods & end MIP-5 at 77.3ft  
bgs

5/17/2011

10:40 take down CAMP equipment as

Per Cristina's recommendation. Therein  
has been intense wind and now we  
are concern for the safety of the equipment

11:14 finish pulling rod back f. 6 hole with

1 bag Bentonite chips reapply asphalt patch and temp

11:21 move equipment to MIP-6 and setup

ReOpen pavement to 9 inches bgs

11:50 Dump Asphalt into 55 gal drum and

talk to Cristina. She has new plans in  
the works for the rest of this week.

12:10 Start MIP-6 at 9 inches below ground surface

Hit at 6 ft bgs. Big spike at 16, at

19 ft bgs the meter maxed out, again at

24 ft maxed out and then stayed there

until 30 ft bgs, the meter continued

to max out and dip down until 40 ft bgs

where we hit refusal

1:10 Sitewalk with Cristina to see some  
new locations, while Zeina cleans up

at MIP-6

1:30 Fill Patch with asphalt MIP-6 (Fill: Bentonite)

2:10 Break for lunch

2:30 Start to clear MIP-7

0-5" Asphalt

(6)

(6)

5-8" fmc S Syp FG

8-36" #1 fmc S E FG

Move from MIP-7 to 4' original boulder at 3'  
MIP-7

6 inches of asphalt

6-10 inches F-MG t fmc S t S

10-24" F-mG and C l S

24-50" C 1 S f-mG

50"-62" C 1 S F G t fmc S

3:40 Start MIP-7 @ 4" bgs

3:42 MIP went in cracked and Bent the

MIP it now must be replaced

This location will be sealed and  
capped tomorrow.

4:19 Finish cleaning up equipment filling and  
patching MIP-7 move Equipment to vehicles

to load and trouble shot / repair MIP.  
8:00 Repairs are finished, all personnel off site

S/17/2011

JM 05/17/2011

5/18/2011

Personnel: A. Eisinger, J. Diamond, Q. Brandt

Weather: Light Rain, Cloudy, 50°, 5-10 mph wind.

Scope: Relocate MIP-7 due to obstruction and then

Clear the first 5 ft by hand and combine  
from there down with the MIP. When  
MIP-7 is complete move to one of the  
3 locations Cristina gave us yesterday  
clear stand the others.

Materials: Lany Equipment, John Deere Gator,  
Geoprade

7:40 On site paper, calibrate GPR.

7:55 J. Diamond at Zebra drives and  
starts to prepare the MIP

8:30 Starts to set up at MIP-7. Finds new  
location and hand clear it to 5 ft

9:20 Finish hand clearing MIP-7, move  
MIP into position and ready equipment

9:33 Start MIP-7 @ 14 inches bgs

10:30 Readings went up from 29-37.5  
at 37.5 End of Boring (Clean up  
Patch and fill hole fill (Bentonite)  
Tamp down Patch (Asphalt)

11:30 Finish cleanup on MIP-7 call  
Cristina to confirm the order of the  
next 3 locations (MIP-1b, MIP-3b, &  
MIP-2b)

(68)

5/18/2011

(69)

11:45 Move to MIP-1b and setup on location

Note: Both of the PTD's are  
wet and not operating properly they  
will be dried out again tonight.

12:40 Patch up and fill location and move to  
another location for MIP-1b due to an  
obstruction in previous location

0-6" Asphalt

6"-37" of m.s. form G t.s. soft moist tan  
37"-56" of m.s. P.S. I.C. R.P.G. wet brown

56"-57" wood chips wet brown

57"-78" D Limes I.F.G. gray brown

1:37 Start MIP-1b at ground surface.  
file Name "MIP1b"

@ 10ft bgs ZCD starts to climb at 21ft bgs  
it returns to normal

2:08 25.6 End of MIP Refusal

2:36 move all equipment to MIP-3b  
open pavement and clear the first 5 ft  
by hand.

0-6" Asphalt

6"-37" G - P m.s. I.C. I Brown

Obstruction in the way. Abandon hole

3:30 lightning + heavy rain Resuming work to day

Thurs

5/18/2011

5/19/2011

Personnel: A. Eisberg, J. Diamond, Q. Brandt

Weather: Cloudy, 53°, wind 0-5 mph.

Scope: Clear and complete MIP-2b and MIP-3b

Materials: CAMP Equipment, John Deere MIP,

Geoprobe

7:50 Arrive On site Zebra on site starting to set up Calibrate Equipment.

8:30 Start to clear MIP-3b  
0-9" Asphalt.

9"-60" (extreme) S s.f. l fm G moist Red Brown

60"-66" Organic Silts s.f.S t twigs & grass moist Black

66"-72" (extreme) S s.f. l fm G moist Red Brown

9:00 Start MIP-3b @ 3" Bgs

@ 37 ft bgs there was a small bump in the readings @ 39 ft bgs another little bump.  
42 ft bgs Another little bump/bit

10:50 End of Boring/MIP @ 81.85 ft bgs  
Start to pull Rods and clean up

11:35 Fill hole with  $\frac{1}{4}$  bag of bentonite chips and  $\frac{1}{4}$  bag of sand and patch with asphalt cold patch & tamp down  $\frac{1}{2}$  bag  
There is a problem with the MIP, John takes it apart to investigate while he does this the last of the equipment is moved to MIP-2b and gets setup.

(20)

(21)

5/19/2011

12:30 - 1:00 Lunch

12:45 Call Cristina grader progress and see if there is any more work to be done after MIP-2b

1:49 Start to clear MIP-2b to 5 ft

0-6" Asphalt

6"-34 ft (imp) S s.f. l fm G red-brown moist  
Abandon hole, observation at 3 ft back fill  
and patch with  $\frac{1}{2}$  bag of asphalt

New location for MIP-2b 18 inches east of  
compression location

0-7" Asphalt

7-12" fmc S s.f.G 1 \$

12"-60" of S s.f.m G 0 \$

2:19 Start MIP-2b from 3" bgs  
29 ft bgs Hit EP continues to go up and down with each foot of progression  
in a upwards trending pattern until 35 ft bgs  
when it goes off the chart, it goes off the chart again at 36 then it reestablishes a rise and fall pattern the same trending downward until 50 ft bgs where it levels off

4:07 End of boring 78.25 P<sub>4</sub> " Rods and  
Start to clean up work area

5:02 Fill hole with bentonite chips and patch top with Asphalt patch & tamp  $\frac{1}{2}$  bag each

5/19/2011

5:15 John Starts to investigate the reason why the heater on the MIP kept turning off and constantly needs to be turned back on.

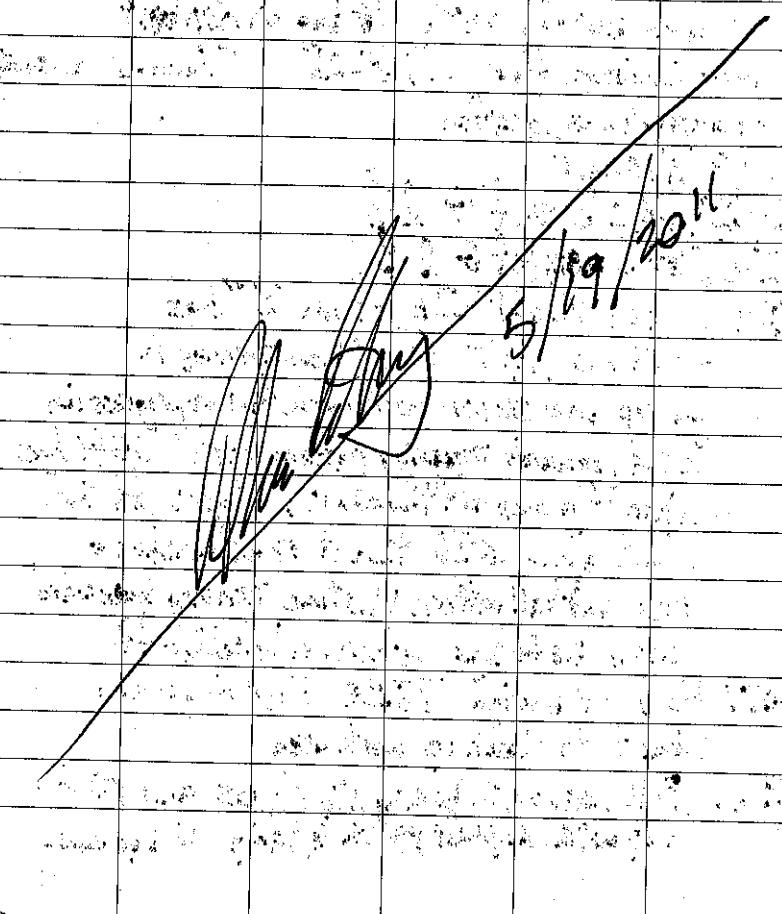
5:50

Pack up the last of the gear

John said that he will figure out what to do with the probe tomorrow

6:00

All Personnel off Site



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(73)

5/20/2011

Personnel: L. Fisberg, J. Pardon, Q. Brandt

Weather: Cloudy, 70°, Rain

Equipment: John Deere Gator Mounted with an MIP Setup, Geoprobe, CPT/MIP component.

Scope: MIP-5b. Located behind the Northfield Bank

7:40 On site Zebra has arrived and is finishing repairs (Tying into new probe). Calibrate equipment

8:45 Call Crismon to determine today's Scope and his final boring locations

8:00 Move equipment into position at MIP-5b

8:30 Start clearing MIP-5b

0-5° Asphalt

5-43° from S of Son G 1 ft E Bank most  
43-60° S of Son G 1 ft G 1 C

9:02 Back fill hole and prepare MIP equipment

9:23 Start MIP-5b @ 3 inches bgs  
ECD spikes and move out the meter at 13 ft

It went down about 10 ft then mixed with the meter again until 22 ft bgs. From 22 ft bgs to 25 ft bgs the ECD spikes up high and then falls. End of coring/MIP

at 25.65 ft bgs. Due to refusal  
Start to clean up at this location  
Prepare to move to MIP-1c

5/20/2011

1028 Move to MTP-1c and start to hand clear the location and set up CPT/PMP equipment

1040 While attempting to clear MTP-1c the hand auger broke, the crew (Zebra) attempted to proceed using the shake bar, this proved unsuccessful, then they used the Rig. They were reminded that there company is liable if they proceed and hit any utilities, they acknowledged this and went on while hammering they hit something hard and discontinued at this location. I have selected another spot to attempt to clear at 10ft further down the transect. 1st attempt was 3.5ft.

11:00 Attempt to clear MTP-1c at a new location. This location proved to be unsuccessful also ending at 2.5ft bgs.

11:10 Move to new location patch of 11 old locations with Asphalt + bedding + back fill materials.

11:15 One of the PID's alarm started to go off. I restarted the fan and it stopped again. Turn off PID

(74)

(75)

5/20/2011

1130 Just finished the 3rd location of 3 ft before hitting an obstruction and discontinuing move to 4th location another 10 ft further on the transect.

1136 MTP-1c has been cleared. The drillers used a macrocore tube to clear the location they tied the rig to hammer it into the ground.

1145 Start MTP-1c @ 3 inches bgs

1220 End at 26.30 ft bgs Nahit's

Start to pull rods

1240 fill hole with bentonite and apply asphalt cold patch to top end tap down

1250 Clean up and move equipment to MTP-1d

105 Quincy went out for Diesel and Asphalt John and I are going to take lunch

120 I'm back and John is sitting in his truck now we are waiting for Quincy to continue work

231 Start MTP-1d hole was cleared with the use of the macrocore SII was dropped back into the hole as soon as it was taken out

at 26 ft bgs is our first hit it is relatively small. There was another small hit at 28 and 31 and 38 and 53,

5/20/2011

413 End of Boring at 63.35  
due to a rupture in the gas line

451 fill hole with bentonite chips  
and tamp. Then fill top with asphalt  
and tamp.

5:00 finish cleaning up

5:30 All personnel off site

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5/23/2011

Personnel: A. Fisberg, C. Ramesciotti, J. Diamond

Quincy Brant, Tom Harn

Weather: Cloudy, 50°

Equipment: CAMP equipment, RAE QRAE II multigas  
LaMotte 2020 Solinst Peristaltic pump, water level  
indicator, inter face meter, Geoprobe

Scope: Get water levels and soil samples  
for locations that we have used the MTP  
at depths specified by Cristina R.

7:30 A. Fisberg on site

7:45 C. Ramesciotti: onsite Discuss scope of  
work for the day

8:00 Calibrate all Equipment

8:30 John Diamond Arrives

10:00 Quincy Brant Arrives with Geoprobe

10:45 Collected Field Blank 80230-FB-110523

4 VOC's, SVOC's, PEST, Metals, mercury.

By: Poring lab provided Deionized water  
from Lab provided container into decontaminated  
bowl and then into sample bottles: 3-70 mL  
Amber Vials for VOC's, 1-250 mL HDPE  
HNO<sub>3</sub> preserved for metals + mercury, 4-1 L  
amber glass unpreserved for PCB, Pesticides,  
SVOC's

5/23/11

(78)

11:10 Start work to Retrieve Samples at  
MIP7

Sample 1205 Sample Time MIP7  
and DLP1. Taken onsite.  
37-38ft bgs (former Sand & L.C. Bfg.

Red Brown moist

1245 Start MIP6; Find water table  
take a soil sample 0-6" Above  
and then a soil sample at 40 ft bgs.

0-6" Asphalt

6"-5 ft (Former S.S. l.C. & FG Red Brown  
Bog) Recovery 55 inches,

5-5.5 Slough

5.5-6 f-m S f-a. f G l\$ Red Brown 40" Recovery

6-10 f-m S a. \$ f G Red Brown

10-11 f-m S a. \$ s.f G Red Brown 20" Rec

11-12 f-m S l. f G Red Brown water table  
at 11"

13:00 Field Blank 80230-FBAQ-110523  
for VOA-Semi VOA-Metals Mercury  
Hg

1335 Sample Time MIP6 at 10ft Ht

14:00 (cont.) MIP6 at 25-30'

Denver reports THAT 2nd rod has been  
defomed. Drawing on now until replacement

5/23/11

(79)

Eqw, present is planned or clear is  
dictated to clean smoother work.

14:15 Clean Beakers for lenses,  
discrete small collection of specimens  
recovered HIGH below

MIP6 + PID Recovery Description

10-15'	370125	70%	DK reddish brown mud coarse 50% silty sand, wet.
--------	--------	-----	---

25-30'	47029	40%	25-28 - DK reddish brown moist f-c sand - silty, moist
--------	-------	-----	---

Soil sample	28-30'	-	DK reddish brown v. stiff silty, cl. clay, moist
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33½ fibs water sample 1653

11:15 ft bgs water sample 1730

For all water samples the turbidity is less than 50 mnt 6/11  
6/10 1/11 Personnel off site

*John Rhy* 5/23/11

80

5/24/11

Personnel A Eibberg, T Horn, J Diamond, Q Brant  
 Weather 80° Partly Cloudy

Equipment: CHTP Equipment, turbidity meter, QRAF  
 Peristaltic pump, water level indicator, interface  
 meter, Geoprobe.

Sloped Recover 2 water samples from MTP5  
 and 2 soil samples from MZ PS. Then  
 follow the specified samples on the  
 sheet (cristina gave for all remaining  
 locations and depths of soil samples)

8:00

8:15

9:00

9:10

9:15

9:23

In Site Zebra is on site setting up  
 T Horn arrives, calibrates equipment  
 and set up continues. receive a call  
 from Mitchell they will be here around  
 1pm to pick up the samples. Call Cristina  
 and leave a message. Tom and I have  
 decided to go forward with sampling the  
 ground water first then taking the  
 soil samples since it is important to  
 sample and get the water samples out quickly.

Reach 32 ft bgs and attempt to sample  
 ground water.  
 Could not sample. There was not enough water  
 try again to sample  
 found water at 24-28 ft starting  
 to sample

81

5/24/11

1055 Water is ready for sampling: turbidity is  
 36, start to sample.

12:49 Sample at 134 bgs and PWP1 @ 1830  
 12:55 Start to sample soil - Turbidity is constant 36  
 MTP5 Soil Sample:

Q-5	Black Mitchell Bay		
5-1	(f) m.s. S s. f G	Dry Red Brown	SM
1-1.5	↓ some		
1-3-2	↓ some		
2-2.5	f G S.s.p.	Moist Red Brown	SM
2-3	↓ some		
3-3.5	f G S.S.l.s	Moist Red Brown	GP
3-5-5	No Recovery		
5-5.5	(f) m.s. S s. f G	Red Brown	SM
5-5-6	f G M.S.l.s	Dry Gray	GP
6-6.5	↓ some		
6-5-7	(f) m.s. S s. f G	Red Brown	SM
7-7.5	f G S.s.m.s. f S	Red Brown	GP
7-5-8	f S S.l.s	Red Brown	SM
8-10	No Recovery		
10-13	f S S.s.f	Red Brown	SM
13-15	No Recovery		
25-16.5	f m.s. S l.s. f G t.s	Red Brown	SP
26.5-30	f f m.s. S s. f G	Red Brown	SM
Sample 10-11 ft and 20-30 ft.			
		12:50 <sup>3</sup>	1:07 <sup>3</sup>

(82)

1:30

Lunch

5/24/11

2:00

Start clean up and move to MIP-1  
Setup

3:00

Rod is jammed in the macrocore and the  
tip is destroyed. Tom has pictures to assist  
working on removing the rod from inside  
the core.

3:35

Soil Sample at MIP-1

165 ft bgs

14-14.2 f/G crushed stone Gray GP

14.2-16.0 ft s.s. of ac S l.G. 1.5 G Red Brown wet

16-16.5 f/mc S 1.5-2 f/G Red Brown wet

16.5-17 (refined) S s. 1 f/G Red Brown moist

Sample (16.5-17)

MIP-2 28-30 ft bgs. the tube crushed

in the Macrocore Rod so what

was retrieved was shake and hammered

out. There was just enough to sample.

25-30 ft s.s. of ac S 1.5 ff G/C Red Brown moist

@ 445

5:30 All personnel off site

5/24/11

(83)

5/25/11  
Personnel: A. Eshberg, J. Pironio; Q. Brandt, T. Horn  
Weather: 80°, SunnyScope: Collect as many soil samples as possible  
in accordance with the ~~soil~~ provided  
program.Equipment: Grapprobe, CAMP Equipment, Peristaltic  
Pump8:00 On site. Zebra On site. Start setup  
By MIP3 and MIP4.9:08 Start at ~~10 ft~~ to send macrocore down to 38 ft bgs  
to retrieve sample. Sampler hit refusal at  
30 ft

9:45 Sample MIP3 @ 30 ft bgs

80230-MIP3-30 &amp; 80231-MIP3-30

10:10 Lab arrives for sample pick up ~~144-2420~~

10:20 Down field Blank AC for yesterday

10:30 Move to MIP4:

11:10 Start work to retrieve sample at 62 ft

Tom Horn Arrives

11:30 Refusal MIP4 @ 20 ft Pull out

Try again a few feet off

11:40 Refusal MIP4 @ 25 ft Pull out

Try again a few feet away from this location

11:47 Refusal @ MIP4 @ 4 ft bgs collect

location as per (Gustins recommendation)  
patch asphalt in all 3 locations.

Location	Depth	Sample #	Date	Color	MSGS
MIP3	30	80230-MIP3-30	6/25/11	Red Brown	SM
MIP3	30	80230-MIP2-50		Red Brown	SM
MIP16	14	80230-MIP16-14		Red Brown	SM
MIF16	9-13.5			P.Brown	SM

MIP5b	22	80230-MIP5b-22	Brown	ML
MIP5b	18-20.5		Brown	CL
MIP5b	20.5-22		Brown	ML

MM MM 5/25/11

Description	Date	Moisture	Notes
lf mcs l \$ & C t fm G	5/25/11	wet	
lf mcs l \$ & C t fm G		wet	
lf mcs l \$ t fm G		wet	
lf mcs s s & C		wet	

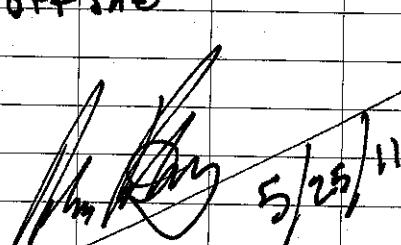
\$ and fmcs l C t fm G	5/25/11	wet	
C a. \$ l fmcs l fm G		wet	
\$ and fmcs l C t fm G		wet	

MM MM 5/25/11

(86)

5/25/11

- 1155 Move to MIP1b and start work  
to retrieve sample from 14 ft  
12:10 Refusal at 2.5 ft move to location  
a few feet away try again  
12:40 Sample at 14 ft bgs and patch holes  
1:00 Lunch  
1:30 Return from lunch / clean up site  
move to Sb MIP5b  
2:20 Refusal on MIP5b at 12 ft bgs  
move to a new location try again  
1:55 Sample @ 22 ft bgs MIP5b  
3:05 Clean up and move to MIP1c  
and try to retrieve sample at 26 ft bgs  
Refusal at 16 ft End day  
Clean up and patch asphalt  
Pack up equipment  
3:45 Tom Horn off site  
4:15 Clean up is completed all personnel  
off site


 5/25/11

(87)

5/26/11

- Personell: A. Lushberg, Lukas Reiss, Q. Brandt,  
Tom Horn  
Weather: 80° Sunny  
Scope: finish the four remaining locations  
by gathering soil samples from there specified  
depths  
Equipment: same as yesterday  
8:00 On site Setup and Calibrate equipment  
8:10 Zebra onsite continue with setup  
8:40 Start to ~~the~~ send down macrocore for a sample  
at MIP3b at depth of 45 ft bgs  
8:55 Pull out Blind probe send down macrocore  
9:00 Tom Horn on site  
9:10 Sample MIP3b @ 45 ft bgs 32 inch rec  
9:15 Field Blank drawn  
9:40 move to MIP2b attempt to recover a sample  
at 40 ft bgs  
10:30 Sample MIP2b @ 40 ft bgs 36 inch rec  
10:50 Move to MIP1d and start working  
towards retrieving a sample @ 37.5 ft bgs  
11:40 MIP1d 33.5 → 37.5 ft bgs 48 inch recovery  
11:55 Start work at MIP1c sample at 26 ft bgs  
12:25 Sample MIP1c @ 26 ft bgs  
12:30 Clean up, Tom Horn off site  
12:50 Lukas Reiss off site  
make new 55 gal drum for soil, and all personnel  
Mark Kieck 5/26/11 off site

(68)

5/26/11

Location	Depth	Square ft	Color	USCS
FB				
MIP3b	40-45	80230-FB-110526	Brown	SP
MIP2b	25-40	80230-MIP2b-40	Brown	SP
MIP1d	36.5-37.5	80230-MIP1d-37.5	RedBrown	SM
MIP1d	33.5-36.5	→	RedBrown	SP
MIP1c	26	80230-MIP1c-26	Red Brown	

Aug 5/26/11

(69)

5/26/11

Description	moisture	notes
down 9' + 9/15	wet	920
f m c S t \$ t f m G	wet	1030
f m G S t \$	wet	1140
refined S s \$ t f G	—	—
f m S l \$ t G	wet	—
refined S l \$ l f m G C C	wet	6 inch recording 12:25

Aug 5/26/11

(10)

6/7/2011

Personnel AE Ishberg, J. Diamond, Q. Brandt  
 Weather 90° F, Mostly Sunny Hazy  
 Scope Clear location MIP5c and perform  
 MIP work at this location + time  
 Allows move to MIP6b followed  
 by MIP7b

7:40 On site Request B on site. Start  
 to clear MIP5c. Calibrate  
 equipment call National Grid  
 there are no markouts on this side  
 of the street.

7:55 Chatscarfield from National Grid  
 gives shows me the markouts  
 in the area and lets me know  
 my dig locators are safe. He  
 recommends removing out one cell  
 This information was relayed to Cristina R.  
 Call Pine Tree lamp on 9 min Rd 200  
 is acting up. they will replace this  
 today

8:10 J. Diamond is on site  
 Contact Cristina let her know  
 how things are going thus far and  
 get location names  
 MIP5c needs to be relocated due  
 to obstructions at 1.5 ft. Seal  
 location with Asphalt and tarp.

8:50

(91) 6/7/2011

9:10 finish patching asphalt move to new  
 location for MIP5c. start to clear it.  
 9:30 Seal in MIP5c

0"-8" Asphalt  
 8"-3.5" fomer S.S. & f.G. most

9:45 Go to ST Bank & Trust talk to the Manager  
 let him know we will be working next  
 to the building this week

9:47 Fill MIP5c @ 3.5' due to obstruction  
 in the hole. Seal location with Asphalt & Tarp

9:50 Select new location for MIP5c  
 0"-8" Asphalt concrete

8"-24" fomer S.S. & f.G. most, Grey Brown  
 24"-36" f.S. & f.G. most L. Red Brown

10:40 Too many rocks to proceed @ MIP5c  
 Back fill and apply Asphalt & tarp and tamp down

10:47 Move to a new location for MIP5c.  
 Call Cristina leave message about progress

10:55 Confirm start new location for MIP5c

0"-8" Asphalt concrete  
 8"-4' f.S. & f.M.G.

12:10 Abandon location move to new  
 location after speaking with Cristina  
 She recommends further back from the  
 street.

92

6/7/2011

12:15 Patch & fill location Tracy  
Down Asphalt  
12:20 Lunch

12:30 Return from lunch and start  
~~new~~ New location for MPP5c  
12:57 Asked to confirm the locations  
of all previous MPP investigations  
on the map

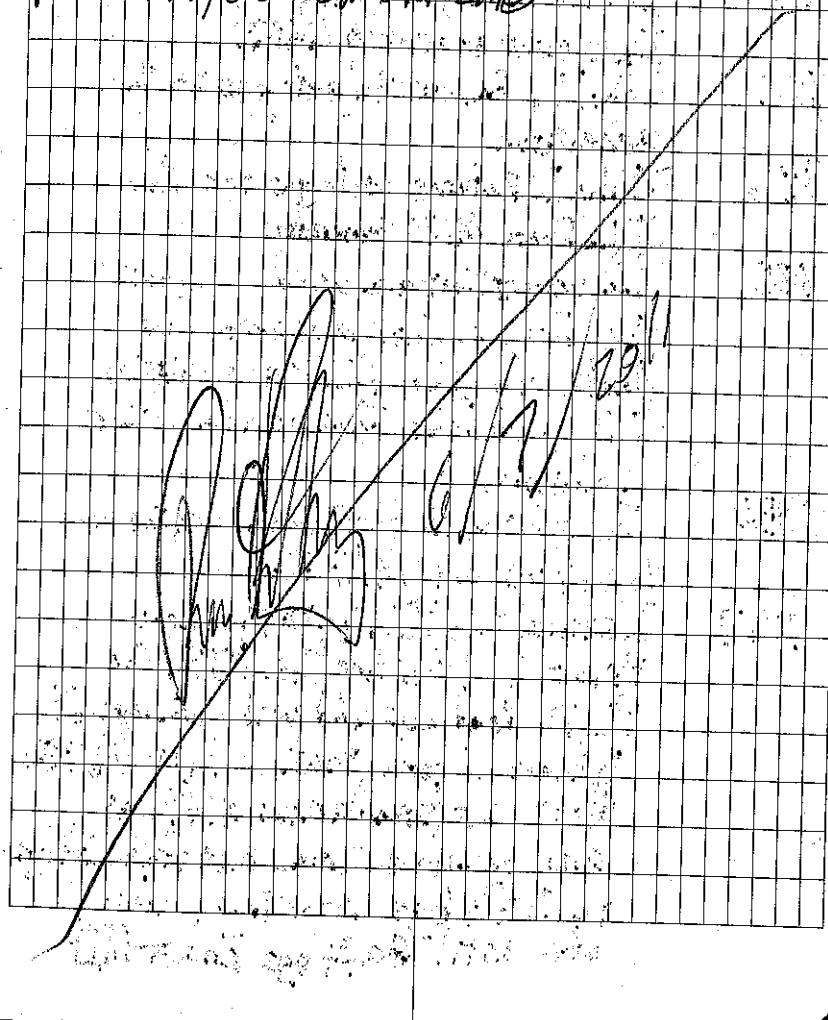
13:36 J. Diamond Starts to clear MPP6b  
leaving MPP5c temporarily.  
MPP6b Cleared started with  
hand Auger then completed with  
Blind Probe, final depth of 66"

14:27 J. Diamond takes the Geoprobe  
to MPP5c and advances the  
blind probe to 61' bgs. John  
says the hole is bad, it twisted  
the rods. He said it will not be good  
for the MPP we must find new  
locations for MPP5c

14:52 Reposition rig for a new location  
for MPP5c Back fill + Asphalt patch  
the last one ~~one~~ prior to this.  
Move to new location for MPP5c  
Back fill and seal this location  
discontinued at 3' due to obstruction

6/7/2011 93

1604 Start New locations for MPP5c  
1615 Location cleared to 5 ft and sealed  
1620 Start to clear MPP7b  
Location cleared to 5 ft  
1706 Clean up and back fill hole MPP7b  
1715 All personnel off site



(94)

6/8/2011

Personnel: K. Esberg, Q. Brandt, J. Diamond  
 Weather: Sunny 95°F

Scope: Start work at MIP5c, then move to MIP6b, then if time allows move to MIP7b.

7:40

On Site Zebra is setting up, Calibrate Equipment and setup LIDAR.

8:00

John D. Ran out to get gas for the generator.

8:16

John D. Returns with gas for the John Deere MIP Generator.

8:35

The screws that are used to mount the device that measures the depth for the MIP are missing. John looks for them or some other solution.

8:50

The solution is Zip ties! Continue to set up on MIP5c.

9:07

I stuck the PID into the hole prior to starting the MIP. The PID read 200 and was climbing still when I took it out. Start MIP5c first hit (small) around 32 ft bgs and continuous small hits from 32 ft bgs to 39 ft bgs after this it levels off until 60 ft bgs End of MIP

(95) 6/8/2011

11:31 End of MIP5c @ 60 ft bgs

12:18 Set up on MIP6b. Full and long MIP5c

12:42 Lunch

13:12 Return from lunch

13:52 Start MIP6b @ 3 inches bgs

We are getting small hits on the LCD from the start every foot until 12 ft bgs

where we get a larger hit but still nothing too large on the small side

Start this at levels out @ 31 ft bgs

There is a small spike. At 40 ft bgs it goes up and plateaus and slowly makes its way back down by 48 ft bgs. End at 62.25

16:02 End of MIP6b @ 62.25. Start to clean up.

16:57 Count the number of locations affected for MIP5c = 7 locations

17:23 Finish Clean up. All personnel off site

6/8/2011

⑥

6/9/2011

- Personnel  
Weather  
Scope
- 100°F humid Sunny  
MIP7b finish The MIP at  
this location then begin an  
area to be determined
- 7:40 All personnel onsite, calibrate  
equipment
- 8:30 Clear MIP7b
- 8:55 Start MIP7b @ 6" bgs EOB  
at 14.65 ft bgs due to obstruction
- 9:08 End of MIP7b @ 14.65 move  
this to a new location and continue
- 9:25 Start to clear a new location for  
MIP7b
- 9:46 Start MIP7b No hits hit  
obstruction @ 19.55 ft bgs.  
End MIP7b @ 19.55 ft bgs
- 10:15 Backfill + seal with Asphalt + tarp down.
- 10:23 Move 10ft to a new location and  
clear and prep work area
- 10:54 Start New location for MIP7b
- 11:18 End MIP7b at 18<sup>ft</sup> 17.95 ft bgs
- 11:22 Call existing letters know we have  
hit Rebusal once again and that  
we are going to move in the opposite  
direction that we have moved last time

⑦

6/9/2011

- 11:35 Lunch
- 12:05 I am back from lunch and scan reading
- 12:10 Start to move into position for MIP7b  
and clear location
- 12:53 Start MIP7b at the new cleared  
location. End at 37.45 ft bgs  
No hits
- 1:55 End MIP7b at 37.45 ft bgs
- 2:54 Move to MIP46 and start to hand  
clear the location
- 0"-3" Asphalt concrete  
3"-45" frac S 1# L C DFG  
45"-60" S 1# L C DFS
- 3:21 Finish hand clearing MIP7b back fill  
and apply Asphalt patch. Handed down
- 3:30 Move to MIP2c and start to clear this  
location
- 0"-3" Asphalt  
3"-24" frac S 1# L C DFG
- 24" took a PIP Reading when pulled it off after  
30 seconds it was at 147 and changing  
14"-60" frac S 1# L C DFG
- 4:30 finish clearing MIP2c and clean up
- 4:53 All personnel off site.

*John R. Kelly*

6/9/2011

98

6/10/2011

Personnel: A. Kusberg, Q. Brandt, J. Dieman

Weather: 85° Sunny

Scope: Clear and complete MIP work on  
MIP4c then if time allows go to  
MIP2c or clear MIP5d

7:30

On S, the Zebra is here setting up  
Calibrate Equipment and call  
Cristina to confirm MIP4c's location  
location has been cleared to  
5ft bgs. JD calibrates and checks  
MIP by giving it something to  
make sure it is working properly  
on the membrane.

9:11

Start MIP4c

9:23

Refusal @ 12.25ft Bgs

9:28

Move to new location start to clear

Start MIP4c

at 2.4 ft bgs first hit

at 3.5 ft there is a very high spike  
at 3.6 ft moved out the other until 4.2 ft  
at 4.2 ft it falls most of the way down to  
baseline and then spikes half way upit continues to go up and down in a  
downward sloping trend until 5.7 ft bgs  
where it turns around and starts to  
go up then we discover there →

6/10/2011

99

→ 1:15 a problem with the heater did we must

Pull out 58.25ft bgs

1:21 pull out @ 58.25ft bgs End of MIP4c

1:53 John D said the reason for that small spike  
May be because the heater gave out.1:24 John Brings the MIP Back to the  
Trailer to work on it Quincy cleanup  
the site and put fills and patches the  
holes

12:26 Move Rig and equipment over to MIP5d

12:30 Take a quick lunch

12:40 Open up MIP5d and start to hand clear

G=4" Asphalt concrete

4"-35" FS with FG dry Red brown

38" boulder Quincy oats for the blind pockets  
advance the rest of the way to 5 feet

1:15-1:45 John + Quincy take lunch

1:45 Return from lunch more equipment in position

2:08 Start MIP5d Hydrogen in the MIP

is too low to proceed John said that  
we will have a fresh tank for Monday

Clean up

3:05 All personnel off site

14/10/2011 6/10/2011

(100)

6/13/2011

Personnel: P. Eisberg, J. Diamond, G. Brady

Weather: Cloudy, Drizzle, 72°

Scope: MIP5d Then MIP4b

7:40. Arrive on site. Start calibration of CAMP Equipment.

8:30. Zebra on site. Start Setup and Switch out Gas canisters.

9:00 Calibrate MIP

9:41 Start MIP5d

6-9 still setting. disregard this.

9 baseline End MIP @ 2.95 ft bgs

10:08 End MIP5d @ 2.95 ft bgs Due to Refusal  
pull Rods

10:10 Call Cristina and confirm that we've moved into a new ISR West and try again.

Weather: Sunny 72°

10:30 Repair MIP, fill and temp hole

11:00 Need to replace MIP. The wires got crossed up too close to the probe. Location has been moved and cleared. It is now ready to start once the MIP is finished.

11:42 Replaced MIP. Replaced move to new MIP5d location and set up

(101)

6/13/2011

12:02 Start MIP5d

Small hit on the PIP @ 2.8 ft bgs

12:55 End of MIP5d. Due to Refusal

(Q) 39.45 ft bgs

1:43 finish cleanup on MIP5d and move to MIP4b

2:05 Take lunch

2:35 Set up on MIP4b

2:52 Start MIP4b

3:01 Refusal (Q) 10.45 ft bgs for MIP4b

move to a new location and trigger

3:23 Move MIP4b to the other side of the fence after talking to Cristina. She recommends we finish the day out with clearing the remaining locations

3:41 Clear MIP4b to 5' bgs and patch with asphalt

3:55 move to MIP5f and start to clear it

4:10 move MIP5f and try again with refusal

4:15 Clear MIP5f to 6ft

4:20 move to MIP5e and clear to five foot bgs

4:30 Finish clearing MIP5e, patch location. Clean up for the day.

