



December 31, 2025
Job No. 41.163256.00

Daniel R. Nierenberg, P.G.
Project Manager, Remedial Bureau B
New York State Department of Environmental Conservation
625 Broadway, 12th Floor
Albany, NY 12233-7016

Re: Supplemental Injection Work Plan
Former Boyle Auto Wreckers, Inc.
1346 Blondell Avenue
Bronx, NY 10461
NYSDEC BCP No. C203089

Dear Mr. Nierenberg:

Tyll Engineering and Consulting PC (TEC) and GZA GeoEnvironmental of New York (GZA) is pleased to submit this Supplemental Injection Work Plan (Work Plan) to the New York State Department of Environmental Conservation (NYSDEC) on behalf of Blondell Equities, LLC (Volunteer/Owner) for the Former Boyle Auto Wreckers Inc. project located at 1346 Blondell Avenue, Bronx, New York (Site). The Site is being remediated under the Brownfield Cleanup Program (BCP) Site No. C203089.

This Work Plan is being submitted as a response to the NYSDEC's Draft Final Engineering Report (FER) Disapproval Letter, dated August 21, 2025, to address concerns of potential groundwater contamination migrating offsite. The Work Plan was prepared in accordance with the September 2019 Decision Document and the May 2019 RAWP. The Work Plan was prepared in accordance with Title 6 New York Codes, Rules, Regulations Part 375 in the NYSDEC Division of Environmental Remediation Technical Guidance for Site Investigation and Remediation (DER-10, May 3, 2010), and all other applicable regulatory and procedural requirements.

The supplemental groundwater remedial activities described in this Work Plan are conditional on the results of the additional groundwater sampling included in the Draft Supplemental Monitoring Well Installation and Sampling Work Plan submitted to NYSDEC on October 13, 2025. GZA will make recommendations for completing this Work Plan after evaluating the effectiveness of the remedial action soil amendments on groundwater concentrations in our groundwater monitoring report. If the additional round of groundwater sampling shows improved groundwater contaminants of concern (COC) concentrations below their respective NYSDEC Technical Operation Guidance Series (TOGS) 1.1.1 Ambient Water Quality Standards (AWQS), then we do not anticipate that the supplemental injections included in this Work Plan will be completed.

BACKGROUND

During a November 14, 2024 meeting, the NYSDEC indicated a minimum of two rounds of groundwater samples would be required to evaluate the performance of the remedy on groundwater quality and to inform the need for Engineering Controls during site management. Four (4) prepacked monitoring wells (MW-1, MW-



2, MW-3, and MW-4) were installed along the perimeter of the Site. The four monitoring wells were completed with a stickup and locking cap at current grade and developed with a submersible pump.

Two rounds of groundwater sampling were performed on November 14, 2024, and January 14, 2025. The November 2024 sampling event included samples from MW-2 and MW-4 submitted for Volatile Organic Compounds (VOCs), Semi-Volatile Organic Compounds (SVOCs), total and dissolved metals, pesticides, polychlorinated biphenyls (PCBs), and per- and polyfluoroalkyl substances (PFAS) laboratory analysis. The January 2025 sampling event only included sampling for VOC and SVOC analysis from MW-1, MW-2, and MW-4. Wells MW-1 and MW-3 were not sampled in November 2024 and MW-3 in January 2025 due to the lack of measurable groundwater in the wells. The laboratory analytical results from November 2024 and January 2025 were compared to the NYSDEC Technical Operation Guidance Series (TOGS) 1.1.1 Ambient Water Quality Standards (AWQS) and Guidance Values and PFAS were compared to NYSDEC Part 375 PFAS Remedial Program, dated April 2023 (Part 375 PFAS).

Groundwater sampling analytical results indicated that PCBs and pesticides were not present in groundwater. The following contaminants were found in exceedance of their respective TOGS 1.1.1 AWQS or Part 375 PFAS limit:

- VOCs - 1,2,4,5-tetramethylbenzene (max. of 10 micrograms per liter [ug/L]), 1,2,4-trimethylbenzene (max. of 170 ug/L), 1,3,5-trimethylbenzene (max. of 5.5 ug/L), benzene (max. of 8.0 ug/L), cis-1,2-dichloroethene (max. of 54 ug/L), ethylbenzene (max. of 110 ug/L), isopropylbenzene (max. of 7.6 ug/L), n-propylbenzene (max. of 14 ug/L), naphthalene (max. of 10 ug/L), o-xylene (max. of 87 ug/L), p/m-xylene (max. of 180 ug/L), sec-butylbenzene (max. of 5.5 ug/L), toluene (max. of 68 ug/L), trichloroethene (max. of 26 ug/L), and vinyl chloride (max. of 2 ug/L)
- SVOCs - benzo(a)anthracene (max. of 0.08 ug/L), benzo(a)pyrene (max. of 0.07 ug/L), benzo(b)fluoranthene (max. of 0.09 ug/L), benzo(k)fluoranthene (max. of 0.04 ug/L), chrysene (max. of 0.08 ug/L), hexachlorobenzene (max. of 0.05 ug/L), and indeno (1,2,3,-cd)pyrene (max. of 0.07 ug/L)
- Dissolved metals, including iron (max. of 6,100 ug/L), manganese (max. of 17,470 ug/L), and sodium (max. of 165,000 ug/L).
- Perfluorooctanesulfonic Acid (PFOS) (max. of 0.0712 ug/L) and Perfluorooctanoic Acid (PFOA) (max. of 0.0431 ug/L) were detected above NYSDEC Part 375 PFAS.

Based on the feedback received from the NYSDEC, new permanent monitoring wells were installed and sampled. Details on the installation of the new monitoring wells are included in *Supplemental Monitoring Well Installation & Sampling Work Plan* dated October 13, 2025 and approved by NYSDEC on October 16, 2025.

The groundwater analytical results from the November 4, 2025, sampling event indicated the following AWQS were exceeded in one or more groundwater samples:

- VOCs:
 - Petroleum-related VOCs, only in GZM-1, 1,2,4,5-tetramethylbenzene (max. of 74 ug/L), 1,2,4-trimethylbenzene (max. of 880 ug/L), 1,3,5-trimethylbenzene (max. of 190 ug/L), benzene (85 ug/L), ethylbenzene (max. of 690 ug/L), isopropylbenzene (max. of 77 ug/L),



n-butylbenzene (max. of 23 ug/L), n-propylbenzene (max. of 190 ug/L), o-xylene (max. of 280 ug/L), p/m-xylene (max. of 1,300 ug/L) and toluene (max. of 25).

- SVOCs:
 - Polycyclic Aromatic Hydrocarbons (PAHs), benzo(a)anthracene (max. 0.27 µg/L, GZM-4), benzo(a)pyrene (max. 0.20 µg/L, GZM-4), benzo(b)fluoranthene (max. 0.26 µg/L, GZM-4), benzo(k)fluoranthene (max. 0.10 µg/L, GZM-4), chrysene (max. of 0.17 µg/L) and indeno(1,2,3-cd)pyrene (max. 0.16 µg/L, GZM-4).
 - Naphthalene was detected with a maximum concentration of 34 µg/L in GZM-1.
- Metals:
 - Four metals were detected above TOGS 1.1.1 AWQS. Iron (max. 626 µg/L-dissolved and max 25,400 µg/L – Total in GZM-2), Lead (max. 83.69 µg/L – Total in GZM-4), Manganese (max. 1,942 µg/L-dissolved and max 1,984 µg/L – Total in GZM-2) and Sodium (max. 265,000 µg/L-dissolved and max. 248,000 µg/L – Total in GZM-2)
- PCBs:
 - No PCBs were detected above TOGS 1.1.1 AWQS.
- PFOA and PFAS:
 - Two compounds were detected above TOGS 1.1.1 AWQS. Perfluorooctanesulfonic acid (PFOS) was detected in groundwater samples, with a maximum concentration of 0.0714 µg/L at GZM-2. Perfluorooctanoic acid (PFOA) was detected above TOGS 1.1.1 AWQS in groundwater samples, with a maximum concentration of 0.0252 µg/L at GZM-4.
- Pesticides:
 - No pesticides were detected above TOGS 1.1.1 AWQS.

The previous sampling results from November 2024 and January 2025 are provided in **Figure 1**. This workplan was prepared in accordance with Title 6 New York Codes, Rules, and Regulations (NYCRR) Part 375, the NYSDEC Division of Environmental Remediation (DER) Technical Guidance for Site Investigation and Remediation (DER-10, May 3, 2010), and other applicable regulatory and procedural requirements.

