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# **Work Plan for Natural Attenuation Monitoring**

Bush Industries, Inc.  
312 Fair Oak Street  
Little Valley, New York

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Project No. 6191

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**Geomatrix**

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## WORK PLAN FOR NATURAL ATTENUATION MONITORING

Bush Industries, Inc.  
312 Fair Oak Street  
Little Valley, New York

### 1.0 INTRODUCTION

#### 1.1 BACKGROUND

Geomatrix Consultants, Inc. (Geomatrix) has been retained by Bush Industries, Inc. (Bush Industries) to prepare this Work Plan for Natural Attenuation Monitoring (Work Plan) for the Bush Industries Property (Site) in Little Valley, New York. The location of the Site is shown on Figure 1. This Work Plan is required by and appended to the Amended and Supplemental Order (File No.: 96-07 R9-4314-96-06) agreed to between Bush Industries and the New York State Department of Environmental Conservation (NYSDEC).

The Site is located within the Little Valley Superfund Site (LVSS). The LVSS extends from the Village of Little Valley south to near the City of Salamanca, a distance of approximately 8 miles, and is currently being addressed by the United States Environmental Protection Agency (USEPA). A Remedial Investigation and Feasibility Study (RI/FS) has been prepared for the USEPA culminating in a Record of Decision (ROD) specifying monitored natural attenuation (MNA) as the remedy for TCE contaminated groundwater measured throughout the LVSS.

The USEPA MNA remedy includes groundwater sampling on properties located throughout the LVSS including the Bush Industries property. Bush Industries has agreed to conduct the MNA sampling on its property. This Work Plan describes the procedures to be used by Bush Industries in implementing the MNA sampling at its property.

#### 1.2 SITE DESCRIPTION

The Site is located at 312 Fair Oak Street in the Village of Little Valley, Cattaraugus County, New York. A topographic map of the Site and surrounding area prepared from a 7.5 minute series U.S. Geological Survey map is presented in Figure 1. The parcels comprising the Site have been purchased over a number of years. The Site is situated on a 9.4 acre lot, and contains three contiguous buildings (see Figure 2).

### **1.3 PREVIOUS SITE INVESTIGATIONS**

Bush Industries has conducted an extensive investigation of groundwater conditions at its Site. Results are documented in the report entitled Groundwater Evaluation Report, prepared by Conestoga-Rovers & Associates (CRA) and dated February 21, 2000. The findings presented the Groundwater Evaluation Report are summarized as follows:

1. The highest concentrations of TCE and its degradation products remain in the interior of the Site. There is a residual low level presence of TCE and its degradation products in the interior of the Site with concentrations in groundwater dropping precipitously along the downgradient flow path.
2. Concentrations of TCE at the downgradient perimeter of the Site are approximately equal to or below the New York State Groundwater criterion.
3. Benzene concentrations below method detection limits were estimated to be above the New York State groundwater criteria of 1 ug/L at two interior well locations (MW-D2 and MW-4) where concentrations of 2J (estimated) were reported for the May 1999 sampling event. Benzene was not above the New York State groundwater criteria in any well during the December 1999 sampling event.
4. This distribution trend (rapidly declining concentrations with distance from the interior of the Site) indicates that natural attenuation processes occur limiting constituent migration and the Site does not pose a significant threat to downgradient groundwater quality.

The Groundwater Evaluation Report was approved by NYSDEC by letter dated March 31, 2000.

In May 2000, Bush Industries submitted the Remediation Report prepared by Geomatrix which recommended implementation of a 5-year annual MNA sampling program at the Site. No action was taken with respect to recommendations of the Remediation Report pending completion of the LVSS RI/FS/ and ROD (see Section 1.1).

### **1.4 PROJECT OBJECTIVES**

The objectives of the natural attenuation monitoring are to:

1. Perform annual monitored natural attenuation (MNA) sampling events
2. Evaluate historic and new analytical data to monitor natural attenuation at the Site

## 2.0 SCOPE OF WORK

The MNA monitoring work to be performed at the Bush Industries Site is specified in the following USEPA documents:

*Final Remedial Action Work Plan for the Little Valley Superfund Site*

Contract Number:68-W-98-214

Prepared by Tetra Tech EC, Inc.

Dated October 2006

*Quality Assurance Project Plan Addendum for the Little Valley Superfund Site*

Contract Number:68-W-98-214

Prepared by Tetra Tech EC, Inc.

Dated September 2006

The USEPA Work Plan specifies the following for the Bush Industries Site:

1. Conduct two annual groundwater sampling events for the following wells:  
MW-D1, MW-D2, MW-2, MW-3, MW-6, and MW-8.
2. Analyze samples for the following MNA analyses: Volatile Organic Chemicals (VOCs), alkalinity, sulfate, sulfide, nitrate, chloride, total organic carbon, ferrous iron, ethane, ethene and methane.
3. Data validation.
4. Data evaluation.

Monitoring well MW-8 was destroyed by a snow plow during the winter of 2005-2006. In accordance with the USEPA Work Plan, this well will be replaced in the program by designated alternate well MW-5 (see Section 3.1.1, below).

The USEPA Work Plan specifies two annual MNA sampling events, the first of which has already been performed (October 2006). The Bush Industries Work Plan presented herein specifies nine annual MNA sampling events at its Site which (counting the October 2006 event conducted by USEPA) will constitute ten consecutive annual sampling events. This is consistent with the recommendations presented in the Bush

Industries Remediation Report (see Section 1.3), except the monitoring period has been extended from 5 annual events to 10 annual events.

## **3.0 PROJECT TASKS**

### **3.1 CONSISTENCY WITH USEPA WORK PLAN**

In order to facilitate direct comparison of the Bush Industries analytical results with results from other wells within the LVSS to be sampled by USEPA, the sampling methods, analytical methods and QA/QC protocols specified by USEPA for the LVSS remediation will be utilized for the Bush Industries MNA monitoring and are incorporated herein.

### **3.2 TASK 1: GROUNDWATER SAMPLING**

#### **3.2.1 Monitoring Wells to be Sampled**

The following Bush Industries monitoring wells will be sampled: MW-D1, MW-D2, MW-2, MW-3, MW-6, and MW-5. The rationale for selection of the six Bush Industries monitoring wells to be samples is presented in the USEPA Work Plan referenced above.

#### **3.2.2 Sample Identification System**

The sample identification system will be in accordance with the USEPA Work Plan. Sample identification will include LVRA for the Remedial Action MNA Program activities at the LVSS, a two digit number for the sampling event, a five letter code for the type of sampling being performed (MNAGW), and the location name. Sampling events or rounds will be numbered in sequence, as applicable. For example a groundwater sample collected from monitoring well MW-5 at the Bush Industries property for the third sampling event of the MNA program would be identified as “LVRA03-MNAGW-MW5.”

Duplicate samples (“blind duplicates”) will be “coded” in such a manner that the laboratory will not be able to determine the corresponding original field sample. An explanation of the duplicate “coding” will be written in the field logbook.