4:50 All personnel off site

*Pm RBS*

6/13/2011

(102)

6/14/2011

Personal: A. Eisberg, J. Diamond, Q. Brandt

Weather: 65°F Scattered clouds

Scope: Complete MIP5e at 50.35ft bgs

Then move on to MIP4b, MIP2c  
and MIP5f

7:35

On site, JD onsite, Calibrate equipment  
Equipment calibrated and ready went for  
drilling/geoprobe

8:12

Quarry and the geoprobe arrive and  
start to set up

8:33

Start MIP5e

8:40

End MIP5e at 8.35ft bgs Refusal  
Move to new location, patch and in hand map  
old location.

9:01

Move location towards forest fire to  
try to avoid buried cabinet shown  
in a drawing running under these parking  
spaces. The Zebra crew uses the  
blind probe to clear the location to 5ft bgs  
and sets up for MIP work

9:06

Start MIP5e

@ 16.35 first small hit we continue  
to see small hits declining in intensity  
up to 32.5 where we hit baseline  
again. Stop at 50.35

(103) 6/14/2011

10:30 End MIP5e at 50.35ft bgs  
Start to pull rods and clean up  
Perform maintenance on the rods  
grind down mushroom edges.11:58 Finish Pulling rods and cleaning up  
Prepare rods for next equipment  
to MIP4b

11:43 Start MIP4b

11:50 End MIP4b at 8.05ft bgs Refusal  
Move to a new location and to there

12:10 Lunch

12:30 I return from lunch and meet with ~~MIP4b~~  
Kevin and discuss location and what we are looking  
for12:50 Rob Zebra Returns from Lunch and starts setup  
on the new location for MIP4b, start to  
clear the location, and test the PEP

0'-8" Asphalt

8"-14" m-c Ss C f G L \$ / Brown moist

14"-43" m-c Ss C f G L \$ Brown wet 3.0 PID

43"-45" Peat / woodchips

45"-60" m-c Ss C S S Peat 10.2 PID

60"-68" Organic Clay and Peat 6.9 PID

3.2 PID

1:35 Start MIP4b

(104)

6/14/2011

144

Bore at 10.95 ft Refusal

Take blind Probe down and hammer  
on Rock to see if we can punch  
through

155

We could not punch through the Rock  
Pull rods and back fill hole, fill with  
Bentonite, seal with Asphalt.

207

Start New location for MIP 46 about  
7ft North of the contingent location  
that was inspected during the geophysical  
investigation

2:15

Stop work to Aida man who has  
fallen on his face. Upon further investigation  
we (Quincy and I) discover he is intoxicated  
quite heavily. We called for an ambulance  
and they are now caring for the man. Quincy  
went to clean up.

Karin leaves

245

302

Start MIP 46

first hit @ 19 ft bgs small spike 2.75

@ 27.5 high spike and it continues  
to spike each foot ~~becoming~~<sup>up</sup> until  
42.5 where there is a high spike@ 44 we had a spike that went  
off the chart.

401

End of MIP 46 @ 44.95 ft bgs

(105)

6/14/2011

4:10 Up date drilling on progress

4:15 continue to cleanup P-11 and patch  
MIP 46 and pack up equipment.

4:45 John Diamond off site

500 All personnel off site

6/14/2011

(106)

6/15/11

Personnel: A. Eishberg, J. Diamond, G. Brundt

Weather: 80°F clear Sunny

Scope: ~~Preform~~ Preform MIP work at MIP5f and MIP2c. If time allows start Soil Sampling.

- 7:45 Onsite John is onsite preparing the MIP for the day. Start to calibrate Quincy Drives and starts to prep the geoprobe.
- 8:12 Start to move equipment to MIP5f
- 8:50 Start MIP5f
- 9:18 End MIP5f Refusal at 12.75 ft bgs  
No hits
- 9:21 Pull rods and decontamination  
move to a new position
- 9:29 Clear new location to 5ft bgs
- 9:40 Start MIP5f
- 9:45 End MIP5f @ 60.00 ft No hits
- 9:50 Patch and fill MIP5f
- 11:33 Move equipment to MIP2c
- 12:17 Patch and fill MIP5f
- 12:30 12:47-1:17 Lunch
- 1:17 Move into position for finish prep work
- 1:30 Start MIP2c
- ① 18 ft we have our first hit (small)  
② 22 ft there was a high spike  
continuous high spikes until

6/15/11

(107)

- 2:00 37.65 ft bgs where it refused out  
the meter and hit Refusal
- 2:25 Start to pull rods and decontaminate
- 2:40 Call Cristina tell her of progress.
- 2:42 Finish pulling rods find out MIP  
needs to be repaired.
- 3:12 Receive call from Cristina She wants  
another MIP2c with new probe MIP2c2  
we need to try to get deeper.
- 3:22 John starts to do Rogers on the MIP
- 3:30 and Quincy clears the new location for MIP2c2
- 4:00 John finishes Rogers to the MIP.  
while I try to contact Cristina,  
John told me the overtime rate for  
MIP work is ridiculously expensive. I  
am however Cristina told me to go  
ahead and do what I need to to get  
MIP2c2 done she did not account for  
the costs of ridiculous overtime. Since  
I have tried several times and cannot  
get her to have decide to call it off  
for the day we will have enough time  
to move to 2nd site. I have rescheduled my  
personal matters to account for the project  
All personnel off site.

John Eishberg

6/15/2011

(108)

6/16/2011

Personnel: A Eisinger, J. Diamond, Q. Brandt, T. Harn

Weather: Hazy, little clouds 80's

Scope: Complete MIP2c2 then sample soils

7:40 Onsite Zebra Onsite, Calibrate

Equipment and prep all other equipment

Move into position on MIP2c2

8:15 Start MIP2c2

first hit 18.25 ft bgs good size hit

off the chart at 20.15 ft bgs

Hit Refuse at 20.15 ft bgs

fill and Seal hole with cold pitch Asphalt

&amp; tamp it down. Cristina asked for one

last try at this location to get as deep as

possible. This new location will be called

MIP2c3.

8:36 Clear MIP2c3

9:05 Start MIP2c3

@ 19.25 high hit

@ 22.05 1 ft goes off the chart again at

24.35 until 440 ft bgs where

the spikes are still high but it actually

comes back into range

@ 43 ft bgs they start becoming smaller

They have been sloping downward

Since 40 ft bgs

6/16/2011

(109)

1027 Tom Harn Arrives on site.

② 6.5ft bgs baseline 66-74 small hits

1050 Draw field blank

Small hit @ 69 ft bgs

11:01 End MIP @ 77.45

11:05 talk to Shirley Ng at Mtnem to schedule

Pickup for Monday 20th

11:25 leave Progress Corp. for Cristina in Vassar

12:00 Discuss Progress with Cristina and potential

Sample locations and methods. She

has decided to go with the blind probe push

down to the sample location then push down

the macrocore to grab the sample over continuous

Sampling.

12:15 Lunch Break

1:45 I am back from lunch. John did not

take lunch he worked on the broken

MIP now he finishes up and goes on lunch

at 1:15

1:45 John Returns I have my stuff set up

for soil sampling and Quincy is nearly ready

With the rest of Zebra's equipment.

2:00 Tom Harn leaves

2:12 Start work on <sup>new</sup> MIP2c3 to recover

Sample for MIP2c

⑩

6/16/200

237 Start advancing rods

Refused at 10ft, have to another location

232 Start new location for MIP2c sample

@ 35ft

325 Sampled MIP2c 3 for sample MIP2c

Full TAL/TCL +30 @ 35 ft bgs

4:10 move to MIP2e to sample at 16ft

Full TAL/TCL +30

16:18 Sample MIP2e @ 16 ft bgs

16:20 Clean up fill and patch holes

16:50 finish paper work with Quincy

17:00 Zebra off site, call Cristina

She has 2 additional locations to

sample MW12s @ 35ft bgs

This is supposed to be a difficult  
location

17:30 Finish work Cristina, All Personnel off  
Site

17:30 6/16/200

Gilman III

6/16/200

6/16/200

water depth (13) Sampled ~~13~~ 0/16/2011  
 MIP2c 30ft - 30.5ft 13  
 MIP2c 30.5ft - 35ft 11  
 MIP3e 12ft - 16ft 0

USCS  
 SM  
 ML  
 SM

Color  
 Red Brown  
 Red Brown  
 Red Brown

Description  
 fm S l s  
 Silt Somewhat Sand  
 Silty Sand and fine Gravel

6/16/2011 Moisture Notes (13)  
 wet  
 wet full T C 1/2 ft 325 18  
 wet 1618  
 full T C 1/2 ft 325 18  
 full T C 1/2 ft 325 18

M W 16/20ft

M W 16/20ft

(14)

6/17/2011

Personnel A. Eberberg, Q. Brandt, J. Diamond

Weather Damp cool 75°F

Scope got all 5 remaining soil samples

7:30 Zebry on site

8:10 Akersberg on site

9:05 Start work on MIP4Ecc sample  
at a depth of 45ft

10:20 Sample 80230-MIP4c-41

10:40 Move to MIP4b and attempt  
to retrieve a sample @ 45 ft 6ops

12:25 Sample MIP4b; 80230-MIP4b-45

13:15 Move to MIP6b and setup

13:30 Go to lunch

13:40 Start to rain, Thunder + lightning

15:00 All personnel off site

work called off due to  
lightning

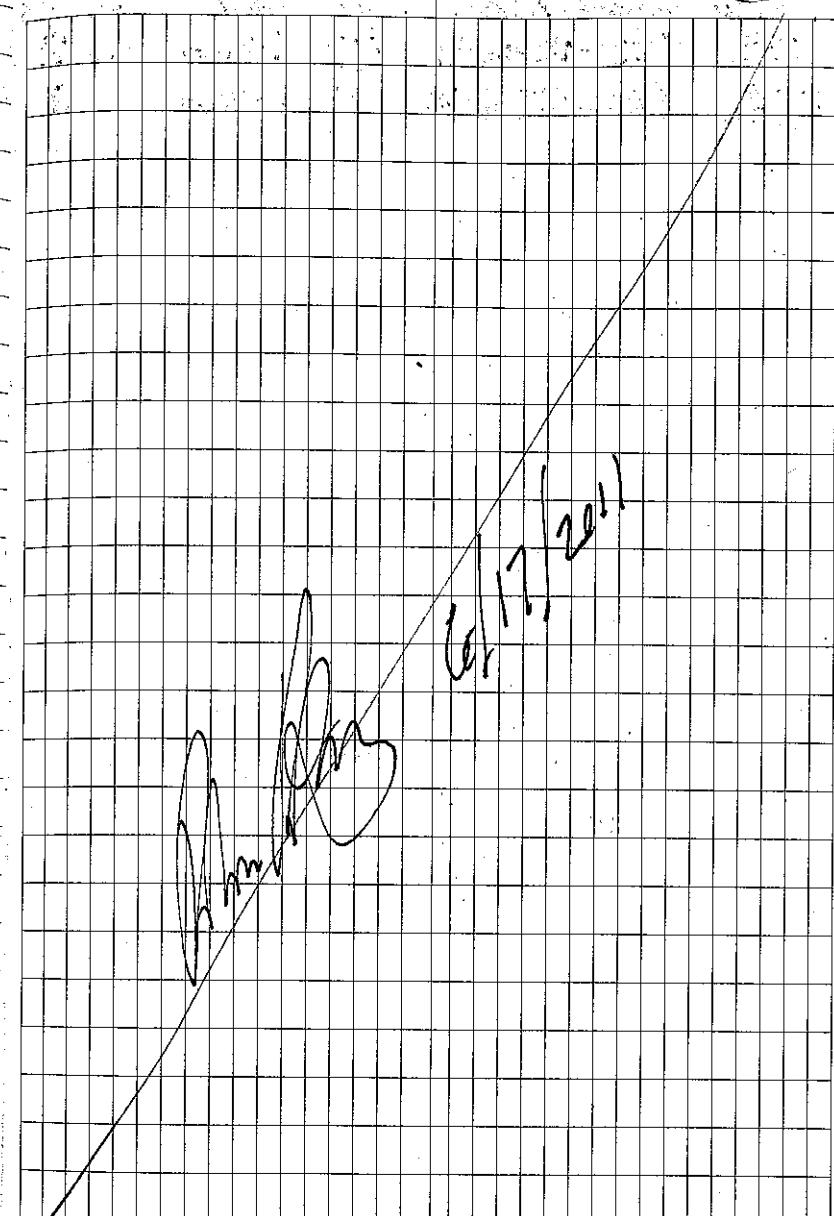
M. Eberberg

6/17/2011

(15)

6/17/2011

W. [unclear]  
[unclear]



(16)	6/17/2011				
Location	Sample Name	Depth	color	PED/HC	
MIP4c	80230-MIP4c-41	41-45	L-Brown	60 SP	
MIP4b	80230-MIP4b-45	41-45	L-Brown	0 SP	

*Collins*  
6/17/2011

(17)	6/17/2011	moisture	100%
		Wet	10:20
		Wet	12:25
			MS/T&FG
			fm S/T&FG

*Collins*  
6/17/2011

(118)

6/23/11

Personnel: R. Eisberg, Terry McKiven  
 Weather: 70°F, Humid, Cloudy, Rain started off  
 Scope: Survey Site

12:00 Arrive on site. Received call that Surveyor will be late, around 1PM

May (2:40) Rec'd. call that Surveyor will be late, perhaps a 1/2 hour or 45 min.

13:30 Call Terry McKiven to ask him how far out he is. He is on the bridge so hopefully he will be here in 10-15 min

13:40 Terry arrives on site. Start to Survey site using GPS

15:00 There is a car in front of Michaels that is over MIP04B, it has been there all day

15:30 All locations are complete except for MIP04B. Monitoring well MW12S has 2 locations shot for it (MW12S and MW12S). The additional location is in case we hit refusal.

15:55 All Personnel off site

6/24/11

(119)

Personnel: AEisberg, Luke Caballero, Quincy Brant  
 Weather: Cloudy, Humid, Drizzle and off 70°F

Scope: Sample MIP5c, MIP6B, MW12S  
 7:40 on site

8:00 Zebra Arrows

9:00 Power Field Block 80230-FB-110624

9:19 Sample @ MIP5c Sample Area  
 80230-MIP5c-36.5

9:42 move to MIP6B and begin to sample

10:13 The drivers have tried 3 locations for MIP6B so far. The last one has bent & 20 ft of these rods

10:57 Sample MIP6b Sample Areas:  
 80230-MIP6b-425 and  
 80230-DUP3-SQ @ 21:00

11:36 finish cleanup and packing up of equipment

11:50 move all equipment to MIP MW12S  
 Start to set up

12:00-12:30 lunch

12:30 Continue to set up on MW12S

12:38 Start MW12S H.T. Rods 2 times

13:00 H.T. Rods with hammer at soft

14:07 Sample MW12S @ 35ft sample  
 80230-MW12S-35

(120)

6/24/201

Location	Depth	Sample #	PID/4888	Color
MIP5c	31.5-32.5	—	1.3 ML	RedBrown
MIP5c	32.5-33.5	—	2.75 M	RedBrown
MIP5c	33.5-36.5	80230-MIP5c-36.5	3.0 SP	RedBrown
MIP6b	41-41.5	80230-MIP6b-42.5	0.8 ML	RedBrown
MIP6b	41.5-42.5	80230-DUP3-SO	S/M	RedBrown
MW12s	34.5-35	80230-MW12s-33	21.0. ML	RedBrown

14:40 go back down MW12s try for entire macrocore from 38-40.

15:20 An additional macrocore has not been successfully retrieved. The tube is stuck in one core and the other was not able to penetrate to 35 ft bgs to start/continue sampling.

16:01 After talking to Cristina and giving an update ~~to~~ I noted I haven't written. The drillers had to retrieve the MW12s sample from the macrocore. By beating it with hammer to shake loose any soil they could, the liner had been crushed inside of the tube and was not allowing the sample to be withdrawn in the manner it is usually taken from the macrocore.

(121)

6/24/201  
Notes

Description	Notes
Silt some Fine Sand	wet
Fm Sand and Silt w/ the gravel	wet
Fine Sand / the Silt trace Gravel wet	9:19
Silt and Fine Sand	moist
Fm Sand some Silt trace Gravel moist	10:07
Silt Some fm Sand trace fm Gravel moist	14:37
16:25 finish cleaning and filling and site cleanup	
16:30 End Day 9/10 personnel off site	

6/24/2011

9/19/11 Water levels and site  
Visit

09:50 S. Kellogg & R. Chenuko  
arrive on site. Meet  
John Diamond of Zebra  
to look at locations in  
Boston Market basement.

10:20 met Kevin Saronwicz of  
NYSDEC and Paul Waddel  
of L.B.H to look at well  
locations

11:30 opened MW-8S and collected  
depth to water

Weather: light rain 2 70°F

12:00 Moved to MW-16S - can't get  
the well open

MW-1 is under water

MW-15S - opened and released  
pressure

12:10 MW-14S bent bolts wait  
open PZS wait open  
took photo

MW-12 bent bolts. Can't open

12:20 MW-13 cluster @ KFC

12:45 MW-2

LBC

well	time	depth to water
MW-8S	11:30	5.01 feet
MW-16S		Cant open
MW-1		underwater
MW-15S	vac	4.63 feet

MW-15S	11:5	3.87	ft
MW-13 → 11D	3.91	2.8	ft
MW-3		2.81	ft
MW-4		2.85	ft
MW-2		2.11	ft
MW-9S	vac	2.49	2.27 ft 2.21
MW-9D	vac	2.27	ft
MW-10D			
MW-10S		2.82	ft
MW-13S		6.31	
MW-13D	vac	6.31	ft 6.21

4:00 pm left site for the day

2:10

1:00 Unloaded equipment at Yonkers  
Warehouse

Seth E. LBC 9/20/11

9/19/11

1230 - General note - wells  
need 10 min to stabilize  
MW-9's took longer  
1300 Car parked on well MW-10D

10/31/2011

Personnel: Alan Klobberg, John Diamond, Lukas Reiss  
Lukke Cava, others

Weather: Clear, Sunny, 40°F

Scope: Do borings in the basement

8:30 Arrive on site. Start Preparing equipment

9:15 Zebra arrives, R.K. calibrates equipment.

10:50 call Seth K. and discuss plan of action

Zebra does not have equipment to  
drill inside today. They are going  
to core through the cement and  
then leave the drilling for tomorrow, T.D.<sup>10/31/2011</sup>

11:00 P-2 Pulled full length of PVC Pipe out on

11:07 P3S 10ft PVC Pulled

11:10 P-3D 20 ft PVC Pulled

11:15 P5 10ft PVC Pulled

11:30 fill P-2, P3S, P-3D, and P-5  
with Portland cement

11:45 DHS could not be located, it is suspected  
that the drive through pad for KFC is built  
over P-4S and P-4D

11:50 L.R. leaves to get more cement

11:55 L.C. Starts Rehab on MW-12S

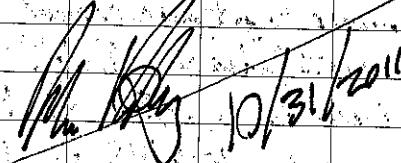
12:07 L.C. moves to MW-15D for Rehab

12:25 L.C. moves to MW-14S for Rehab

12:27 L.R. Returns and helps w/MR Rehab

10/31/2011

- 1:00 finish MW-14S  
1:24 finish MW-15D  
1:36 finish MW-12S  
1:50 look at Remankey 3 Rehab locations  
9nd cuttings in the basement  
2:00 try to open wells to determine  
what is wrong with them  
2:38 Start work on MW-8S  
3:06 finish MW-8S Rehab  
3:15 Geoprobe runs out of fuel  
we try filling it and starting  
it  
3:25 Geoprobe has quit in the fuel line  
air must be bleed out so file  
can make it to the engine  
after much trouble starting  
we finally got the rig started  
6:15 finish cleaning all personnel off site

  
John Kirby 10/31/2011

11/1/2011

Personel: Mike Kirby, Steve Vosilla, Lakes Reiss

weather: Sunny 45°F

Scope: Sample Soil and water under the slab  
in the basement area of the former Paul Miller  
Dry Cleaners

- 7:30 on site start calibrations of YSI,  
Turbidity meter, Multi-RDE  
9:05 Zebra drives woodcass five days week  
9:15 Pour the Trip blank PM-TB-11/2011  
9:45 Pour Field Blank PM-FB-SO 11/2011  
9:50 Pour Field Blank PM-FB-AQ 11/2011  
10:50 Start work on B-2  
10:55 Sample B-2 A ~~PM~~ S.C. 1 ft 6"  
3 inches at concrete slab  
11:03 Sample B-2 B ~~PM~~ S.C. 8 ft 6" 5ft  
11:17 Sample B-4 ~~PM~~ S.C. 3 ft 6" 6"  
11:22 Sample B-4 B ~~PM~~ S.C. 6 ft 6" 5ft  
11:50 Attempt B-4 Retrieval at 15 ft 6" of slab.  
Attempt B-3: Retrieval at 15 ft 6" of slab  
12:15 Sample B-4 R ~~PM~~ fm G.s. fine S  
12:17 Sample B-3 R ~~PM~~ fm G.s. fine S  
Samples B-3 and B-4 were taken from  
the soil between the floor of the vault  
and the slab 1.5 ft below the floor of  
the vault. later conclusions were filed in make

11/1/2011

12:33 Sample B-5 A fSS, STC 6"

12:35 Sample B-5 B fSS, STC 6"

12:30 Sample B-6 A CS, fSS 6"

12:34 Sample B-6 B fSS, fLNB 5F+

All locations were met with refusal:

B-1 Refusal at 5ft

B-2 Refusal at 5ft

B-3 Refusal at 1.5ft

B-4 Refusal at 1.5ft

B-5 Refusal at 6ft

B-6 Refusal at 5ft

At B-2 and B-3 there is potential  
for there to be another slab below the  
ground in side the vault where we were  
drilling.

14:00 Fall out drill and other equipment

Prep for groundwater sampling

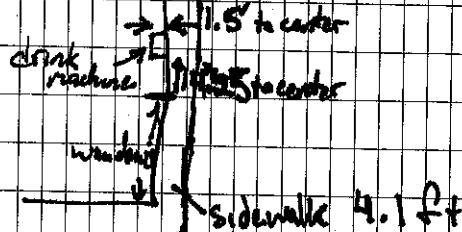
Ricky C. talked to me on the phone he said  
that he would like us to go back in to the  
basement and do an additional location on the  
opposite side of the pad that we did B-1  
next to. In TC's email prior he  
wanted to do an angle boring from outside  
the building after that I told the rollers  
we were done drilling in the basement

11/1/2011

The drivers did not bring their portable pump  
with them today they only have a check  
valve. I talked to Ricky about what to do  
and he told me I can figure something out.  
What I figured out was we will have to wait  
for the portable pump tomorrow.

3:00 Go outside to figure out the location of  
the truck machine so I can prep/gear  
for the drilling by running the calculations  
tonight.

Attack wall



3:20 Head to warehouse for sample management

4:10 Arrive Street working on samples

All Soil Samples are 1x2oz Amber jars

Water Samples for field blank are 3x10ml  
Amber Vials. Trip blanks are 2x40ml

Amber Vials. All samples are to be evaluated for VOC's.  
Samples are named as following:

11/1/2011

B-1a	" 6" bgs	PM-SB-1a-A-11/2011	11:01
B-1a	" B 5 ft bgs	PM-SB-1a-B-11/2011	11:22
B-2	" A" 6" bgs	PM-SB-2-A-11/2011	10:55
B-2	" B" 5 ft bgs	PM-SB-2-B-11/2011	11:03
B-3	" A" 6" bgs	PM-SB-3-A-11/2011	12:17
<del>B-3</del>	Refused at 1.5 ft bgs		
B-4	" A" 6" bgs	PM-SB-4-A-11/2011	12:15
<del>B-4</del>	Refused at 1.5 ft bgs		
B-5	" A" 6" bgs	PM-SB-5-A-11/2011	12:33
B-5	" B" 6 ft bgs	PM-SB-5-B-11/2011	12:35
B-6	" A" 6" bgs	PM-SB-6-A-11/2011	13:30
B-6	" B" 5 ft bgs	PM-SB-6-B-11/2011	13:34
Trip Blank	PM-TB-11/2011		9:15
Field Blank	PM Soils	PM-FB-SO-11/2011	9:45

11/1/2011

11/2/2011

Personnel: A Eisinger, Steve Vosilla, Lukas Reiss  
 Weather: Clear Sunny 45°F

Scope: Drilled Soil samples in the basement on the opposite side of B-1a from the pit dug to location B-1a. Sample the water in all the borings in the basement. Perform 2 directional borings outside the Boston Market to get soil samples under the soda machine.

6:30 onsite get live cropping equipment

7:00 Calibrate Equipment. YSI 7000D, Turbidity meter

7:15 Trip Blank. PM-TB-11/2011

7:30 Field Blank Soils. PM-FB-SO-11/2011

7:45 Field Blank Aggregates. PM-FB-AQ-11/2011

7:55 continue calibration of YSI, Turbidity meter

8:45 Drillers Arrive. We start to prepare

for the soil and water in the basement

9:00 Start Rebar on MW-1

9:45 Finish Rebar on MW-1. Then  
 start work Rebar on MW-16S

10:15 Finish Rebar on MW-16S

10:30 Start moving equipment into basement  
 for B-1b

11:10 SAMPLE PM-SB-1b-A-11/2011 fSoil 6"  
 take Duplicate Sample

11:17 Sample PM-SB-1b-B-11/2011 fSoil fGDC 6"

11/2/2011

- 12:00 Prepare for water sampling  
12:30 Sample PM-AB-2-1122011  
12:35 Have outside to setup on B-7  
13:55 Sample PM-AB-7-B-1122011 at 15ft  
B-7 B  
0-2 ft. No Recovery  
2-3 ft. Concrete  
3-3.5 ft. f.S a. C. wet  
3.5-5 fmG coarse fine s  
5-8.5 fB sB lfG lC  
8.5-9 fmG  
9-12 f.S s.C lS  
10-15 f.m.S s.C fmG  
EOF Refused at 15ft  
13:58 B-7 A refusal at 3.5ft  
14:15 Sample PM-AB-1b-1122011  
15:00 10 Sample PM-AB-7-1122011  
16:15 off site head to warehouse for  
Sample management.

*J. J. Kelly* 11/2/2011

11/3/2011

Personnel Alan Culberg, Steve Vassallo

Weather: Clear Sunny 50°F

Scope: finish the final 3 locations on the basement  
for water sampling. Check all rehabilitated  
wells Seth reported that Ricky could not  
open some due to the size of the internal  
Well seal being to big to get out of the flesh  
meat.

7:30 am site talk to Seth about todays scope

8:15 Zebra on site go over scope

8:30 Pour trip blank

8:45 Mr. Field Blank not enough vials for samples  
and blank, called Seth to confirm. OK

9:00 Sample PM-AB-1a-1132011  
Take Duplicate

10:45 Sample PM-AB-6-1132011

11:00 have trailer open and check accessibility  
of all wells Rehabilitated

11:20 Sample PM-AB-5-1132011

11:30 Turn in key to Boston Market have  
Trailer paint patches on the sidewalk

11:45 Recheck personally that all wells are accessible

12:10 Finish Rechecking wells all wells  
Open completely allowing access to  
the well entirely.

11/3/2011

12:30 off site Head to warehouse  
for sample management

\*Upon talking to Ricky I realized that we forgot  
to examine P-1 this will need to be done  
next time we are in the field. Also  
the P-4s and P-4d ~~we~~ could not be located  
they may be under the KFC Drive through  
pad. ~~so~~ P-1 was to be examined and  
there was water in this well it has  
to be kept aside never think about rusted.  
P-4s and P-4d were to be removed.

Also Measure Locations in the basement  
of sampling tubes

11/13/2011

4/4/12 for C

0740 AC + Cd Kikko mixed

6.45 for SLS tests

Partly sunny, 52°

0742 Min. Bar 30.00? 117.45 SLS

from US Environmental - Reeds 112 pm

a fire call on 100 pm 1st flr.

- P140 C12

0742 Return at 100-16 1A pm

0744 PTL at 100-16 9.82

0755 Lower 16 9.55 + 100-16 1-2 9.60 ±

0758 100-145 P113 200 pm (new)

Depth zone 0.6 m

A few J. 75 (7.75)

0824 5.2ft + 16 S Fully

0835 Start 16S Plus

0900 Close to 145

0917 Start 145 fully

0922 16 S new 16 9.82 - end fully

0947 Return P113 20.41 pm

0951 New 125 return 6.63'

0957 Lower 100-16 1-2 100-125

1100 Completed fully + 100-125

1100-135 New 8.89 P113 46 pm

- Start 100 S Big

1200 3 S 100 + 125 Raking (fully dug out)

4/4/12 for C

4/4/12 Rec

- Motor started  $\rightarrow$  rig at 1105 @ 1107  
1216 Start 13S Fall  
1243 - RTW at 11D - 6.42' PIB:0.  
1303 Stop test at 10S PIB:  
1308 - New PS PIB 10.7 m.  
RTW 7.49' down X0.  
1339 - 11D  $\rightarrow$  Fall down start rising  
~1435 End rise at PS  
RTW at 15D 7.15' PIB 61.  
~1450 End 11D Rises slow to 15D  
1530 - 15D RTW 6.17' rapid rising  
and acc. "fall" back. Repeated  
tests using slowslug method  
1547 At 9S - PIB 2.7 m.  
RTW 4.76

1604 - Find 1.95 fall. - rigs

Leave site.

Summary - 11D falls & rising  
head tests on wells:

18	13 S
8S	11 D
14S	12 S
9S	15 D

Data on In-Situ Level Tull 700 ft

4/4/12 FOCL

## Drum log

Drum #	Date	Contents	Amount Full
1	5/19/11	Asphalt + surface soil	80% 5/26/11
2	5/23/11	Purge, H <sub>2</sub> O	½ 5/26/11
3	5/26/11	Soil (from cores)	15% 5/26/11

# DRUM LOG PG 1

## DRUM Date Contents Full?

~~2008~~

2008 9/1/08 Asphalt + water  $\frac{3}{4}$

30 9/1/08 Asphalt, surface 6 in  $\frac{1}{2}$  ft.

31 9/1/08 Asphalt concrete surface 6 in  $\frac{1}{2}$  ft.

32 09/08/2008 dev. water MW 9D full

33 09/08/2008 dev. water MW 9D ~~full~~ full

34 09/08/2008 dev water MW 13D full

35 09/08/2008 dev water MW 13D full

36 09/08/2008 dev water MW 13S full

37 09/09/2008 dev water MW 15D full

38 09/09/2008 dev water MW 15D full

39 09/09/2008 " MW 16S  $\frac{1}{3}$  full

40 09/29/2008 dev water MW 10DS full

41 09/29/2008 " MW 9S  $\frac{1}{3}$  full

42 09/29/2008 plaster  $\frac{1}{4}$  full

3 new drums for water

## CURVE TABLES

### HOW TO USE CURVE TABLES

Table I. contains Tangents and Externals to a  $1^\circ$  curve. Tan. and Ext. to any other radius may be found nearly enough, by dividing the Tan. or Ext. opposite the given Central Angle by the given degree of curve.

To find Deg. of Curve, having the Central Angle and Tangent: Divide Tan. opposite the given Central Angle by the given Tangent.

To find Deg. of Curve, having the Central Angle and External: Divide Ext. opposite the given Central Angle by the given External.

To find Nat. Tan. and Nat. Ex. Sec. for any angle by Table I.: Tan. or Ext. of twice the given angle divided by the radius of a  $1^\circ$  curve will be the Nat. Tan. or Nat. Ex. Sec.

#### EXAMPLE

Wanted a Curve with an Ext. of about 12 ft. Angle of Intersection or I. P. =  $23^\circ 20'$  to the R. at Station 542+72.

Ext. in Tab. I opposite  $23^\circ 20'$  = 120.87.  
 $120.87 \div 12 = 10.07$ . Say a  $10^\circ$  Curve.

Tan. in Tab. I opp.  $23^\circ 20'$  = 1183.1  
 $1183.1 \div 10 = 118.31$ .

Correction for A.  $23^\circ 20'$  for a  $10^\circ$  Cur. = 0.16  
 $118.31 + 0.16 = 118.47$  = corrected Tangent.

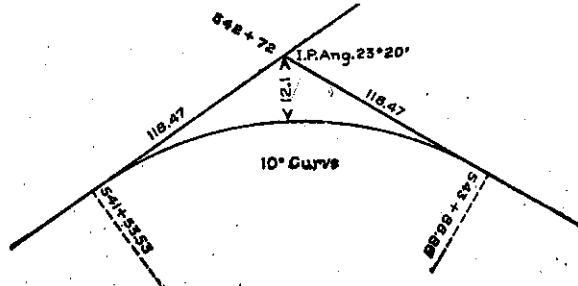
(If corrected Ext. is required find in same way)  
 Ang.  $23^\circ 20' = 23.33^\circ \div 10 = 2.3333$  = L. C.

$2^\circ 19\frac{1}{2}'$ = def. for sta.	542	I. P. = sta.	542+72
$4^\circ 49\frac{1}{2}'$ = " " "	+50	Tan. =	1 .18.47
$7^\circ 19\frac{1}{2}'$ = " " "	543	B. C. = sta.	541+53.53
$9^\circ 49\frac{1}{2}'$ = " " "	+50	L. C. =	2 .33.33
$11^\circ 40'$ = " " "	543+	E. C. = Sta.	543+86.86
	86.86		

$$100 - 53.53 = 46.47 \times 3' (\text{def. for 1 ft. of } 10^\circ \text{ Cur.}) = 139.41' = \\ 2^\circ 19\frac{1}{2}' = \text{def. for sta. 542}.$$

Def. for 50 ft. =  $2^\circ 30'$  for a  $10^\circ$  Curve.

Def. for 36.86 ft. =  $1^\circ 50\frac{1}{2}'$  for a  $10^\circ$  Curve.



## Appendix F

### Slug Test Results

## **OBJECTIVE**

Slug tests were performed at the Paul Miller Site to estimate the hydraulic conductivity (K) of the strata. Wells tested include six “shallow” wells, screened approximately 25 to 35 feet below ground surface (bgs), and two deep wells, screened approximately 60 to 70 feet bgs. The shallow strata exhibit the highest contaminant concentrations on site, estimation of the hydraulic conductivity in these wells will allow a more effective review of remedial options during the feasibility study stage. The two deeper wells were tested because understanding underlying hydraulic conductivity beneath the most contaminated zone will help evaluate plume behavior for different remedial alternatives.

## **PROCEDURE**

Slug tests were performed using a weighted cylinder to displace the water and a pressure transducer to measure and record water level changes over time. At each well tested, the pressure transducer was first deployed and programmed to record depth to water on a logarithmic time basis (water levels were recorded at a rate of four times per second at the start of the test, and the time interval between measurements increased logarithmically throughout the test). Manual water levels were measured periodically to confirm the accuracy of the pressure transducer.

The slug was tied to a rope and lowered to a depth just above the static water level. A few seconds after activating the data logger, the slug was rapidly lowered into the water, displacing the water level upward to initiate the falling head test. After the water level recovered to the static level (i.e. minimum 90 percent recovery), the data logger was turned off and reprogrammed for a new test. A few seconds after re-activating the data logger the slug was rapidly removed from the well, lowering the water level to initiate the rising head test. When the water level returned to the static level, the data logger was turned off and the equipment was removed from the well, decontaminated, and moved to the next well.

Slug test analysis was performed with the aid of Aqtesolv Pro 4.0® (Aqtesolv®), a commercially available software package designed specifically for aquifer test analysis. Aqtesolv® offers a range of analytical solutions that correspond to different aquifer types. Based upon the observed stratigraphy and static water levels, which are fairly close for wells with different screen depths, analytical solutions for unconfined wells were used to analyze the tests. The total thickness of the aquifer is unknown; for purposes of slug test analysis the wells were assumed to be installed within an aquifer whose base is at a depth of 70 feet bgs (i.e. the bottom screen depth of the deep wells). The saturated thickness was assumed to be the water column above 70 feet bgs based upon the static water level depth at each well.

With the exception of MW-15D, the slug tests were analyzed by the Bouwer and Rice (1076) method for unconfined aquifers. This is based on a semi-log plot of time versus hydraulic head, or displacement. Time is plotted on the linear X axis and head difference is plotted on the logarithmic Y-axis. Data plotted in this fashion should yield a curve with a distinct straight line portion that is used to calculate hydraulic conductivity of the formation material. With the exception of MW-15D, the test data were compatible with this method when plotted on semilogarithmic graphs.

