ANNEX F % LIPIDS DATA SHEET

Shealy Environmental Services, Inc.

SDG Narrative 12/12/2016 Case 46613 SDG BD6Q5 SOW: SOM02.3 Contract Number: EPW14035 Solicitation: 2901 MA: 2672.0

PEST/Aroclor	Water sample concentration ug/L = $\frac{(Ax)(Vt)(DF)(GPC)}{(CD)(Tt)(Tt)}$
Equation	(CF)(V_0)(Vi)
	Soil sample concentration (ug/Kg) = $\frac{(A_x)(V_i)(DF)(GPC)}{(\overline{CF})(V_i)(W_i)(D)}$
	Where
	Ar is the response (peak area) of the compound to be measured.
	$\overline{\mathrm{CF}}$ is the mean calibration factor from the initial calibration (area/ng).
	DF is the dilution factor.
e (11)	$GPC = V_{in}/V_{out}$: GPC factor.
	V _{in} is the volume of extract loaded onto GPC column.
	Vour is the volume of extract collected after GPC cleanup.
	V ₁ is volume of the concentrated extract in uL. (If no GPC cleanup is performed, then $V_t = 1000$ µL. If GPC cleanup is performed, then $V_t = V_{ext}$.).
	V_t is the volume of the extract injected in uL.
	V _a : Volume of water extracted in mL.
	Ws is the weight of sample extracted in g
	$D = \frac{100 - \%Moisture}{100 - \%Moisture}$
	100

Columns	PEST/Aroclor DB-35MS 30m x 0.32mm x 0.25um	
	PEST/Aroclor DB-XLB 30m x 0.32mm x 0.50um	

Sample Receiving

The seven fish samples in this SDG were received in sealed shipping container on November 22, 2016 with temperature of 1.9°C.

All temperatures are determined using a calibrated Fluke 66 IR thermometer.

For a cross reference of laboratory ID to EPA Sample Numbers, please see SDG Cover Page on page 3 of this CSF file.

Sample BD6P0 is listed on the COC but not received.

Sample BD6S2 is a field duplicate of sample BD6R5.

The Fish information sheet contains site information. There is no conflict of interest for the lab.

Due to limited amount of samples, %Solid could not be performed for the following samples: BD6R4, BD6R5, BD6R7, BD6R8, BD6R9 and BD6S0.

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The samples were stored in freezer at below -10C after receipt and processed in accordance to MA 2672.0

Lab Sample ID	EPA Sample ID	%Lipids	%Solids
RK22011-001	BD6Q5	0.383	21.8
RK22011-002	BD6Q6	0.367	21.7
RK22011-003	BD6Q7	1.44	22.9
RK22011-004	BD6Q8	0.824	22.1
RK22011-005	BD6S0	2.98	N/A
RK22011-006	BD6Q9	1.01	22.1
RK22011-007	BD6R3	1.22	19.8
RK22011-008	BD6R4	1.09	N/A
RK22011-009	BD6R5	1.62	N/A
RK22011-010	BD6R6	0.714	20.1
RK22011-011	BD6R7	1.26	N/A
RK22011-012	BD6R8	1.81	N/A
RK22011-013	BD6R9	1.69	N/A
RK22011-014	BD6S2	0.917	20.3

Aroclor Fraction

1.6ug each of Aroclor 1016 and 1260 were added to matrix spike and matrix spike duplicate samples. 0.4ug each of Aroclor 1016 and 1260 were added to laboratory control sample.

With the approval from Region 2, 1:1 Acetone/Methylene chloride mix was used as extraction solvent, instead of 1:1 Acetone/Hexane as required in MA 2672.0, so that GPC cleanup can be performed on the extracts.

Due to limited amount of samples for some of the samples, all samples extracts in this SDG were concentrated to 2mL final volume after GPC cleanup to meet required CRQLs.

See the attached Manual Integration Report for a listing of all manual integrations associated with the samples and standards in this SDG. Unless otherwise noted manual integrations were performed due to incorrect auto integration.