#### **3.2.3 Sample Collection**

The wells will be sampled using low flow methodology. The following Standard Operating Procedures (SOPs) obtained from the USEPA Work Plan will be used during sample collection:

- SOP # 1: Groundwater Sampling (Low Flow Purge Procedure)

- SOP # 2: Field Parameter Measurement
- SOP # 3: Water Level Measurements
- SOP # 4: Decontamination (Non-disposable Sampling Equipment)
- SOP # 5: Decontamination (Field Instrumentation-Probes, Water Quality Meters)

These SOPs are included in Attachment 1.

### **3.2.4 Sample Custody**

Immediately following sample collection, each sample container will be marked with the following information:

- Sample Identification
- Project Number
- Date/Time
- Sample Type
- Analyses to be performed
- Preservation Method
- Sampler's Initials

After all sample identification information has been recorded, each sample label will be covered with waterproof clear plastic tape to preserve its integrity. All samples will be recorded and tracked under strict chain-of-custody (COC) protocols.

## **3.3 TASK 2: GROUNDWATER ANALYSES**

As indicated above, the analytical parameter list and methods will be in accordance with the USEPA Work Plan. Groundwater samples will be analyzed for the following parameters: VOCs, alkalinity, sulfate, sulfide, nitrate, chloride, total organic carbon, ferrous iron, ethane, ethene and methane. The analytical program is summarized in Table 1 (from the USEPA Work Plan).

Groundwater samples will be analyzed by a laboratory deemed acceptable by NYSDEC. The selected laboratory(ies) will be subject to NYSDEC concurrence prior to being contracted to perform the analyses.

Quality Assurance and Quality Control (QA/QC) protocols will be as described in Section 4.0.

### **3.4      TASK 3: DATA VALIDATION**

The analytical results obtained for the laboratory(ies) will undergo a systematic data validation to provide assurance that the data is adequate for its intended use. The validation will be performed by personnel who have appropriate training and/or experience in performing data validation for the analyses of interest associated with this project.

Validation will be performed based on an evaluation of method specific QC information (such as holding times, calibration records, laboratory and field blanks, duplicate precision, and surrogate and matrix spike recoveries), the most current version of the USEPA Region 2 Data Validation SOPs ([www.epa.gov/region02/desa/hsw/sops.htm](http://www.epa.gov/region02/desa/hsw/sops.htm)), the most current version of the EPA National Functional Guidelines ([www.epa.gov/superfund/programs/clp/guidance.htm](http://www.epa.gov/superfund/programs/clp/guidance.htm)) and the best professional judgment of the validator. If USEPA has not defined acceptance criteria for a specific parameter or methodology (e.g., alkalinity, methane), the data will be reviewed/validated based on an evaluation of method-specific QC data and the best professional judgment of the validator.

Non-conforming QA/QC Results will be evaluated with respect to their implications for data reliability and usability following EPA Region 2 SOPs, EPA National Functional Guidelines and the best professional judgment of the validator. Qualifiers (as applicable) will be added to the data results.

### **3.5      TASK 4: DATA EVALUATION AND REPORTING**

#### **3.5.1    Annual Summary Reports**

The results of the Bush Industries annual MNA sampling will be evaluated and reported to the NYSDEC in annual summary reports. These reports will include the following:

- An evaluation of whether natural attenuation is occurring according to expectations (i.e., concentrations are decreasing over time).
- An evaluation of whether expansion of the plume is minimal to nonexistent.
- An evaluation of whether any additional receptors are impacted.
- An evaluation of whether any new releases of contaminants to the environment are detected (i.e. spike in concentrations).
- An evaluation of whether cleanup objective levels are being attained.
- Tables of current and historical volatile organic compounds (VOC) and MNA/water quality sampling data for the monitoring wells and piezometers sampled during the event.
- Maps showing the sampling locations.
- Water Table or piezometric head maps.
- Time-series plots of VOC concentrations of each of the Bush Industries monitoring wells on individual plots.
- Executive summary, text and conclusions.

### **3.5.2 Final Report**

After completion of the tenth consecutive groundwater MNA sampling event, a Final Report would be submitted to NYSDEC which will assess trends in chemical concentrations and present recommendations concerning:

1. The need for and/or requirements of continued monitoring; and
2. The need for and/or nature of continued prohibitions of on-site groundwater use.

#### **4.0 PROJECT QUALITY ASSURANCE AND QUALITY CONTROL**

Quality assurance and quality control protocols will be in accordance with the USEPA Quality Assurance Project Plan (USEPA QAPP) referenced in Section 2.0. The QA objectives and QA/QC protocols for the MNA sampling are summarized on the following tables (from the USEPA QAPP).

Table 1. QA Objectives for Field Investigation Data

Table 2. Reference Limits

Table 3. Sample Collection and Analysis Protocols

Table 4. Summary of Analytical QC Procedure Checks, Frequencies, Acceptance Criteria and Corrective actions for Field Screening and Laboratory Sample Analyses

## 5.0 PROJECT SCHEDULE

The first annual MNA sampling event at the Bush Industries Site was conducted by USEPA in October 2006 in accordance with the USEPA Work Plan and QAPP (and therefore in accordance with this Work Plan). The schedule for completion of the program is as follows:

| <i>Activity</i>            | <i>Target Completion Date</i>                               |
|----------------------------|---|
| 2006 MNA Sampling          | October 2006  |
| 2007 MNA Sampling          | Week of September 24, 2007                                  |
| 2007 Annual Summary Report | 90 days after completion of 2007 annual sampling            |
| 2008 MNA Sampling          | Coincident with USEPA 2008 LVSS MNA Sampling (if conducted) |
| 2008 Annual Summary Report | 90 days after completion of 2008 annual sampling            |
| 2009 MNA Sampling          | Coincident with USEPA 2009 LVSS MNA Sampling (if conducted) |
| 2009 Annual Summary Report | 90 days after completion of 2009 annual sampling            |
| 2010 MNA Sampling          | Coincident with USEPA 2010 LVSS MNA Sampling (if conducted) |
| 2010 Annual Summary Report | 90 days after completion of 2010 annual sampling            |
| 2011 MNA Sampling          | Coincident with USEPA 2011 LVSS MNA Sampling (if conducted) |
| 2011 Annual Summary Report | 90 days after completion of 2011 annual sampling            |
| 2012 MNA Sampling          | Coincident with USEPA 2012 LVSS MNA Sampling (if conducted) |
| 2012 Annual Summary Report | 90 days after completion of 2012 annual sampling            |

|                            |  |
|----------------------------|--|
| 2013 MNA Sampling          | Coincident with USEPA 2013 LVSS<br>MNA Sampling (if conducted) |
| 2013 Annual Summary Report | 90 days after completion of<br>2013 annual sampling            |
| 2014 MNA Sampling          | Coincident with USEPA 2014 LVSS<br>MNA Sampling (if conducted) |
| 2014 Annual Summary Report | 90 days after completion of<br>2014 annual sampling            |
| 2015 MNA Sampling          | Coincident with USEPA 2015<br>LVSS MNA Sampling (if conducted) |
| Final Report               | 90 days after completion of<br>2015 annual sampling            |