The slug test data collected at MW-15D exhibited an oscillatory response and was not compatible with the Bouwer and Rice method. In addition, the well returned to static very rapidly. A second set of falling and rising tests were performed, using a slower slug motion, and the rapid oscillatory response was

confirmed. Due to the oscillatory response, the data from MW-15D was analyzed by the method of Springer and Gelhar (1991), which was specifically developed for slug tests exhibiting the oscillatory response.

## RESULTS

The slug test results are summarized on Table 1, along with a summary of the strata screened at each well. In several cases, more than one K value is provided by the Bouwer and Rice method for a given test. This is because in some cases the automated fit performed by Aqtesolv® did not select the best fit for the line. In such cases, the manual fit capability in Aqtesolv® was used to calculate a more appropriate value based on the data. The best results computed for each well are noted on the table as “preferred fit”. For MW-15d, the results for the initial test, which were performed with the typical rapid slug motion, are preferred over the second set of tests, when the slug was inserted/removed more slowly as discussed above.

Lithologic logs of the indicate that the geologic deposits at the Paul Miller site are predominantly fine to medium sand, with silty sand lenses, and coarse sand and gravel locally (e.g. MW-11d). In some cases blow counts were low (e.g. a maximum of 15 blows per 6 inches at MW-15d), indicating fairly loose soils; in some cases the soils exhibited higher blow counts indicating more compact soils (e.g. blow counts generally at least 30 per six inches at MW-14S). The calculated hydraulic conductivities range from 0.05 ft/day (MW-14s – falling and rising head tests) to 69 feet per day (MW-15d – falling head test). The relative hydraulic conductivities generally compare favorably with the strata screened at each well, as summarized below:

- MW-15d exhibited the highest hydraulic conductivity at 59-69 ft/day; this is consistent with its strata – medium sand exhibiting low blow counts.
- MW-11d, MW-12s and MW-13s exhibited hydraulic conductivity values ranging from 3 to 6 ft/day; MW-11d and MW-13s both primarily screen sand with blow counts slightly higher than MW-16d, consistent with the lower K observed. MW-12S is screened in more compact sand and silt, and a lower K value would not have been surprising for this well.
- MW-8s, MW-9s and MW-16s exhibited K values of less than 1 ft/day. MW-9s is screened in silty medium sand and MW-16s is screened in fine sand and silty sand; strata at these wells exhibited low blow counts and the lower K values are consistent with the finer aquifer materials. MW-8s is screened in fine sand and sandy gravel with clay layers exhibiting somewhat higher blow counts, both consistent with the lower K values exhibited.
- MW-14s exhibited the lowest K value, 0.05 ft/day. This well is screened in silty sand and sandy silt, and its blow counts were among the highest of the wells tested, which are both consistent with the low K.

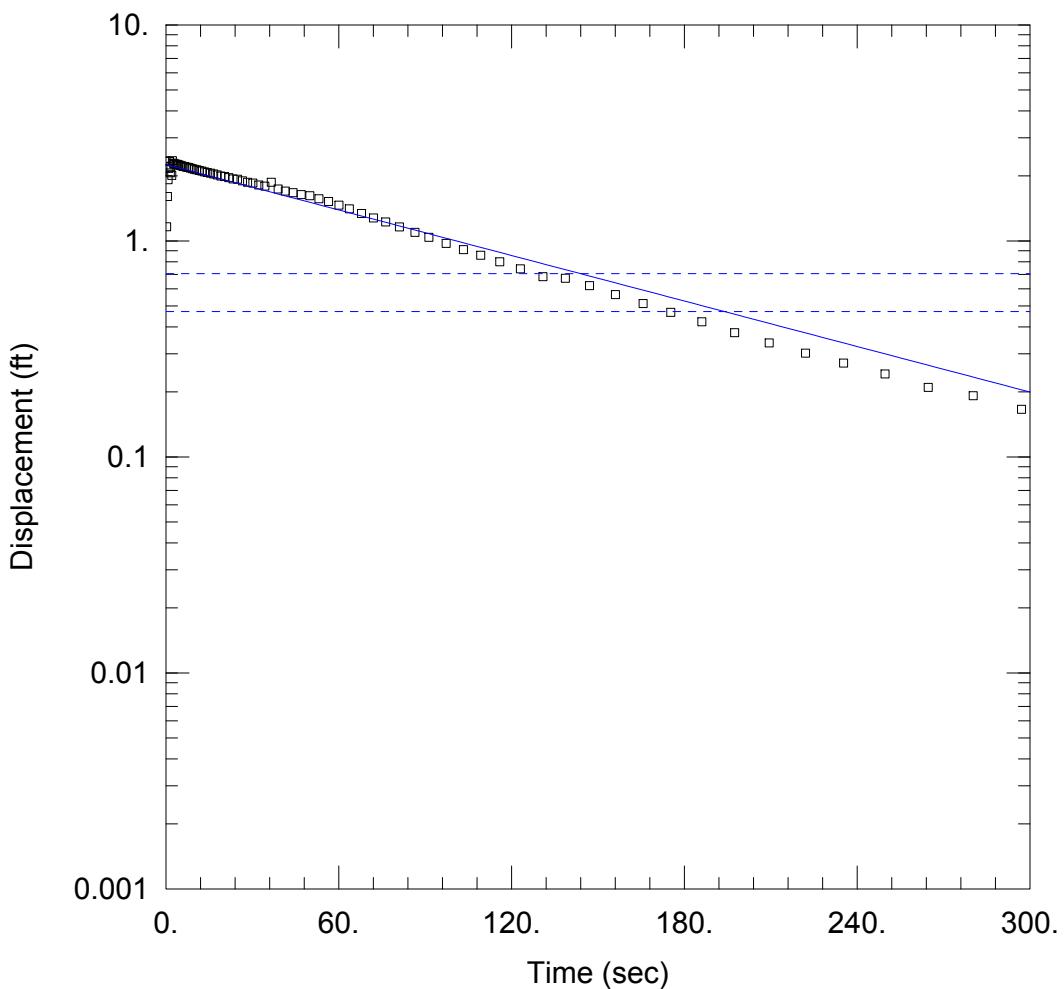
## **Sensitivity Analysis**

Due to uncertainty regarding aquifer thickness, select Aqtesolv® files were revised to recalculate the hydraulic conductivity based upon an aquifer thickness of 125 feet (approximately double the thickness used for the calculations discussed above). The results indicate that the calculations are not significantly affected by variations in aquifer thickness. For example, the falling head test at MW-15d, which yielded a result of 69.0 ft/day by the Springer-Gelhar method yielded a K value of 52.7 ft/day, well within an order of magnitude of the original result. Changing the aquifer thickness for the falling head test at MW-12s resulted in a K value of 3.5 ft/day, nearly identical to the original result.

## **References**

Bouwer, H. and R.C. Rice, 1976. A slug test method for determining hydraulic conductivity of unconfined aquifers with completely or partially penetrating wells, Water Resources Research, vol. 12, no. 3, pp. 423-428.

Springer, R.K. and L.W. Gelhar, 1991. Characterization of large-scale aquifer heterogeneity in glacial outwash by analysis of slug tests with oscillatory response, Cape Cod, Massachusetts, U.S. Geol. Surv. Water Res. Invest. Rep. 91-4034, pp. 36-40.



### FALLING

Data Set: C:\...\8s falling.aqt  
 Date: 04/16/12

Time: 13:29:08

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

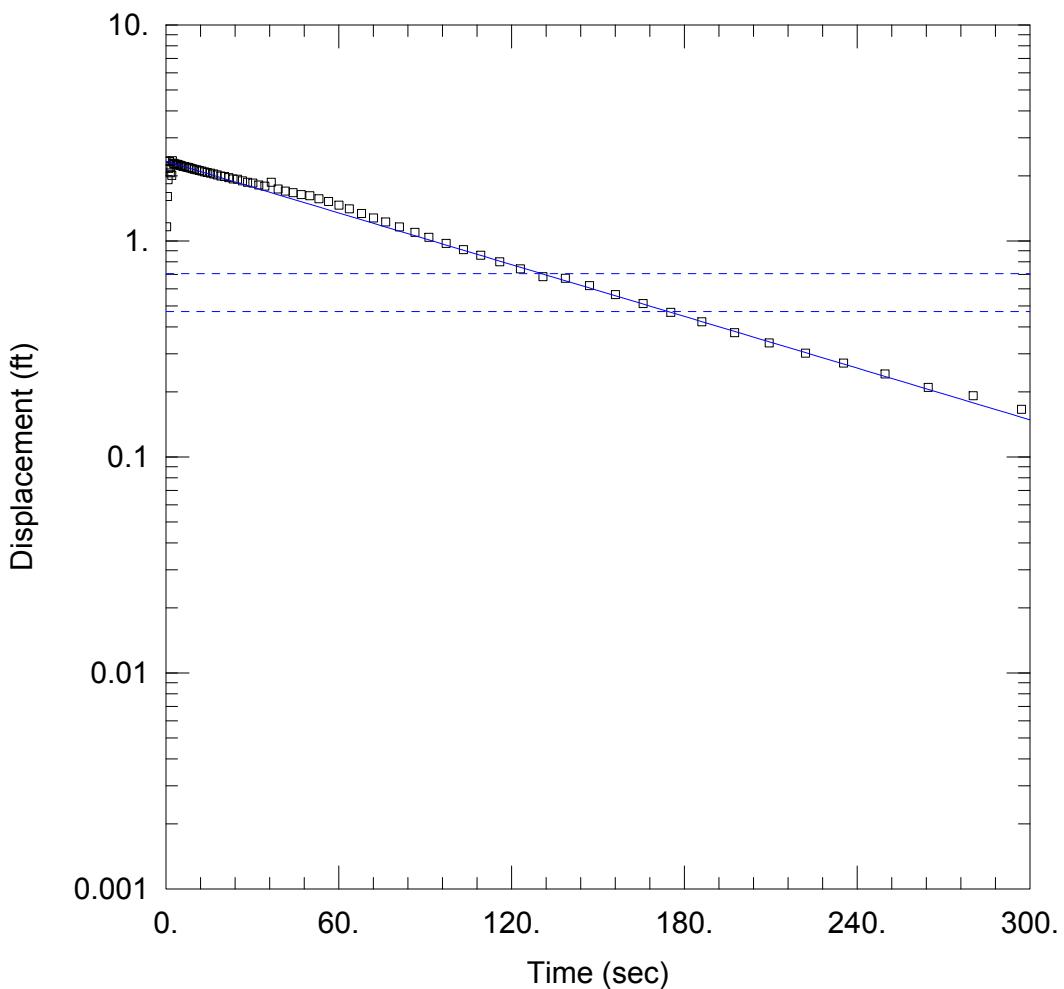
Saturated Thickness: 62.51 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-8s)

Initial Displacement: <u>2.353 ft</u>	Static Water Column Height: <u>27.51 ft</u>
Total Well Penetration Depth: <u>27.51 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.6198 ft/day</u>	y0 = <u>2.259 ft</u>



### FALLING

Data Set: C:\...\8s falling.aqt  
 Date: 04/16/12

Time: 10:51:54

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

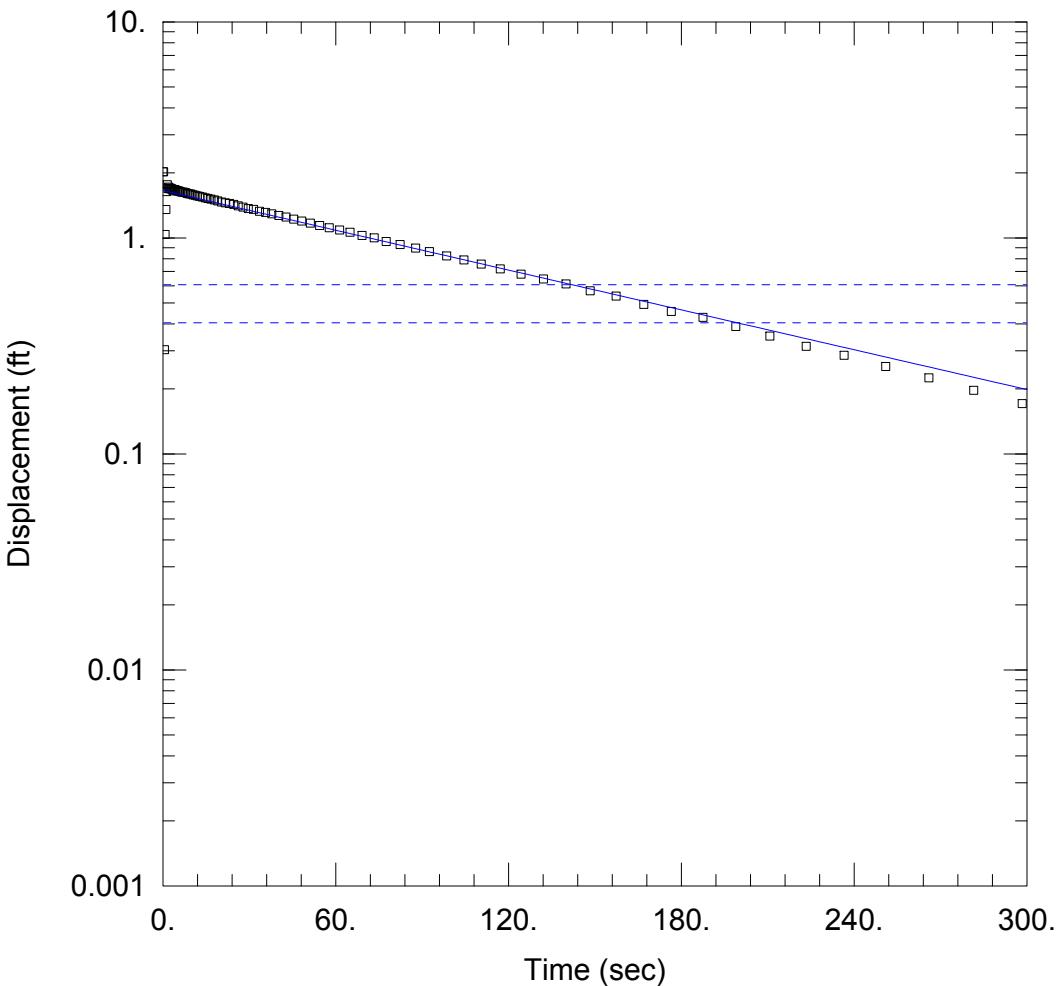
Saturated Thickness: 62.51 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-8s)

Initial Displacement: <u>2.353 ft</u>	Static Water Column Height: <u>27.51 ft</u>
Total Well Penetration Depth: <u>27.51 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.7044 ft/day</u>	y0 = <u>2.338 ft</u>



### RISING

Data Set: C:\...\8s rising.aqt  
 Date: 04/16/12

Time: 12:42:49

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

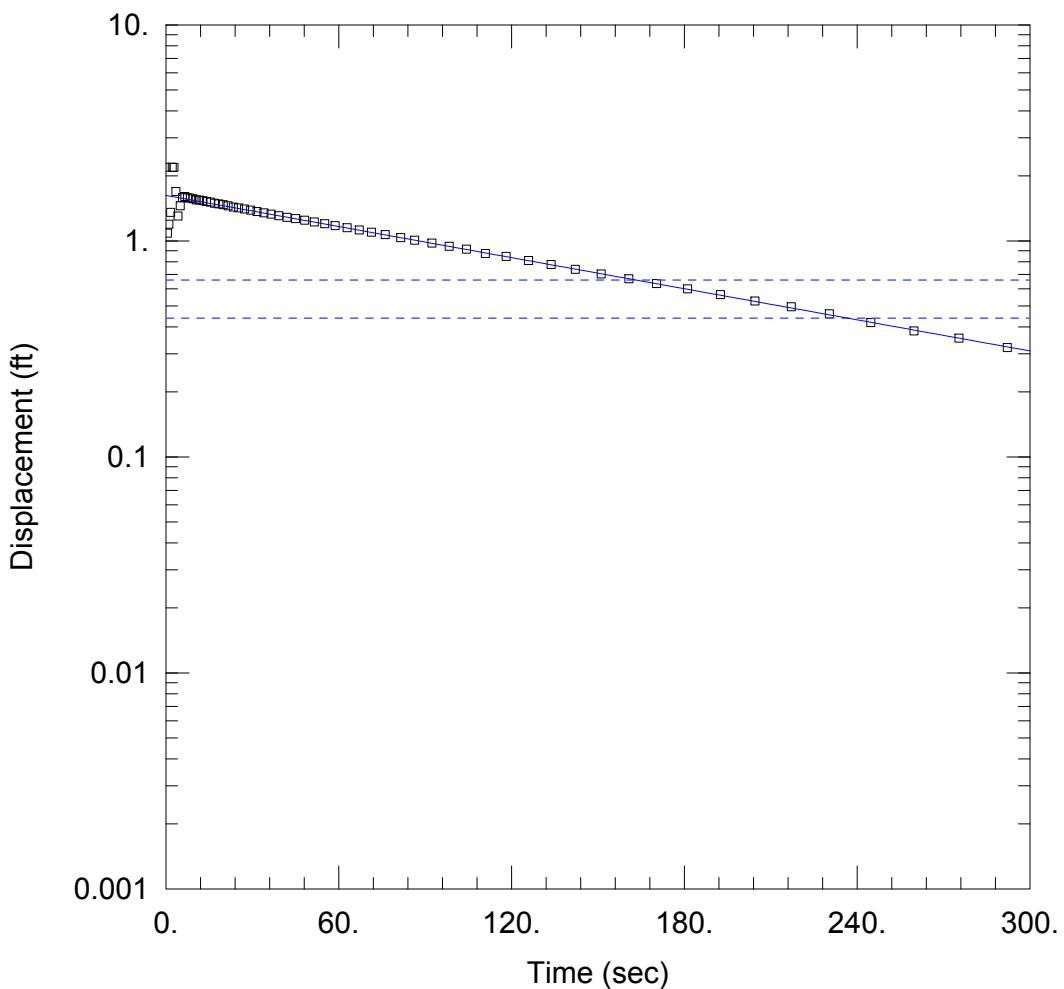
Saturated Thickness: 62.51 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-8s)

Initial Displacement: <u>2.024 ft</u>	Static Water Column Height: <u>27.51 ft</u>
Total Well Penetration Depth: <u>27.51 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.5424 ft/day</u>	y0 = <u>1.659 ft</u>



### FALLING

Data Set: C:\...\9s falling.aqt  
 Date: 04/16/12

Time: 10:54:11

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

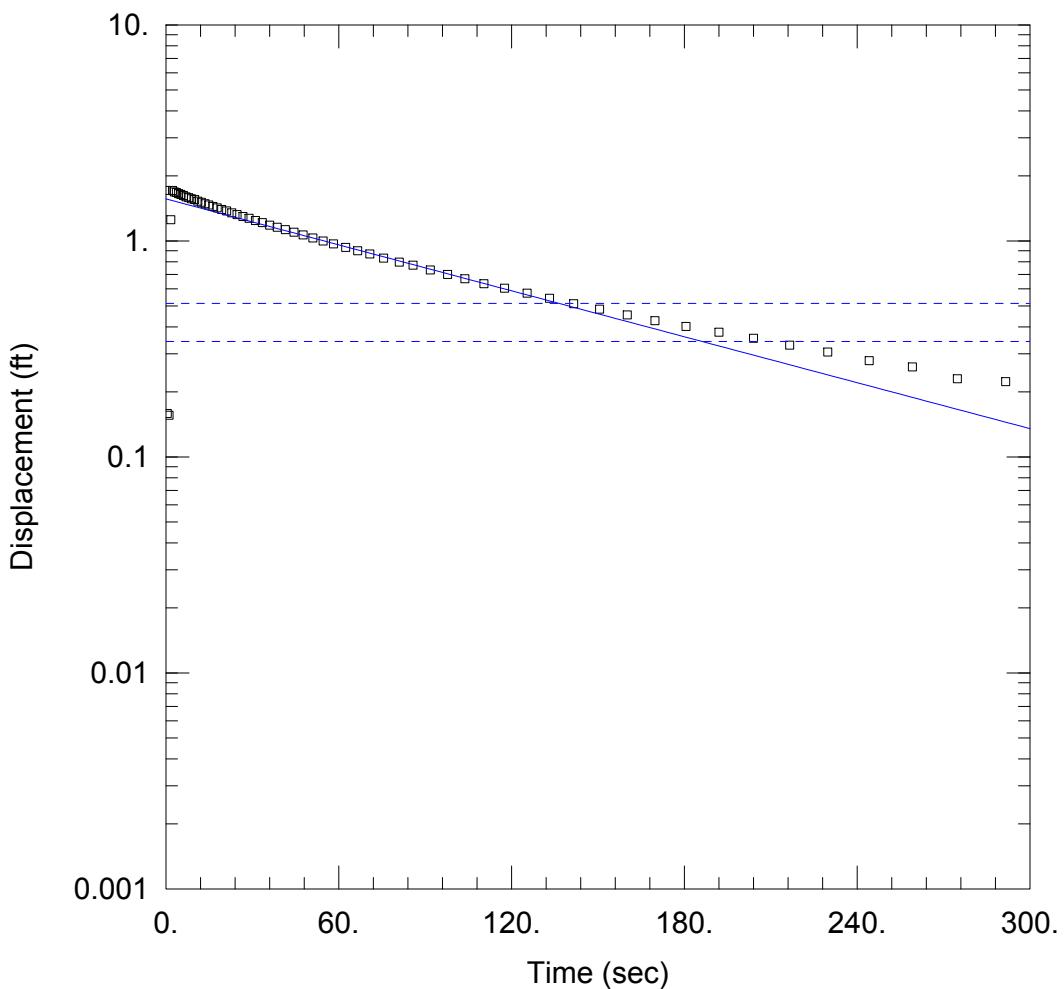
Saturated Thickness: 65.24 ft      Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-9s)

Initial Displacement: <u>2.195</u> ft	Static Water Column Height: <u>30.24</u> ft
Total Well Penetration Depth: <u>30.24</u> ft	Screen Length: <u>10.</u> ft
Casing Radius: <u>0.0833</u> ft	Well Radius: <u>0.33</u> ft

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.4293</u> ft/day	y0 = <u>1.624</u> ft



### RISING

Data Set: C:\...\9s rising.aqt  
 Date: 04/16/12

Time: 13:30:39

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

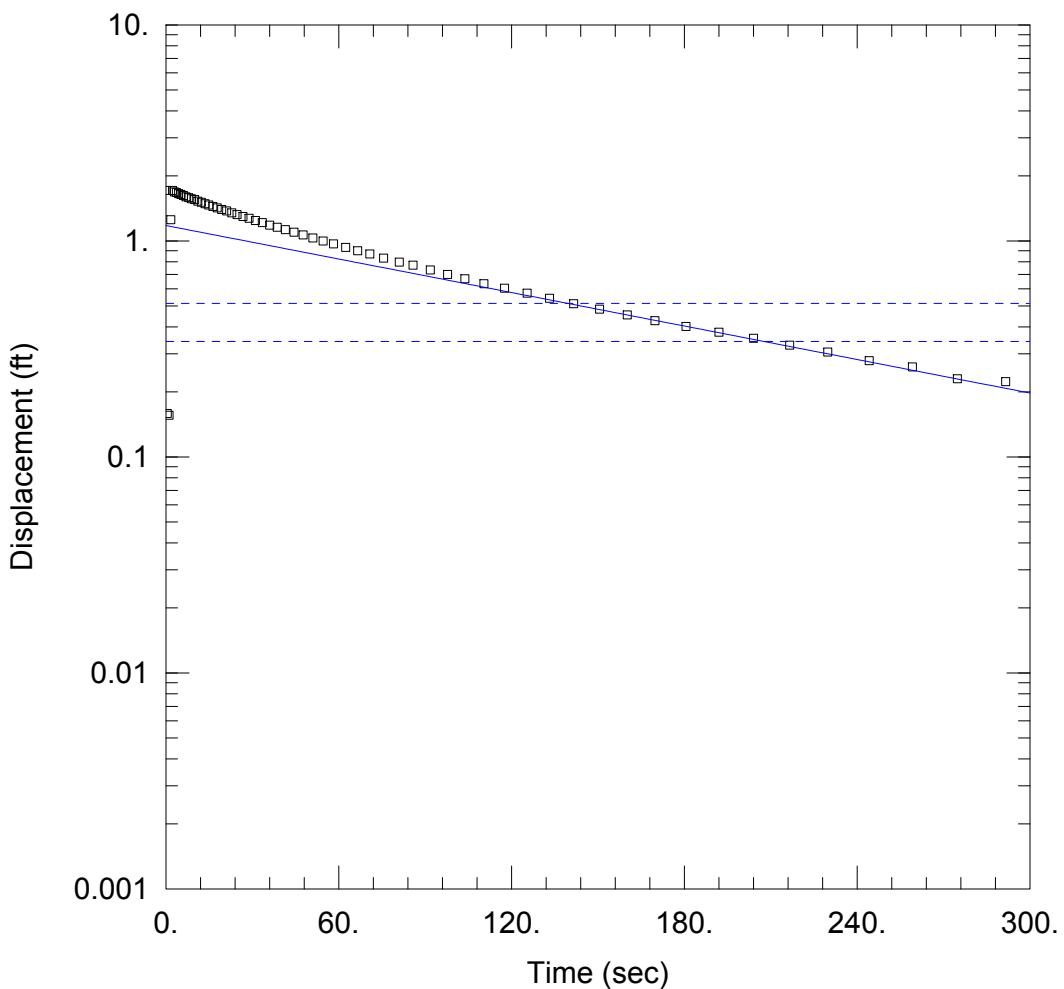
Saturated Thickness: 65.24 ft      Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-9s)

Initial Displacement: <u>1.712</u> ft	Static Water Column Height: <u>30.24</u> ft
Total Well Penetration Depth: <u>30.24</u> ft	Screen Length: <u>10.</u> ft
Casing Radius: <u>0.0833</u> ft	Well Radius: <u>0.33</u> ft

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.6342</u> ft/day	y0 = <u>1.565</u> ft



### RISING

Data Set: C:\...\9s rising.aqt  
 Date: 04/16/12

Time: 11:42:54

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

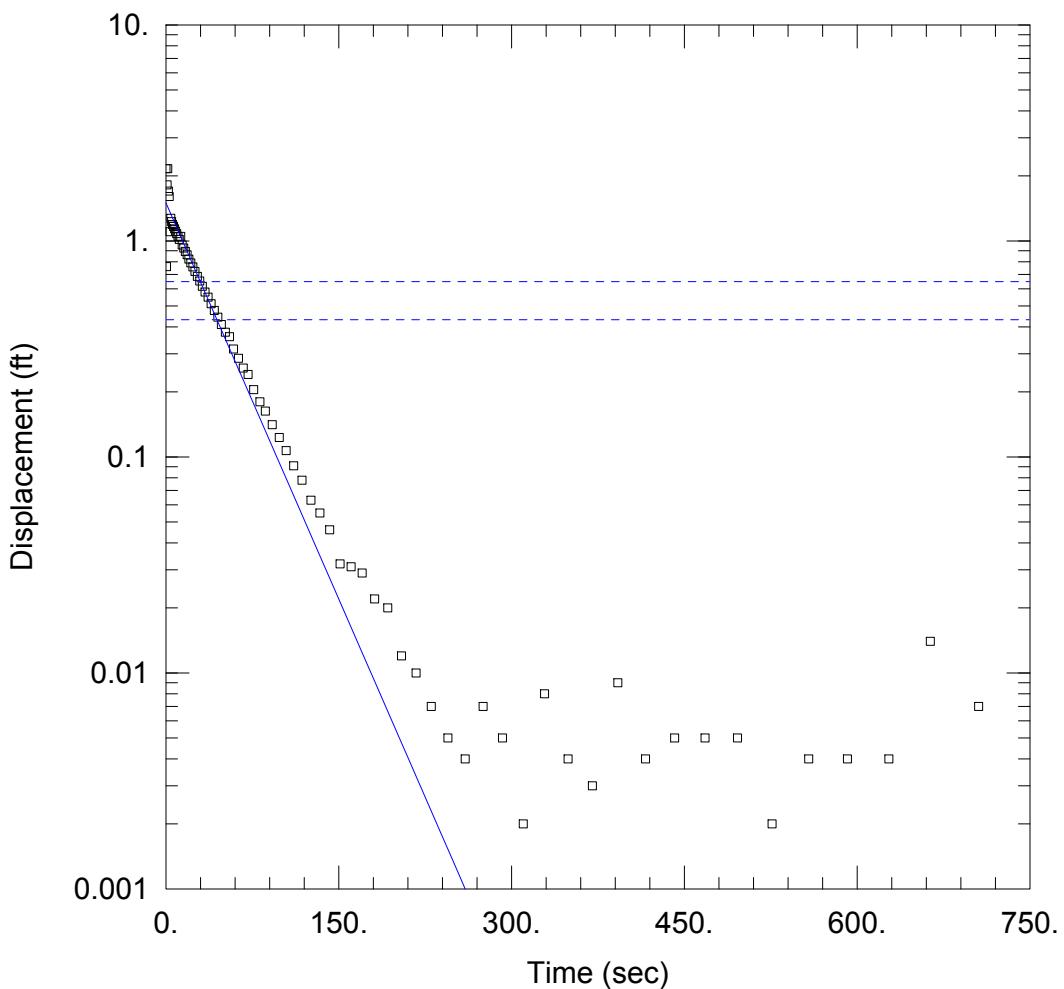
Saturated Thickness: 65.24 ft      Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-9s)

Initial Displacement: <u>1.712</u> ft	Static Water Column Height: <u>30.24</u> ft
Total Well Penetration Depth: <u>30.24</u> ft	Screen Length: <u>10.</u> ft
Casing Radius: <u>0.0833</u> ft	Well Radius: <u>0.33</u> ft

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.4625</u> ft/day	y0 = <u>1.178</u> ft



### FALLING

Data Set: C:\...\11d falling.aqt  
 Date: 04/16/12

Time: 11:44:45

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

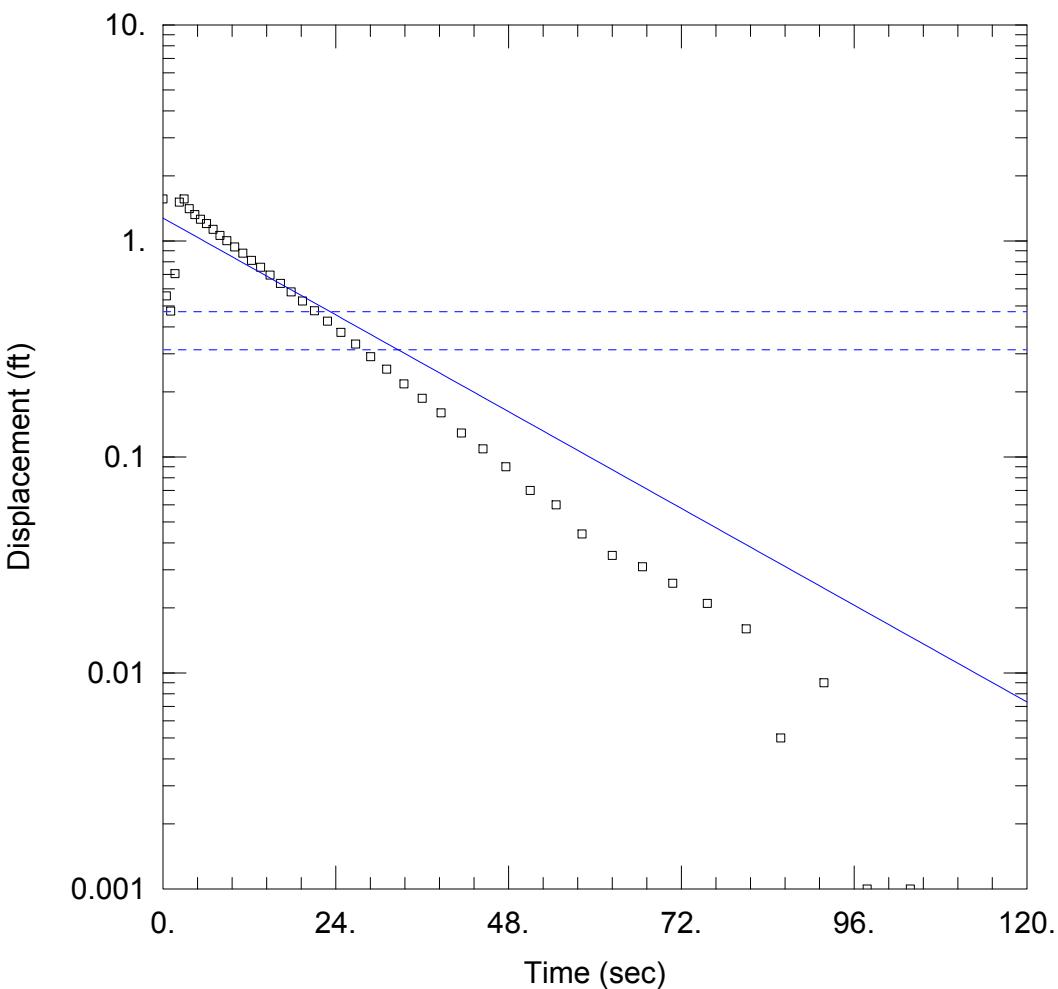
Saturated Thickness: 63.58 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (New Well)

Initial Displacement: <u>2.159 ft</u>	Static Water Column Height: <u>63.58 ft</u>
Total Well Penetration Depth: <u>63.58 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>3.076 ft/day</u>	y0 = <u>1.498 ft</u>



### RISING

Data Set: C:\...\11d rising.aqt  
 Date: 04/16/12

Time: 13:32:23

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

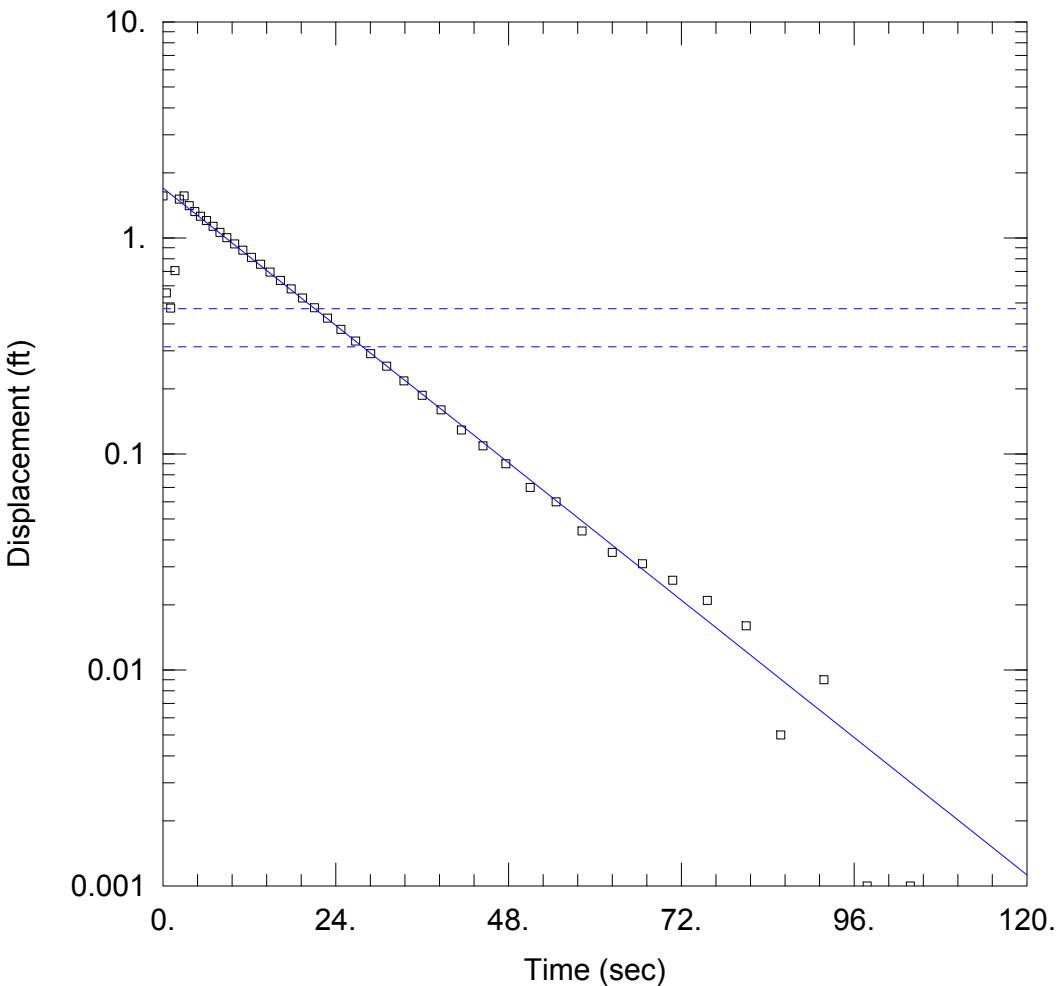
Saturated Thickness: 63.58 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (New Well)