As per the SOW, an example calculation is attached for Tetrachloro-m-xylene on the DB-35MS column in sample BD6Q5.

ANNEX G TRIP REPORT

SAMPLING TRIP REPORT

Site Name: CERCLIS ID Number: Sampling Date: CLP Case Number: Site Location: Richardson Hill Rd LF Site NYD 00 17-20 October 2016 46613 Sidney, New York

1. Laboratories Receiving Samples:

Sample Type	Laboratory Code	Name and Address of Laboratory
Surface Water and Sediment for PCBs	SHEALY	106 Vantage Point Drive West Columbia, SC 29172
Fish Tissue Samples for PCBs	SHEALY	106 Vantage Point Drive West Columbia, SC 29172

2. Sample Dispatch Data:

The total numbers of environmental samples delivered to the laboratory via OSCAR Station were 7 Surface Water and Sediment samples. This number included BD6M4 a duplicate water sample of BD6M3 & BD6S5 a duplicate sediment sample of BD6S54. A rinsate blank (RB) was not needed due to the sampling was collect via direct filled and no equipment was used. Sample numbers BD6M3 is the MS/MSDs for Surface Water and BD6S4 is the MS/MSDs for Sediment samples during this sampling event. The total numbers of individual fishes collected is 84. These fish samples are awaiting a lab assignment. Until we receive a lab assignment these fishes will be secured and maintain frozen in the USEPA Laboratory in Edison, NJ.

On October 17, 2016, Robert Finke, Charren Cabaroy, James Kurtenbach, Robert Morrell and I traveled to the site and met with personnel at the treatment plant and sign-in. We then conducted a recon of the South Pond (GP) and the Herrick Hollow Creek (HC). We observed that there was flowing water and that there was a possibility of collect fish samples. After the recon we went back to the plant to sign-out but it was closed and we were not able to sign-out.

On October 18, 2016, met with Ronald Chiarello from O'Brien & Gere (OBG) at the Plant. The sign-in sheet was taped to the door of the plant. We all sign-in and tape the sign-in sheet back on to the door. Mr. Chiarello, from OBG was assigned to assist us during the event. We traveled to South Pond and while gearing up to collect samples, we met Representatives from NYDEP and the Site RPM (Pam Tames). Both the RPM and the NYDEP representative observed the sampling procedures. DEP requested that we collect additional fish samples for analysis. We stated we would try to collect additional fish samples with the understating that the creek was very low and the number of fish samples could be limited. At South Pond we collected 10 Creek

Chub and 14 Pumpkin Seed Sunfish. These fish samples were divided into the number of sample groups requested by DEP and the Duplicate Blind Sample as well as the MS/MSD sample requirement. We also, collected Sediment and Water samples from South Pond. After collecting and processing the fish samples we took a lunch break and at the time DEP representative departed. After lunch we returned to the site and collected Sediment and Water samples from the furthermost downstream point in Herrick Hollow Creek (HC). At is location we collected Creek Chubs and Brook Trouts. The size and number of the Trouts collect were small. The two largest Trout collected were 181 mm and 158 mm all others were under 150 mm in size. We collected a total of 9 Trouts ranging from 125 to 181. We also, collected Sediment and Water samples from HC6. Samples were process, secured and place immediately in the sample cooler with ice.

On October 19, 2016, we met with Ronald Chiarello at the Plant. Signed the sign-in sheet and then, went to sampling location HC5. At HC5 we collected 10 Creek Chub and after processing the fishes, we then went to sampling location HC4. At HC4 we collected 9 Creek Chub and after processing the fishes, we then went to sampling location HC3. At HC3 we collected 13 Creek Chub and after processing the fishes, we then went to the last sampling location HC2 on Herrick Hollow Creek. At HC2 we collected 10 Creek Chub these were also process. At all the Fish sample location we also collect Sediment and Water samples. After processing all the samples, they were secured and place immediately in the sample cooler with ice.