**TABLE 1**  
**QA OBJECTIVES FOR FIELD INVESTIGATION DATA**  
**BUSH INDUSTRIES SITE**

| <b>Parameter</b>              | <b>Measurement</b> | <b>Matrix</b> | <b>Method</b>            | <b>Units</b>        | <b>Precision</b>                        | <b>Accuracy</b>                | <b>PQL or Sensitivity</b>                      | <b>Completeness</b> |
|-------------------------------|--------------------|---------------|--------------------------|---------------------|---|--------------------------------|--|---------------------|
| Water Level                   | Screening          | Aqueous       | Direct Field Measurement | feet                | ± 0.01                                  | N/A                            | ± 0.01   | 95%                 |
| pH                            | Screening          | Aqueous       | Direct Field Measurement | Std. Units          | ± 0.1                                   | N/A                            | ± 0.1  | 95%                 |
| Temperature                   | Screening          | Aqueous       | Direct Field Measurement | °C                  | ± 0.1                                   | N/A                            | ± 0.1  | 95%                 |
| Specific Conductivity         | Screening          | Aqueous       | Direct Field Measurement | umhos/cm or mS/cm   | ± 1% of full scale*                     | N/A                            | ± 0.001  | 95%                 |
| Dissolved Oxygen              | Screening          | Aqueous       | Direct Field Measurement | mgO <sub>2</sub> /L | ± 3%                                    | N/A                            | ± 0.1  | 90%                 |
| Oxidation-Reduction Potential | Screening          | Aqueous       | Direct Field Measurement | mV                  | ± 10                                    | N/A                            | ± 1  | 90%                 |
| Turbidity                     | Screening          | Aqueous       | Direct Field Measurement | NTU                 | ± 2                                     | N/A                            | ± 2  | 90%                 |
| Low Concentration TCL VOCs    | Definitive         | Aqueous       | SOM01.1                  | µg/L                | Compound Specific (± 24% RPD; advisory) | Compound Specific (59 - 172%R) | Compound Specific (0.5-20 µg/L); see Table 3-6 | 90%                 |
| Total Organic Carbon          | Definitive         | Aqueous       | SW-846 method 9060       | mg/L                | ± 25% RPD                               | N/A                            | 1 mg/L   | 90%                 |
| Alkalinity                    | Definitive         | Aqueous       | MCAWW Method 310.1       | mg/L                | ± 25% RPD                               | N/A                            | 1 mg/L   | 90%                 |
| Sulfate                       | Definitive         | Aqueous       | EPA 300.1                | mg/L                | ± 20% RPD                               | 75-125%R                       | 1 mg/L   | 90%                 |

| <b>Parameter</b> | <b>Measurement</b> | <b>Matrix</b> | <b>Method</b>                  | <b>Units</b> | <b>Precision</b> | <b>Accuracy</b> | <b>PQL or Sensitivity</b> | <b>Completeness</b> |
|------------------|--------------------|---------------|--------------------------------|--------------|------------------|-----------------|---------------------------|---------------------|
| Sulfide          | Definitive         | Aqueous       | MCAWW<br>Method 376.1          | mg/L         | ± 25% RPD        | N/A             | 1 mg/L                    | 95%                 |
| Nitrate          | Definitive         | Aqueous       | EPA 300.1                      | mg/L         | ± 20% RPD        | 75-125%R        | 0.05 mg/L                 | 90%                 |
| Chloride         | Definitive         | Aqueous       | EPA 300.1                      | mg/L         | ± 20% RPD        | 75-125%R        | 1 mg/L                    | 90%                 |
| Ferrous Iron     | Definitive         | Aqueous       | Std. Methods<br>3500Fe-D       | µg/L         | ± 25% RPD        | N/A             | 10 µg/L                   | 90%                 |
| Ethane           | Definitive         | Aqueous       | GC/FID (SW-846<br>Method 3810) | µg/L         | ± 25% RPD        | N/A             | 5 µg/L                    | 90%                 |
| Ethene           | Definitive         | Aqueous       | GC/FID (SW-846<br>Method 3810) | µg/L         | ± 25% RPD        | N/A             | 5 µg/L                    | 90%                 |
| Methane          | Definitive         | Aqueous       | GC/FID (SW-846<br>Method 3810) | µg/L         | ± 25% RPD        | N/A             | 5 µg/L                    | 90%                 |

**NOTES:**

Method References:

SOM01.1 = USEPA Contract Laboratory Program Statement of Work for Multi-Media, Multi-Concentration Organics (May 2005 or latest revision).

MCAWW = Methods for Chemical Analysis of Water and Wastes, March 1983.

Std. Methods = Standard Methods for the Examination of Water and Wastewater, 20th Edition (January 2000).

SW-846 = Test Methods for Evaluating Solid Waste, Physical/Chemical Methods (November 1986, revised through November 2000 via updates I through IVB).

EPA 300.1 = Determination of Inorganic Anions in Drinking Water by Ion Chromatography, Revision 1 (27 April 1999).

Acronyms/abbreviations include:

%R = Percent Recovery

FID = Flame Ionization Detector

N/A = Not Applicable

PQL = Practical Quantitation Limit

VOCs = Volatile Organic Compounds

\*Precision dependent on meter and scale.

 GC = Gas Chromatograph  
 NTU = Nephelometric Turbidity Units  
 RPD = Relative Percent Difference  
 SD = Standard Deviation  
 TCL = Target Compound List

| <b>Analyte</b>                           | <b>CAS Number</b> | <b>Project Screening Criteria/Action Limit</b> | <b>Project Quantitation Limit</b> |
|--|-------------------|--|-----------------------------------|
| <b>LOW CONCENTRATION TCL VOCs (µg/L)</b> |                   |  |                                   |
| Dichlorodifluoromethane                  | 75-71-8           | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| Chloromethane                            | 74-87-3           | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| Vinyl Chloride                           | 75-01-4           | 2 (EPA MCL); 2 (NYSDEC)                        | 0.5                               |
| Bromomethane                             | 74-83-9           | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| Chloroethane                             | 75-00-3           | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| Trichlorofluoromethane                   | 75-69-4           | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| 1,1-Dichloroethene                       | 75-35-4           | 7 (EPA MCL); 2 (NYSDEC)                        | 0.5                               |
| 1,1,2-Trichloro-1,2,2-trifluoroethane    | 76-13-1           | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| Acetone                                  | 67-64-1           | -- (EPA MCL); 50 (NYSDEC)                      | 5                                 |
| Carbon Disulfide                         | 75-15-0           | -- (EPA MCL); 60 (NYSDEC)                      | 0.5                               |
| Methyl Acetate                           | 79-20-9           | -- (EPA MCL); -- (NYSDEC)                      | 0.5                               |
| Methylene Chloride                       | 75-09-2           | 5 (EPA MCL); 5 (NYSDEC)                        | 0.5                               |
| trans-1,2-Dichloroethene                 | 156-60-5          | 100 (EPA MCL); 5 (NYSDEC)                      | 0.5                               |
| Methyl ter-Butyl Ether                   | 1634-04-4         | 20 (EPA MCL); 10 (NYSDEC)                      | 0.5                               |
| 1,1-Dichloroethane                       | 75-34-3           | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| cis-1,2-Dichloroethene                   | 156-59-2          | 70 (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| 2-Butanone                               | 78-93-3           | -- (EPA MCL); 50 (NYSDEC)                      | 5                                 |
| Chloroform                               | 67-66-3           | 80 (EPA MCL); 7 (NYSDEC)                       | 0.5                               |
| 1,1,1-Trichloroethane                    | 71-55-6           | 200 (EPA MCL); 5 (NYSDEC)                      | 0.5                               |
| Cyclohexane                              | 110-82-7          | -- (EPA MCL); -- (NYSDEC)                      | 0.5                               |
| Carbon Tetrachloride                     | 56-23-5           | 5 (EPA MCL); 5 (NYSDEC)                        | 0.5                               |
| Benzene                                  | 71-43-2           | 5 (EPA MCL); 1 (NYSDEC)                        | 0.5                               |
| 1,2-Dichloroethane                       | 107-06-2          | 5 (EPA MCL); 0.6 (NYSDEC)                      | 0.5                               |
| 1,4-Dioxane                              | 123-91-1          | -- (EPA MCL); -- (NYSDEC)                      | 20                                |
| Trichloroethene                          | 79-01-6           | 5 (EPA MCL); 5 (NYSDEC)                        | 0.5                               |
| Methylcyclohexane                        | 108-87-2          | -- (EPA MCL); -- (NYSDEC)                      | 0.5                               |