PERMEABLE REACTIVE BARRIER INJECTIONS

A Permeable Reactive Barrier (PRB) will be injected into the subsurface to stabilize and degrade petroleum related VOCs and SVOCs in the groundwater before they can migrate offsite. PRBs typically utilize a stabilizing compound to capture and prevent migration of specific contaminants while also introducing reagents to accelerate the degradation of the target compounds. GZA proposes to utilize PetroFix[®] manufactured by Regenesis[®]. PetroFix[®] has a dual function to both stabilize contaminants and stimulate degradation via naturally occurring microbes. It uses a Colloidal Activated Carbon (CAC) to adsorb hydrocarbons dissolved in the groundwater and prevent the migration downgradient of the injection while also introducing additional anaerobic electron acceptors that promote the microbial degradation of hydrocarbons. The CAC coats soil surfaces and remains in place following the injection. This allows groundwater to flow through the area without disturbing the natural pathways of flow, but sorbs VOCs onto the PetroFix[®] surface, preventing plume migrations beyond the treatment area. The electron acceptors, nitrate and sulfate, are included as part of PetroFix[®], and the characteristics of the colloidal



activated carbon enhance the bioremediation of VOCs the sorb to the PetroFix® activated carbon surfaces.

The proposed blend of PetroFix® will include a 50%/50% mix of both ammonium sulfate and sodium nitrate as the anaerobic electron receptors. The introduction of ammonium sulfate into the impacted groundwater will provide nitrogen, a key nutrient for microbes, and sulfate which has a higher redox potential than methanogenesis and is a preferential degradation pathway for sulfate reducer microbes. The introduction of sodium nitrate into the impacted groundwater will also provide nitrogen as a nutrient for microbes and nitrate which has the highest redox potential of the anaerobic degradation pathways via denitrifier microbes. By providing key nutrients and the necessary electron receptors for degradation through two separate redox processes, the subsurface will have more favorable conditions for a faster microbial degradation of the hydrocarbons captured in the CAC and prevent downgradient migration of hydrocarbon contaminants present in the groundwater at the Site. Please see Regenesis PetroFix® information included in **Attachment A**.

Previous chemical treatment of soils and groundwater included the mixing of Regenesis ORC Advanced® at the bottom of excavation to enhance the aerobic degradation of contaminants. PetroFix® was selected as PRB treatment for this area due to its effectiveness in both aerobic and anaerobic environments as a polishing treatment for any migrating contamination in groundwater. PetroFix® does include electron acceptors for anaerobic bioremediation; however, the product is fully compatible with aerobic conditions. The activated carbon component will continue to adsorb hydrocarbons under aerobic and anaerobic conditions. The adsorption and concentration of hydrocarbons on the PetroFix® will further stimulate aerobic bioremediation through the promotion of biofilms and concentration of resources. The electron acceptors provided by PetroFix® may not be fully utilized immediately but will provide alternative degradation pathways as oxygen is consumed from the aerobic degradation and anaerobic conditions develop. Some sulfate and nitrate can still be utilized in an aerobic system as isolated pockets of more reduced conditions will exist.

GZA and Regenesis® developed the proposed dosing of PetroFix® based on the desired target area and the levels of contamination observed during the November 2025 groundwater sampling event. The target treatment area will be the northern corner of the property along Ponton Avenue and the rear of the building as shown on **Figure 2**. A total of 8,800 pounds of PetroFix® CAC and 440 pounds of PetroFix® electron acceptor blend will be mixed with 23,338 gallons of water for injection into the subsurface. The quantity of PetroFix® was calculated using the manufacturer's modeling software, PlumeForce, which determined the volume based on the groundwater velocity, influxing compounds, bioremediation, including the formation of daughter products, back diffusion, treatment area lithology, and time. Based on observations from the remedial investigations, subsurface soils encountered were comprised of silty sand and silty clay in the vadose and smear zones, underlain by a sandy layer in the phreatic zone. The model assumes a highly reduced environment with a dissolved oxygen content less than 0.5 mg/L, breakthrough of the least adsorptive compounds (vinyl chloride and ethene) are projected in approximately 3,600 days after the application. The software assumed a continuous influx of all recorded contaminants at current concentrations over a ten-year time window, assuming a seepage velocity introducing new contamination at 150 feet per year.