On October 20, 2016, met with Ronald Chiarello at the Plant. Signed the sign-in sheet and placed the sign-in sheet as requested in the plants mail box. Mr Chiarello then guided us to the two sample location TC1 and TC2. As we arrived at TC1 it started to rain hard. Due to the rain it was not safe to use the electro-fishing gear. By 10:00 AM, it was still raining hard and now it was thundering. The weather condition was not safe to sample and I made a safety call, ending the sampling event without collecting samples from TC1 and TC2. We packed up and departed the area, returning to Edison, NJ with all samples secured and on ice.

At Edison, we off-loaded the fish samples and place them is the EPA laboratory sample freezer. They will be there, secured and frozen until we are notify as to where to ship them. As for the Sediment and Water samples, these were pack in two cooler with ice and shipped to Shealy Environmental Services an EPA CLP laboratory for analysis. These samples were shipped overnight express via UPS and were received the next day.

On November 21, 2016, at Edison, NJ we received the dry ice needed to ship the frozen fish samples to the laboratories for analysis. The fish samples were taken from the EPA laboratory samples freezer and packed with dry ice in a two coolers, and secured for shipping. One cooler was shipped to NYSDEC Hale Creek Field Station with sample group# 2017-FS-HC1-GP-002-PS (CLP#BD6R1) for analysis. The other cooler was shipped to Shealy Environmental Services an EPA CLP laboratory with all the other fish samples for analysis. These samples were shipped overnight express via UPS and were received the next day.

3. Sample Descriptions:

And the second			SEDIMENT SA		-	
Sample Location	Sample#	CLP#	DTG	Analysis ²	Laboratory	Remarks
South Pond HC1	2017-SED-HC1-SP1	BD6S4	10/18/16-1020	PCBs Aroclors	SHEALY	MS/MSD
South Pond HC1	2017-SEDSP2	BD6S5	10/18/16-1022	PCBs Aroclors	SHEALY	Dupl of BD6S4
HC2	2017-SED-HC2	BD6S6	10/19/16-1625	PCBs Aroclors	SHEALY	
HC3	2017-SED-HC3	BD6S7	10/19/16-1420	PCBs Aroclors	SHEALY	
HC4	2017-SED-HC4	BD6S8	10/19/16-1115	PCBs Aroclors	SHEALY	