| <i>Analyte</i>   | <i>CAS Number</i>   | <i>Project Screening Criteria/Action Limit</i> | <i>Project Quantitation Limit</i> |
|--|---------------------|--|-----------------------------------|
| <b>LOW CONCENTRATION TCL VOCs (<math>\mu\text{g/L}</math>) (continued)</b> |                     |  |                                   |
| 1,2-Dichloropropane  | 78-87-5             | 5 (EPA MCL); 1 (NYSDEC)                        | 0.5                               |
| Bromodichloromethane   | 75-27-4             | 80 (EPA MCL); 50 (NYSDEC)                      | 0.5                               |
| cis-1,3-Dichloropropene  | 10061-01-5          | -- (EPA MCL); 0.4 (NYSDEC)                     | 0.5                               |
| 4-Methyl-2-Pentanone   | 108-10-1            | -- (EPA MCL); -- (NYSDEC)                      | 5                                 |
| Toluene  | 108-88-3            | 1000 (EPA MCL); 5 (NYSDEC)                     | 0.5                               |
| trans-1,3-Dichloropropene  | 10061-02-6          | -- (EPA MCL); 0.4 (NYSDEC)                     | 0.5                               |
| 1,1,2-Trichloroethane  | 79-00-5             | 5 (EPA MCL); 1 (NYSDEC)                        | 0.5                               |
| Tetrachloroethylene  | 127-18-4            | 5 (EPA MCL); 5 (NYSDEC)                        | 0.5                               |
| 2-Hexanone   | 591-78-6            | -- (EPA MCL); 50 (NYSDEC)                      | 5                                 |
| Dibromochloromethane   | 124-48-1            | 80 (EPA MCL); 50 (NYSDEC)                      | 0.5                               |
| 1,2-Dibromoethane  | 106-93-4            | -- (EPA MCL); -- (NYSDEC)                      | 0.5                               |
| Chlorobenzene  | 108-90-7            | 100 (EPA MCL); 5 (NYSDEC)                      | 0.5                               |
| Ethylbenzene   | 100-41-4            | 700 (EPA MCL); 5 (NYSDEC)                      | 0.5                               |
| o-Xylene   | 95-47-6             | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| m,p-Xylene   | 108-38-3 / 106-42-3 | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| Styrene  | 100-42-5            | 100 (EPA MCL); 5 (NYSDEC)                      | 0.5                               |
| Bromotform   | 75-25-2             | 80 (EPA MCL); 50 (NYSDEC)                      | 0.5                               |
| Isopropylbenzene   | 98-82-8             | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| 1,1,2,2-Tetrachloroethane  | 79-34-5             | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| 1,3-Dichlorobenzene  | 541-73-1            | -- (EPA MCL); 3 (NYSDEC)                       | 0.5                               |
| 1,4-Dichlorobenzene  | 106-46-7            | 75 (EPA MCL); 3 (NYSDEC)                       | 0.5                               |
| 1,2-Dichlorobenzene  | 95-50-1             | 600 (EPA MCL); 3 (NYSDEC)                      | 0.5                               |
| 1,2-Dibromo-3-chloropropane  | 96-12-8             | 0.2 (EPA MCL); 0.04 (NYSDEC)                   | 0.5                               |
| 1,2,4-Trichlorobenzene   | 120-82-1            | 70 (EPA MCL); 5 (NYSDEC)                       | 0.5                               |
| 1,2,3-Trichlorobenzene   | 87-61-6             | -- (EPA MCL); 5 (NYSDEC)                       | 0.5                               |

**TABLE 2**  
**REFERENCE LIMITS**  
**BUSH INDUSTRIES SITE**

| WQ/MNA PARAMETER            | Analyte | Project Screening Criteria/Action Limit                          | Project Quantitation Limit |
|-----------------------------|---------|--|----------------------------|
| Total Organic Carbon (mg/L) |         | NC (EPA MCL); NC (NYSDEC); >20 (MNA Tech Protocol)               | 1                          |
| Alkalinity (mg/L)           |         | NC (EPA MCL); NC (NYSDEC); >2x background (MNA Tech Protocol)    | 1                          |
| Sulfate (mg/L)              |         | 250* (EPA MCL); 250 (NYSDEC); >20 (MNA Tech Protocol)            | 1                          |
| Sulfide (mg/L)              |         | NC (EPA MCL); NC (NYSDEC); >1 (MNA Tech Protocol)                | 1                          |
| Nitrate (mg/L)              |         | 10 (EPA MCL); 10 (NYSDEC); <1 (MNA Tech Protocol)                | 0.05                       |
| Chloride (mg/L)             |         | 250* (EPA MCL); 250 (NYSDEC); >2x background (MNA Tech Protocol) | 1                          |
| Ferrous Iron (µg/L)         |         | NC (EPA MCL); NC (NYSDEC); >1 (MNA Tech Protocol)                | 10                         |
| Methane (µg/L)              |         | NC (EPA MCL); NC (NYSDEC); >0.5 (MNA Tech Protocol)              | 5                          |
| Ethane (µg/L)               |         | NC (EPA MCL); NC (NYSDEC); >0.01 (MNA Tech Protocol)             | 5                          |
| Ethene (µg/L)               |         | NC (EPA MCL); NC (NYSDEC); >0.01 (MNA Tech Protocol)             | 5                          |

**NOTES:**

\*indicates value is a secondary drinking water regulation criterion.

NC indicates no criteria available.

EPA Criteria from 2004 Edition of the Drinking Water Standards and Health Advisories. EPA 822-R-04-005. Winter 2004.

NYSDEC Values are from Ambient Water Quality Standards and Guidance Values and Groundwater Effluent Limitations, June 1998;

Errata Sheet for the June 1998 TOGS 1.1.1, January 1999; and April 2000 Addendum to the June 1998 TOGS 1.1.1, April 2000.

MNQA Tech Protocol values are from Wiedemeier, T., M. Swanson, et.al. Technical Protocol for Evaluating Natural Attenuation of Chlorinated Solvents in Ground Water. EPA/600/R-98/128. September 1998.