Initial Displacement: <u>1.566 ft</u>	Static Water Column Height: <u>63.58 ft</u>
Total Well Penetration Depth: <u>63.58 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
$K = 4.697 \text{ ft/day}$	$y_0 = 1.276 \text{ ft}$



### RISING

Data Set: C:\...\11d rising.aqt  
 Date: 04/16/12

Time: 11:45:33

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

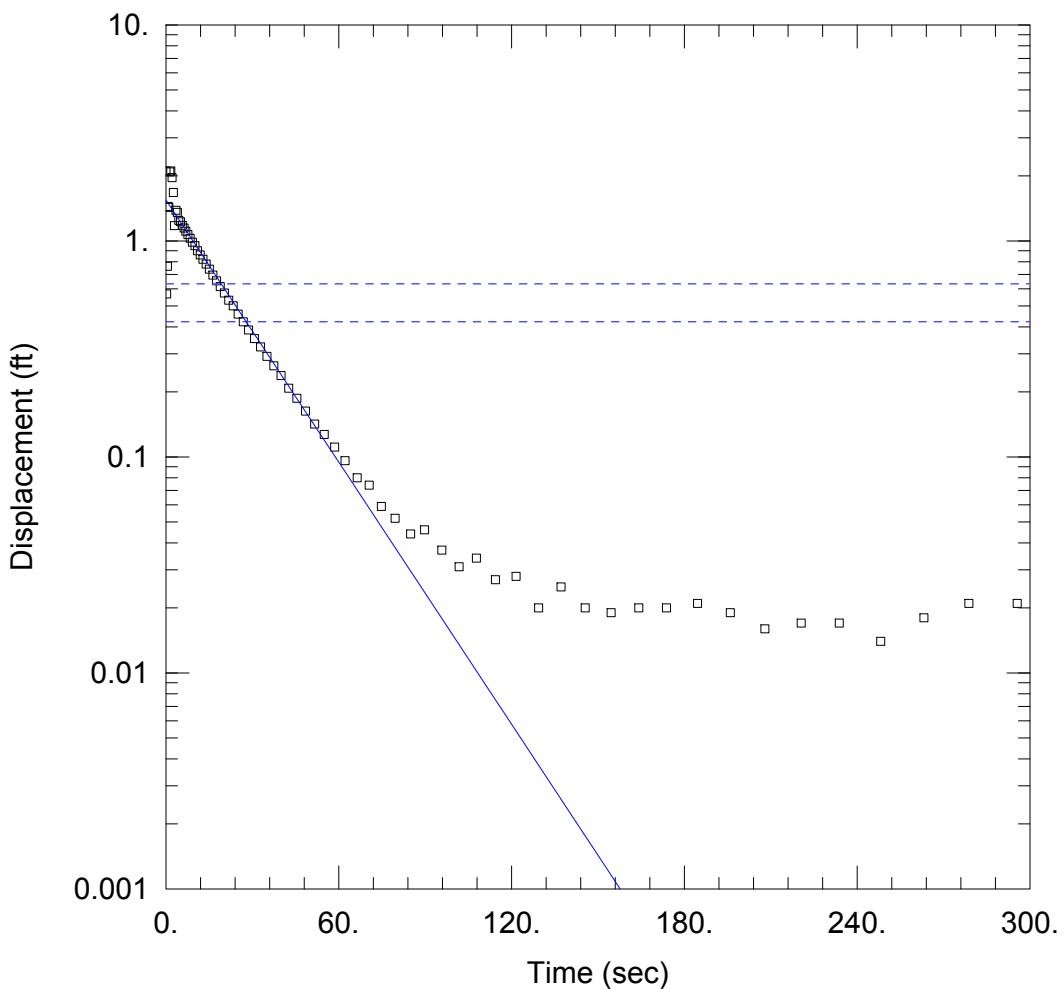
Saturated Thickness: 63.58 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (New Well)

Initial Displacement: 1.566 ft Static Water Column Height: 63.58 ft  
 Total Well Penetration Depth: 63.58 ft Screen Length: 10. ft  
 Casing Radius: 0.0833 ft Well Radius: 0.33 ft

### SOLUTION

Aquifer Model: Unconfined Solution Method: Bouwer-Rice  
 $K = 6.666 \text{ ft/day}$   $y_0 = 1.702 \text{ ft}$



### FALLING

Data Set: C:\...\12s falling.aqt  
 Date: 04/16/12

Time: 13:00:20

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

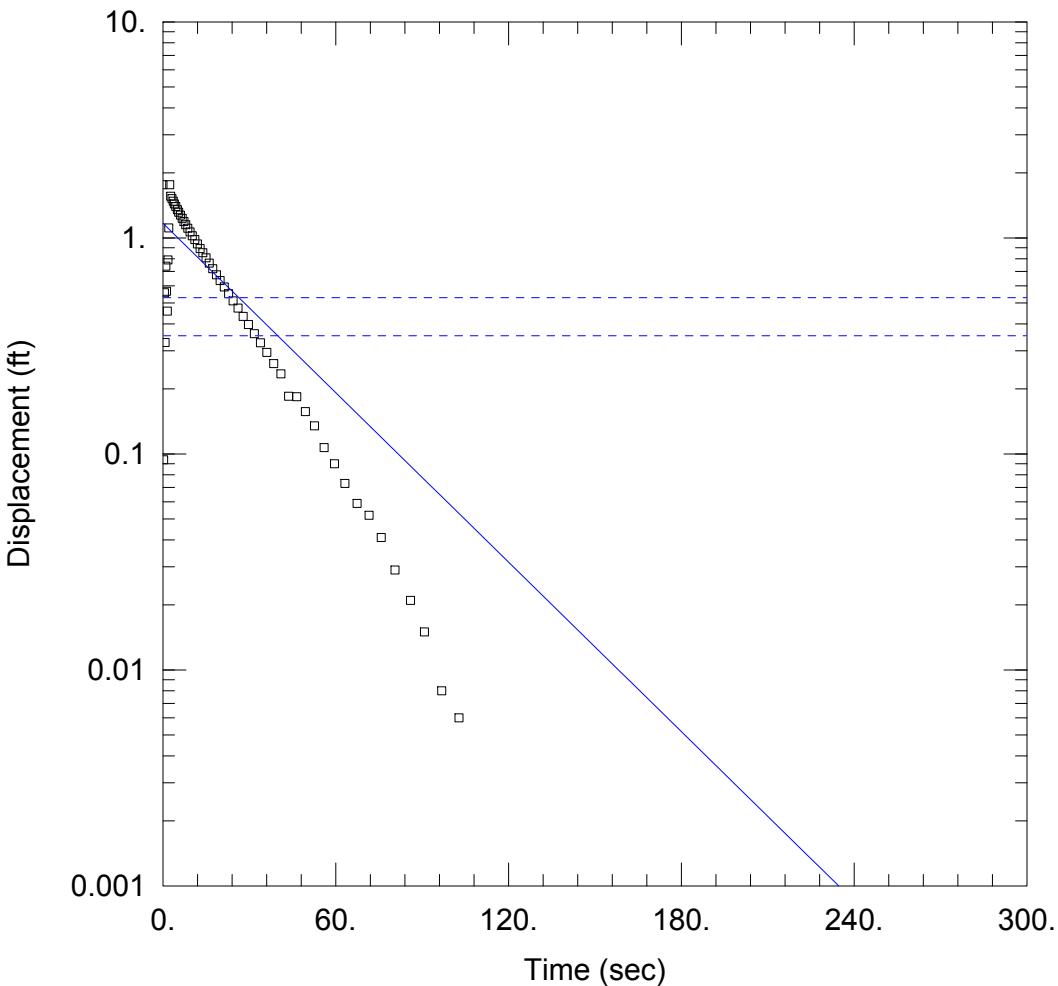
Saturated Thickness: 63.37 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-12s)

Initial Displacement: <u>2.11 ft</u>	Static Water Column Height: <u>28.37 ft</u>
Total Well Penetration Depth: <u>28.37 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>3.585 ft/day</u>	y0 = <u>1.545 ft</u>



### RISING

Data Set: C:\...\12s rising.aqt  
 Date: 04/16/12

Time: 13:34:37

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

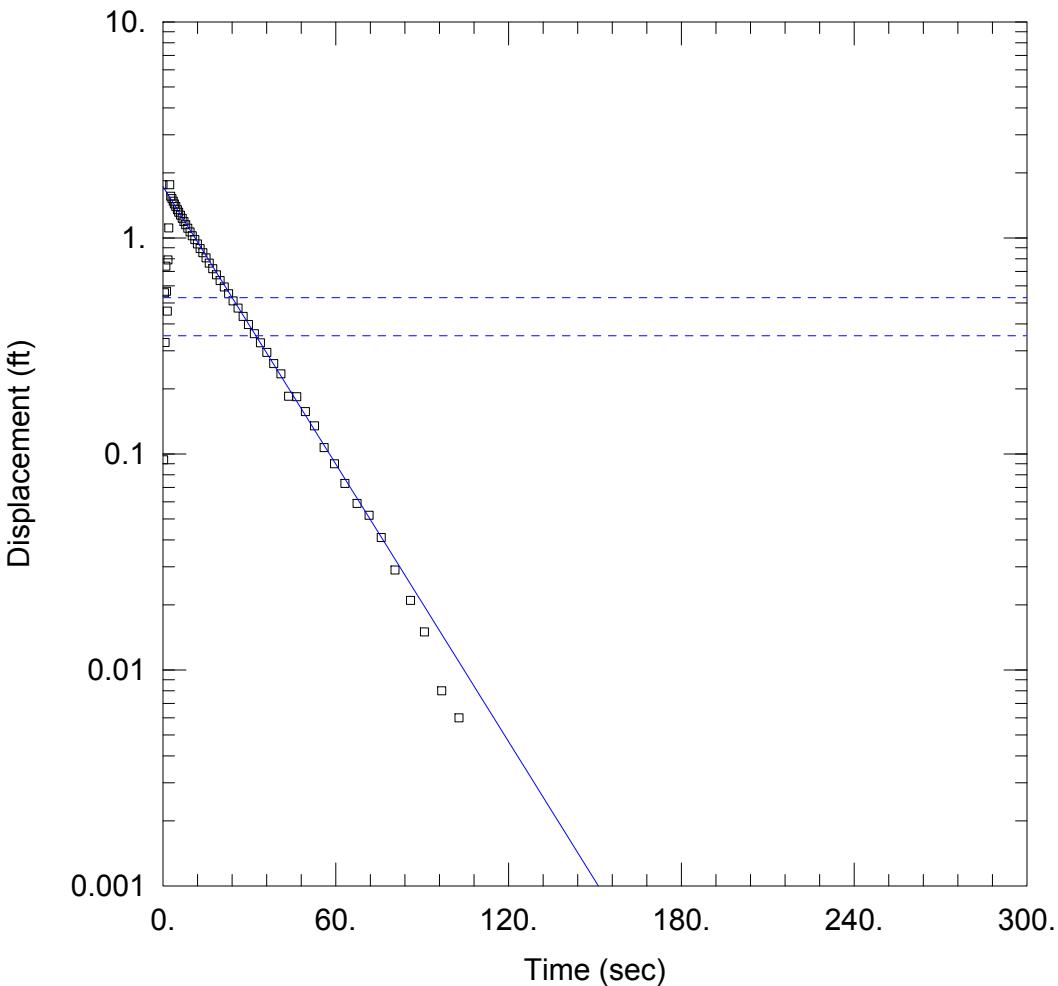
Saturated Thickness: 63.37 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-12s)

Initial Displacement: <u>1.763 ft</u>	Static Water Column Height: <u>28.37 ft</u>
Total Well Penetration Depth: <u>28.37 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>2.318 ft/day</u>	y0 = <u>1.169 ft</u>



### RISING

Data Set: C:\...\12s rising.aqt  
 Date: 04/16/12

Time: 11:47:05

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

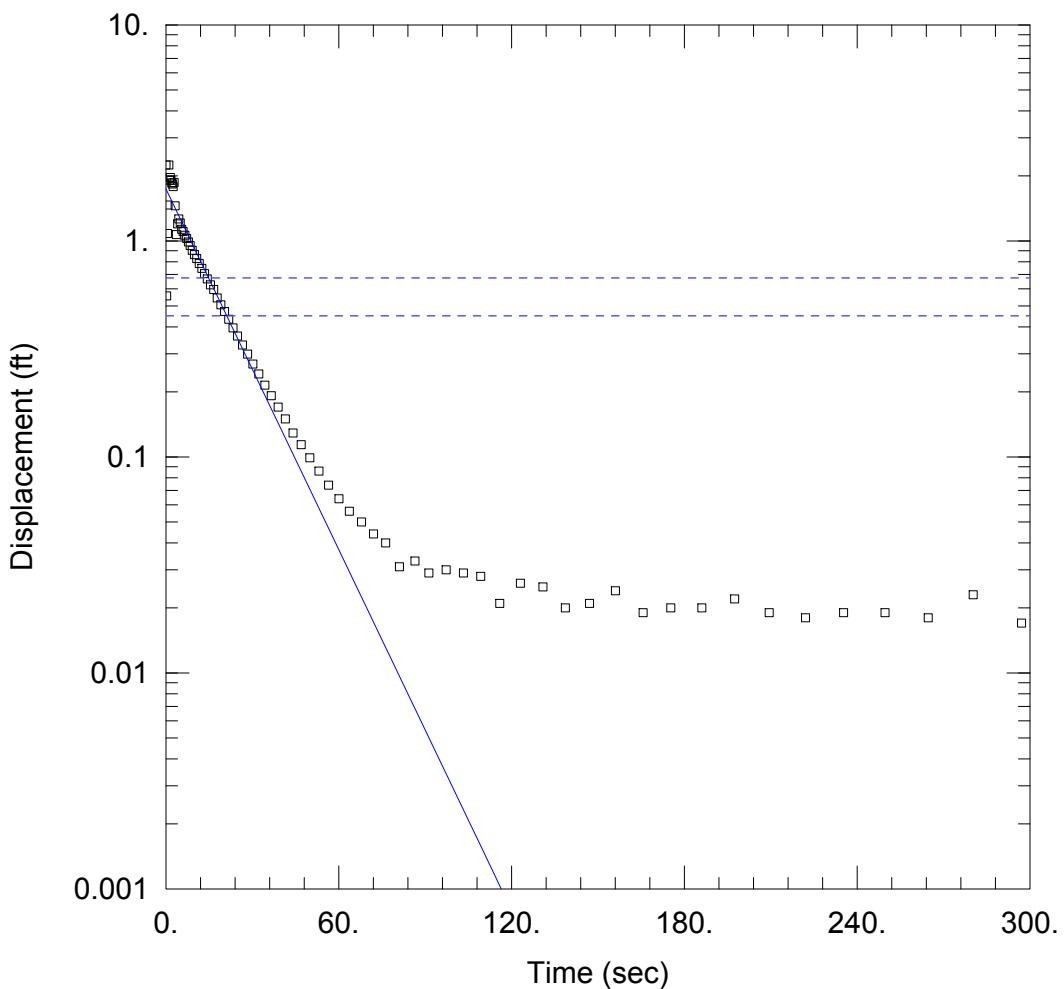
Saturated Thickness: 63.37 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-12s)

Initial Displacement: <u>1.763 ft</u>	Static Water Column Height: <u>28.37 ft</u>
Total Well Penetration Depth: <u>28.37 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>3.797 ft/day</u>	y0 = <u>1.73 ft</u>



### FALLING

Data Set: C:\...\13s falling.aqt  
 Date: 04/16/12

Time: 11:48:28

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

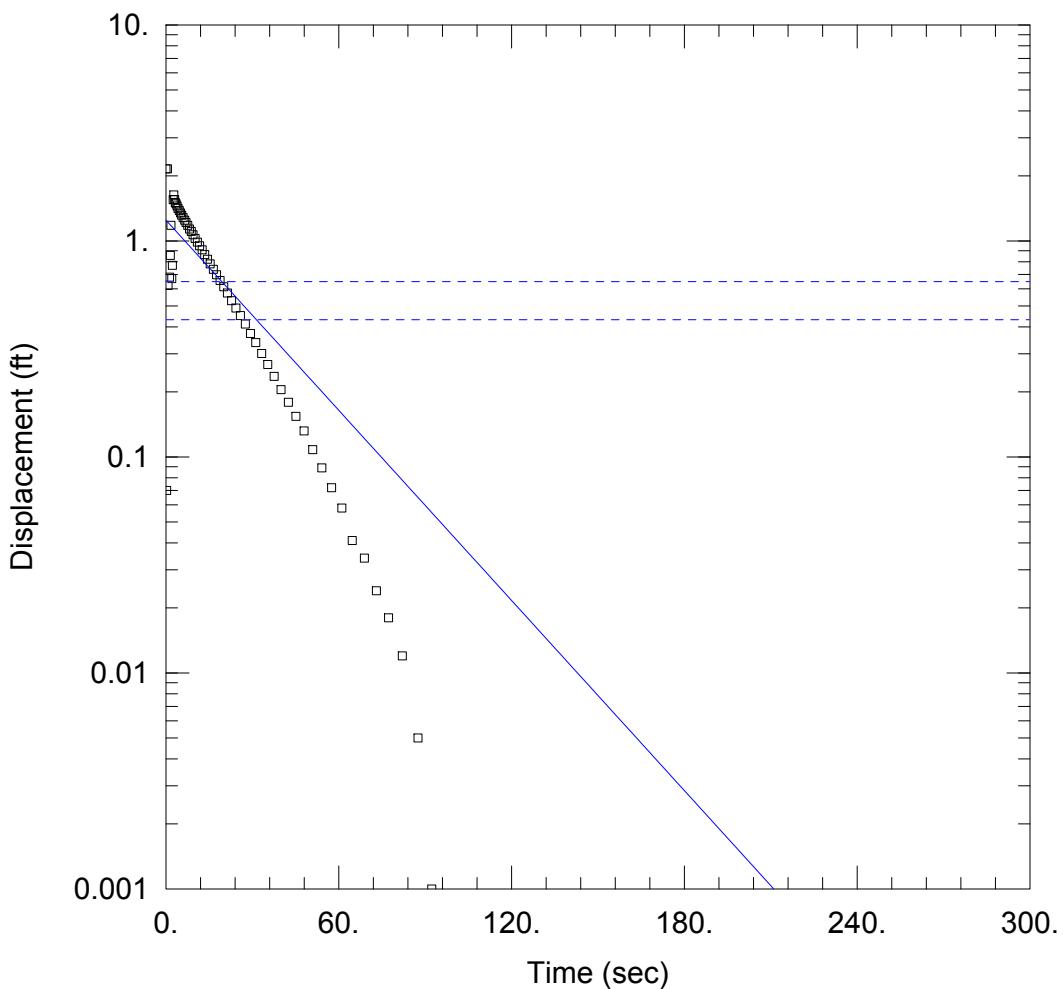
Saturated Thickness: 61.11 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-13s)

Initial Displacement: <u>2.247 ft</u>	Static Water Column Height: <u>26.11 ft</u>
Total Well Penetration Depth: <u>26.11 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
$K = 4.874 \text{ ft/day}$	$y_0 = 1.738 \text{ ft}$



### RISING

Data Set: C:\...\13s rising.aqt  
 Date: 04/16/12

Time: 13:36:28

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

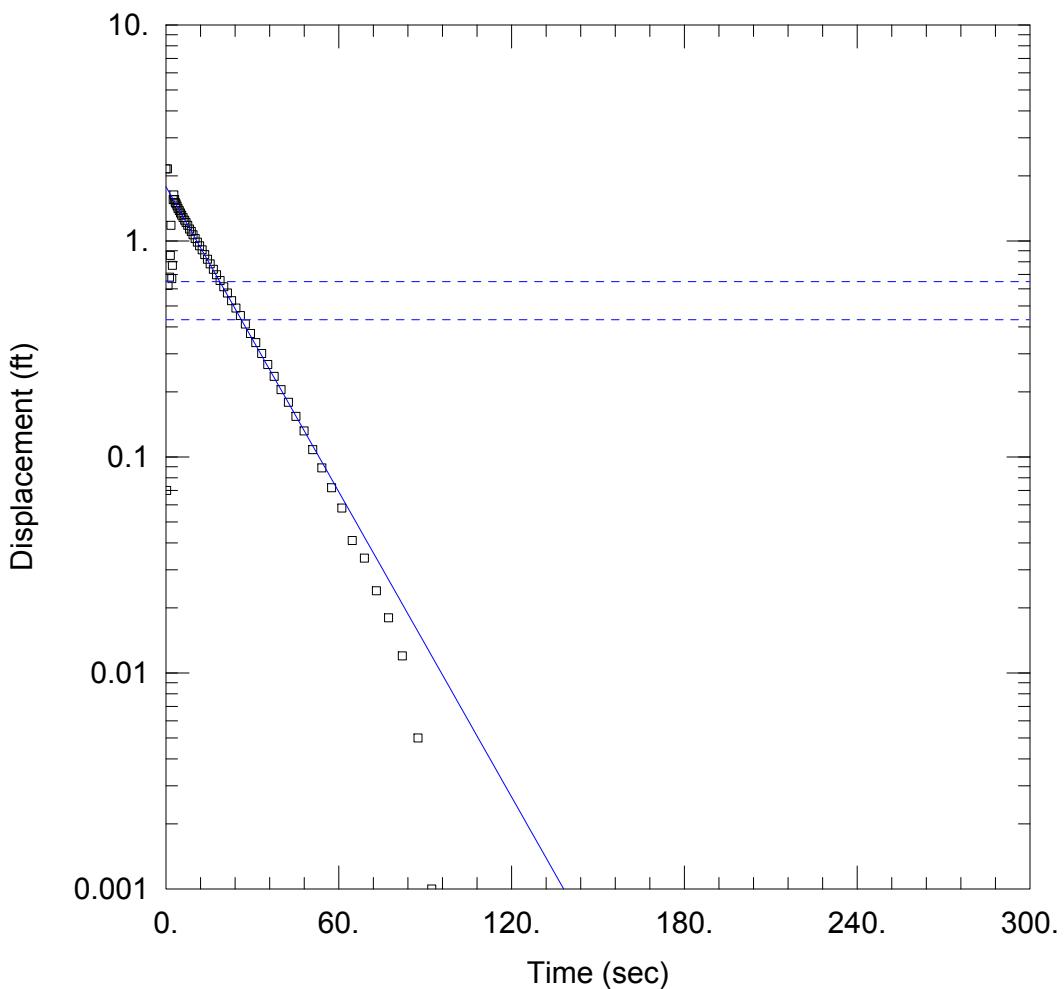
Saturated Thickness: 61.11 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-13s)

Initial Displacement: <u>2.158 ft</u>	Static Water Column Height: <u>26.11 ft</u>
Total Well Penetration Depth: <u>26.11 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>2.57 ft/day</u>	y0 = <u>1.247 ft</u>



### RISING

Data Set: C:\...\13s rising.aqt  
 Date: 04/16/12

Time: 11:49:02

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

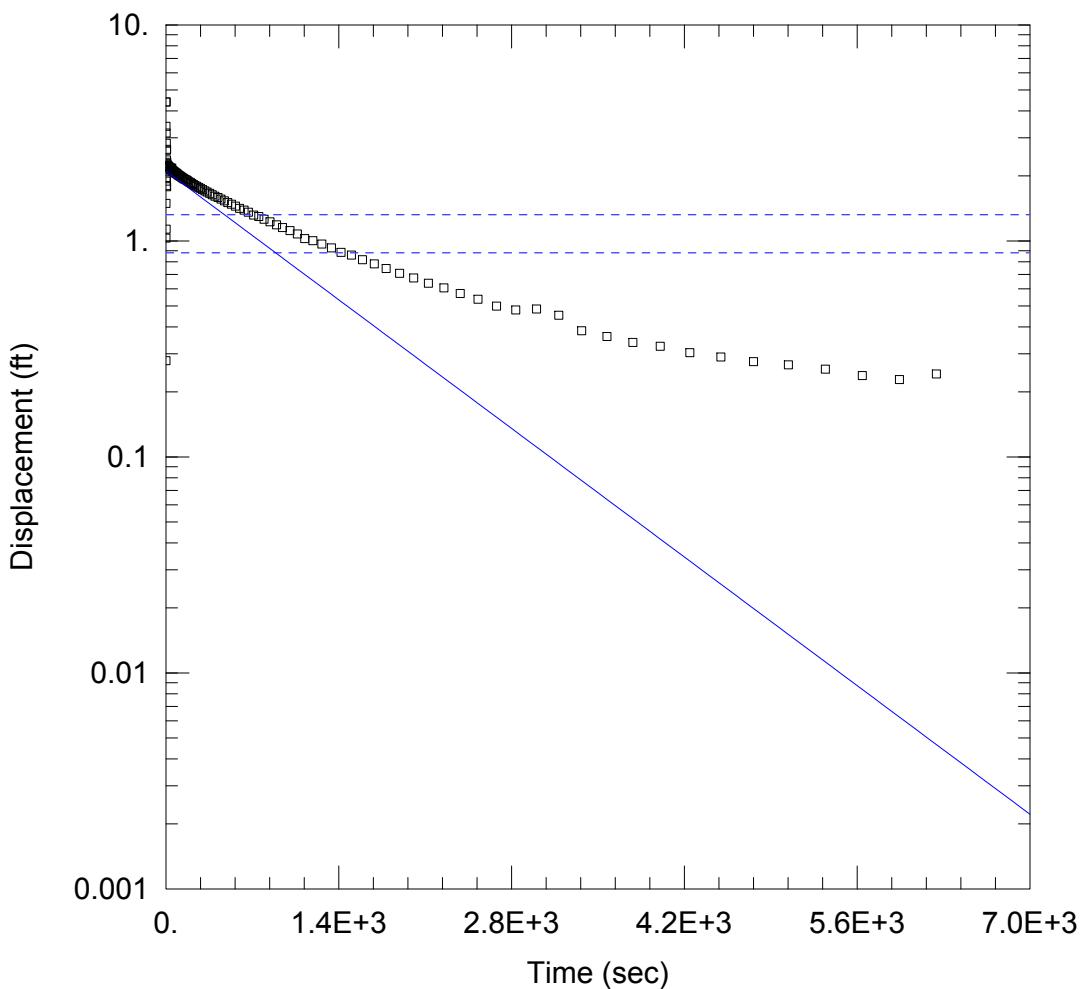
Saturated Thickness: 61.11 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-13s)

Initial Displacement: <u>2.158 ft</u>	Static Water Column Height: <u>26.11 ft</u>
Total Well Penetration Depth: <u>26.11 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>4.125 ft/day</u>	y0 = <u>1.783 ft</u>



### FALLING

Data Set: C:\...\14s falling.aqt  
 Date: 04/16/12

Time: 13:41:11

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

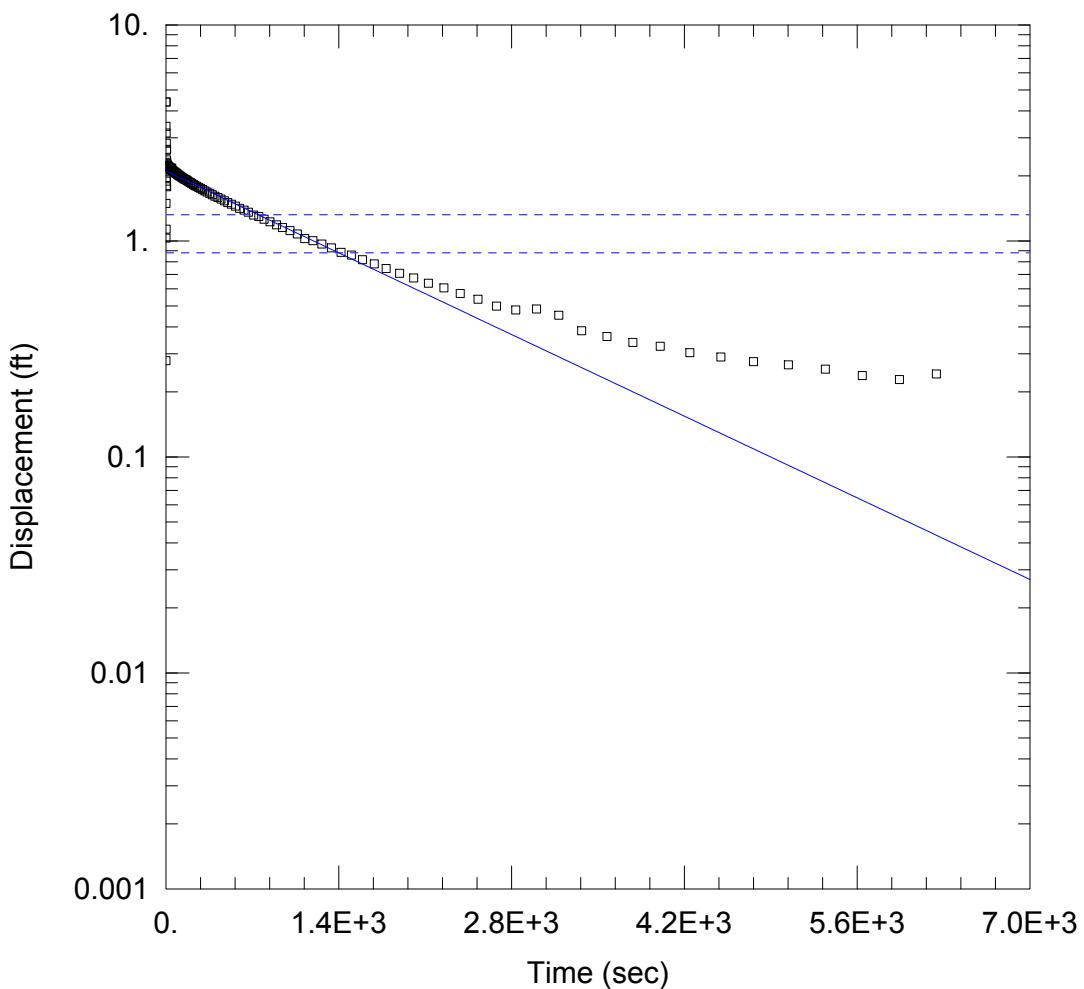
Saturated Thickness: 62.25 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-14s)

Initial Displacement: <u>4.402 ft</u>	Static Water Column Height: <u>26.25 ft</u>
Total Well Penetration Depth: <u>26.25 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.07448 ft/day</u>	y0 = <u>2.102 ft</u>



### FALLING

Data Set: C:\...\14s falling.aqt  
 Date: 04/16/12

Time: 11:50:15

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

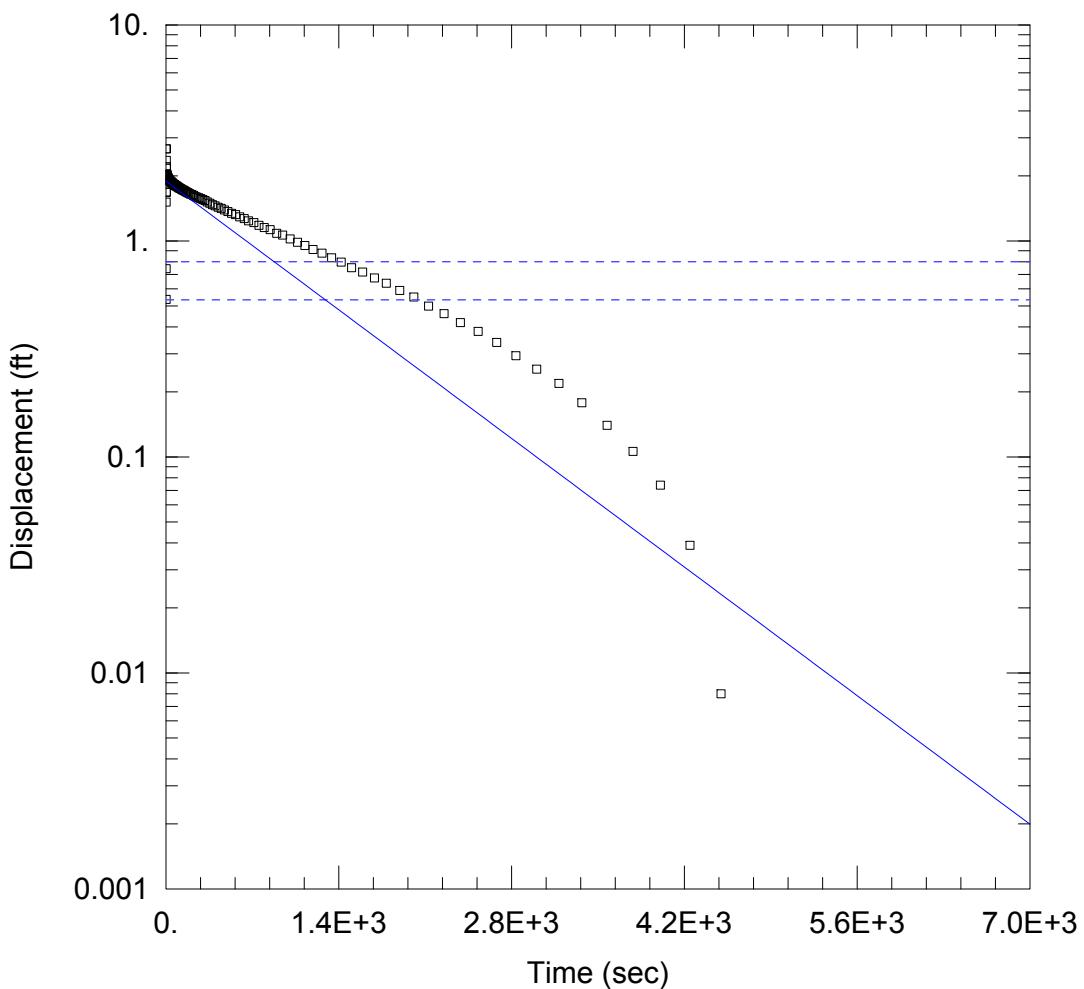
Saturated Thickness: 62.25 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-14s)

Initial Displacement: <u>4.402 ft</u>	Static Water Column Height: <u>26.25 ft</u>
Total Well Penetration Depth: <u>26.25 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.04729 ft/day</u>	y0 = <u>2.102 ft</u>