HC5	2017-SED-HC5	BD6S9	10/19/16-0945	PCBs Aroclors	SHEALY	
HC6			1			
нсо	2017-SED-HC6	BD6T0	10/18/16-1535	PCBs Aroclors	SHEALY	
<u>c 1</u>			RFACE WATER			
Sample Location	Sample#	CLP#	DTG	Analysis ²	Laboratory	Remarks
South Pond HC1	2017-WAT-HC1-SP1	BD6M3	10/18/16-1020	PCBs Aroclors	SHEALY	MS/MSD
South Pond HC1	2017-WAT-SP2	BD6M4	10/18/16-1022	PCBs Aroclors	SHEALY	Dupl of BD6M3
HC2	2017-WAT-HC2	BD6M5	10/19/16-1610	PCBs Aroclors	SHEALY	•
HC3	2017-WAT-HC3	BD6M6	10/19/16-1420	PCBs Aroclors	SHEALY	
HC4	2017-WAT-HC4	BD6M7	10/19/16-1100	PCBs Aroclors	SHEALY	
HC5	2017-WAT-HC5	BD6M8	10/19/16-0930	PCBs Aroclors	SHEALY	
HC6	2017-WAT-HC6	BD6M9	10/18/16-1535			
nco				PCBs Aroclors	SHEALY	
FISH SAMPLES (Creek Chub)						Length(mm)/Weight(g)
Sample Location	Sample#	CLP#	DTG	Analysis ²	Laboratory	Remarks
South Pond	2017-FS-HC1-GP-	BD6N0	10/18/16-1020	PCBs Aroclors	SHEALY	123/22
HC1	001-SEC-1-CC	220110	10/10/10 1020	1 CD5 / Hotoris	STILLIET	123/22
South Pond HC1	2017-FS-HC1-GP- 002-SEC-1-CC	BD6N1	10/18/16-1022	PCBs Aroclors	SHEALY	127/22
South Pond HC1	2017-FS-HC1-GP- 003-SEC-1-CC	BD6N2	10/19/16-1610	PCBs Aroclors	SHEALY	121/20
South Pond	2017-FS-HC1-GP-	BD6N3	10/19/16-1420	PCBs Aroclors	SHEALY	113/16
HC1	004-SEC-1-CC	DDOINS	10/19/10-1420	r CDS Alociois	SHEALI	115/10
South Pond	2017-FS-HC1-GP-	BD6N4	10/19/16-1100	PCBs Aroclors	SHEALY	76/6
HC1	005-SEC-1-CC 2017-FS-HC1-GP-	-	10/10/16 0020	DOD- Amelian	OUPALX	7516
	005-SEC-2-CC		10/19/16-0930	PCBs Aroclors	SHEALY	75/6
	2017-FS-HC1-GP-		10/18/16-1020	PCBs Aroclors	SHEALY	66/4
	005-SEC-3-CC		10/10/10 1020	I CD3 / HOCIOIS	SHEALT	00/4
	2017-FS-HC1-GP-		10/18/16-1022	PCBs Aroclors	SHEALY	66/4
	005-SEC-4-CC					
	2017-FS-HC1-GP-		10/19/16-1610	PCBs Aroclors	SHEALY	. 70/4
100	005-SEC-5-CC					
HC2	2017-FS-HC2-GP-	BD6N5	10/19/16-1100	PCBs Aroclors	SHEALY	122/21
HC2	001-SEC-1-CC 2017-FS-HC2-GP-	BD6N6	10/10/16 0020	DCDg Angeler	CHEAT V	117/16
	002-SEC-1-CC	DUOINO	10/19/16-0930	PCBs Aroclors	SHEALY	117/16
	2017-FS-HC2-GP-	1	10/18/16-1535	PCBs Aroclors	SHEALY	126/22
		1	10/10/10/10/10/00	- CDS AUDUIS	SILCALI	120/22