**TABLE 3**  
**SAMPLE COLLECTION AND ANALYSIS PROTOCOLS**  
**BUSH INDUSTRIES SITE**

| Sample Type | Matrix | Sampling Device                        | Samples <sup>(1/2)</sup> | No. of Samples <sup>(1/2)</sup> | Parameter   | Sample Container <sup>(3/4)</sup>         | Sample Preservation  | Analytical Method <sup>(5)</sup>       | PQL                               | Holding Time <sup>(6)</sup> |
|-------------|--------|--|--------------------------|---------------------------------|---|---|--|--|-----------------------------------|-----------------------------|
| Groundwater | Water  | Positive Displacement Submersible Pump | 6                        | 6                               | pH; temperature; specific conductivity DO; ORP; turbidity [Field Screening] | NA  | NA   | Direct Field Measurement Following SOP | NA                                | Analyze Immediately         |
|             |        |  | 6                        | 6                               | Low Concentration TCL Volatile Organic Compounds [CLP Lab]                  | (4) 40 mL VOA vials w/Teflon lined septum | 1:1 HCl to pH<2; Cool to 4°C                                   | SOM01.1                                | Compound specific (0.5 - 20 µg/L) | 10 days                     |
|             |        |  | 6                        | 6                               | Total Organic Carbon [DESA Lab]   | (1) 1 L amber glass                       | H <sub>2</sub> SO <sub>4</sub> to pH<2; Cool to 4°C            | SW-846 Method 9060                     | 1 mg/L                            | 28 days*                    |
|             |        |  | 6                        | 6                               | Alkalinity [DESA Lab]   | (1) 1 L polyethylene                      | Cool to 4°C  | MCAWW Method 310.1                     | 1 mg/L                            | 14 days*                    |
|             |        |  | 6                        | 6                               | Sulfate [DESA Lab]  | (1) 1 L polyethylene                      | Cool to 4°C  | EPA 300.1                              | 1 mg/L                            | 28 days*                    |
|             |        |  | 6                        | 6                               | Sulfide [DESA Lab]  | (1) 1 L polyethylene                      | NaOH to pH >12; 4 drops of zinc acetate per liter; Cool to 4°C | MCAWW Method 376.1                     | 1 mg/L                            | 7 days*                     |
|             |        |  | 6                        | 6                               | Nitrate [DESA Lab]  | (1) 1 L polyethylene                      | Cool to 4°C  | EPA 300.1                              | 0.05 mg/L                         | 48 hours*                   |
|             |        |  | 6                        | 6                               | Chloride [DESA Lab]   | (1) 1 L polyethylene                      | Cool to 4°C  | EPA 300.1                              | 1 mg/L                            | 28 days*                    |
|             |        |  | 6                        | 6                               | Ferrous Iron [Sub Lab]  | (1) 100 mL amber glass                    | 2mL HCl; Cool to 4°C   | Std. Methods Method 3500Fe-D           | 10 µg/L                           | 24 hours*                   |
|             |        |  | 6                        | 6                               | Ethane [Sub Lab]  | (5) 40-mL VOA vials w/Teflon lined septum | Cool to 4°C  | GC/FID (SW-846 Method 3810)            | 5 µg/L                            | 7 days*                     |
|             |        |  | 6                        | 6                               | Ethene [Sub Lab]  | (5) 40-mL VOA vials w/Teflon lined septum | Cool to 4°C  | GC/FID (SW-846 Method 3810)            | 5 µg/L                            | 7 days*                     |
|             |        |  | 6                        | 6                               | Methane [Sub Lab]   | (5) 40-mL VOA vials w/Teflon lined septum | Cool to 4°C  | GC/FID (SW-846 Method 3810)            | 5 µg/L                            | 7 days*                     |

**TABLE 3**  
**SAMPLE COLLECTION AND ANALYSIS PROTOCOLS**  
**BUSH INDUSTRIES SITE**

| Sample Type | Matrix | Sampling Device  | No. of Samples <sup>(1/2)</sup> | Parameter                                | Sample Container <sup>(3/4)</sup>         | Sample Preservation          | Analytical Method <sup>(5)</sup> | PQL                               | Holding Time <sup>(6)</sup> |
|-------------|--------|--|---------------------------------|--|---|------------------------------|----------------------------------|-----------------------------------|-----------------------------|
| Field Blank | Water  | Collected Rinsate Passed Over/Through Sampling Equipment | 1                               | TCL Volatile Organic Compounds [CLP Lab] | (4) 40-mL VOA vials w/Teflon lined septum | 1:1 HCl to pH<2; Cool to 4°C | SOM01.1                          | Compound specific (0.5 - 20 µg/L) | 10 days                     |
| Trip Blank  | Water  | Direct Fill of Sample Bottles                            | 1                               | TCL Volatile Organic Compounds [CLP Lab] | (4) 40-mL VOA vials w/Teflon lined septum | 1:1 HCl to pH<2; Cool to 4°C | SOM01.1                          | Compound specific (0.5 - 20 µg/L) | 10 days                     |
|             |        |  | 6                               | Ethane [Sub Lab]                         | (5) 40-mL VOA vials w/Teflon lined septum | Cool to 4°C                  | GC/FID (SW-846 Method 3810)      | 5 µg/L                            | 7 day*                      |
|             |        |  | 6                               | Ethane [Sub Lab]                         | (5) 40-mL VOA vials w/Teflon lined septum | Cool to 4°C                  | GC/FID (SW-846 Method 3810)      | 5 µg/L                            | 7 days*                     |
|             |        |  | 6                               | Methane [Sub Lab]                        | (5) 40-mL VOA vials w/Teflon lined septum | Cool to 4°C                  | GC/FID (SW-846 Method 3810)      | 5 µg/L                            | 7 days*                     |

**NOTES:**

- The number in parentheses in the "No. of Samples" column denotes the number of duplicate samples.
- The number of field, trip and DI water blanks is estimated based on the approximate number of days in the field for each type of sampling during the MNA Program events.
- The number in parentheses in the "Sample Container" column denotes the number of containers needed. Additional volume must be sent for laboratory QA/QC sample analyses.
- All bottles will comply with OSWER Directive 9240.0-05A: "Specifications and Guidance for Obtaining Contaminant-Free Sample Containers", EPA 540/R-93/051, December 1992.
- Method References:  
 SOM01.1 = USEPA Contract Laboratory Program Statement of Work for Multi-Media, Multi-Concentration Organics (May 2005 or latest revision).  
 MCAWW = Methods for Chemical Analysis of Water and Wastes, March 1983.  
 Std. Methods = Standard Methods for the Examination of Water and Wastewater, 20th Edition (January 2000).  
 SW-846 = Test Methods for Evaluating Solid Waste, Physical/Chemical Methods (November 1986, revised through November 2000 via Updates I through IVB).
- EPA300.1 = Determination of Inorganic Anions in Drinking Water by Ion Chromatography, Revision I (27 April 1999),  
 EPA/600/R-98/128 = Technical Protocol for Evaluating Natural Attenuation of Chlorinated Solvents in Groundwater (September 1998).
- All holding times listed are from Verified Time of Sample Receipt (VTSR) unless noted otherwise (\* denotes from time of sample collection).
- Acronyms/Abbreviations used:  
 CLP = Contract Laboratory Program  
 DO = Dissolved Oxygen  
 PQL = Practical Quantitation Limit  
 TCL = Target Compound List
- DESA = Division of Environmental Science and Assessment  
 ORP = Oxidation-Reduction Potential  
 Sub Lab = Non-RAS Subcontract Laboratory  
 VOA = Volatile Organic Analysis