### RISING

Data Set: C:\...\14s rising.aqt  
 Date: 04/16/12

Time: 13:38:29

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

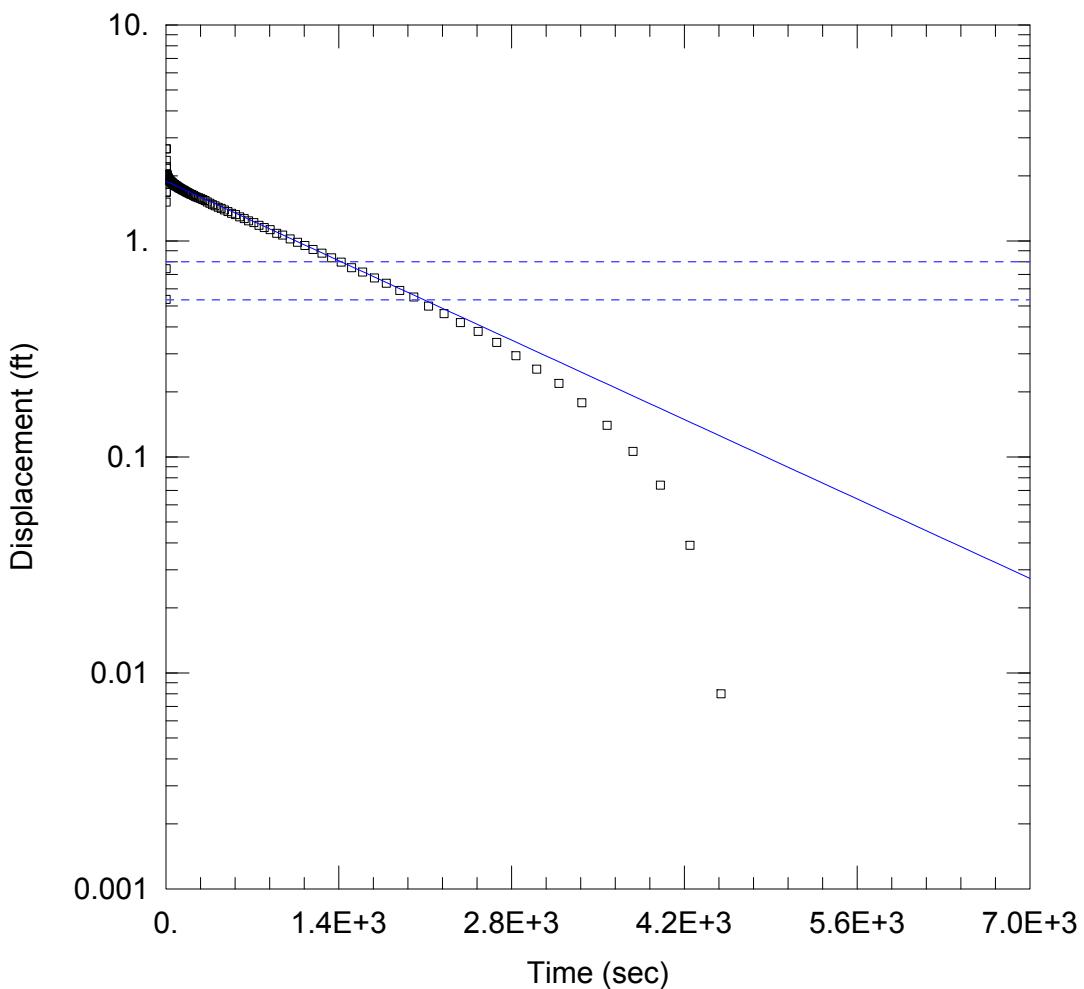
Saturated Thickness: 62.25 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-14s)

Initial Displacement: <u>2.669 ft</u>	Static Water Column Height: <u>26.25 ft</u>
Total Well Penetration Depth: <u>26.25 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.07448 ft/day</u>	y0 = <u>1.889 ft</u>



### RISING

Data Set: C:\...\14s rising.aqt  
 Date: 04/16/12

Time: 11:50:57

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

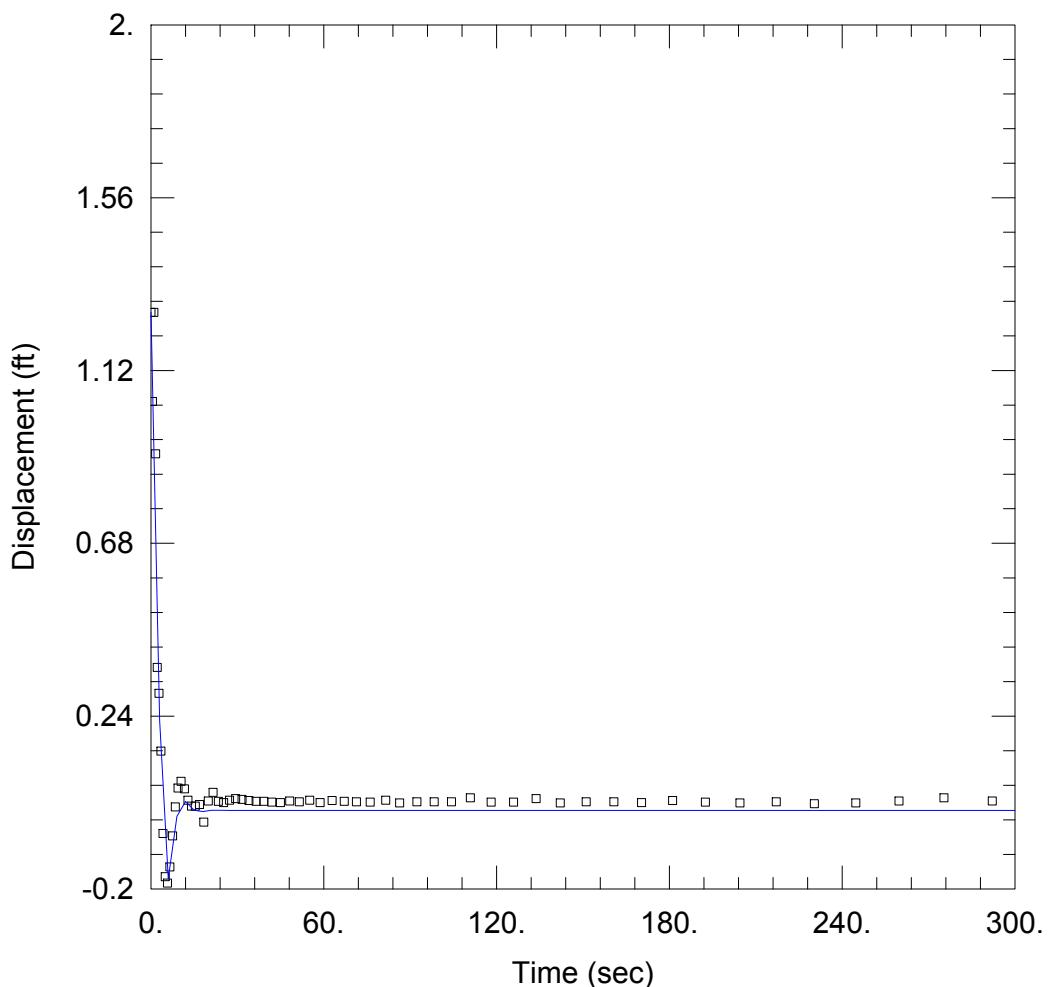
Saturated Thickness: 62.25 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-14s)

Initial Displacement: <u>2.669 ft</u>	Static Water Column Height: <u>26.25 ft</u>
Total Well Penetration Depth: <u>26.25 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.04601 ft/day</u>	y0 = <u>1.889 ft</u>



### FALLING

Data Set: C:\...\15d falling1.aqt  
 Date: 04/16/12

Time: 11:52:12

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

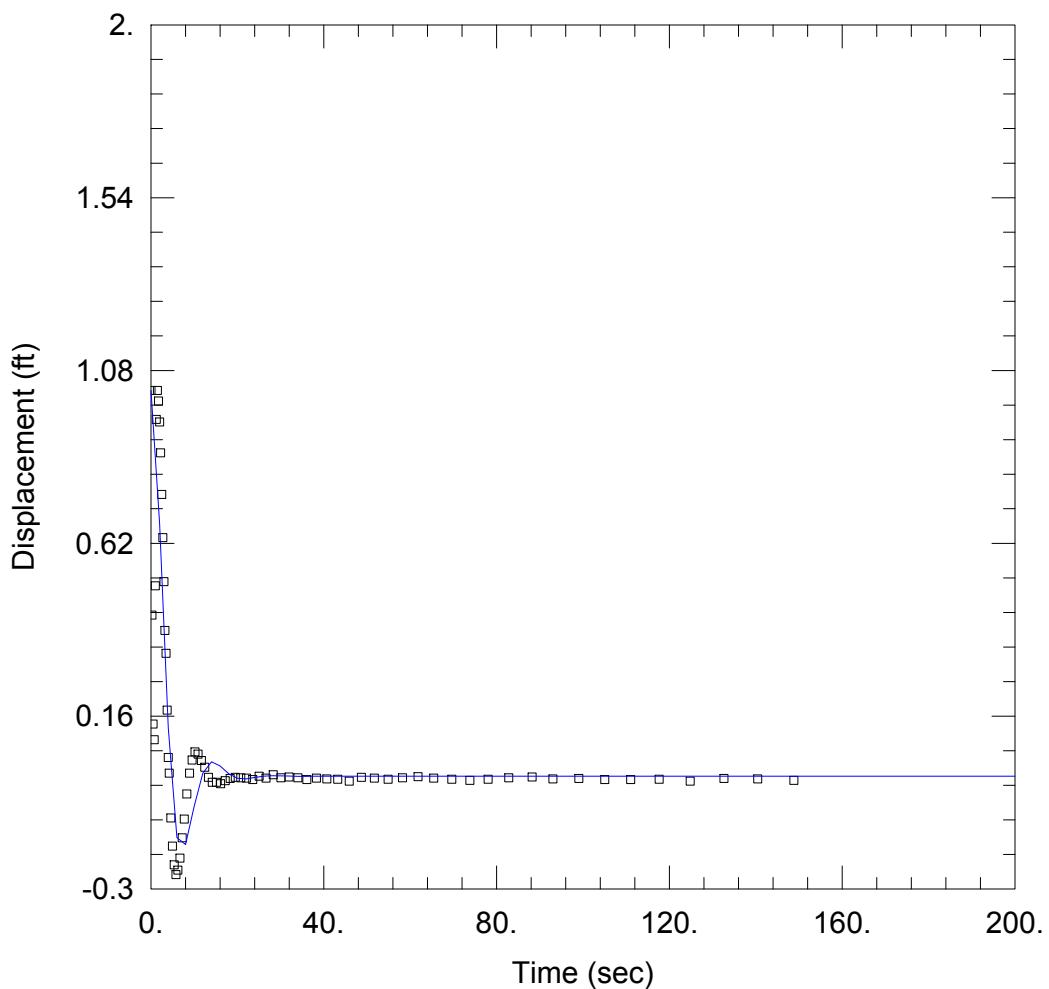
Saturated Thickness: 62.85 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-16s)

Initial Displacement: <u>1.268 ft</u>	Static Water Column Height: <u>62.85 ft</u>
Total Well Penetration Depth: <u>62.85 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Springer-Gelhar</u>
K = <u>69. ft/day</u>	Le = <u>74.21 ft</u>



### RISING

Data Set: C:\...\15d rising1.aqt  
 Date: 04/16/12

Time: 11:54:18

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

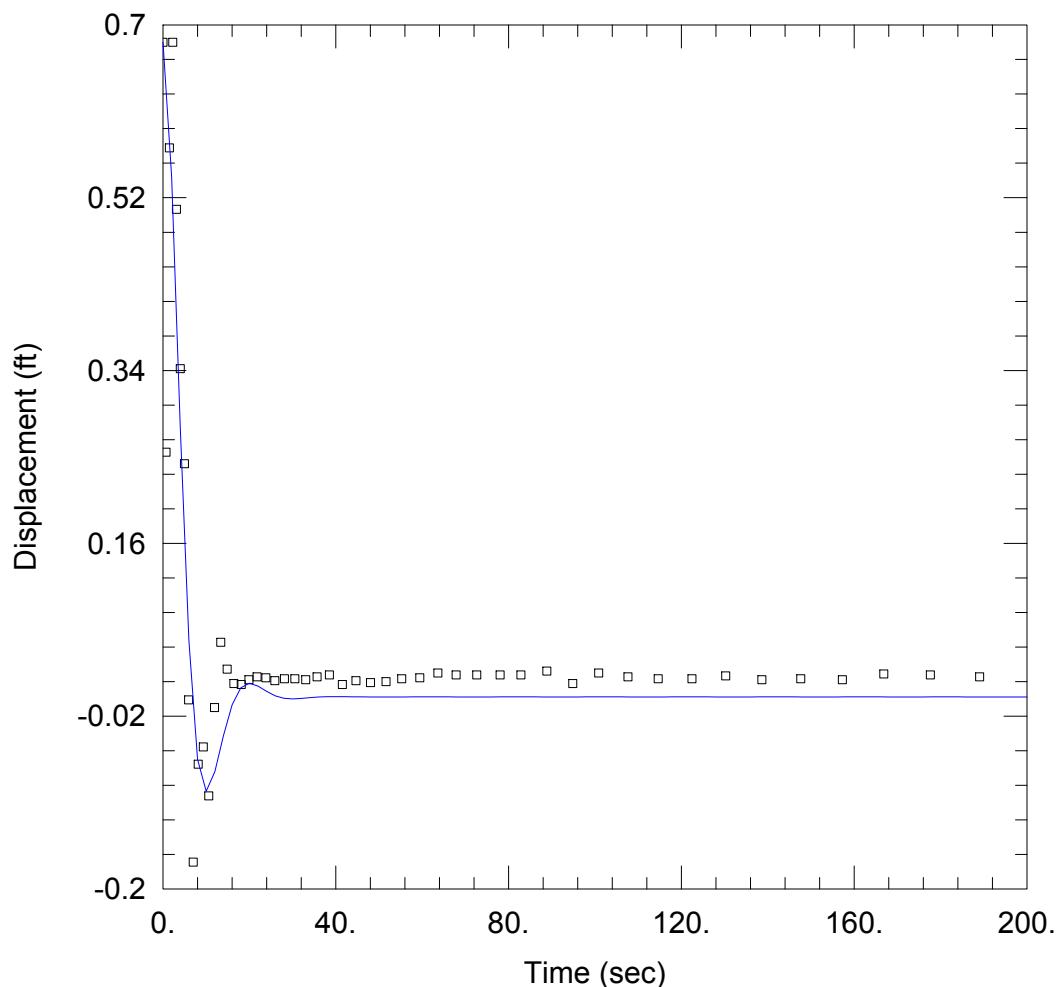
Saturated Thickness: 62.85 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-16s)

Initial Displacement: <u>1.027 ft</u>	Static Water Column Height: <u>62.85 ft</u>
Total Well Penetration Depth: <u>62.85 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Springer-Gelhar</u>
K = <u>59.53 ft/day</u>	Le = <u>129.2 ft</u>



### FALLING - 2

Data Set: C:\...\15d falling2.aqt  
 Date: 04/16/12

Time: 11:53:27

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

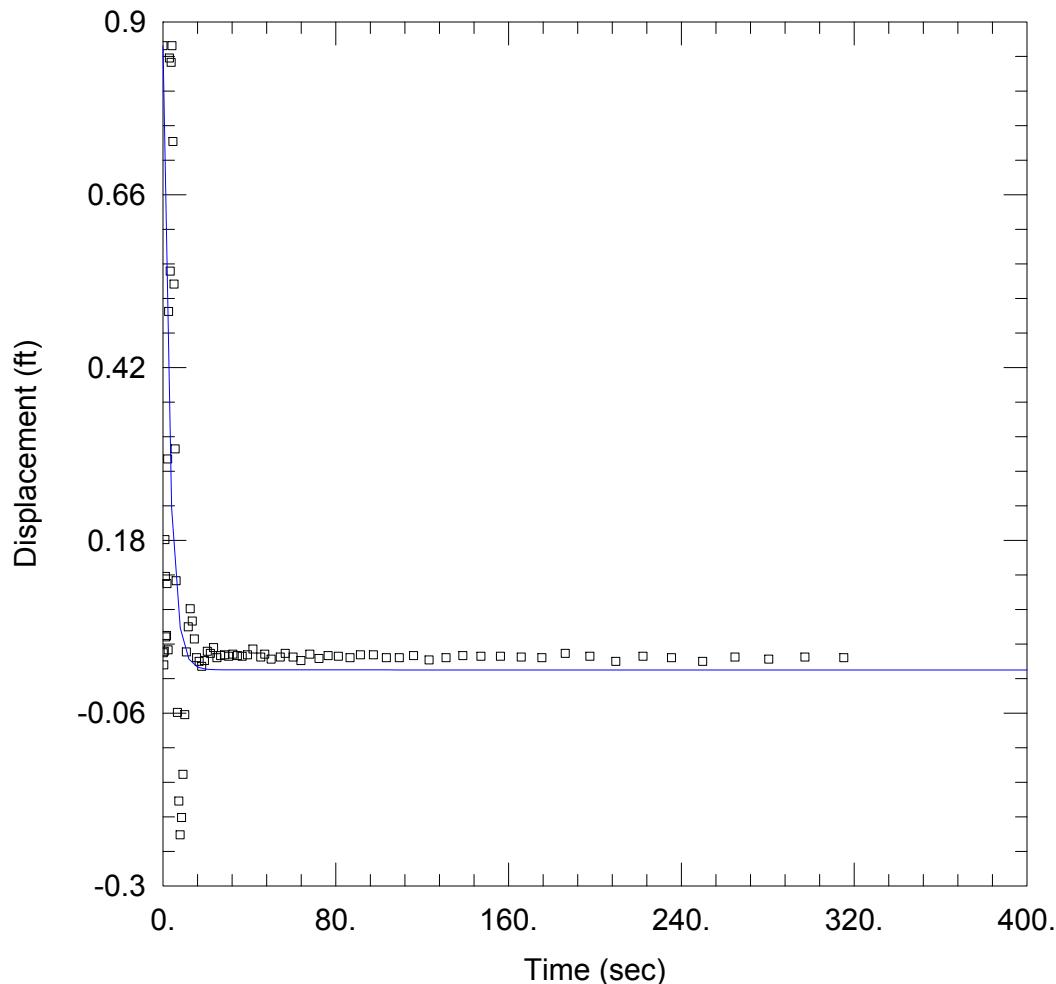
Saturated Thickness: 62.85 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-16s)

Initial Displacement: <u>0.682 ft</u>	Static Water Column Height: <u>62.85 ft</u>
Total Well Penetration Depth: <u>62.85 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Springer-Gelhar</u>
K = <u>38.74 ft/day</u>	Le = <u>235.5 ft</u>



### RISING-2

Data Set: C:\...\15d rising2.aqt  
 Date: 04/16/12

Time: 11:55:15

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

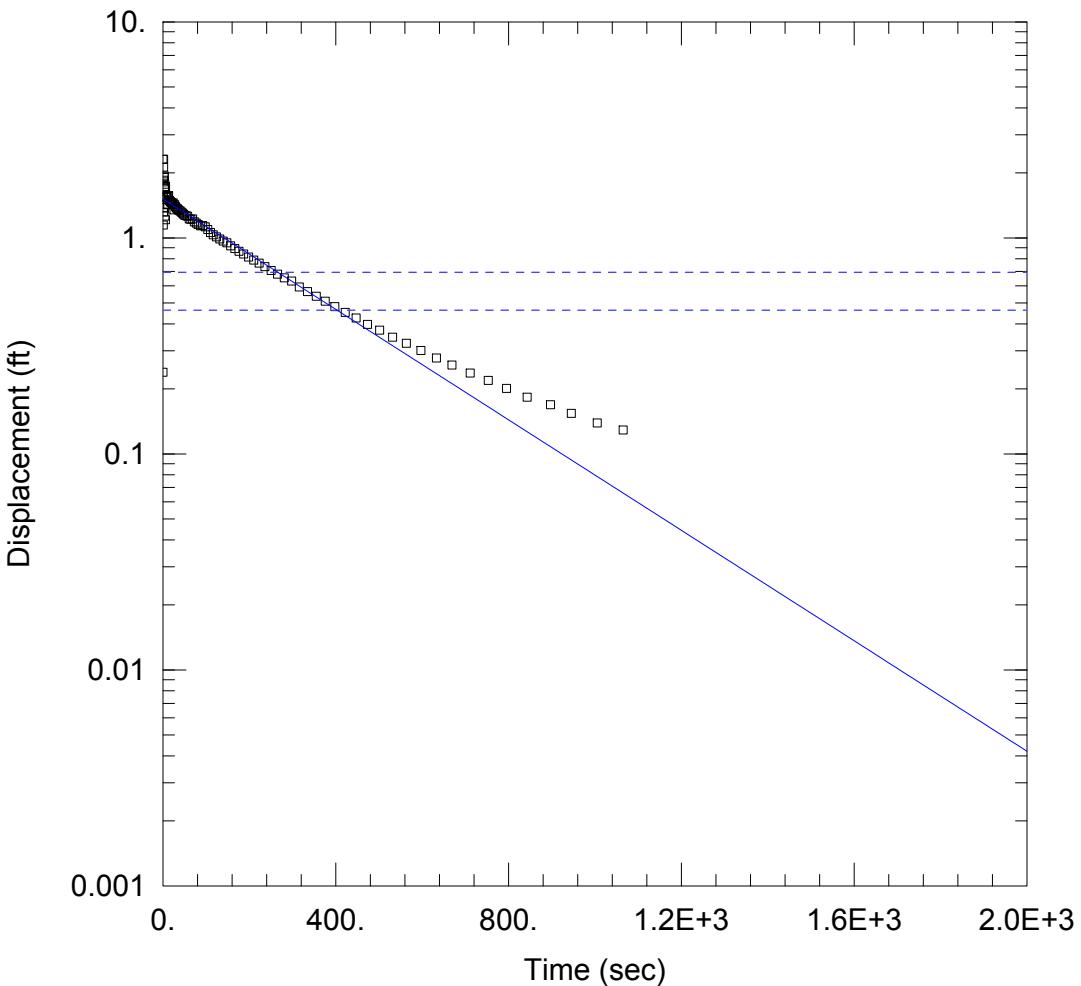
Saturated Thickness: 62.85 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-16s)

Initial Displacement: <u>0.867 ft</u>	Static Water Column Height: <u>62.85 ft</u>
Total Well Penetration Depth: <u>62.85 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Springer-Gelhar</u>
K = <u>37.14 ft/day</u>	Le = <u>0.1 ft</u>



### FALLING

Data Set: C:\...\16s falling.aqt  
 Date: 04/16/12

Time: 11:56:31

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

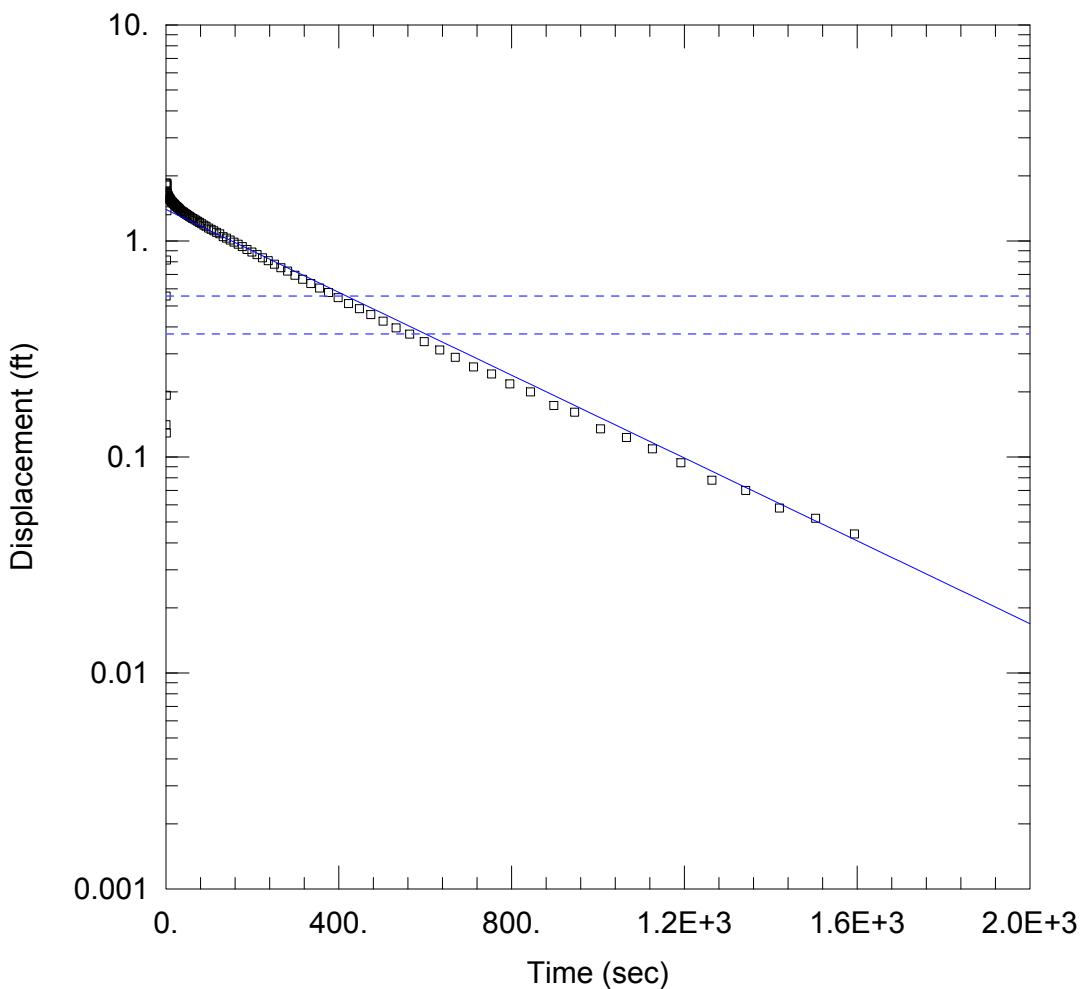
Saturated Thickness: 60.18 ft Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-16s)

Initial Displacement: <u>2.313 ft</u>	Static Water Column Height: <u>25.18 ft</u>
Total Well Penetration Depth: <u>25.18 ft</u>	Screen Length: <u>10. ft</u>
Casing Radius: <u>0.0833 ft</u>	Well Radius: <u>0.33 ft</u>

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.2228 ft/day</u>	y0 = <u>1.516 ft</u>



### RISING

Data Set: C:\...\16s rising.aqt  
 Date: 04/16/12

Time: 11:57:33

### PROJECT INFORMATION

Company: CDM  
 Client: NYSDEC  
 Project: Site 2-43-018  
 Location: Paul Miller Site  
 Test Well: MW-11d  
 Test Date: 3/28/07

### AQUIFER DATA

Saturated Thickness: 60.18 ft      Anisotropy Ratio (Kz/Kr): 1.

### WELL DATA (MW-16s)

Initial Displacement: <u>1.853</u> ft	Static Water Column Height: <u>25.18</u> ft
Total Well Penetration Depth: <u>25.18</u> ft	Screen Length: <u>10.</u> ft
Casing Radius: <u>0.0833</u> ft	Well Radius: <u>0.33</u> ft

### SOLUTION

Aquifer Model: <u>Unconfined</u>	Solution Method: <u>Bouwer-Rice</u>
K = <u>0.167</u> ft/day	y0 = <u>1.397</u> ft

Slug Test Results Summary  
Former Paul Miller Dry Cleaners  
Port Richmond, NY

Location	Test	Stickup feet	Total Well Depth (bottom of screen) feet bgs	Well Depth feet bgs	Depth to Water feet TIC	Aquifer Base feet bgs	Aquifer Thickness Feet	Top of Screen feet bgs	Depth to top of Screen from water level Feet	Screen Length Feet	Effective Screen Length Feet	K ft/day	Method	Remarks	Strata Notes
MW-8s	FH	0	35	35	7.49	70	62.51	25	17.51	10	10	0.62	Bouwer & Rice		
MW-8s	FH - manual fit	0	35	35	7.49	70	62.51	25	17.51	10	10	0.70	Bouwer & Rice	Preferred fit	
MW-8s	RH	0	35	35	7.49	70	62.51	25	17.51	10	10	0.54	Bouwer & Rice	Preferred fit	
MW-9s	FH	0	35	35	4.76	70	65.24	25	20.24	10	10	0.43	Bouwer & Rice	Preferred fit	
MW-9s	RH	0	35	35	4.76	70	65.24	25	20.24	10	10	0.63	Bouwer & Rice		
MW-9S	RH - manual fit	0	35	35	4.76	70	65.24	25	20.24	10	10	0.46	Bouwer & Rice		
MW-11d	FH	0	70	70	6.42	70	63.58	60	53.58	10	10	3.1	Bouwer & Rice	Preferred fit	
MW-11d	RH	0	70	70	6.42	70	63.58	60	53.58	10	10	4.7	Bouwer & Rice		
MW-11d	RH - manual fit	0	70	70	6.42	70	63.58	60	53.58	10	10	6.66	Bouwer & Rice	Preferred fit	
MW-12s	FH	0	35	35	6.63	70	63.37	25	18.37	10	10	3.6	Bouwer & Rice	Preferred fit	
MW-12s	RH	0	35	35	6.63	70	63.37	25	18.37	10	10	2.3	Bouwer & Rice		
MW-12s	RH - manual fit	0	35	35	6.63	70	63.37	25	18.37	10	10	3.8	Bouwer & Rice	Preferred fit	
MW-13s	FH	0	35	35	8.89	70	61.11	25	16.11	10	10	4.9	Bouwer & Rice	Preferred fit	
MW-13s	RH	0	35	35	8.89	70	61.11	25	16.11	10	10	2.6	Bouwer & Rice		
MW-13s	RH - manual fit	0	35	35	8.89	70	61.11	25	16.11	10	10	4.1	Bouwer & Rice	Preferred fit	
MW-14s	FH	0	34	34	7.75	70	62.25	24	16.25	10	10	0.07	Bouwer & Rice		
MW-14s	FH - manual	0	34	34	7.75	70	62.25	24	16.25	10	10	0.05	Bouwer & Rice	Preferred fit	
MW-14s	RH	0	34	34	7.75	70	62.25	24	16.25	10	10	0.07	Bouwer & Rice		
MW-14s	RH - manual fit	0	34	34	7.75	70	62.25	24	16.25	10	10	0.05	Bouwer & Rice	Preferred fit	
MW-15d	FH	0	70	70	7.15	70	62.85	60	52.85	10	10	69.0	Springer-Gelhar	Preferred Test	
MW-15d	RH	0	70	70	7.15	70	62.85	60	52.85	10	10	59.5	Springer-Gelhar	Preferred Test	
MW-15d	FH2	0	70	70	7.15	70	62.85	60	52.85	10	10	38.7	Springer-Gelhar	Slow insertion	
MW-15d	RH2	0	70	70	7.15	70	62.85	60	52.85	10	10	37.1	Springer-Gelhar	Slow insertion	
MW-16s	FH	0	35	35	9.82	70	60.18	25	15.18	10	10	0.22	Bouwer & Rice	Preferred fit	
MW-16s	RH	0	35	35	9.82	70	60.18	25	15.18	10	10	0.17	Bouwer & Rice	Preferred fit	

TIC - measurement from top of inner casing

bgs - below ground surface

RH - rising head

FH - falling head

## Appendix G

### Indoor Air Analytical Results

# Galson Laboratories Data Package

Client : Camp, Dresser & McKee, Inc.  
Project : NYDEC  
SDG : L235614

Data package for samples received:  
03/14/11





Ms. Cristina Ramacciotti  
Camp, Dresser & McKee, Inc.  
110 Fieldcrest Avenue  
6th Floor  
Edison, NJ 08837

April 04, 2011

DOH ELAP# 11626

Account# 14621

Login# L235614

Dear Ms. Ramacciotti:

Enclosed are the analytical results for the samples received by our laboratory on March 14, 2011. All test results meet the quality control requirements of AIHA and NELAC unless otherwise stated in this report. All samples on the chain of custody were received in good condition unless otherwise noted.

Results in this report are based on the sampling data provided by the client and refer only to the samples as they were received at the laboratory. Unless otherwise requested, all samples will be discarded 14 days from the date of this report.

Please contact Tonya Lancaster at (877) 482-5227, if you would like any additional information regarding this report.

Thank you for using Galson Laboratories.

Sincerely,

**Galson Laboratories**

A handwritten signature in black ink that reads "Mary G. Unangst". The signature is fluid and cursive, with "Mary" and "G." being more stylized and "Unangst" having a more formal, printed-like appearance.

Mary G. Unangst  
Laboratory Director

Enclosure(s)



## LABORATORY ANALYSIS REPORT

6601 Kirkville Road Client : Camp, Dresser & McKee, Inc.  
East Syracuse, NY 13057 Site : Paul Miller  
(315) 432-5227 Project No. : NYDEC  
FAX: (315) 437-0571 Date Sampled : 11-MAR-11 Account No.: 14621  
www.galsonlabs.com Date Received : 14-MAR-11 Login No. : L235614  
Date Analyzed : 18-MAR-11  
Report ID : 684937

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**Perchloroethylene**

<u>Sample ID</u>	<u>Lab ID</u>	<u>Time</u> <u>minutes</u>	<u>Front</u> <u>ug</u>	<u>Back</u> <u>ug</u>	<u>Total</u> <u>ug</u>	<u>Conc</u> <u>ug/m3</u>
PMBANK1	L235614-1	1440	<0.06	<0.06	<0.06	<1
PMBANKDUP	L235614-2	1440	<0.06	<0.06	<0.06	<1
PMAMBAIR	L235614-3	1440	<0.06	<0.06	<0.06	<1

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

---

Level of quantitation: 0.06 ug  
Analytical Method : mod. NYS DOH 311-9  
OSHA PEL (TWA) : 100 ppm  
Collection Media : M3M-3520

Submitted by: mln  
Approved by : nkp  
Date : 21-MAR-11 NYS DOH # : 11626  
QC by: Tony D'Amico

---

< -Less Than	mg -Milligrams	m3 -Cubic Meters	kg -Kilograms
> -Greater Than	ug -Micrograms	l -Liters	NS -Not Specified
NA -Not Applicable	ND -Not Detected	ppm -Parts per Million	



## LABORATORY FOOTNOTE REPORT

6601 Kirkville Road  
East Syracuse, NY 13057  
(315) 432-5227  
FAX: (315) 437-0571  
[www.galsonlabs.com](http://www.galsonlabs.com)

Client Name : Camp, Dresser & McKee, Inc.  
Site : Paul Miller  
Project No. : NYDEC

Date Sampled : 11-MAR-11 Account No.: 14621  
Date Received: 14-MAR-11 Login No. : L235614  
Date Analyzed: 18-MAR-11

---

Unless otherwise noted below, all quality control results associated with the samples were within established control limits.

Unrounded results are carried through the calculations that yield the final result and the final result is rounded to the number of significant figures appropriate to the accuracy of the analytical method. Please note that results appearing in the columns preceding the final result column may have been rounded in order to fit the report format and therefore, if carried through the calculations, may not yield an identical final result to the one reported.

The stated LOQs for each analyte represent the demonstrated LOQ concentrations prior to correction for desorption efficiency (if applicable).

L235614 (Report ID: 684937):  
Total ug corrected for a desorption efficiency of 103%.  
SOPs: GC-SOP-12(3), GC-SOP-16(5), GC-SOP-9(4)

---

< -Less Than mg -Milligrams m³ -Cubic Meters kg -Kilograms  
> -Greater Than ug -Micrograms l -Liters NS -Not Specified  
NA -Not Applicable ND -Not Detected ppm -Parts per Million

---



**INVOICE**

**Galson Laboratories**

Login No : L235614  
Project : STANDARD  
PO No : 089780230TASK3A  
Site : Paul Miller

Invoice No : 284524  
Invoice Date : 04-APR-11  
Account No : 14621  
Client Project : NYDEC

Bill To : Ms. Cristina Ramacciotti  
Camp, Dresser & McKee, Inc.  
110 Fieldcrest Avenue  
6th Floor  
Edison NJ 08837

Report To : Ms. Cristina Ramacciotti  
TERMS: NET 30 DAYS  
Finance charges will be applied based on the terms and conditions of sale.

<b>Sample #</b>	<b>Client ID</b>	<b>Media</b>	<b>Analysis</b>	<b>Unit Price</b>
L235614-1	PMBANK1	M3M-3520	IG-PERC311-9/Perchloroethylene	\$ 65.00
L235614-1	PMBANK1	OPTIONS	IG-DELIVERABLES/Deliverables	\$ 0.00
L235614-2	PMBANKDUP	M3M-3520	IG-PERC311-9/Perchloroethylene	\$ 65.00
L235614-3	PMAMBAIR	M3M-3520	IG-PERC311-9/Perchloroethylene	\$ 65.00
			Subtotal :	\$ 195.00
			20 % Deliverables :	+ \$ 39.00
			Sampling Media from Order# 196013 :	+ \$ 20.00
			Amount Due --->	\$ 254.00

---

Please Remit To: Galson Laboratories, P.O. Box 8000, Dept 684, Buffalo, NY 14267  
Phone: 888-432-5227

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## **Internal Chain of Custody**

## GALSON LABORATORIES INTERNAL CHAIN OF CUSTODY

PAGE 1 of 1

Lab ID.	Client ID	Matrix	Storage	Sequence No.	Date/Time	Action	Tech. Name	Department		
L235614		Camp, Dresser & McKee, Inc.		14621	14-MAR-11					
L235614-1	PMBANK1	M3M-3520	IG-Low Level Frz							
				1	03/14/2011 12:28	Check In	CMARCHETTI	Login		
				2	03/16/2011 04:54	Check Out	RROGERS	GC Prep		
L235614-1	PMBANK1	OPTIONS	IG-Freezer							
				1	03/14/2011 12:28	Check In	CMARCHETTI	Login		
				2	03/16/2011 04:54	Check Out	RROGERS	GC Prep		
L235614-2	PMBANKDUP	M3M-3520	IG-Low Level Frz							
				1	03/14/2011 12:28	Check In	CMARCHETTI	Login		
				2	03/16/2011 04:54	Check Out	RROGERS	GC Prep		
L235614-3	PMAMBAIR	M3M-3520	IG-Low Level Frz							
				1	03/14/2011 12:28	Check In	CMARCHETTI	Login		
				2	03/16/2011 04:54	Check Out	RROGERS	GC Prep		

## **GC ANALYSIS**

Camp, Dresser & McKee, Inc.