HC2	2017-FS-HC2-GP-	BD6N7	10/18/16-1535	PCBs Aroclors	SHEALY	116/16
	003-SEC-1-CC					
	2017-FS-HC2-GP-		10/19/16-1100	PCBs Aroclors	SHEALY	112/16
	003-SEC-2-CC					
HC2	2017-FS-HC2-GP-	BD6N8	10/19/16-0930	PCBs Aroclors	SHEALY	117/18
	004-SEC-1-CC					
	2017-FS-HC2-GP-		10/18/16-1535	PCBs Aroclors	SHEALY	105/12
	004-SEC-2-CC					
HC2	2017-FS-HC2-GP-	BD6N9	10/18/16-1535	PCBs Aroclors	SHEALY	100/13
	005-SEC-1-CC					
	2017-FS-HC2-GP-		10/18/16-1535	PCBs Aroclors	SHEALY	76/9
a	005-SEC-2-CC					
	2017-FS-HC2-GP-		10/19/16-1100	PCBs Aroclors	SHEALY	80/6
	005-SEC-3-CC					
HC3	2017-FS-HC3-GP-	BD6P0	10/19/16-0930	PCBs Aroclors	SHEALY	129/18
	001-SEC-1-CC					
	2017-FS-HC3-GP-	-	10/18/16-1535	PCBs Aroclors	SHEALY	109/11
	001-SEC-2-CC					
НС3	2017-FS-HC3-GP-	BD6P1	10/18/16-1535	PCBs Aroclors	SHEALY	120/18
nes	002-SEC-1-CC	DDOIT	10/10/10 1000			
	2017-FS-HC3-GP-		10/19/16-1100	PCBs Aroclors	SHEALY	120/20
	002-SEC-2-CC					
HC3	2017-FS-HC3-GP-	BD6P2	10/19/16-0930	PCBs Aroclors	SHEALY	113/14
nes	003-SEC-1-CC	DD012		1 CD5 / HOURS		
	2017-FS-HC3-GP-	-	10/18/16-1535	PCBs Aroclors	SHEALY	100/12
	003-SEC-2-CC		10/10/10-1555		STILLE	100/12
НСЗ	2017-FS-HC3-GP-	BD6P3	10/18/16-1535	PCBs Aroclors	SHEALY	111/14
HC3	and a second second state the second second	BD0F5	10/10/10-1555	T CDS AIOCIOIS	SILALI	111/14
	004-SEC-1-CC		10/18/16-1535	PCBs Aroclors	SHEALY	99/11
	2017-FS-HC3-GP-		10/18/10-1555	FCBS AIOCIOIS	SILALI	99/11
1102	004-SEC-2-CC	BD6P4	10/18/16-1535	PCBs Aroclors	SHEALY	83/7
HC3	2017-FS-HC3-GP-	BD0P4	10/18/10-1355	FCDS AIOCIOIS	SILALI	85/7
	005-SEC-1-CC	_	10/19/16-0930	PCBs Aroclors	SHEALY	74/5
	2017-FS-HC3-GP-		10/19/10-0930	FCDS AIOCIOIS	SILALI	7475
	005-SEC-2-CC		10/18/16-1535	PCBs Aroclors	SHEALY	77/5
	2017-FS-HC3-GP-		10/18/10-1333	PCDS AIOCIOIS	SHEALI	1115
	005-SEC-3-CC		10/10/10 1525	DCDs Amalans	SHEALY	66/4
	2017-FS-HC3-GP-	S	10/18/16-1535	PCBs Aroclors	SHEALY	00/4
	005-SEC-4-CC		10/10/16 1100	DCD A lan	OUPALY	50/2
	2017-FS-HC3-GP-		10/19/16-1100	PCBs Aroclors	SHEALY	59/3
	005-SEC-5-CC	DDCDE	10/10/16 0000		CUEALV	122/22
HC4	2017-FS-HC4-GP-	BD6P5	10/19/16-0930	PCBs Aroclors	SHEALY	133/22
	001-SEC-1-CC		10/10/16 1525			105/11
HC4	2017-FS-HC4-GP-	BD6P6	10/18/16-1535	PCBs Aroclors	SHEALY	105/11
	002-SEC-1-CC		10/10/16 1505		CHEAL M	102/10
	2017-FS-HC4-GP-		10/18/16-1535	PCBs Aroclors	SHEALY	123/19
	002-SEC-2-CC		10/10/14 6 1 50 5			105/00
HC4	2017-FS-HC4-GP-	BD6P7	10/18/16-1535	PCBs Aroclors	SHEALY	125/20
	003-SEC-1-CC		10/11/11/1		OTTO A CAN	10(/12
	2017-FS-HC4-GP-		10/18/16-1535	PCBs Aroclors	SHEALY	106/12
	003-SEC-2-CC					115/15
HC4	2017-FS-HC4-GP-	BD6P8	10/19/16-0930	PCBs Aroclors	SHEALY	117/15
	004-SEC-1-CC					
	2017-FS-HC4-GP-		10/18/16-1535	PCBs Aroclors	SHEALY	112/14
	004-SEC-2-CC					
HC4	2017-FS-HC4-GP-	BD6P9	10/18/16-1535	PCBs Aroclors	SHEALY	111/11