**TABLE 4**  
**SUMMARY OF ANALYTICAL QC PROCEDURE CHECKS, FREQUENCIES, ACCEPTANCE CRITERIA,  
 AND CORRECTIVE ACTIONS FOR FIELD SCREENING AND LABORATORY SAMPLE ANALYSES  
 BUSH INDUSTRIES SITE**

| Parameter                  | Method                             | QC Procedure                | Data Quality Indicators (DQIs)      | Frequency  | Acceptance Criteria                             | Corrective Action   |
|----------------------------|------------------------------------|-----------------------------|-------------------------------------|--|---|---|
| Total Organic Carbon       | SW-846                             | Method Blank                | Accuracy                            | 1 per event  | no constituent >CRQL                            | suspend analysis until source rectified                     |
|                            | Method 9060                        | Laboratory Duplicate Sample | Precision                           | 1 per event  | ± 25% RPD                                       | reanalyze   |
| Alkalinity                 | MCAWW Method 310.1                 | Method Blank                | Accuracy                            | 1 per event  | no constituent >CRQL                            | suspend analysis until source rectified                     |
|                            |                                    | Laboratory Duplicate Sample | Precision                           | 1 per event  | ± 25% RPD                                       | reanalyze   |
| Sulfide                    | MCAWW Method 376.1                 | Method Blank                | Accuracy                            | 1 per event  | no constituent >CRQL                            | suspend analysis until source rectified                     |
|                            |                                    | Laboratory Duplicate Sample | Precision                           | 1 per event  | ± 25% RPD                                       | reanalyze   |
| Sulfate, Nitrate, Chloride | EPA 300.1                          | Laboratory Reagent Blank    | Accuracy                            | 1 per event  | no analyte >PQL                                 | suspend analysis until source rectified                     |
|                            |                                    | Laboratory Fortified Blank  | Accuracy                            | 1 per event  | 85 - 115%R (unless MRL <10xPQL, then 75 - 125%) | check calculations, suspend analysis until source rectified |
| Surrogate Compounds        | Laboratory Fortified Sample Matrix | Accuracy                    | 1 per event                         | 75 - 125%R   | quality outliers as "matrix induced bias"       | reanalyze affected samples                                  |
|                            | Surrogate Compounds                | Accuracy                    | all samples                         | 90 - 115%R   |   |   |
| Ferrous Iron               | Laboratory Duplicate Sample        | Precision                   | 1 per event or 1 per analysis batch | ±20% RPD (MRL to 10x MRL); ±10% RPD (10x MRL to highest calibration level) | qualify affected samples                        | suspend analysis until source rectified                     |
|                            | Std. Methods (3500Fe-D)            | Method Blank                | Accuracy                            | 1 per event  | no constituent >CRQL                            | reanalyze   |
|                            | Laboratory Duplicate Sample        | Precision                   | 1 per event                         | ± 25% RPD  |   |   |

**TABLE 4**  
**SUMMARY OF ANALYTICAL QC PROCEDURE CHECKS, FREQUENCIES, ACCEPTANCE CRITERIA,  
 AND CORRECTIVE ACTIONS FOR FIELD SCREENING AND LABORATORY SAMPLE ANALYSES  
 BUSH INDUSTRIES SITE**

| Parameter                     | Method                         | QC Procedure  | Data Quality Indicators (DQIs) | Frequency                      | Acceptance Criteria  | Corrective Action  |
|-------------------------------|--------------------------------|---|--------------------------------|--------------------------------|--|--|
| Methane, Ethane, Ethene       | GC/FID (SW-846<br>Method 3810) | Method Blank  | Accuracy                       | 1 per event                    | no constituent >CRQL   | suspend analysis until source rectified                        |
|                               |                                | Laboratory Duplicate Sample   | Precision                      | 1 per event                    | $\pm 25\%$ RPD   | reanalyze  |
| Low Concentration<br>TCL VOCs | OLC03.2                        | Method Blank  | Accuracy                       | 1 every 12 hours               | no constituent >CRQL   | suspend analysis until source rectified                        |
|                               |                                | Deuterated Monitoring Compounds   | Accuracy                       | all samples                    | compound specific (full range: 37 - 171%R)                           | check calculations and instruments, reanalyze affected samples |
|                               |                                | Internal Standards  | Accuracy;<br>Sensitivity       | all samples                    | $\pm 40\%$ of response area;<br>$\pm 120$ sec retention time shift   | check calculations and instruments, reanalyze affected samples |
|                               |                                | Matrix Spike/Matrix Spike Duplicate (optional; not required for EPA Region 2) | Accuracy; Precision            | if requested, then 1 per event | advisory; compound specific (full range: 61 - 145%R, $\pm 14\%$ RPD) | flag outliers  |

## NOTES:

## Method References:

SOM01.1 = USEPA Contract Laboratory Program Statement of Work for Multi-Media, Multi-Concentration Organics (May 2005 or latest revision).

MCAWW = Methods for Chemical Analysis of Water and Wastes, March 1983.

Std. Methods = Standard Methods for the Examination of Water and Wastewater, 20th Edition (January 2000).

SW-846 = Test Methods for Evaluating Solid Waste, Physical/Chemical Methods (November 1986, revised through November 2000 via Updates I through IVB).

EPA 300.1 = Determination of Inorganic Anions in Drinking Water by Ion Chromatography, Revision I (27 April 1999).

EPA/600/R-98/128 = Technical Protocol for Evaluating Natural Attenuation of Chlorinated Solvents in Groundwater (September 1998).

Acronyms/Abbreviations used:

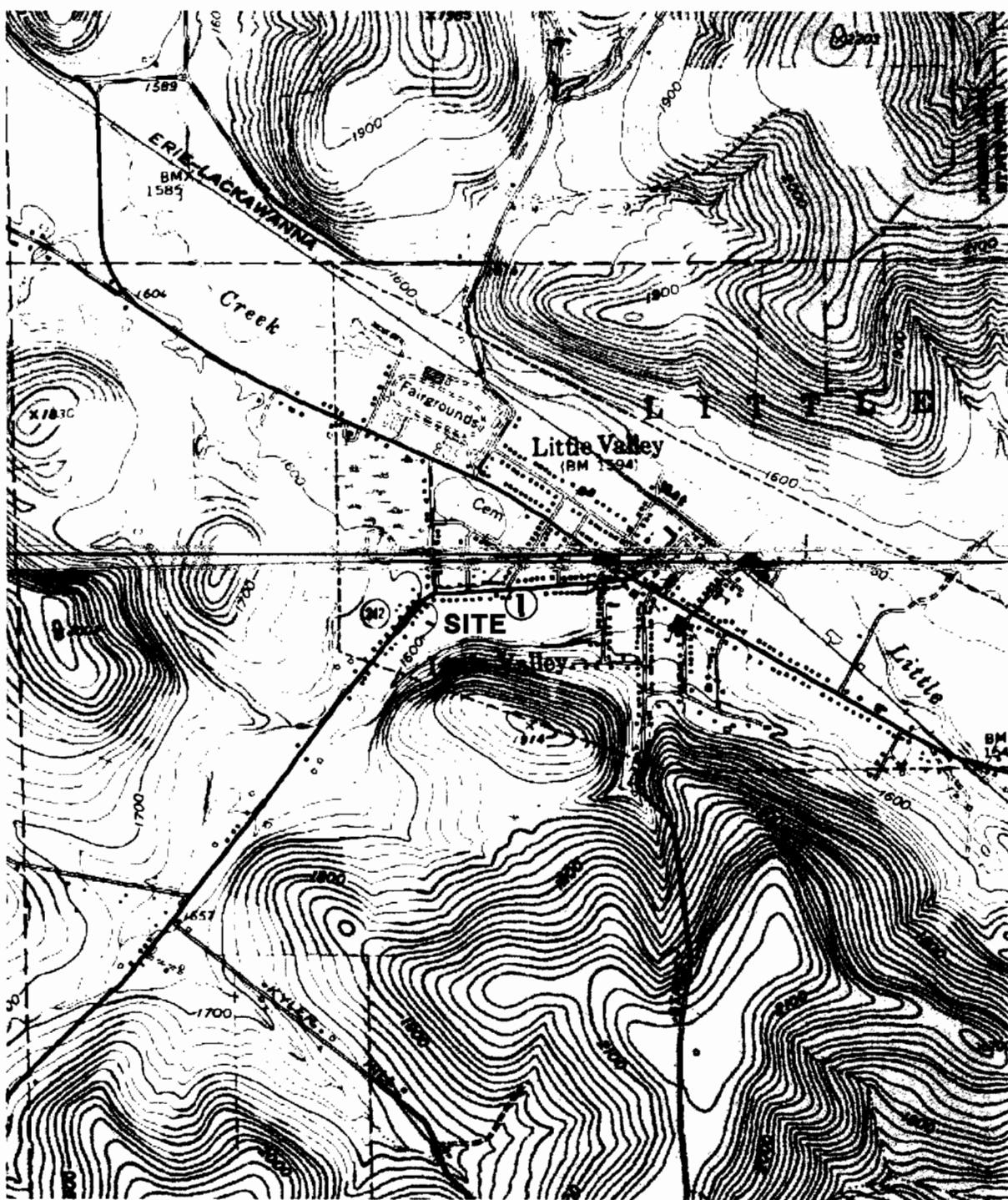
%R = Percent Recovery

MRL = Minimum Reporting Level

RPD = Relative Percent Difference

TCL = Target Compound List

CRQL = Contract Required Quantitation Limit  
 PQL = Practical Quantitation Limits  
 SD = Standard Deviation  
 VOCs = Volatile Organic Compounds



SOURCE:

USGS CATTARAUGUS AND LITTLE  
VALLEY, NEW YORK QUADRANGLES.

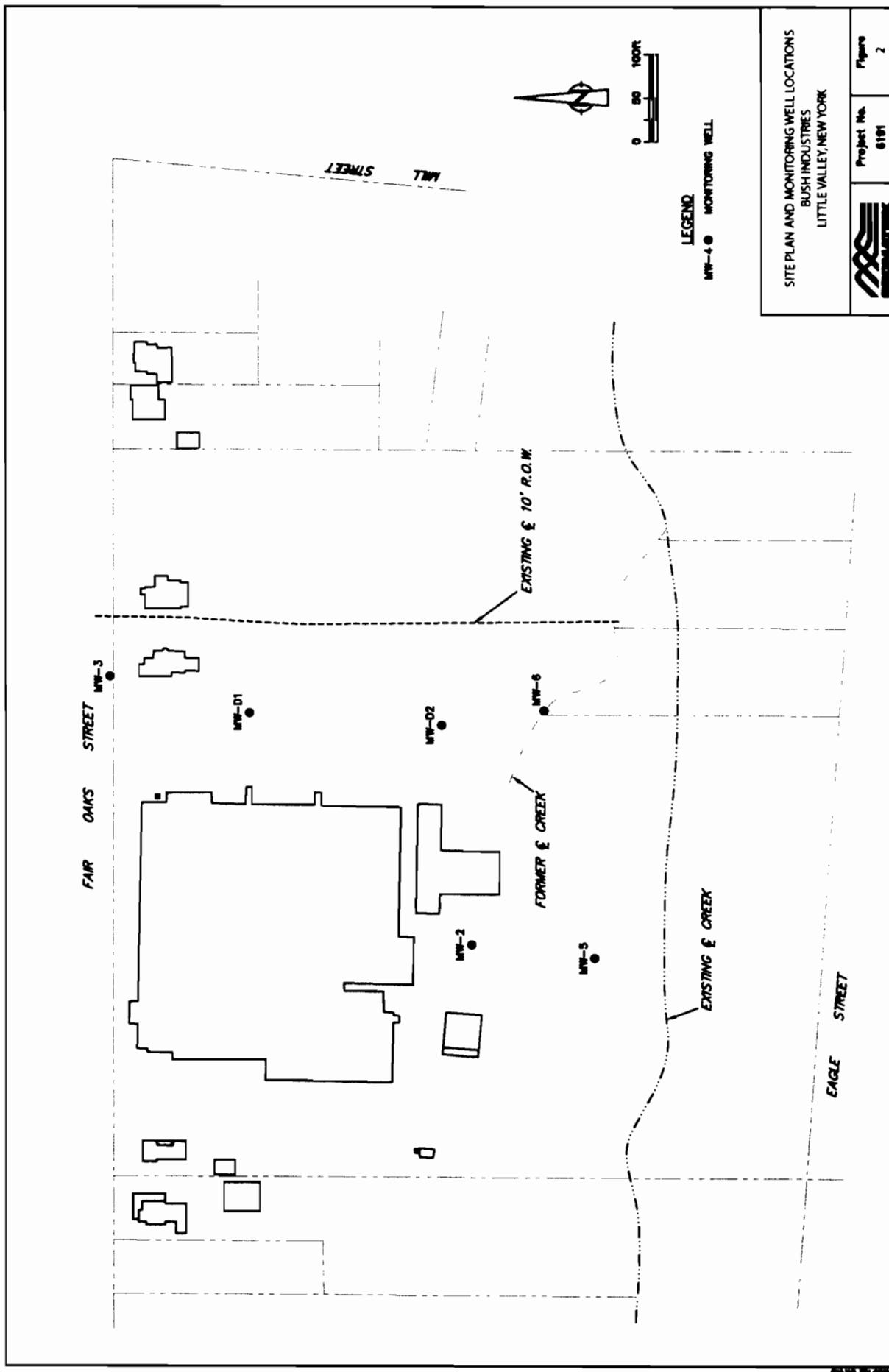


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FLEXPAC C:\\EDWARD\\MAPS\\  
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SITE LOCATION  
BUSH INDUSTRIES  
LITTLE VALLEY, NEW YORK

Project No.  
6191  
Figure  
1



**ATTACHMENT 1**

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**Standard Operating Procedures**

**ATTACHMENT A**

**STANDARD OPERATING PROCEDURES**

- SOP #1      Groundwater Sampling [Low Flow Purge Procedure]**
- SOP #2      Field Parameter Measurement**
- SOP #3      Water Level Measurements**
- SOP #4      Decontamination [Non-disposable Chemical Sampling Equipment]**
- SOP #5      Decontamination [Field Instrumentation - Probes, Water Quality Meters, etc.]**