Package: L235614

Project: NYDEC

Galson Laboratories received 3 samples for Perchloroethylene analysis on March 14, 2011. The samples were assigned to Galson Login: L235614. Samples were analyzed by Gas Chromatography (GC) following method(s):

Perchloroethylene	mod. NYS DOH 311-9
-------------------	--------------------

Galson SOP(s): GC-SOP-12, GC-SOP-16, and GC-SOP-9.

Sample ID	Date Sampled	Date Analyzed
L235614-1 (PMBANK1)	March 11, 2011	March 18, 2011
L235614-2 (PMBANKDUP)	March 11, 2011	March 18, 2011
L235614-3 (PMAMBAIR)	March 11, 2011	March 18, 2011

The samples were prepared on March 16, 2011 and were associated with analytical workgroup WG194433 and preparation workgroup WG194153. All Continuing Calibration Verification (CCV) standards, the Detection Limit Standard (DLS), the Blank Spike (BS), and the Blank Spike Duplicate (BSD) were within the established percent recovery control limits.

The samples were analyzed on instrument HP23. The initial calibration for HP23 was performed on Mar. 18, 2011 and the range was approximately from 0.03 to 2.0ug/mL.

#### Quality Control Summary

- The continuing calibration check standards consist of the CCV, which is a standard prepared from a separate source than the initial calibration. The control limits are statistically determined and are listed on the associated CCV recovery forms.
- The detection limit standard, or DLS, is a standard prepared at the concentration that is equivalent to the LOQ. The control limits are 70 to 130%.
- Blanks consist of the eluent/diluent blank and media blanks obtained from the laboratory supply.
- Blank spikes are prepared in-house; the media is spiked with the analyte(s) of interest, then prepared and analyzed with the associated samples. The blank spike control limits are statistically determined and are listed on the associated BS/BSD recovery forms.
- The CCV, DLS, and BS/BSD recoveries are reported on their respective Recovery Report Forms.

**Title:** Created from worklist V:\WG19443320110318112229.wle  
**Datasource:** CHROMDB  
**Location:** HP23\Sequences\201103  
**Timebase:** HP23  
**# Samples:** 17

**Created:** 03/18/2011  
**Last Update:** 03/18/2011

No.	Name:	Comment	Samp Type	Pos.	Inj. Date/Time	inj. Vol.	Dilution	DE Vol	Lot Corr	Program	Method
1	CS2	CS2	IBLANK		1 03/18/2011 11:26	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
2	CS2	CS2	IBLANK		1 03/18/2011 11:39	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
3	WG194433-1	WG194433-1,IH196262-3,1	DLS		8 03/18/2011 11:52	2	1	1	0	HP23_PERC_MI	HP23_20110318_PEI
4	WG194433-2	WG194433-2,IH196262-2,1	CCV		9 03/18/2011 12:05	2	1	1	0	HP23_PERC_MI	HP23_20110318_PEI
5	WG194153-2	WG194153-2,WG194433,STD2BADGE	EBLANK		10 03/18/2011 12:18	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
6	WG194153-3A	WG194153-3A,WG194433,STD2BADGE	MBLANK		11 03/18/2011 12:30	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
7	WG194153-3B	WG194153-3B,WG194433,STD2BADGE	MBLANK		12 03/18/2011 12:43	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
8	WG194153-4	WG194153-4,IH196262-1,1,STD2BADGE	BS		13 03/18/2011 12:56	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
9	WG194153-5	WG194153-5,IH196262-1,1,STD2BADGE	BSD		14 03/18/2011 13:09	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
10	CS2	CS2	IBLANK		1 03/18/2011 13:22	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
11	L235614-1A	L235614-1A,WG194433,STD2BADGE	SAMP		15 03/18/2011 13:34	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
12	L235614-1B	L235614-1B,WG194433,STD2BADGE	SAMP		16 03/18/2011 13:47	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
13	L235614-2A	L235614-2A,WG194433,STD2BADGE	SAMP		17 03/18/2011 14:00	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
14	L235614-2B	L235614-2B,WG194433,STD2BADGE	SAMP		18 03/18/2011 14:13	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
15	L235614-3A	L235614-3A,WG194433,STD2BADGE	SAMP		19 03/18/2011 14:26	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
16	L235614-3B	L235614-3B,WG194433,STD2BADGE	SAMP		20 03/18/2011 14:39	2	1	2	0	HP23_PERC_MI	HP23_20110318_PEI
17	WG194433-3	WG194433-3,IH196262-2,1	CCV		21 03/18/2011 14:51	2	1	1	0	HP23_PERC_MI	HP23_20110318_PEI

## INITIAL/CONTINUING CALIBRATION REPORT

Client : Camp, Dresser & McKee, Inc.  
 Account No: 14621  
 Login No. : L235614

Lab Sample ID Type Spike Lot # Instrument Analysis Date	Limits (%)	WG194433-2			WG194433-3					
		True Value (ug)	Found (ug)	Recovery (%)	True Value (ug)	Found (ug)	Recovery (%)	True Value ( ))	Found ( ))	Recovery (%)
TETRACHLOROETHYLENE	80.0-120.	0.987	1.03	104.	0.987	0.987	99.9			

## DETECTION LIMIT STANDARD RECOVERY REPORT

Client : Camp, Dresser & McKee, Inc.  
 Account No: 14621  
 Login No. : L235614

Lab Sample ID Type Spike Lot # Instrument Analysis Date	Limits (%)	WG194433-1 DLS IH196262-3 HP23 Mar 18, 2011 11:52								
		True Value (ug)	Found (ug)	Recovery (%)	True Value ( ))	Found ( ))	Recovery (%)	True Value ( ))	Found ( ))	Recovery (%)
TETRACHLOROETHYLENE	70.0-130.	0.0300	0.0317	106.						

METHOD BLANK REPORT

**Client** Camp, Dresser & McKee, Inc.  
**Account No.** 14621  
**Login No.** L235614

Lab Sample ID	Type	Instrument	Analysis Date	Analysis Time	LOQ (ug)	Found (ug)					
TETRACHLOROETHYLENE		WG194153-3 MBLANK HP23 03/18/11 12:30			0.06	<0.06					

Client Name: Camp, Dresser & McKee, Inc.  
Client Acct: 14621

Login: L235614

QC Sample	Type	Analysis Date	Instrument	Media
WG194153-4	BS	Mar 18, 2011 12:56	HP23	M3M-3520
WG194153-5	BSD	Mar 18, 2011 13:09	HP23	M3M-3520

Parameter	True Value ug	WG194153-4			WG194153-5			QC Limits	
		BS ug	% Rec #	BSD ug	% Rec #	RPD #	Rec	RPD	
TETRACHLOROETHYLENE	1.97	1.93	97.6	1.95	98.9	1.3	75.0-125	20.0	

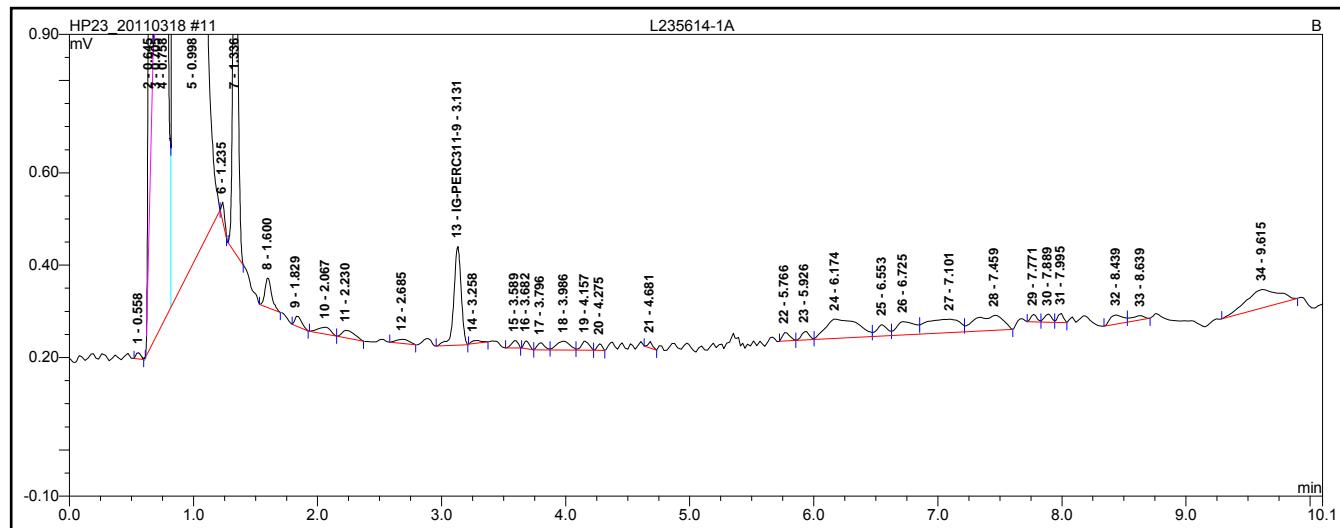
# Column to be used to flag recovery and RPD values with an asterisk.  
\* Values outside of QC limits.

COMMENTS: \_\_\_\_\_

## Raw Data

**L235614-1A****L235614-1A,WG194433,STD2BADGE**

Sample Name:	L235614-1A	Injection Volume:	2.0
Vial Number:	15	Channel:	B
Sample Type:	SAMP	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 13:34		
Run Time (min):	10.10		

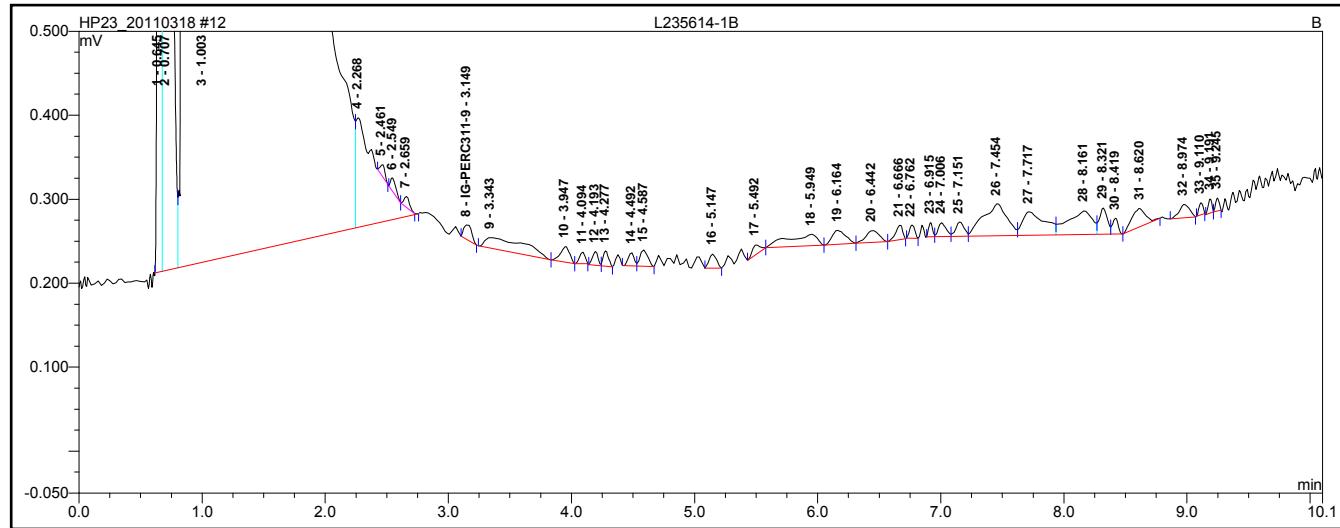


No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.56	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
2	n.a.	0.65	0.021	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
3	n.a.	0.71	0.009	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
4	n.a.	0.76	0.238	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
5	n.a.	1.00	45.251	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
6	n.a.	1.24	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
7	n.a.	1.34	0.039	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
8	n.a.	1.60	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
9	n.a.	1.83	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
10	n.a.	2.07	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
11	n.a.	2.23	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
12	n.a.	2.69	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
13	IG-PERC311-9	3.13	0.015	XXLOff	3.546E-1	2.801E-2	2.820E+0	-0.0047	1	2.0	-0.0094
14	n.a.	3.26	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
15	n.a.	3.59	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
16	n.a.	3.68	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
17	n.a.	3.80	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
18	n.a.	3.99	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
19	n.a.	4.16	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
20	n.a.	4.28	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
21	n.a.	4.68	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
22	n.a.	5.77	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
23	n.a.	5.93	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
24	n.a.	6.17	0.013	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
25	n.a.	6.55	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
26	n.a.	6.73	0.005	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
27	n.a.	7.10	0.009	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
28	n.a.	7.46	0.009	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
29	n.a.	7.77	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
30	n.a.	7.89	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
31	n.a.	8.00	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
32	n.a.	8.44	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
33	n.a.	8.64	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
34	n.a.	9.61	0.012	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
<b>Total:</b>			45.653								

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		0.015	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	2	n.a.

**L235614-1B****L235614-1B,WG194433,STD2BADGE**

Sample Name:	L235614-1B	Injection Volume:	2.0
Vial Number:	16	Channel:	B
Sample Type:	SAMP	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 13:47		
Run Time (min):	10.10		

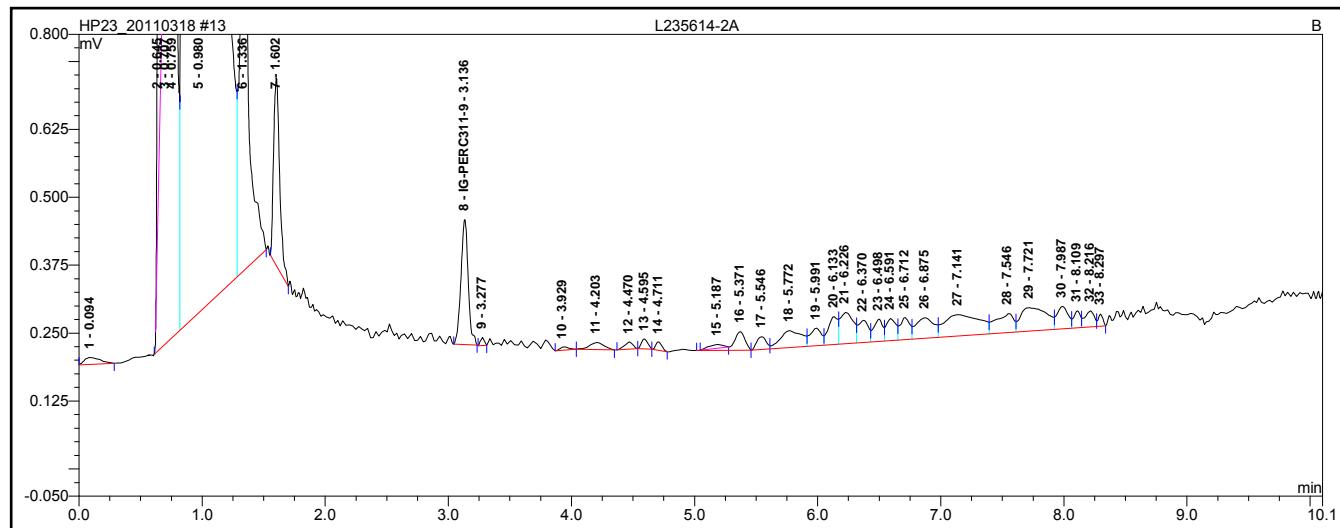


No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.65	0.031	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
2	n.a.	0.71	0.075	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
3	n.a.	1.00	50.304	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
4	n.a.	2.27	0.026	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
5	n.a.	2.46	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
6	n.a.	2.55	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
7	n.a.	2.66	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
8	1G-PERC311-9	3.15	0.001	XXLOff	3.546E-1	2.801E-2	2.820E+0	-0.0095	1	2.0	-0.0189
9	n.a.	3.34	0.006	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
10	n.a.	3.95	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
11	n.a.	4.09	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
12	n.a.	4.19	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
13	n.a.	4.28	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
14	n.a.	4.49	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
15	n.a.	4.59	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
16	n.a.	5.15	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
17	n.a.	5.49	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
18	n.a.	5.95	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
19	n.a.	6.16	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
20	n.a.	6.44	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
21	n.a.	6.67	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
22	n.a.	6.76	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
23	n.a.	6.92	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
24	n.a.	7.01	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
25	n.a.	7.15	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
26	n.a.	7.45	0.009	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
27	n.a.	7.72	0.006	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
28	n.a.	8.16	0.006	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
29	n.a.	8.32	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
30	n.a.	8.42	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
31	n.a.	8.62	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
32	n.a.	8.97	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
33	n.a.	9.11	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
34	n.a.	9.19	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
35	n.a.	9.24	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
<b>Total:</b>			<b>50.497</b>								

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		0.001	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	2 n.a.	

**L235614-2A****L235614-2A,WG194433,STD2BADGE**

Sample Name:	L235614-2A	Injection Volume:	2.0
Vial Number:	17	Channel:	B
Sample Type:	SAMP	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 14:00		
Run Time (min):	10.10		

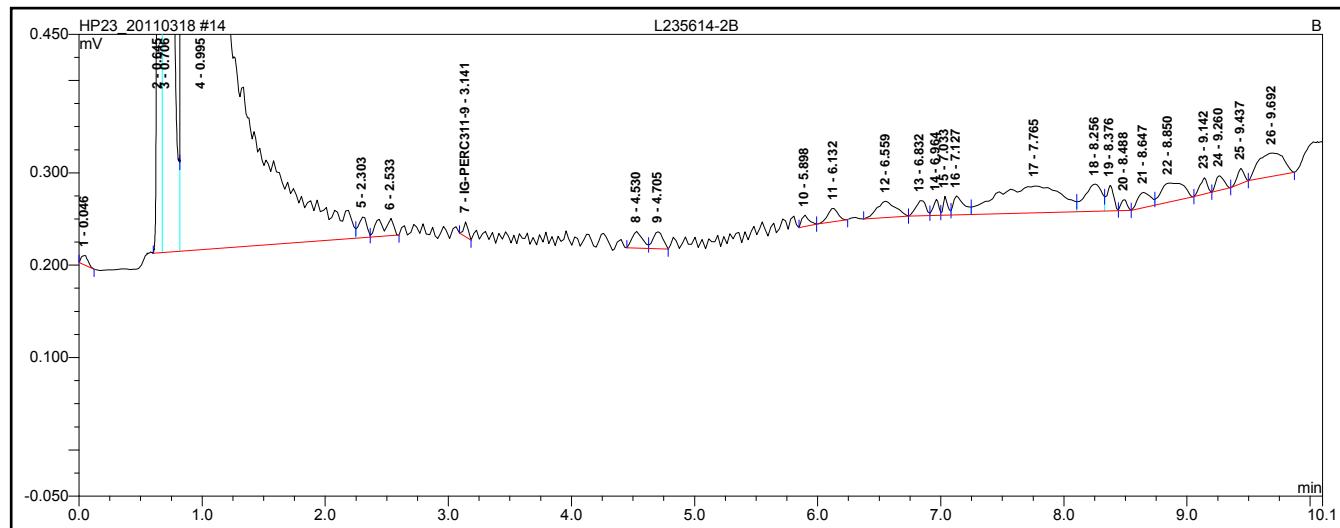


No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.09	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
2	n.a.	0.64	0.019	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
3	n.a.	0.71	0.011	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
4	n.a.	0.76	0.249	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
5	n.a.	0.98	45.714	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
6	n.a.	1.34	0.076	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
7	n.a.	1.60	0.020	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
8	IG-PERC311-9	3.14	0.016	XXLOff	3.546E-1	2.801E-2	2.820E+0	-0.0044	1	2.0	-0.0087
9	n.a.	3.28	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
10	n.a.	3.93	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
11	n.a.	4.20	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
12	n.a.	4.47	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
13	n.a.	4.59	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
14	n.a.	4.71	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
15	n.a.	5.19	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
16	n.a.	5.37	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
17	n.a.	5.55	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
18	n.a.	5.77	0.006	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
19	n.a.	5.99	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
20	n.a.	6.13	0.005	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
21	n.a.	6.23	0.007	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
22	n.a.	6.37	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
23	n.a.	6.50	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
24	n.a.	6.59	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
25	n.a.	6.71	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
26	n.a.	6.88	0.006	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
27	n.a.	7.14	0.013	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
28	n.a.	7.55	0.006	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
29	n.a.	7.72	0.011	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
30	n.a.	7.99	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
31	n.a.	8.11	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
32	n.a.	8.22	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
33	n.a.	8.30	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
<b>Total:</b>			<b>46.201</b>								

Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
	0.016	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	2 n.a.	

**L235614-2B****L235614-2B,WG194433,STD2BADGE**

Sample Name:	L235614-2B	Injection Volume:	2.0
Vial Number:	18	Channel:	B
Sample Type:	SAMP	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 14:13		
Run Time (min):	10.10		

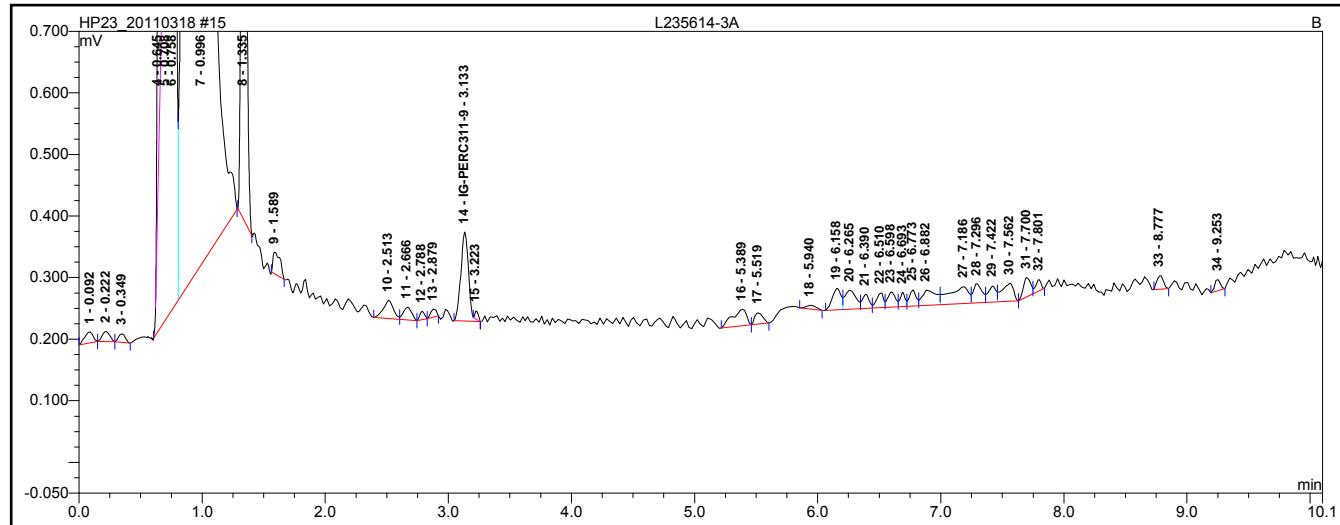


No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.05	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
2	n.a.	0.65	0.035	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
3	n.a.	0.71	0.081	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
4	n.a.	0.99	45.325	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
5	n.a.	2.30	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
6	n.a.	2.53	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
7	IG-PERC311-9	3.14	0.001	XXLOff	3.546E-1	2.801E-2	2.820E+0	-0.0097	1	2.0	-0.0194
8	n.a.	4.53	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
9	n.a.	4.71	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
10	n.a.	5.90	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
11	n.a.	6.13	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
12	n.a.	6.56	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
13	n.a.	6.83	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
14	n.a.	6.96	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
15	n.a.	7.03	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
16	n.a.	7.13	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
17	n.a.	7.76	0.018	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
18	n.a.	8.26	0.005	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
19	n.a.	8.38	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
20	n.a.	8.49	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
21	n.a.	8.65	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
22	n.a.	8.85	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
23	n.a.	9.14	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
24	n.a.	9.26	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
25	n.a.	9.44	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
26	n.a.	9.69	0.006	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
<b>Total:</b>		45.501									

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		0.001	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	2	n.a.

**L235614-3A****L235614-3A,WG194433,STD2BADGE**

Sample Name:	L235614-3A	Injection Volume:	2.0
Vial Number:	19	Channel:	B
Sample Type:	SAMP	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 14:26		
Run Time (min):	10.10		



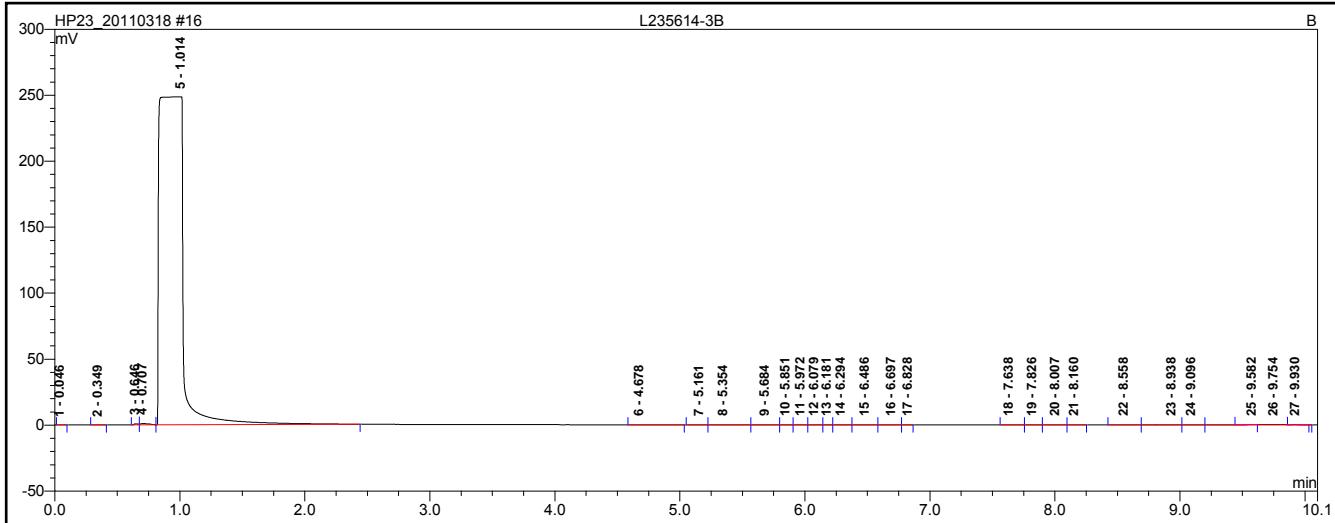
No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.09	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
2	n.a.	0.22	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
3	n.a.	0.35	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
4	n.a.	0.65	0.013	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
5	n.a.	0.71	0.011	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
6	n.a.	0.76	0.174	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
7	n.a.	1.00	45.266	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
8	n.a.	1.33	0.049	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
9	n.a.	1.59	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
10	n.a.	2.51	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
11	n.a.	2.67	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
12	n.a.	2.79	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
13	n.a.	2.88	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
14	IG-PERC311-9	3.13	0.010	XXLOff	3.546E-1	2.801E-2	2.820E+0	-0.0062	1	2.0	-0.0124
15	n.a.	3.22	0.000	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
16	n.a.	5.39	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
17	n.a.	5.52	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
18	n.a.	5.94	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
19	n.a.	6.16	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
20	n.a.	6.27	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
21	n.a.	6.39	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
22	n.a.	6.51	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
23	n.a.	6.60	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
24	n.a.	6.69	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
25	n.a.	6.77	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
26	n.a.	6.88	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
27	n.a.	7.19	0.005	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
28	n.a.	7.30	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
29	n.a.	7.42	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
30	n.a.	7.56	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
31	n.a.	7.70	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
32	n.a.	7.80	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
33	n.a.	8.78	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
34	n.a.	9.25	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.

Total: 45.576

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		0.010	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	2 n.a.	

**L235614-3B****L235614-3B,WG194433,STD2BADGE**

Sample Name:	L235614-3B	Injection Volume:	2.0
Vial Number:	20	Channel:	B
Sample Type:	SAMP	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 14:39		
Run Time (min):	10.10		



No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.05	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
2	n.a.	0.35	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
3	n.a.	0.65	0.030	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
4	n.a.	0.71	0.080	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
5	n.a.	1.01	53.036	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
6	n.a.	4.68	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
7	n.a.	5.16	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
8	n.a.	5.35	0.005	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
9	n.a.	5.68	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
10	n.a.	5.85	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
11	n.a.	5.97	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
12	n.a.	6.08	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
13	n.a.	6.18	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
14	n.a.	6.29	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
15	n.a.	6.49	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
16	n.a.	6.70	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
17	n.a.	6.83	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
18	n.a.	7.64	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
19	n.a.	7.83	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
20	n.a.	8.01	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
21	n.a.	8.16	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
22	n.a.	8.56	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
23	n.a.	8.94	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
24	n.a.	9.10	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
25	n.a.	9.58	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
26	n.a.	9.75	0.022	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
27	n.a.	9.93	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
<b>Total:</b>			<b>53.212</b>								

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug

Continuing Calibration Standards

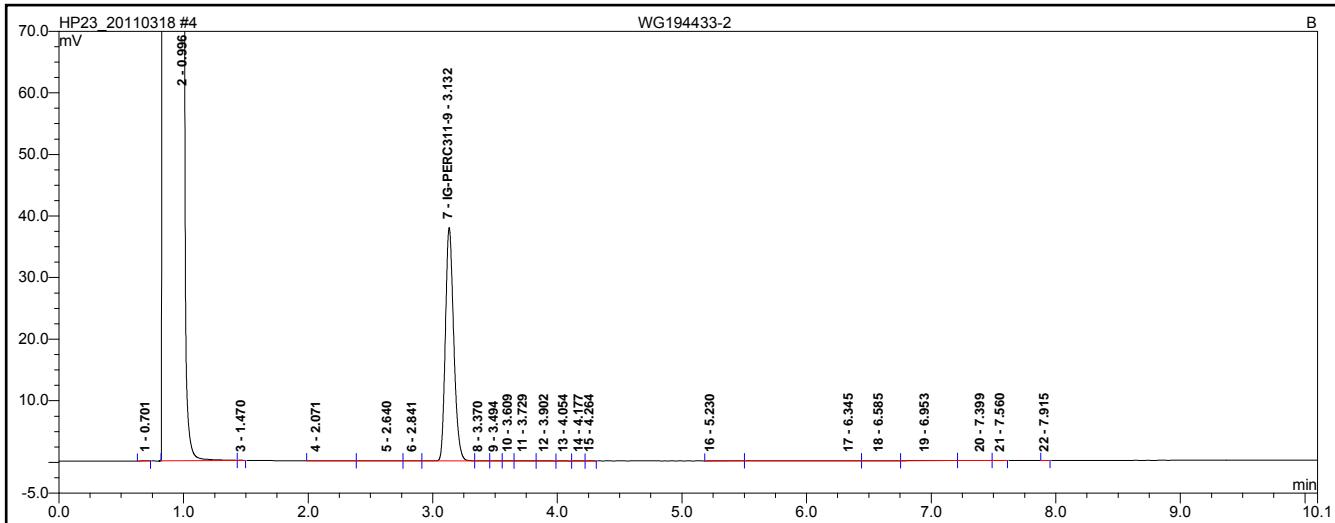
and

Continuing Blank Data

WG194433-2

WG194433-2,IH196262-2,1

<b>Sample Name:</b>	<b>WG194433-2</b>	<b>Injection Volume:</b>	<b>2.0</b>
<b>Vial Number:</b>	<b>9</b>	<b>Channel:</b>	<b>B</b>
<b>Sample Type:</b>	<b>CCV</b>	<b>Lot Correction:</b>	<b>0 ug</b>
<b>Control Program:</b>	<b>HP23_PERC_METHOD_A</b>	<b>Standard Method:</b>	<b>External</b>
<b>Quantif. Method:</b>	<b>HP23_20110318_PERC311-9</b>		
<b>Recording Time:</b>	<b>3/18/2011 12:05</b>		
<b>Run Time (min):</b>	<b>10.10</b>		

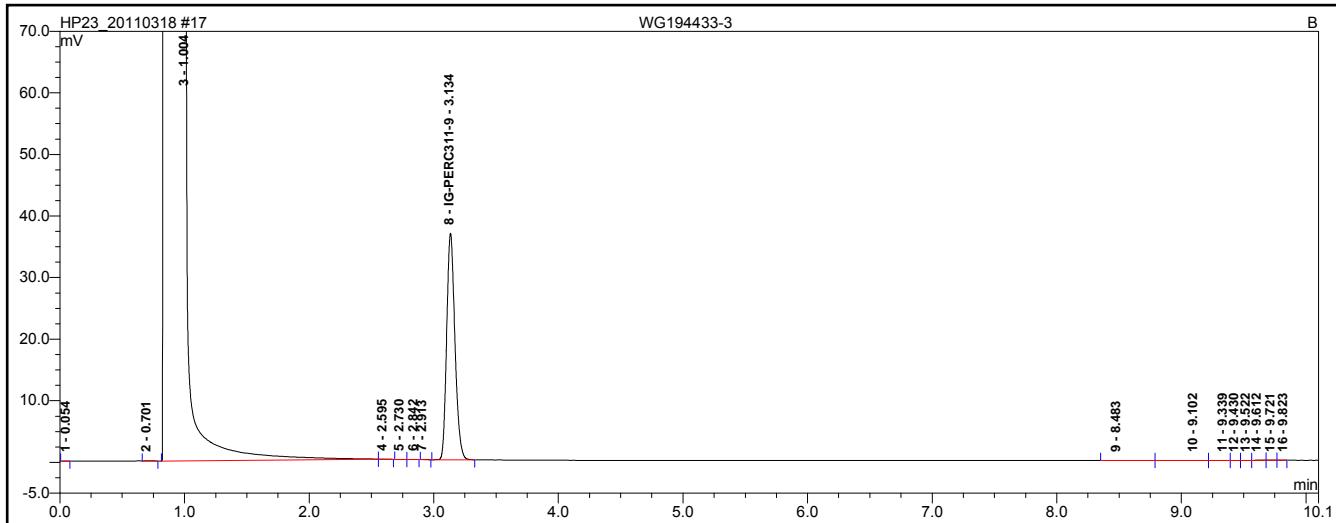


**Total:** 48.373

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		2.921	XXOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	1	n.a.

**WG194433-3****WG194433-3,IH196262-2,1**

Sample Name:	WG194433-3	Injection Volume:	2.0
Vial Number:	21	Channel:	B
Sample Type:	CCV	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 14:51		
Run Time (min):	10.10		



No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.05	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
2	n.a.	0.70	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
3	n.a.	1.00	48.393	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
4	n.a.	2.60	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
5	n.a.	2.73	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
6	n.a.	2.84	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
7	n.a.	2.91	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
8	IG-PERC311-9	3.13	2.811	XXLOff	3.546E-1	2.801E-2	2.820E+0	0.9868	1	1.0	0.9868
9	n.a.	8.48	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
10	n.a.	9.10	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
11	n.a.	9.34	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
12	n.a.	9.43	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
13	n.a.	9.52	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
14	n.a.	9.61	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
15	n.a.	9.72	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
16	n.a.	9.82	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.