	005-SEC-1-CC	1	1	Т	1	
	2017-FS-HC4-GP-	_	10/10/16 1525			
	005-SEC-2-CC		10/18/16-1535	PCBs Aroclors	SHEALY	110/13
HC5	2017-FM-HC5-GP- 001-SEC-1-CC	BD6Q0	10/18/16-1535	PCBs Aroclors	SHEALY	134/20 MS/MSD
	2017-FM-HC5-GP- 001-SEC-2-CC		10/18/16-1535	PCBs Aroclors	SHEALY	137/24
a. K	2017-FM-HC5-GP- 002-SEC-1-CC		10/18/16-1535	PCBs Aroclors	SHEALY	136/23
HC5	2017-FS-HC5-GP- 003-SEC-1-CC	BD6Q2	10/18/16-1535	PCBs Aroclors	SHEALY	120/17
	2017-FS-HC5-GP- 003-SEC-2-CC		10/18/16-1535	PCBs Aroclors	SHEALY	108/13
HC5	2017-FS-HC5-GP- 004-SEC-1-CC	BD6Q3	10/18/16-1535	PCBs Aroclors	SHEALY	114/14
	2017-FS-HC5-GP- 004-SEC-2-CC		10/18/16-1535	PCBs Aroclors	SHEALY	94/9
HC5	2017-FS-HC5-GP- 005-SEC-1-CC	BD6Q4	10/18/16-1535	PCBs Aroclors	SHEALY	95/9
	2017-FS-HC5-GP- 005-SEC-2-CC]	10/18/16-1535	PCBs Aroclors	SHEALY	88/7
	2017-FS-HC5-GP- 005-SEC-3-CC		10/18/16-1535	PCBs Aroclors	SHEALY	89/7
HC6	2017-FS-HC6-GP- 001-SEC-1-CC	BD6Q5	10/18/16-1535	PCBs Aroclors	SHEALY	157/38
HC6	2017-FS-HC6-GP- 002-SEC-1-CC	BD6Q6	10/18/16-1535	PCBs Aroclors	SHEALY	120/18
	2017-FS-HC6-GP- 002-SEC-2-CC		10/18/16-1535	PCBs Aroclors	SHEALY	107/14
HC6	2017-FS-HC6-GP- 003-SEC-1-CC	BD6Q7	10/18/16-1535	PCBs Aroclors	SHEALY	119/16
	2017-FS-HC6-GP- 003-SEC-2-CC		10/18/16-1535	PCBs Aroclors	SHEALY	105/12
HC6	2017-FD-HC6-GP- 004-SEC-1-CC	BD6Q8/ BD6S0	10/18/16-1535	PCBs Aroclors	SHEALY	141/28 DUPLICATE
	2017-FD-HC6-GP- 004-SEC-2-CC		10/18/16-1535	PCBs Aroclors	SHEALY	113/16
HC6	2017-FS-HC6-GP- 005-SEC-1-CC	BD6Q9	10/18/16-1535	PCBs Aroclors	SHEALY	120/18
	2017-FS-HC6-GP- 005-SEC-2-CC		10/18/16-1535	PCBs Aroclors	SHEALY	116/16