### **Groundwater Sampling [Low Flow Purge Procedure] (SOP #1)**

1. Check and record the condition of the well for any damage or evidence of tampering.
2. Remove the well cap.
3. Measure well headspace with a PID or FID and record the reading in the field logbook.
4. Measure the depth to water as stated in SOP#3, and record the measurement in the field logbook. Do not measure the depth to the bottom of the well at this time (to avoid disturbing any sediment that may have accumulated); see Step 18.
5. Lay out plastic sheeting and place the monitoring, purging and sampling equipment on the sheeting. To avoid cross-contamination, do not let any downhole equipment touch the ground.
6. Re-check and record the depth to water after approximately 5 minutes at the well location. If the measurement has changed more than 0.01 foot, check and record the measurement again, then begin well purging.
7. Attach and secure the Teflon-lined polyethylene tubing to the low-flow submersible pump. As the pump is slowly lowered into the well, secure the safety drop cable, tubing, and electrical lines to each other using nylon stay-ties placed approximately 5 feet apart.
8. Set the pump at approximately the middle of the screen and/or the best depth based on the stratigraphy of the well. Be careful not to place the pump intake less than 2 feet above the bottom of the well as this may cause mobilization of any sediment present in the bottom of the well.
9. Start pumping the well at 0.2 to 0.5 liters per minute.
10. Monitor the water level in the well periodically during pumping - ideally the pump rate should equal the well recharge rate with little or no water level drawdown in the well (drawdown shall be 0.3 foot or less). There should be at least 1 foot of water over the pump intake so there is no risk of the pump suction being broken, or entrapment of air in the sample. Record the pumping rate adjustments and depth(s) to water in the logbook. Pumping rates should, if needed, be reduced to the minimum capabilities of the pump (0.1 to 0.2 liters per minute) to avoid purging the well dry. However, if the recharge rate of the well is very low and the well is purged dry, then wait until the well has recharged to a sufficient level and collect the appropriate volume of sample with the submersible pump.
11. Purge the well at a low flow rate (from 0.2 to 0.5 liters per minute). During purging, monitor the field parameters (temperature, pH, turbidity, ORP, specific conductivity, and DO) approximately every 3 to 5 minutes. A flow-through cell will be used to monitor the field parameters (SOP #2). Begin measuring field parameters after the flow-through cell has been "flushed" with groundwater twice.

### Groundwater Sampling [Low Flow Purge Procedure] (SOP #1) [cont'd]

12. The well is considered stabilized and ready for sample collection when the indicator parameters have stabilized for three consecutive readings, as follows:
  - 0.1 for pH
  - 3 percent for specific conductance
  - 10 percent for dissolved oxygen
  - 10 percent for turbidity
  - 10 mV for Eh

Dissolved oxygen and turbidity usually require the longest time to achieve stabilization. The pump must not be removed from the well between purging and sampling.
13. Once the field parameters have stabilized, collect the samples directly from the end of the tubing. Volatiles and analyses that degrade by aeration must be collected first.
14. Fill the sample bottles by allowing the pump discharge to flow gently down the inside of the bottle with minimal turbulence. Cap each bottle as it is filled.
15. Preserve and label the samples, and record them on the chain of custody. Place immediately into a cooler for shipment and maintain at 4°C.
16. The filling and preservation procedures will be:
  - VOCs - Determine the amount of 1:1 HCl preservative required to adjust the pH of the sample to less than 2 in an extra 40 ml glass vial. Add this volume to the empty 40 ml vials prior to sampling. Fill each container with sample to just overflowing so that no air bubbles are entrapped inside. If effervescence occurs, submit the sample without preservative and note on the chain of custody form.
  - Other Parameters - Fill each container and preserve immediately as required. To test for pH, pour a minimal portion of sample onto broad range pH paper to verify that the appropriate pH level has been obtained.
17. Carefully remove the pump assembly from the well. The Teflon-lined polyethylene tubing will be dedicated to each well. The tubing should be placed in a large plastic garbage bag, sealed, and labeled with the appropriate well identification number.

**Groundwater Sampling [Low Flow Purge Procedure] (SOP #1) [cont'd]**

18. After sampling is complete, measure the total depth of the well.
19. Close and lock the well.

## **Field Parameter Measurement (SOP #2)**

Field parameters will be monitored during purging of the monitoring wells and prior to surface water sampling, utilizing a Horiba® U-22 water quality meter or equivalent. Measurements will be conducted in accordance with the manufacturer's instructions and the following procedure:

1. Calibrate the water quality meter as per manufacturer's instructions.
2. For low flow purging of the monitoring wells:
  - Attach a flow-through cell to the Teflon-lined polyethylene tubing. Position the water quality meter probe in the flow-through cell. Begin purging the monitoring well, following SOP#1.
  - After the cell has been "flushed" at least twice, begin monitoring the field parameters, and continue approximately every 3 to 5 minutes during purging. All water quality measurements will be recorded in the appropriate field logbook or on a well purge data sheet.
  - When the indicator parameters have stabilized for three consecutive readings (see Step 12 of SOP #1), the well is considered stabilized and ready for sample collection. Remove the flow-through cell from the tubing.
3. For surface water sampling, measure field parameters immediately prior to sample collection by positioning the probe directly in the water body.
4. Decontaminate the probe of the water quality meter between wells/surface water locations (SOP #5).
5. Record water quality measurements in the appropriate field logbook, noting well identification or surface water location, sample date and time, and observations.

### **Decontamination [Non-disposable Chemical Sampling Equipment] (SOP #4)**

Decontamination of non-disposable sampling equipment used to collect samples for chemical analyses (i.e., scoops, trowels, bowls, split-spoons, etc.) will be conducted as described below:

1. Alconox detergent and potable water scrub.
2. Potable water rinse.
3. 10 percent nitric acid rinse (ultra pure grade) when sampling for inorganics. Carbon steel split-spoons will be rinsed with a 1 percent nitric acid solution to avoid stripping of metals.
4. Deionized water rinse.
5. Methanol rinse followed by a hexane rinse (solvents are pesticide grade or better) for equipment involved in the sampling of organics.
6. Deionized water rinse (volume at least five times amount of solvent used in rinse step above).
7. Air dry.
8. Wrap or cover exposed ends of equipment with aluminum foil for transport and handling.

Decontamination of sampling equipment will be kept to a minimum in the field and, wherever possible, dedicated disposable sampling equipment will be used. Decontamination fluids will be stored in DOT-approved 55-gallon drums or in an on-site storage tank (liquids only) until disposal. Personnel directly involved in equipment decontamination will wear appropriate protective clothing, as stated in the HASP.

### **Decontamination [Field Instrumentation - Probes, Water Quality Meters, etc.] (SOP #5)**

Field instrumentation (such as interface probes, water quality meters, etc.) will be decontaminated between sample locations by rinsing with deionized water. If visible contamination still exists on the equipment after the rinse, an Alconox detergent scrub will be added, and the probe thoroughly rinsed again.

Any decontamination fluids generated will be stored in DOT-approved 55-gallon drums or in an on-site storage tank (liquids only) until disposal. Personnel directly involved in equipment decontamination will wear appropriate protective clothing, as stated in the HASP.