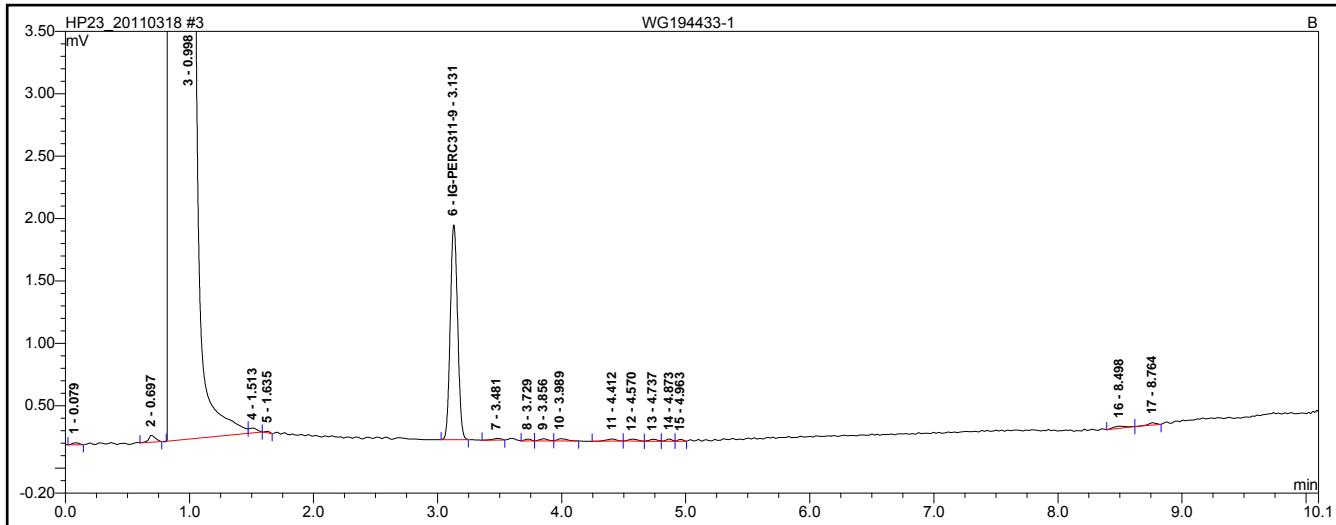
Total: 51.222

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		2.811	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	1	n.a.

**WG194433-1**

WG194433-1,IH196262-3,1

Sample Name:	WG194433-1	Injection Volume:	2.0
Vial Number:	8	Channel:	B
Sample Type:	DLS	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 11:52		
Run Time (min):	10.10		

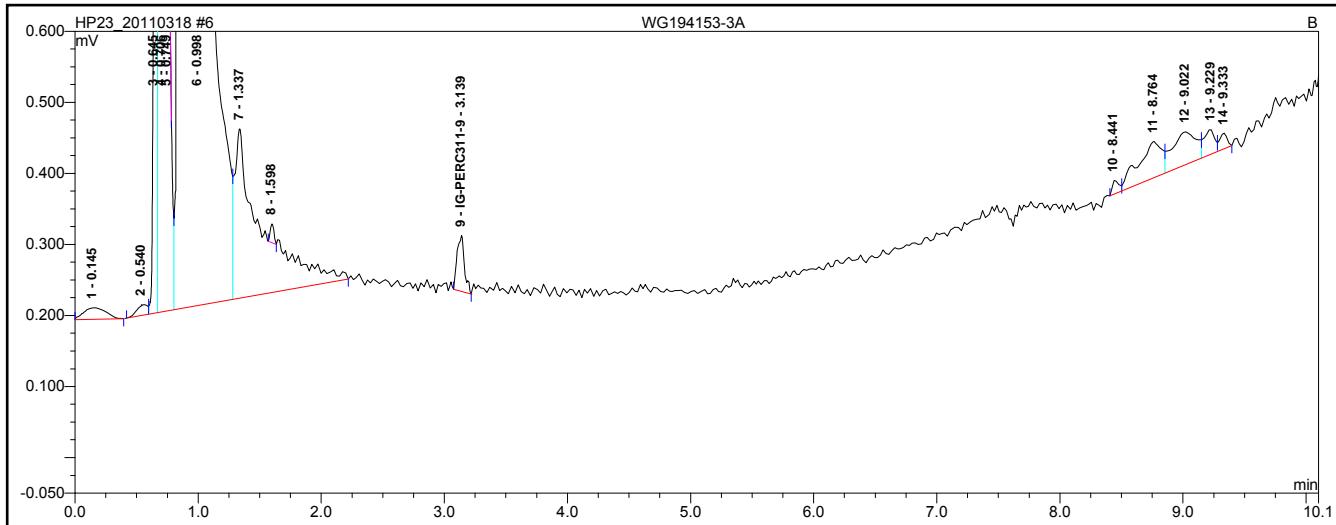


No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.08	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
2	n.a.	0.70	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
3	n.a.	1.00	45.472	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
4	n.a.	1.51	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
5	n.a.	1.63	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
6	IG-PERC311-9	3.13	0.117	XXLOff	3.546E-1	2.801E-2	2.820E+0	0.0317	1	1.0	0.0317
7	n.a.	3.48	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
8	n.a.	3.73	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
9	n.a.	3.86	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
10	n.a.	3.99	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
11	n.a.	4.41	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
12	n.a.	4.57	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
13	n.a.	4.74	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
14	n.a.	4.87	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
15	n.a.	4.96	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
16	n.a.	8.50	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
17	n.a.	8.76	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
Total:			45.612								

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		0.117	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	1	n.a.

**WG194153-3A****WG194153-3A,WG194433,STD2BADGE**

Sample Name:	WG194153-3A	Injection Volume:	2.0
Vial Number:	11	Channel:	B
Sample Type:	MBLANK	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 12:30		
Run Time (min):	10.10		

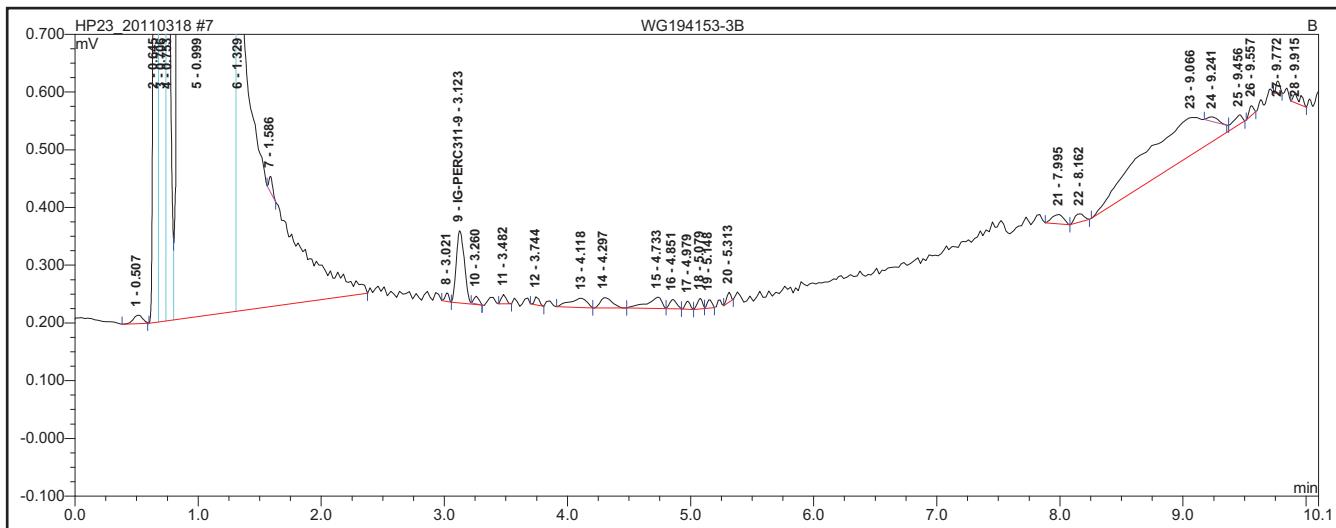


No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.15	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
2	n.a.	0.54	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
3	n.a.	0.65	0.022	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
4	n.a.	0.71	0.087	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
5	n.a.	0.75	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
6	n.a.	1.00	45.366	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
7	n.a.	1.34	0.061	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
8	n.a.	1.60	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
9	IG-PERC311-9	3.14	0.005	XXLOff	3.546E-1	2.801E-2	2.820E+0	-0.0080	1	2.0	-0.0160
10	n.a.	8.44	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
11	n.a.	8.76	0.011	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
12	n.a.	9.02	0.011	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
13	n.a.	9.23	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
14	n.a.	9.33	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
<b>Total:</b>			<b>45.578</b>								

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		0.005	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	2	n.a.

**WG194153-3B****WG194153-3B,WG194433,STD2BADGE**

Sample Name:	WG194153-3B	Injection Volume:	2.0
Vial Number:	12	Channel:	B
Sample Type:	MBLANK	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 12:43		
Run Time (min):	10.10		



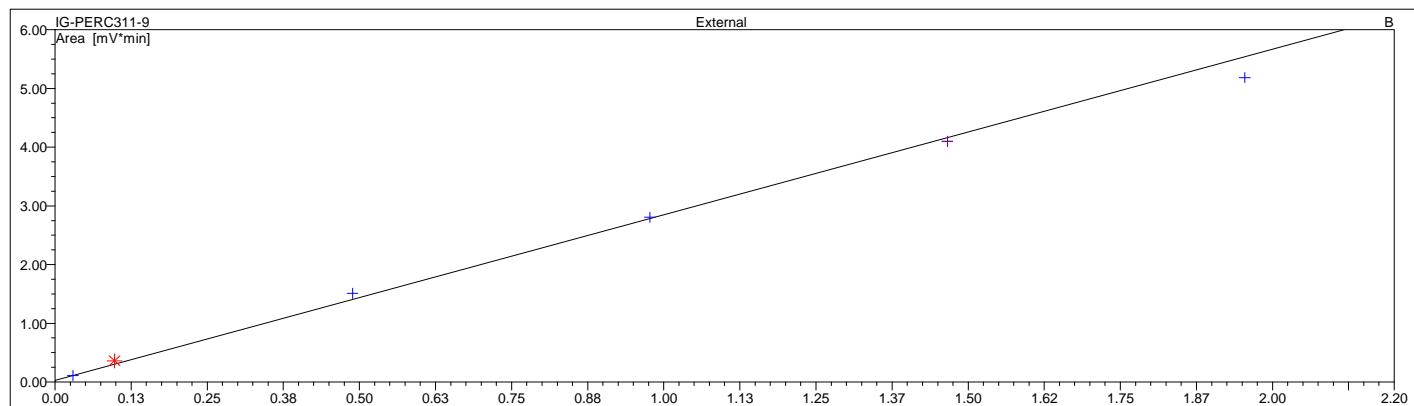
No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.51	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
2	n.a.	0.64	0.044	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
3	n.a.	0.71	0.058	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
4	n.a.	0.75	0.036	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
5	n.a.	1.00	45.973	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
6	n.a.	1.33	0.161	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
7	n.a.	1.59	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
8	n.a.	3.02	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
9	IG-PERC311-9	3.12	0.009	XXLOff	3.546E-1	2.801E-2	2.820E+0	-0.0066	1	2.0	-0.0132
10	n.a.	3.26	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
11	n.a.	3.48	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
12	n.a.	3.74	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
13	n.a.	4.12	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
14	n.a.	4.30	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
15	n.a.	4.73	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
16	n.a.	4.85	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
17	n.a.	4.98	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
18	n.a.	5.08	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
19	n.a.	5.15	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
20	n.a.	5.31	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
21	n.a.	7.99	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
22	n.a.	8.16	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
23	n.a.	9.07	0.048	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
24	n.a.	9.24	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
25	n.a.	9.46	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
26	n.a.	9.56	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
27	n.a.	9.77	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
28	n.a.	9.92	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
<b>Total:</b>			<b>46.353</b>								

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		0.009	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	2 n.a.	

## Initial Calibration Data

**Method:** HP23\_20110318\_PERC311-9  
**Program:** HP23\_PERC\_METHOD\_A  
**Sequence:** HP23\_20110318\_PERC311-9  
**Path:** HP23\Methods

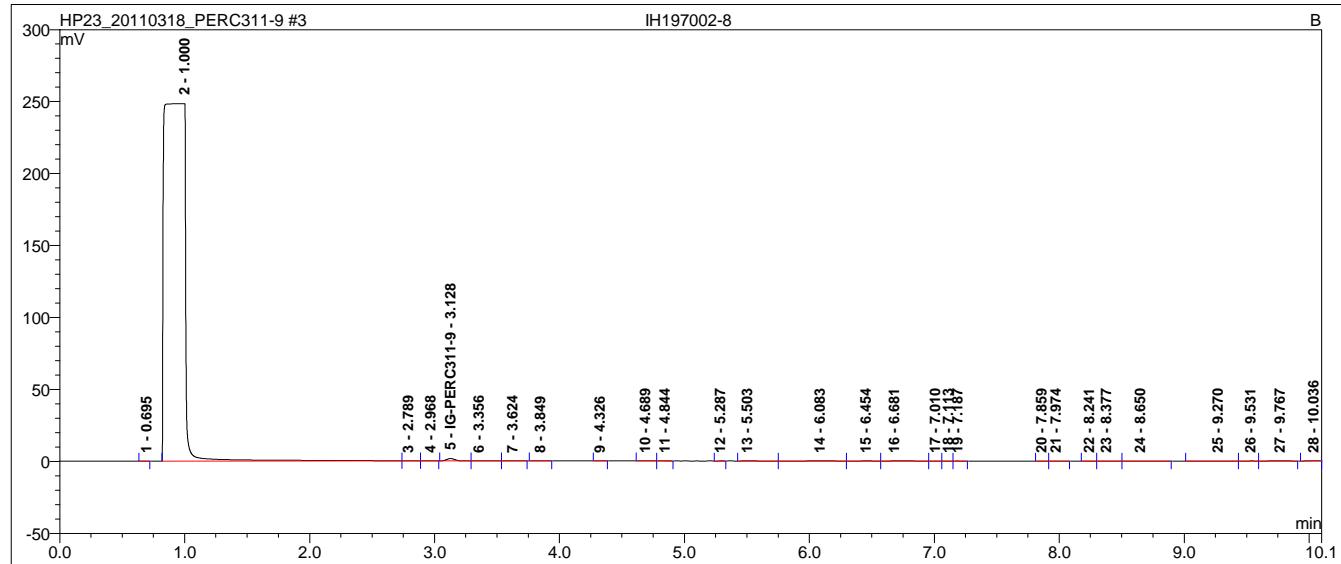
Name	Cal.Mode	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Std.Dev.	Rel.Std.Dev. %	R-Square
IG-PERC311-9	Fixed	XXLOff	3.546E-1	2.801E-2	2.820E+0	0.011	8.881	0.9973



Calibration Levels	Calibration Sequence	Target Amount	Area mV*min	Amnt.Dev. (rel) %	RF-Value Conc/Area	Offset b	Slope m	n Point Status
		IG-PERC311-9	B	PERC311-9	PERC311-9	IG-PERC311-9	IG-PERC311-9	IG-PERC311-9
IH197002-8	HP23\Methods\HP23_20110318_PERC311-9	0.0293	0.110	-0.3	3.546E-1	2.801E-2	2.820E+0	Enabled
IH197002-7	HP23\Methods\HP23_20110318_PERC311-9	0.0977	0.362	21.1	3.546E-1	2.801E-2	2.820E+0	Disabled
IH197002-6	HP23\Methods\HP23_20110318_PERC311-9	0.4886	1.510	7.5	3.546E-1	2.801E-2	2.820E+0	Enabled
IH197002-5	HP23\Methods\HP23_20110318_PERC311-9	0.9772	2.806	0.8	3.546E-1	2.801E-2	2.820E+0	Enabled
IH197002-4	HP23\Methods\HP23_20110318_PERC311-9	1.4658	4.098	-1.5	3.546E-1	2.801E-2	2.820E+0	Enabled
IH197002-3	HP23\Methods\HP23_20110318_PERC311-9	1.9545	5.185	-6.4	3.546E-1	2.801E-2	2.820E+0	Enabled

**IH197002-8**

Sample Name:	IH197002-8	Injection Volume:	2.0
Vial Number:	2	Channel:	B
Sample Type:	ICAL	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 9:32		
Run Time (min):	10.10		



No.	Peak Name	Comment	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	n.a.	0.69	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
2	n.a.	n.a.	1.00	46.756	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
3	n.a.	n.a.	2.79	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
4	n.a.	n.a.	2.97	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
5	IG-PERC311-9		3.13	0.110	XXLOff	3.546E-1	2.801E-2	2.820E+0	0.0292	1	1.0	0.0292
6	n.a.	n.a.	3.36	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
7	n.a.	n.a.	3.62	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
8	n.a.	n.a.	3.85	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
9	n.a.	n.a.	4.33	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
10	n.a.	n.a.	4.69	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
11	n.a.	n.a.	4.84	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
12	n.a.	n.a.	5.29	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
13	n.a.	n.a.	5.50	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
14	n.a.	n.a.	6.08	0.005	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
15	n.a.	n.a.	6.45	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
16	n.a.	n.a.	6.68	0.005	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
17	n.a.	n.a.	7.01	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
18	n.a.	n.a.	7.11	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
19	n.a.	n.a.	7.19	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
20	n.a.	n.a.	7.86	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
21	n.a.	n.a.	7.97	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
22	n.a.	n.a.	8.24	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
23	n.a.	n.a.	8.38	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
24	n.a.	n.a.	8.65	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
25	n.a.	n.a.	9.27	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
26	n.a.	n.a.	9.53	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
27	n.a.	n.a.	9.77	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
28	n.a.	n.a.	10.04	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.

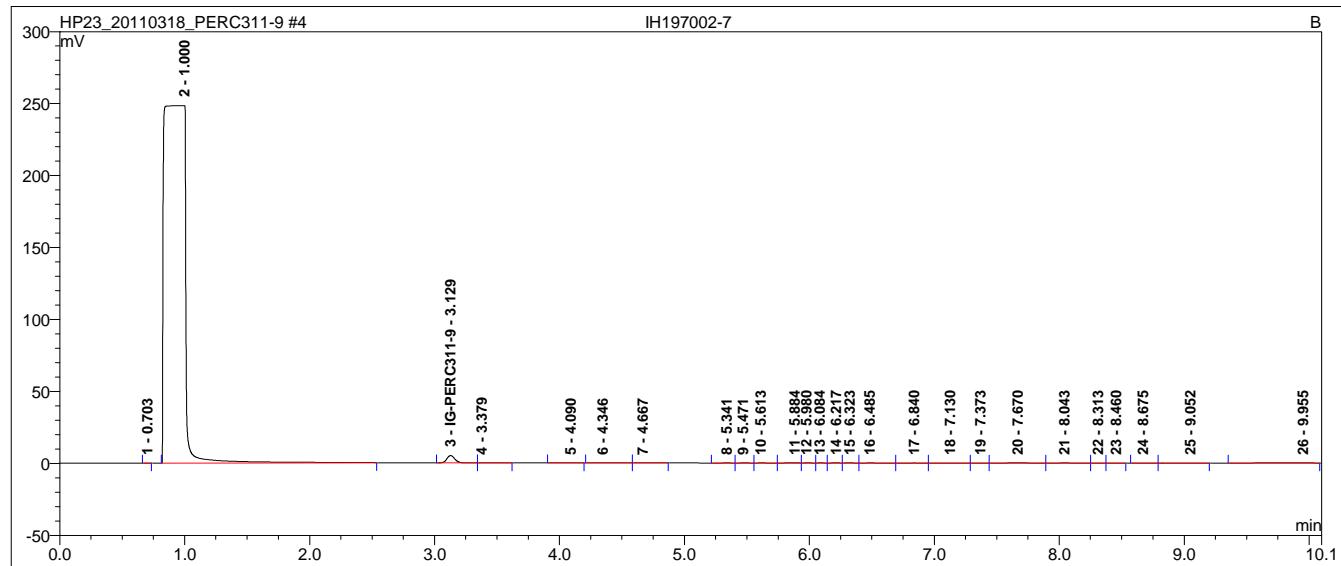
Total: 46.913

Group Name	Group Total ug
	0.0292

**IH197002-7**

not used - dev &gt;20%

Sample Name:	IH197002-7	Injection Volume:	2.0
Vial Number:	3	Channel:	B
Sample Type:	ICAL	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 9:45		
Run Time (min):	10.10		



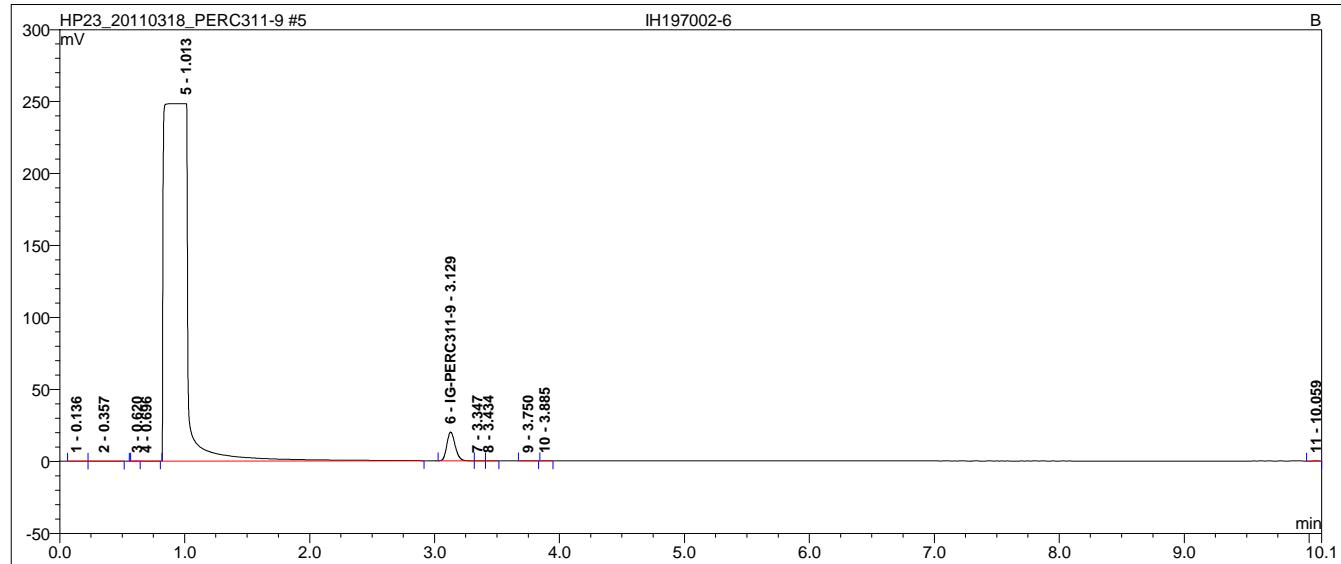
No.	Peak Name	Comment	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	n.a.	0.70	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
2	n.a.	n.a.	1.00	47.356	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
3	IG-PERC311-9		3.13	0.362	XXLOff	3.546E-1	2.801E-2	2.820E+0	0.1183	1	1.0	0.1183
4	n.a.	n.a.	3.38	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
5	n.a.	n.a.	4.09	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
6	n.a.	n.a.	4.35	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
7	n.a.	n.a.	4.67	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
8	n.a.	n.a.	5.34	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
9	n.a.	n.a.	5.47	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
10	n.a.	n.a.	5.61	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
11	n.a.	n.a.	5.88	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
12	n.a.	n.a.	5.98	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
13	n.a.	n.a.	6.08	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
14	n.a.	n.a.	6.22	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
15	n.a.	n.a.	6.32	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
16	n.a.	n.a.	6.49	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
17	n.a.	n.a.	6.84	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
18	n.a.	n.a.	7.13	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
19	n.a.	n.a.	7.37	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
20	n.a.	n.a.	7.67	0.007	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
21	n.a.	n.a.	8.04	0.005	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
22	n.a.	n.a.	8.31	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
23	n.a.	n.a.	8.46	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
24	n.a.	n.a.	8.67	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
25	n.a.	n.a.	9.05	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
26	n.a.	n.a.	9.96	0.005	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.

Total: 47.773

Group Name	Group Total ug
	0.1183

**IH197002-6**

Sample Name:	IH197002-6	Injection Volume:	2.0
Vial Number:	4	Channel:	B
Sample Type:	ICAL	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 9:58		
Run Time (min):	10.10		

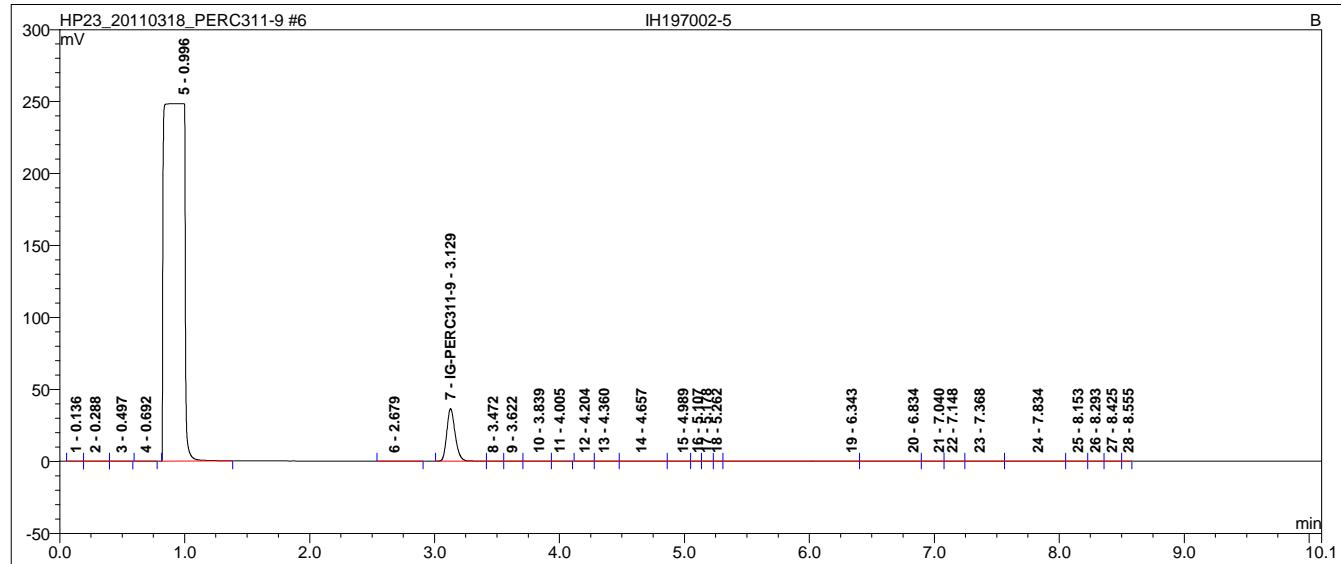


No.	Peak Name	Comment	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	n.a.	0.14	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
2	n.a.	n.a.	0.36	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
3	n.a.	n.a.	0.62	0.000	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
4	n.a.	n.a.	0.70	0.006	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
5	n.a.	n.a.	1.01	53.332	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
6	1G-PERC311-9		3.13	1.510	XXLOff	3.546E-1	2.801E-2	2.820E+0	0.5254	1	1.0	0.5254
7	n.a.	n.a.	3.35	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
8	n.a.	n.a.	3.43	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
9	n.a.	n.a.	3.75	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
10	n.a.	n.a.	3.89	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
11	n.a.	n.a.	10.06	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
Total:				54.856								

	Group Name	Group Total ug
		0.5254

**IH197002-5**

Sample Name:	IH197002-5	Injection Volume:	2.0
Vial Number:	5	Channel:	B
Sample Type:	ICAL	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 10:11		
Run Time (min):	10.10		

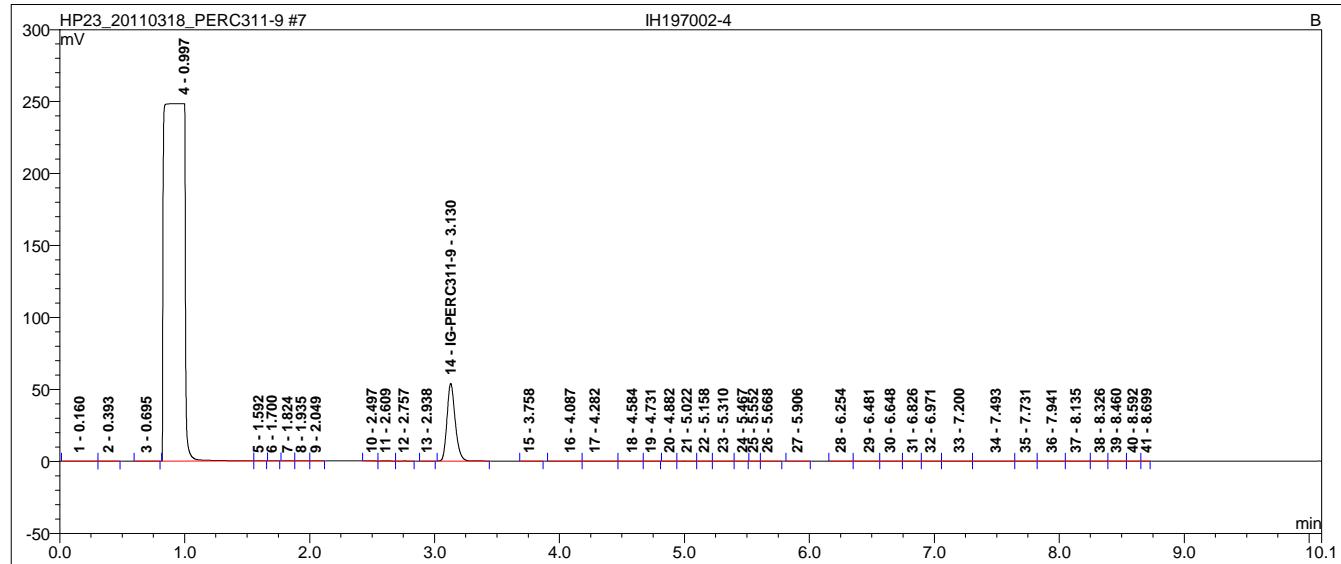


No.	Peak Name	Comment	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	n.a.	0.14	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
2	n.a.	n.a.	0.29	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
3	n.a.	n.a.	0.50	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
4	n.a.	n.a.	0.69	0.007	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
5	n.a.	n.a.	1.00	45.497	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
6	n.a.	n.a.	2.68	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
7	IG-PERC311-9		3.13	2.806	XXLOff	3.546E-1	2.801E-2	2.820E+0	0.9851	1	1.0	0.9851
8	n.a.	n.a.	3.47	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
9	n.a.	n.a.	3.62	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
10	n.a.	n.a.	3.84	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
11	n.a.	n.a.	4.00	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
12	n.a.	n.a.	4.20	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
13	n.a.	n.a.	4.36	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
14	n.a.	n.a.	4.66	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
15	n.a.	n.a.	4.99	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
16	n.a.	n.a.	5.11	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
17	n.a.	n.a.	5.18	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
18	n.a.	n.a.	5.26	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
19	n.a.	n.a.	6.34	0.021	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
20	n.a.	n.a.	6.83	0.010	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
21	n.a.	n.a.	7.04	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
22	n.a.	n.a.	7.15	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
23	n.a.	n.a.	7.37	0.006	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
24	n.a.	n.a.	7.83	0.008	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
25	n.a.	n.a.	8.15	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
26	n.a.	n.a.	8.29	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
27	n.a.	n.a.	8.42	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
28	n.a.	n.a.	8.55	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
<b>Total:</b>				48.392								

	Group Name	Group Total ug
		0.9851

**IH197002-4**

Sample Name:	IH197002-4	Injection Volume:	2.0
Vial Number:	6	Channel:	B
Sample Type:	ICAL	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 10:24		
Run Time (min):	10.10		



No.	Peak Name	Comment	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	n.a.	0.16	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
2	n.a.	n.a.	0.39	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
3	n.a.	n.a.	0.70	0.006	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
4	n.a.	n.a.	1.00	45.485	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
5	n.a.	n.a.	1.59	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
6	n.a.	n.a.	1.70	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
7	n.a.	n.a.	1.82	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
8	n.a.	n.a.	1.93	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
9	n.a.	n.a.	2.05	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
10	n.a.	n.a.	2.50	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
11	n.a.	n.a.	2.61	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
12	n.a.	n.a.	2.76	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
13	n.a.	n.a.	2.94	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
14	1G-PERC311-9		3.13	4.098	XXLOff	3.546E-1	2.801E-2	2.820E+0	1.4431	1	1.0	1.4431
15	n.a.	n.a.	3.76	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
16	n.a.	n.a.	4.09	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
17	n.a.	n.a.	4.28	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
18	n.a.	n.a.	4.58	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
19	n.a.	n.a.	4.73	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
20	n.a.	n.a.	4.88	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
21	n.a.	n.a.	5.02	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
22	n.a.	n.a.	5.16	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
23	n.a.	n.a.	5.31	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
24	n.a.	n.a.	5.47	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
25	n.a.	n.a.	5.55	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
26	n.a.	n.a.	5.67	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
27	n.a.	n.a.	5.91	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
28	n.a.	n.a.	6.25	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
29	n.a.	n.a.	6.48	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
30	n.a.	n.a.	6.65	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
31	n.a.	n.a.	6.83	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
32	n.a.	n.a.	6.97	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
33	n.a.	n.a.	7.20	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
34	n.a.	n.a.	7.49	0.005	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
35	n.a.	n.a.	7.73	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
36	n.a.	n.a.	7.94	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
37	n.a.	n.a.	8.13	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.

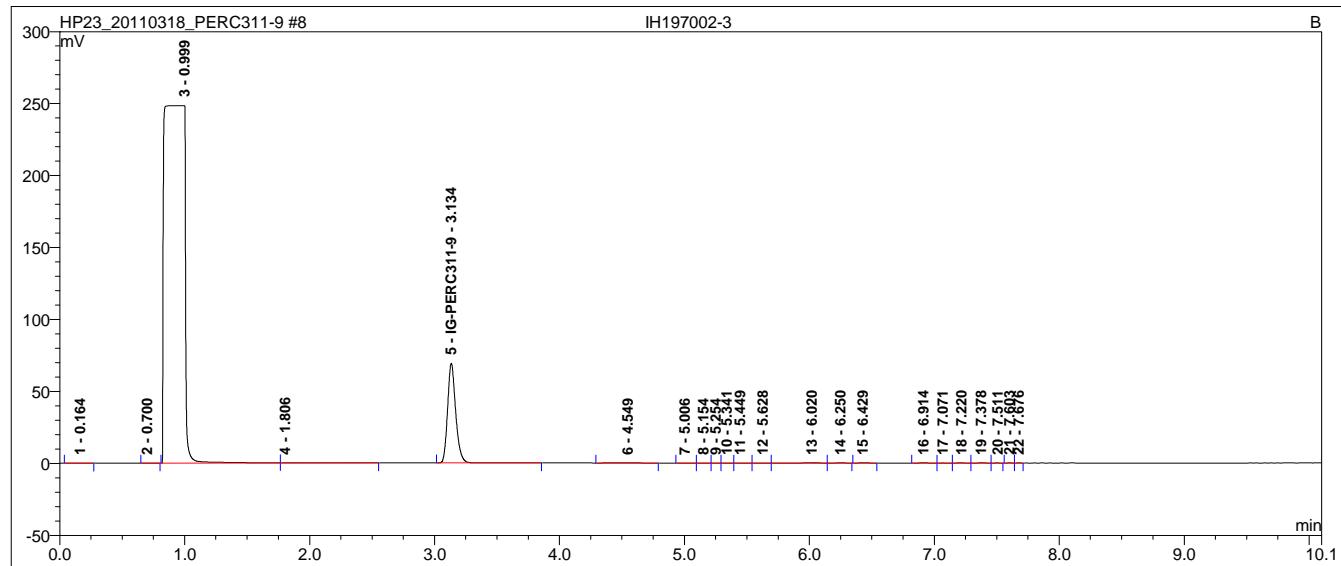
38	n.a.	n.a.	8.33	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
39	n.a.	n.a.	8.46	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
40	n.a.	n.a.	8.59	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
41	n.a.	n.a.	8.70	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.