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Sample Location	Sample#	CLP#	DTG	Analysis ²	Laboratory	Remarks
South Pond HC1	2017-FM-HC1-GP- 001-SEC-1-PS	BD6R0	10/19/16-1420	PCBs Aroclors	SHEALY	132/46 MS/MSD
	2017-FM-HC1-GP- 001-SEC-2-PS		10/19/16-1100	PCBs Aroclors	SHEALY	125/38
	2017-FM-HC1-GP- 001-SEC-3-PS		10/19/16-0930	PCBs Aroclors	SHEALY	114/26
	2017-FM-HC1-GP- 001-SEC-4-PS		10/18/16-1535	PCBs Aroclors	SHEALY	103/18
	2017-FM-HC1-GP- 001-SEC-5-PS	-	10/18/16-1020	PCBs Aroclors	SHEALY	117/32
South Pond IC1	2017-FS-HC1-GP- 002-SEC-1-PS	BD6R1	10/18/16-1022	PCBs Aroclors	SHEALY	117/28 SHIPPED TO NYSDEC
	2017-FS-HC1-GP- 002-SEC-2-PS		10/19/16-1610	PCBs Aroclors	SHEALY	91/14
	2017-FS-HC1-GP- 002-SEC-3-PS		10/19/16-1420	PCBs Aroclors	SHEALY	110/26
	2017-FS-HC1-GP- 002-SEC-4-PS	-	10/19/16-1100	PCBs Aroclors	SHEALY	83/12
	2017-FS-HC1-GP- 002-SEC-5-PS	_	10/19/16-1100	PCBs Aroclors	SHEALY	82/10
	2017-FD-HC1-GP- 003-SEC-1-PS	BD6R2/ BD6S1	10/19/16-0930	PCBs Aroclors	SHEALY	90/14 DUPLICATE
	2017-FD-HC1-GP- 003-SEC-2-PS		10/18/16-1020	PCBs Aroclors	SHEALY	98/18
	2017-FD-HC1-GP- 003-SEC-3-PS	-	10/18/16-1022	PCBs Aroclors	SHEALY	90/12
	2017-FD-HC1-GP- 003-SEC-4-PS	1	10/19/16-1610	PCBs Aroclors	SHEALY	80/10
	2017-FD-HC1-GP- 003-SEC-5-PS		10/19/16-1420	PCBs Aroclors	SHEALY	78/8
	005 520 5 1 5	FI	SH SAMPLES (Brook Trout)		
Sample Location	Sample#	CLP#	DTG	Analysis ²	Laboratory	Remarks
HC6	2017-PS-HC6-GP- 001-SEC-1-BT	BD6R3	10/18/16-1535	PCBs Aroclors	SHEALY	181/54 MS/MSD
HC6	2017-PS-HC6-GP- 001-SEC-2-BT	BD6R6	10/18/16-1535	PCBs Aroclors	SHEALY	158/40
HC6	2017-PS-HC6-GP- 001-SEC-3-BT	BD6R7	10/18/16-1535	PCBs Aroclors	SHEALY	148/30
HC6	2017-PS-HC6-GP- 002-SEC-1-BT	BD6R4	10/18/16-1535	PCBs Aroclors	SHEALY	139/30
HC6	2017-PS-HC6-GP- 002-SEC-2-BT	BD6R8	10/18/16-1535	PCBs Aroclors	SHEALY	137/28
HC6	2017-PS-HC6-GP- 002-SEC-3-BT	BD6R9	10/18/16-1535	PCBs Aroclors	SHEALY	125/26
HC6	2017-PS-HC6-GP- 003-SEC-1-BT	BD6R5/ BD6S2	10/18/16-1535	PCBs Aroclors	SHEALY	134/24 DUPLICATE
	2017-PS-HC6-GP-		10/18/16-1535	PCBs Aroclors	SHEALY	129/20
	2017-PS-HC6-GP-		10/18/16-1535	PCBs Aroclors	SHEALY	126/20
			Notes			
locations) – (GF trout)	003-SEC-2-BT	ps) – (SEC-1,2,3	10/18/16-1535 Notes ge species, PS= predat 3,4,or 5= individual fit	PCBs Aroclors	SHEALY	126/20 - (HC1,2,3,4,5,6= Stat

4. Sampling Personnel:

Name	Organization	Site Duties
Michael A. Mercado	USEPA Region II DESA/HWSB	Project Manager/Sample Management
James Kurtenbach	USEPA Region II DESA/MAB	Aquatic Biologist
Robert Finke	USEPA Region II DESA/HWSB	Field Personnel
Robert Morrell	USEPA Region II DESA/MAB	Field Personnel
Charren Cabaroy	USEPA Region II DESA/HWSB	Field Personnel
Amelia Jackson	USEPA Region II DESA/HWSB	Quality Assurance Officer

5. Additional Comments:

Fish samples collected were:

A total of 9 Brook Trouts, 15 Pumpkin Seed Sunfish, and 60 Creek Chubs collected from South Pond and Herrick Hollow Creek.

Each individual fish was labeled by placing a sample identification tag in a plastic bag with the fish and are currently in the EPA DESA Laboratory being kept frozen until a Laboratory was identified to do the analysis.

Sediment samples collected were:

A total of 7 Sediment were collect from South Pond and Herrick Hollow Creek.

Surface Water samples collected were:

A total of 7 Surface Water samples were collect from South Pond and Herrick Hollow Creek.

Report Prepared By:

Michael A. Mercado

Date November 30, 2016