Total: 49.646

Group Name	Group Total ug
	1.4431

**IH197002-3**

Sample Name:	IH197002-3	Injection Volume:	2.0
Vial Number:	7	Channel:	B
Sample Type:	ICAL	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 10:57		
Run Time (min):	10.10		



No.	Peak Name	Comment	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	n.a.	0.16	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
2	n.a.	n.a.	0.70	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
3	n.a.	n.a.	1.00	45.697	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
4	n.a.	n.a.	1.81	0.059	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
5	IG-PERC311-9		3.13	5.185	XXLOff	3.546E-1	2.801E-2	2.820E+0	1.8288	1	1.0	1.8288
6	n.a.	n.a.	4.55	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
7	n.a.	n.a.	5.01	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
8	n.a.	n.a.	5.15	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
9	n.a.	n.a.	5.25	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
10	n.a.	n.a.	5.34	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
11	n.a.	n.a.	5.45	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
12	n.a.	n.a.	5.63	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
13	n.a.	n.a.	6.02	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
14	n.a.	n.a.	6.25	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
15	n.a.	n.a.	6.43	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
16	n.a.	n.a.	6.91	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
17	n.a.	n.a.	7.07	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
18	n.a.	n.a.	7.22	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
19	n.a.	n.a.	7.38	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
20	n.a.	n.a.	7.51	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
21	n.a.	n.a.	7.60	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.
22	n.a.	n.a.	7.68	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	1.0	n.a.

Total: 50.97

	Group Name	Group Total ug
		1.8288

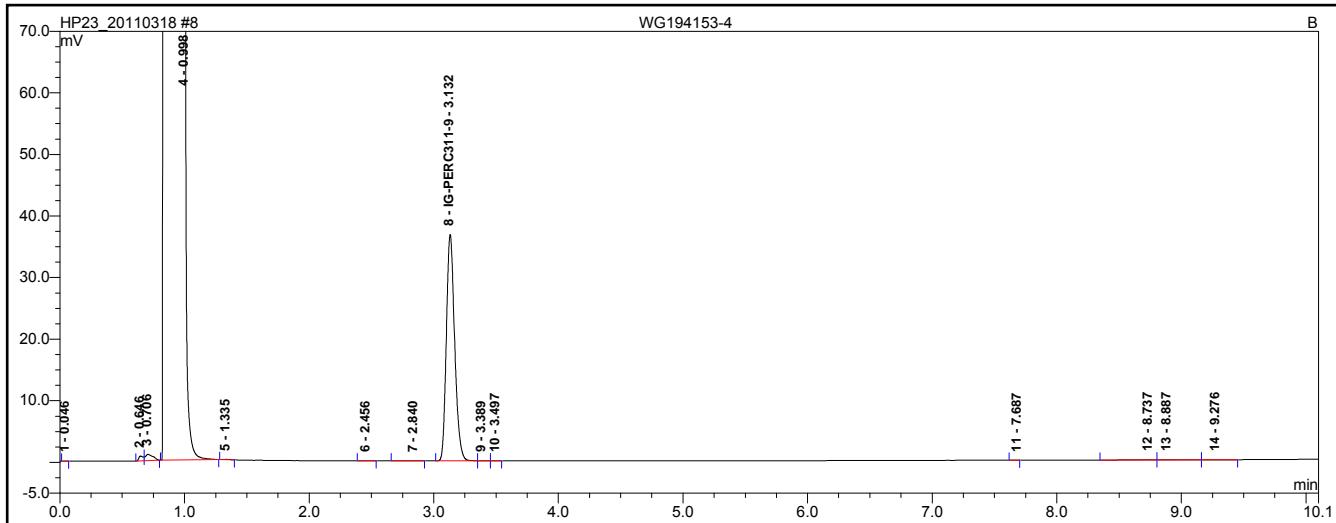
Blank Spike

Blank Spike Duplicate

Data

**WG194153-4****WG194153-4,IH196262-1,1,STD2BADGE**

Sample Name:	WG194153-4	Injection Volume:	2.0
Vial Number:	13	Channel:	B
Sample Type:	BS	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 12:56		
Run Time (min):	10.10		

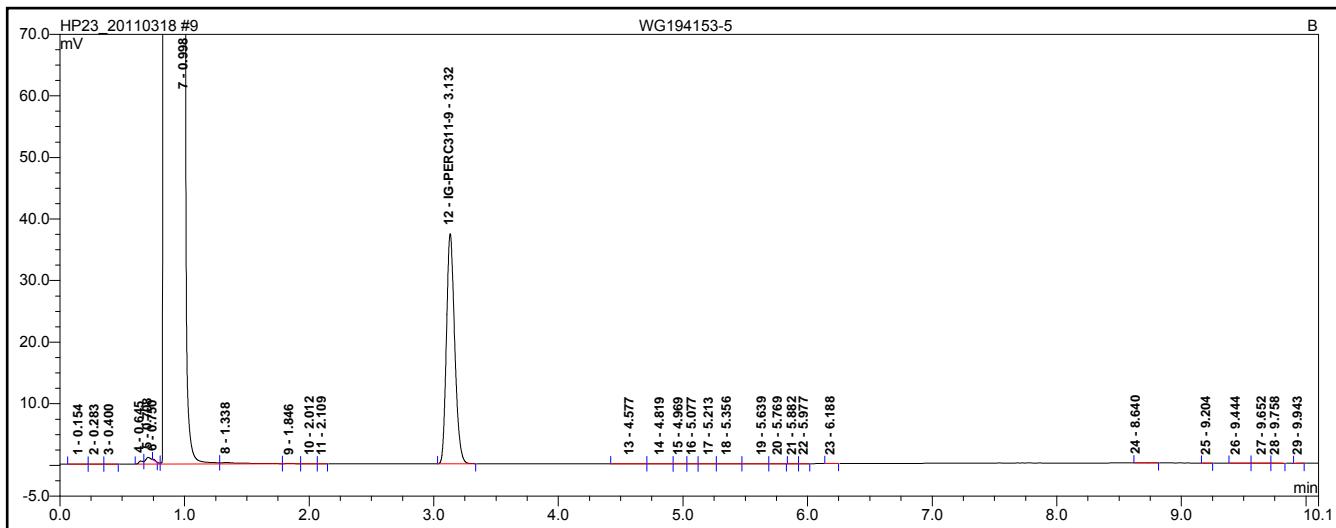


No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.05	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
2	n.a.	0.65	0.031	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
3	n.a.	0.71	0.073	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
4	n.a.	1.00	45.353	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
5	n.a.	1.34	0.004	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
6	n.a.	2.46	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
7	n.a.	2.84	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
8	IG-PERC311-9	3.13	2.826	XXLOff	3.546E-1	2.801E-2	2.820E+0	0.9923	1	2.0	1.9845
9	n.a.	3.39	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
10	n.a.	3.50	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
11	n.a.	7.69	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
12	n.a.	8.74	0.013	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
13	n.a.	8.89	0.007	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
14	n.a.	9.28	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
Total:			48.314								

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		2.826	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	2	n.a.

**WG194153-5****WG194153-5,IH196262-1,1,STD2BADGE**

Sample Name:	WG194153-5	Injection Volume:	2.0
Vial Number:	14	Channel:	B
Sample Type:	BSD	Lot Correction:	0 ug
Control Program:	HP23_PERC_METHOD_A	Standard Method:	External
Quantif. Method:	HP23_20110318_PERC311-9		
Recording Time:	3/18/2011 13:09		
Run Time (min):	10.10		



No.	Peak Name	RT min	Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Conc ug/mL	*Dilution	*DeVol mL	Total ug
1	n.a.	0.15	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
2	n.a.	0.28	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
3	n.a.	0.40	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
4	n.a.	0.65	0.019	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
5	n.a.	0.71	0.080	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
6	n.a.	0.75	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
7	n.a.	1.00	45.157	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
8	n.a.	1.34	0.041	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
9	n.a.	1.85	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
10	n.a.	2.01	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
11	n.a.	2.11	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
12	IG-PERC311-9	3.13	2.864	XXLOff	3.546E-1	2.801E-2	2.820E+0	1.0057	1	2.0	2.0115
13	n.a.	4.58	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
14	n.a.	4.82	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
15	n.a.	4.97	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
16	n.a.	5.08	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
17	n.a.	5.21	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
18	n.a.	5.36	0.003	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
19	n.a.	5.64	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
20	n.a.	5.77	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
21	n.a.	5.88	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
22	n.a.	5.98	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
23	n.a.	6.19	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
24	n.a.	8.64	0.002	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
25	n.a.	9.20	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
26	n.a.	9.44	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
27	n.a.	9.65	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
28	n.a.	9.76	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.
29	n.a.	9.94	0.001	n.a.	n.a.	n.a.	n.a.	n.a.	1	2.0	n.a.

Total: 48.191

	Group Name	Group Area mV*min	Cal.Type	RF-Value Conc/Area	Offset b	Slope m	Group Conc ug/mL	*Dilution	*DeVol mL	Group Total ug
		2.864	XXLOff	3.546E-1	2.801E-2	2.820E+0	n.a.	1	2 n.a.	

## Prep Data



# Galson Sample Prep Report

Print Date: 03/28/2011 13:04:53

Department: 32

Workgroup: WG194153

Sample Number:

Sample Number	Sample Type	Sample Part	Media Type	Analyte		Product Code		CAS Number		Dispenser ID	Desorption Start Time	Desorption Stop Time	#1 Prep Analyst	#2 Prep Analyst	Prep Date	Timer Used	Shaken	Comments
				Perchloroethylene		IG-PERC311-9		127-18-4										
L235614-1	SAMP	A	OPTIONS	0228	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
L235614-1	SAMP	B	OPTIONS	0228	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
L235614-2	SAMP	A	M3M-3520	0228	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
L235614-2	SAMP	B	M3M-3520	0228	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
L235614-3	SAMP	A	M3M-3520	0228	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
L235614-3	SAMP	B	M3M-3520	0228	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
WG194153-1	IBLANK	A	M3M-3520		1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
WG194153-1	IBLANK	B	M3M-3520		1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
WG194153-2	EBLANK	A	M3M-3520		1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
WG194153-2	EBLANK	B	M3M-3520		1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
WG194153-3	MBLANK	A	M3M-3520	0235	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
WG194153-3	MBLANK	B	M3M-3520	0235	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
WG194153-4	BS	A	M3M-3520	651	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
WG194153-4	BS	B	M3M-3520	651	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
WG194153-5	BSD	A	M3M-3520	651	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															
WG194153-5	BSD	B	M3M-3520	651	1	32	2 ml	IH19580 2	CS2 W/UNDECANE	D761	03/16/2011 06:25:00	03/16/2011 06:55:00	RROGERS		16-MAR-11		Y	
			SOP Name: GC-SOP-9															



## QC Spiking Report

Print Date March 28, 2011 1:06 PM

Workgroup WG194153

Sample	QC Type	Desc	Reference Sample	Standard Used	Pipette No	Amount Spiked	Spike Units	Spiked Analyst	Media Lot	Comment
WG194153-1	IBLANK	Instrument Blank	CS2							
	33	M3M-3520	IG-PERC311-9	Perchloroethylene						
WG194153-2	EBLANK	Eluent Blank	CS2							
	33	M3M-3520	IG-PERC311-9	Perchloroethylene						
WG194153-3	MBLANK	Media Blank	OVM						0235	
	33	M3M-3520	IG-PERC311-9	Perchloroethylene						
WG194153-4	BS	Blank Spike		IH196262-1	G55	20	ul	MRH	0235	
	33	M3M-3520	IG-PERC311-9	Perchloroethylene						
WG194153-5	BSD	Blank Spike Dup	WG194153-4	IH196262-1	G55	20	ul	MRH	0235	
	33	M3M-3520	IG-PERC311-9	Perchloroethylene						

## Standards Prep Logs



## Galson Chemical Inventory Report

03/28/2011 14:15

Ref	Description	Location	Dept	Date Received	Expire Date	Dispose Date	D
11529	UNDECANE	SC5	24GC	01/30/2009	01/30/2012		
	<b>Vendor</b> Fisher	<b>Manufacturer</b> ACROS		<b>Lot No</b> B0126569			
	<b>Comments</b> Cat#14066-1000 100mls						
Cas Num		Chemical Name		% Purity	Conc(ug/mL)		
1120-21-4		Undecane		99.5			



## Galson Chemical Inventory Report

03/28/2011 14:16

Ref	Description	Location	Dept	Date Received	Expire Date	Dispose Date	D
15573	CS2 517	BUNKER	24GC	02/09/2011	02/09/2014		
<b>Vendor</b>	Sigma-Aldrich	<b>Manufacturer</b>	SIGMA ALDRICH	<b>Lot No</b>	03496LMV		
<b>Comments</b>							
Cas Num		Chemical Name		% Purity	Conc(ug/mL)		
	75-15-0	CARBON DISULFIDE		99.96			



## Galson Chemical Inventory Report

03/28/2011 14:17

Ref	Description	Location	Dept	Date Received	Expire Date	Dispose Date	D
14920	TETRACHLOROETHYLENE	SC3-S4-4	24GC	10/13/2010	10/11/2011		
	<b>Vendor</b> Spex Certiprep	<b>Manufacturer</b> SPEX		<b>Lot No</b> C1101007019			
	<b>Comments</b>						
	Cas Num	Chemical Name		% Purity	Conc(ug/mL)		
	127-18-4	TETRACHLOROETHYLENE		99			



## Galson Chemical Inventory Report

03/28/2011 14:17

Ref	Description	Location	Dept	Date Received	Expire Date	Dispose Date	D
15575	CS2 517	BUNKER	24GC	02/09/2011	02/09/2014		
<b>Vendor</b>	Sigma-Aldrich	<b>Manufacturer</b>	SIGMA ALDRICH	<b>Lot No</b>	03496LMV		
<b>Comments</b>							
Cas Num		Chemical Name		% Purity	Conc(ug/mL)		
	75-15-0	CARBON DISULFIDE		99.96			



## Galson Chemical Inventory Report

03/28/2011 14:18

Ref	Description	Location	Dept	Date Received	Expire Date	Dispose Date	D
13129	TETRACHLOROETHYLENE	SC3-S4-4	24GC	12/17/2009	12/17/2012		
	<b>Vendor</b> Sigma-Aldrich	<b>Manufacturer</b> SIGMA ALDRICH		<b>Lot No</b> 14196BK			
	<b>Comments</b>						
	Cas Num	Chemical Name		% Purity	Conc(ug/mL)		
	127-18-4	TETRACHLOROETHYLENE		99.99			



## Galson Laboratories - Standards Report

Lot: IH195802

03/28/2011

IH195802	CS2 W/UNDECANE	Analyst: MHERMANN Prep Date: 03/11/2011 Expiration Date: 01/30/2012			
Final Volume: 1.000.00mL					
/11529	UNDECANE	Initial Conc: 37.00ug/mL	Initial Weight: 0.0370g	Prep/Purc. Date: 01/30/2009	Expiration Date: 01/30/2012
				Initial Volume: 50.00uL	
	<u>CAS Number</u>	<u>Description</u>		<u>Final Conc</u>	
	1120-21-4	Undecane		36.82ug/mL	
/15573	CS2 517	Initial Conc: 1,263,136.84ug/m	Initial Weight: 1263.1368g	Prep/Purc. Date: 02/09/2011	Expiration Date: 02/09/2014
				Initial Volume: 999,950.00uL	
	<u>CAS Number</u>	<u>Description</u>		<u>Final Conc</u>	
	75-15-0	CARBON DISULFIDE		1262631.59ug/mL	



## Galson Laboratories - Standards Report

Lot: IH197002

03/28/2011

IH197002 /14920	Final Volume:2.00mL  IG-PERC311-9 --- CURVE/DE/LOQ	Analyst:MHERMANN Prep Date:03/18/2011 Expiration Date:06/18/2011		
/IH196702 /IH196702/11529	TETRACHLOROETHYLENE  <u>CAS Number</u> 127-18-4  CS2/UNDECANE(diluent) UNDECANE  <u>CAS Number</u> 1120-21-4  CS2 517	Initial Conc:9,870.98ug/mL  <u>Description</u> TETRACHLOROETHYLENE  Initial Conc:37.00ug/mL  <u>Description</u> Undecane  Initial Conc:1,263,136.84ug/m  <u>Description</u> CARBON DISULFIDE	Initial Weight:0.0197g  Analyst:RROGERS  Initial Weight:0.0370g  Initial Weight:1263.1368g	Prep/Purc. Date:10/13/2010 Expiration Date:10/11/2011  <u>Final Conc</u> 9772.27ug/mL  Prep/Purc. Date:03/17/2011 Expiration Date:01/30/2012 Prep/Purc. Date:01/30/2009 Expiration Date:01/30/2012  <u>Final Conc</u> 18.26ug/mL  Prep/Purc. Date:02/09/2011 Expiration Date:02/09/2014  <u>Final Conc</u> 626098.64ug/mL
IH197002-1 /IH197002/14920	50X DILUTION  TETRACHLOROETHYLENE  <u>CAS Number</u> 127-18-4  CS2/UNDECANE(diluent) UNDECANE  <u>CAS Number</u> 1120-21-4  CS2 517	Initial Conc:9,772.27ug/mL  <u>Description</u> TETRACHLOROETHYLENE  Initial Conc:18.26ug/mL  <u>Description</u> Undecane  Initial Conc:626,098.64ug/mL  <u>Description</u> CARBON DISULFIDE	Initial Weight:0.0197g  Analyst:RROGERS  Initial Weight:0.0370g  Initial Weight:1263.1368g	Prep/Purc. Date:10/13/2010 Expiration Date:10/11/2011  <u>Final Conc</u> 195.45ug/mL  Prep/Purc. Date:03/17/2011 Expiration Date:01/30/2012 Prep/Purc. Date:01/30/2009 Expiration Date:01/30/2012  <u>Final Conc</u> 0.37ug/mL  Prep/Purc. Date:02/09/2011 Expiration Date:02/09/2014  <u>Final Conc</u> 12521.97ug/mL
IH197002-2 /IH197002-5/IH197002-2 /IH197002-1/IH197002/1 4920	1000X DILUTION  TETRACHLOROETHYLENE  <u>CAS Number</u> 127-18-4  CS2/UNDECANE(diluent)	Initial Conc:195.45ug/mL  <u>Description</u> TETRACHLOROETHYLENE  Initial Conc:9.77ug/mL  <u>Description</u> Undecane	Initial Weight:0.0197g  Analyst:RROGERS  Initial Weight:0.0370g  Initial Weight:1263.1368g	Prep/Purc. Date:10/13/2010 Expiration Date:10/11/2011  Prep/Purc. Date:03/17/2011 Expiration Date:01/30/2012  Prep/Purc. Date:01/30/2009 Expiration Date:01/30/2012  Prep/Purc. Date:03/17/2011 Expiration Date:01/30/2012

/IH197002-5/IH197002-2	UNDECANE		Prep/Purc. Date:01/30/2009 Expiration Date:01/30/2012
/IH197002-1/IH197002/I			
H196702/11529			
		Initial Conc:0.37ug/mL	Initial Volume:50.00uL
	<u>CAS Number</u>	<u>Description</u>	<u>Final Conc</u>
	1120-21-4	Undecane	0.018ug/mL
/IH197002-5/IH197002-2	CS2 517		Prep/Purc. Date:02/09/2011 Expiration Date:02/09/2014
/IH197002-1/IH197002/I			
H196702/15575			
		Initial Conc:12,521.97ug/mL	Initial Volume:50.00uL
	<u>CAS Number</u>	<u>Description</u>	<u>Final Conc</u>
	75-15-0	CARBON DISULFIDE	626.1ug/mL
IH197002-3	LEV 6		Analyst:MHERMANN Prep Date:03/18/2011 Expiration Date:04/18/2011
Final Volume:1,000.00uL			
/IH197002-1/IH197002/1	TETRACHLOROETHYLENE		Prep/Purc. Date:10/13/2010 Expiration Date:10/11/2011
4920			
		Initial Conc:195.45ug/mL	Initial Weight:0.0197g
	<u>CAS Number</u>	<u>Description</u>	<u>Initial Volume:</u> 10.00uL
	127-18-4	TETRACHLOROETHYLENE	<u>Final Conc</u> 1.95ug/mL
/IH197002-1/IH197002/I	CS2/UNDECANE(diluent)		Prep/Purc. Date:03/17/2011 Expiration Date:01/30/2012
H196702			
/IH197002-1/IH197002/I	UNDECANE		Prep/Purc. Date:01/30/2009 Expiration Date:01/30/2012
H196702/11529			
		Initial Conc:0.37ug/mL	Initial Weight:0.0370g
	<u>CAS Number</u>	<u>Description</u>	<u>Initial Volume:</u> 10.00uL
	1120-21-4	Undecane	<u>Final Conc</u> 0.0037ug/mL
/IH197002-1/IH197002/I	CS2 517		Prep/Purc. Date:02/09/2011 Expiration Date:02/09/2014
H196702/15575			
		Initial Conc:12,521.97ug/mL	Initial Weight:1263.1368g
	<u>CAS Number</u>	<u>Description</u>	<u>Initial Volume:</u> 10.00uL
	75-15-0	CARBON DISULFIDE	<u>Final Conc</u> 125.22ug/mL
IH197002-4	LEV 5		Analyst:MHERMANN Prep Date:03/18/2011 Expiration Date:04/18/2011
Final Volume:2,000.00uL			
/IH197002-1/IH197002/1	TETRACHLOROETHYLENE		Prep/Purc. Date:10/13/2010 Expiration Date:10/11/2011
4920			
		Initial Conc:195.45ug/mL	Initial Weight:0.0197g
	<u>CAS Number</u>	<u>Description</u>	<u>Initial Volume:</u> 15.00uL
	127-18-4	TETRACHLOROETHYLENE	<u>Final Conc</u> 1.47ug/mL
/IH197002-1/IH197002/I	CS2/UNDECANE(diluent)		Prep/Purc. Date:03/17/2011 Expiration Date:01/30/2012
H196702			
/IH197002-1/IH197002/I	UNDECANE		Prep/Purc. Date:01/30/2009 Expiration Date:01/30/2012
H196702/11529			
		Initial Conc:0.37ug/mL	Initial Weight:0.0370g
	<u>CAS Number</u>	<u>Description</u>	<u>Initial Volume:</u> 15.00uL
	1120-21-4	Undecane	<u>Final Conc</u> 0.0027ug/mL
/IH197002-1/IH197002/I	CS2 517		Prep/Purc. Date:02/09/2011 Expiration Date:02/09/2014
H196702/15575			

		Initial Conc:12,521.97ug/mL	Initial Weight:1263.1368g	Initial Volume:15.00uL
	<u>CAS Number</u>	<u>Description</u>	<u>Final Conc</u>	
	75-15-0	CARBON DISULFIDE	93.91ug/mL	
IH197002-5	LEV 4		Analyst:MHERMANN	Prep Date:03/18/2011 Expiration Date:04/18/2011
Final Volume:1,000.00uL				
/IH197002-5/IH197002-2	TETRACHLOROETHYLENE		Prep/Purc. Date:10/13/2010	Expiration Date:10/11/2011
/IH197002/14920				
		Initial Conc:9.77ug/mL	Initial Weight:0.0197g	Initial Volume:100.00uL
	<u>CAS Number</u>	<u>Description</u>	<u>Final Conc</u>	
	127-18-4	TETRACHLOROETHYLENE	0.98ug/mL	
/IH197002-5/IH197002-2	CS2/UNDECANE(diluent)		Analyst:RROGERS	Prep/Purc. Date:03/17/2011 Expiration Date:01/30/2012
/IH197002/IH196702				
/IH197002-5/IH197002-2	UNDECANE		Prep/Purc. Date:01/30/2009	Expiration Date:01/30/2012
/IH197002/IH196702/11				
529				
		Initial Conc:0.02ug/mL	Initial Weight:0.0370g	Initial Volume:100.00uL
	<u>CAS Number</u>	<u>Description</u>	<u>Final Conc</u>	
	1120-21-4	Undecane	0.0018ug/mL	
/IH197002-5/IH197002-2	CS2 517		Prep/Purc. Date:02/09/2011	Expiration Date:02/09/2014
/IH197002/IH196702/15				
575				
		Initial Conc:626.10ug/mL	Initial Weight:1263.1368g	Initial Volume:100.00uL
	<u>CAS Number</u>	<u>Description</u>	<u>Final Conc</u>	
	75-15-0	CARBON DISULFIDE	62.61ug/mL	
IH197002-6	LEV 3		Analyst:MHERMANN	Prep Date:03/18/2011 Expiration Date:04/18/2011
Final Volume:1,000.00uL				
/IH197002-2/IH197002/1	TETRACHLOROETHYLENE		Prep/Purc. Date:10/13/2010	Expiration Date:10/11/2011
4920				
		Initial Conc:9.77ug/mL	Initial Weight:0.0197g	Initial Volume:50.00uL
	<u>CAS Number</u>	<u>Description</u>	<u>Final Conc</u>	
	127-18-4	TETRACHLOROETHYLENE	0.49ug/mL	
/IH197002-2/IH197002/I	CS2/UNDECANE(diluent)		Analyst:RROGERS	Prep/Purc. Date:03/17/2011 Expiration Date:01/30/2012
H196702				
/IH197002-2/IH197002/I	UNDECANE		Prep/Purc. Date:01/30/2009	Expiration Date:01/30/2012
H196702/11529				
		Initial Conc:0.02ug/mL	Initial Weight:0.0370g	Initial Volume:50.00uL
	<u>CAS Number</u>	<u>Description</u>	<u>Final Conc</u>	
	1120-21-4	Undecane	0.00091ug/mL	
/IH197002-2/IH197002/I	CS2 517		Prep/Purc. Date:02/09/2011	Expiration Date:02/09/2014
H196702/15575				
		Initial Conc:626.10ug/mL	Initial Weight:1263.1368g	Initial Volume:50.00uL
	<u>CAS Number</u>	<u>Description</u>	<u>Final Conc</u>	
	75-15-0	CARBON DISULFIDE	31.3ug/mL	
IH197002-7	LEV 2		Analyst:MHERMANN	Prep Date:03/18/2011 Expiration Date:04/18/2011
Final Volume:1,000.00uL				
/IH197002-2/IH197002/1	TETRACHLOROETHYLENE		Prep/Purc. Date:10/13/2010	Expiration Date:10/11/2011
4920				

		Initial Conc:9.77ug/mL	Initial Weight:0.0197g	Initial Volume:10.00uL
		<u>CAS Number</u> 127-18-4	<u>Description</u> TETRACHLOROETHYLENE	<u>Final Conc</u> 0.098ug/mL
/IH197002-2/IH197002/I H196702	CS2/UNDECANE(diluent)		Analyst:RROGERS	Prep/Purc. Date:03/17/2011 Expiration Date:01/30/2012
/IH197002-2/IH197002/I H196702/11529	UNDECANE			Prep/Purc. Date:01/30/2009 Expiration Date:01/30/2012
		Initial Conc:0.02ug/mL	Initial Weight:0.0370g	Initial Volume:10.00uL
		<u>CAS Number</u> 1120-21-4	<u>Description</u> Undecane	<u>Final Conc</u> 0.00018ug/mL
/IH197002-2/IH197002/I H196702/15575	CS2 517			Prep/Purc. Date:02/09/2011 Expiration Date:02/09/2014
		Initial Conc:626.10ug/mL	Initial Weight:1263.1368g	Initial Volume:10.00uL
		<u>CAS Number</u> 75-15-0	<u>Description</u> CARBON DISULFIDE	<u>Final Conc</u> 6.26ug/mL
IH197002-8 4920	LEV 1		Analyst:MHERMANN	Prep Date:03/18/2011 Expiration Date:04/18/2011
		Final Volume:1.000.00uL		
/IH197002-5/IH197002/I H196702	TETRACHLOROETHYLENE			Prep/Purc. Date:10/13/2010 Expiration Date:10/11/2011
		Initial Conc:0.98ug/mL	Initial Weight:0.0197g	Initial Volume:30.00uL
		<u>CAS Number</u> 127-18-4	<u>Description</u> TETRACHLOROETHYLENE	<u>Final Conc</u> 0.029ug/mL
/IH197002-5/IH197002/I H196702	CS2/UNDECANE(diluent)		Analyst:RROGERS	Prep/Purc. Date:03/17/2011 Expiration Date:01/30/2012
/IH197002-5/IH197002/I H196702/11529	UNDECANE			Prep/Purc. Date:01/30/2009 Expiration Date:01/30/2012
		Initial Conc:0.00ug/mL	Initial Weight:0.0370g	Initial Volume:30.00uL
		<u>CAS Number</u> 1120-21-4	<u>Description</u> Undecane	<u>Final Conc</u> 0.000055ug/mL
/IH197002-5/IH197002/I H196702/15575	CS2 517			Prep/Purc. Date:02/09/2011 Expiration Date:02/09/2014
		Initial Conc:62.61ug/mL	Initial Weight:1263.1368g	Initial Volume:30.00uL
		<u>CAS Number</u> 75-15-0	<u>Description</u> CARBON DISULFIDE	<u>Final Conc</u> 1.88ug/mL



## Galson Laboratories - Standards Report

Lot: IH196702

03/28/2011

IH196702	CS2/UNDECANE	Analyst: RROGERS Prep Date:03/17/2011 Expiration Date:01/30/2012			
/11529	Final Volume:1.000.00mL UNDECANE	Initial Conc:37.00ug/mL	Initial Weight:0.0370g	Prep/Purc. Date:01/30/2009	Expiration Date:01/30/2012 Initial Volume:50.00uL
	<u>CAS Number</u> 1120-21-4	<u>Description</u> Undecane		<u>Final Conc</u> 36.82ug/mL	
/15575	CS2 517	Initial Conc:1,263,136.84ug/m	Initial Weight:1263.1368g	Prep/Purc. Date:02/09/2011	Expiration Date:02/09/2014 Initial Volume:999,950.00uL
	<u>CAS Number</u> 75-15-0	<u>Description</u> CARBON DISULFIDE		<u>Final Conc</u> 1262631.59ug/mL	



## Galson Laboratories - Standards Report

Lot: IH196262

03/28/2011

IH196262	IG-PERC311-9 --- CCV/DLS	Analyst: MHERMANN Prep Date: 03/15/2011 Expiration Date: 06/15/2011
Final Volume: 2.01mL /13129	TETRACHLOROETHYLENE	Initial Conc: 9,874.56ug/mL Initial Weight: 0.0198g <u>CAS Number</u> 127-18-4 <u>Description</u> TETRACHLOROETHYLENE <u>Prep/Purc. Date</u> : 12/17/2009 <u>Expiration Date</u> : 12/17/2012 <u>Initial Volume</u> : 12.20uL <u>Final Conc</u> 9873.57ug/mL
/IH195802	CS2 W/UNDECANE	Initial Final Volume: 1,000.00 mL <u>Analyst</u> : MHERMANN Initial Weight: 1.2543g <u>Prep/Purc. Date</u> : 03/11/2011 <u>Expiration Date</u> : 01/30/2012 <u>Initial Volume</u> : 993.11uL
/IH195802/11529	UNDECANE	Initial Conc: 37.00ug/mL Initial Weight: 0.0370g <u>CAS Number</u> 1120-21-4 <u>Description</u> Undecane <u>Prep/Purc. Date</u> : 01/30/2009 <u>Expiration Date</u> : 01/30/2012 <u>Initial Volume</u> : 50.00uL <u>Final Conc</u> 18.23ug/mL
/IH195802/15573	CS2 517	Initial Conc: 1,263,136.84ug/m Initial Weight: 1263.1368g <u>CAS Number</u> 75-15-0 <u>Description</u> CARBON DISULFIDE <u>Prep/Purc. Date</u> : 02/09/2011 <u>Expiration Date</u> : 02/09/2014 <u>Initial Volume</u> : 999,950.00uL <u>Final Conc</u> 625355.86ug/mL
IH196262-1	100X DILUTION	Analyst: MHERMANN Prep Date: 03/15/2011 Expiration Date: 04/15/2011
Final Volume: 1,000.00uL /IH196262/13129	TETRACHLOROETHYLENE	Initial Conc: 9,873.57ug/mL Initial Weight: 0.0198g <u>CAS Number</u> 127-18-4 <u>Description</u> TETRACHLOROETHYLENE <u>Prep/Purc. Date</u> : 12/17/2009 <u>Expiration Date</u> : 12/17/2012 <u>Initial Volume</u> : 10.00uL <u>Final Conc</u> 98.74ug/mL
/IH196262/IH195802	CS2 W/UNDECANE	Initial Final Volume: 1,000.00 mL <u>Analyst</u> : MHERMANN Initial Weight: 1.2543g <u>Prep/Purc. Date</u> : 03/11/2011 <u>Expiration Date</u> : 01/30/2012 <u>Initial Volume</u> : 10.00uL
/IH196262/IH195802/11 529	UNDECANE	<u>Prep/Purc. Date</u> : 01/30/2009 <u>Expiration Date</u> : 01/30/2012
/IH196262/IH195802/15 573	CS2 517	Initial Conc: 18.23ug/mL Initial Weight: 0.0370g <u>CAS Number</u> 1120-21-4 <u>Description</u> Undecane <u>Prep/Purc. Date</u> : 02/09/2011 <u>Expiration Date</u> : 02/09/2014 <u>Initial Volume</u> : 10.00uL <u>Final Conc</u> 0.18ug/mL
IH196262-2	CCV(C15)	Analyst: MHERMANN Prep Date: 03/15/2011 Expiration Date: 04/15/2011
Final Volume: 1,999.97uL /IH196262-2/IH196262-1 /IH196262/13129	TETRACHLOROETHYLENE	Initial Conc: 625,355.86ug/mL Initial Weight: 1263.1368g <u>CAS Number</u> 75-15-0 <u>Description</u> CARBON DISULFIDE <u>Prep/Purc. Date</u> : 12/17/2009 <u>Expiration Date</u> : 12/17/2012 <u>Initial Volume</u> : 10.00uL <u>Final Conc</u> 6253.56ug/mL
		Initial Conc: 98.74ug/mL Initial Weight: 0.0198g <u>CAS Number</u> 127-18-4 <u>Description</u> TETRACHLOROETHYLENE <u>Initial Volume</u> : 20.00uL <u>Final Conc</u> 0.99ug/mL

/IH196262-2/IH196262-1	CS2 W/UNDECANE	Analyst:MHERMANN	Prep/Purc. Date:03/11/2011 Expiration Date:01/30/2012
/IH196262/IH195802	Initial Final Volume:1,000.00 mL	Initial Weight:1.2543g	Initial Volume:20.00uL
/IH196262-2/IH196262-1	UNDECANE		Prep/Purc. Date:01/30/2009 Expiration Date:01/30/2012
/IH196262/IH195802/11			
529			
	Initial Conc:0.18ug/mL	Initial Weight:0.0370g	Initial Volume:20.00uL
	<u>CAS Number</u> 1120-21-4	<u>Description</u> Undecane	<u>Final Conc</u> 0.0018ug/mL
/IH196262-2/IH196262-1	CS2 517		Prep/Purc. Date:02/09/2011 Expiration Date:02/09/2014
/IH196262/IH195802/15			
573			
	Initial Conc:6,253.56ug/mL	Initial Weight:1263.1368g	Initial Volume:20.00uL
	<u>CAS Number</u> 75-15-0	<u>Description</u> CARBON DISULFIDE	<u>Final Conc</u> 62.54ug/mL
IH196262-3	DLS	Analyst:MHERMANN	Prep Date:03/15/2011 Expiration Date:04/15/2011
Final Volume:2.043.55uL			
/IH196262-2/IH196262/1	TETRACHLOROETHYLENE		Prep/Purc. Date:12/17/2009 Expiration Date:12/17/2012
3129			
	Initial Conc:0.99ug/mL	Initial Weight:0.0198g	Initial Volume:62.00uL
	<u>CAS Number</u> 127-18-4	<u>Description</u> TETRACHLOROETHYLENE	<u>Final Conc</u> 0.030ug/mL
/IH196262-2/IH196262/I	CS2 W/UNDECANE	Analyst:MHERMANN	Prep/Purc. Date:03/11/2011 Expiration Date:01/30/2012
H195802	Initial Final Volume:1,000.00 mL	Initial Weight:1.2543g	Initial Volume:62.00uL
/IH196262-2/IH196262/I	UNDECANE		Prep/Purc. Date:01/30/2009 Expiration Date:01/30/2012
H195802/11529			
	Initial Conc:0.00ug/mL	Initial Weight:0.0370g	Initial Volume:62.00uL
	<u>CAS Number</u> 1120-21-4	<u>Description</u> Undecane	<u>Final Conc</u> 0.000055ug/mL
/IH196262-2/IH196262/I	CS2 517		Prep/Purc. Date:02/09/2011 Expiration Date:02/09/2014
H195802/15573			
	Initial Conc:62.54ug/mL	Initial Weight:1263.1368g	Initial Volume:62.00uL
	<u>CAS Number</u> 75-15-0	<u>Description</u> CARBON DISULFIDE	<u>Final Conc</u> 1.9ug/mL