The depth of the injections for the PRB will be to target the top 10 feet of the groundwater table. Based on the depth of groundwater observed during the monitoring well installation and sampling performed by GZA in November 2025, the injections depths will vary from approximately 16 feet below ground surface (bgs) along Ponton Avenue and from approximately 6 to 16 feet bgs along the rear of the building. The varying depths of the injections are based on the changes in surface elevations observed along the



PRB footprint and targets the groundwater table at elevations 6.21 feet to -3.79 feet in the North American Vertical Datum 1988 (NAVD88).

Once these injection depths are confirmed across the PRB footprint, an injection contractor will be subcontracted to perform the injections. The subcontractor will mobilize a Direct-Push Technology (DPT) type drill rig to advance rods to the target injection depth and will inject chemicals through the hollow rods. Regenesis recommends incorporating a bottom-up injection method that uses multiple port retractable screens for the application. The radius of influence is best determined by using the test kit provided with each shipment of PetroFix which identifies the colorimetric changes to the groundwater. Smaller batches of the proposed injection blend will be mixed in a dedicated mixing tank, ensuring the ratio of CAC-to-electron acceptor-to-water is maintained. Each batch will be sufficiently mixed utilizing a manufacturer's recommended method.

Injections will be performed at 6-foot on-center spacing, totaling 46 injection points along the PRB alignment illustrated on **Figure 2**. The spacing of the injection points was determined to ensure even coverage. Each injection point will have approximately 173.9 pounds of CAC, 8.7 pounds of electron acceptor, and 326 gallons of water injected into it. The PetroFix® will be delivered to the injection wells using chemically compatible hoses and connections secured to the camlock on each DPT injection point head, and chemically-compatible positive displacement pumps. A pressure gauge and flow meter (or equivalent measurement of volume in the mix tank) will be used to monitor the flow rate and pressure during injection; Regenesis recommends injecting under low pressure to promote natural flow of PetroFix® into the permeable zones and prevent fracturing of the soil. Injections are proposed for a 10-foot depth interval between estimated EL. 92.0 to 82.0 feet based on the previous groundwater elevations observed during the January 2025 sampling (or just above the water table to 10 feet below the top of the water table). Injections will be started at the bottom of the proposed injection elevation and will be raised in 2-foot increments with 35 gallons per increment throughout the injection process to evenly distribute the PetroFix® vertically. If necessary, GZA will adjust injection pressures depending on the density of the subsurface conditions (i.e., with slower flow rates and higher pressures in denser formations).

This remedial design assumed that fines comprise 20-40% of the soil surfaces with a seepage velocity estimated at 150 feet per year. With an injection dosing of PetroFix® at 19,394 mg/L, this falls into range, making downgradient mobilization of the treatment material unlikely as the soil surfaces are anticipated to contain enough surface area to allow the dosage of treatment material at a relatively low anticipated seepage velocity.

PetroFix® can remain suspended in the groundwater and, therefore, be mobile in the subsurface following injections for a brief period. PetroFix® is designed to have limited mobility in soil surfaces because its carbon particles adhere to soil surfaces naturally in six to twelve weeks. Specific site conditions may prolong this period, such as groundwater velocity, high PetroFix® application dosages, and low available soil surface area. Site data provided to Regenesis indicated no anticipated complications that may prolong this period of suspension. To ensure the treatment material remains within the anticipated target area, visual observations in the pre-existing GZM-1 and GZM-2 monitoring wells and a new monitoring well location will be completed during post-injection groundwater sampling. Changes in the suspended PetroFix® concentration or turbidity by an increasing black or grey color in these downgradient areas would indicate PetroFix® remains mobile and is still being transported downgradient. Additionally, the performance of the PRB could be determined by the reduction of VOC concentrations in the monitoring wells. The installation and sampling of an additional monitoring well will be used to monitor post-injection conditions and assess the migration of contamination.



Potential contingency plans after the initial application would include PRB bioremediation enhancement and PRB expansion. An expansion would consist of adding a second row to the barrier if a breakthrough is observed. This would provide a new amount of PetroFix® and increase the adsorption spaces if the PRB becomes saturated with VOCs. Another option would be the restoration of the PetroFix® adsorption spaces by stimulating biotic or abiotic destruction of VOCs after the initial injection event. Chlorinated VOCs are less adsorptive than petroleum hydrocarbons (PHCs), therefore, chlorinated VOCs would be the first to breakthrough the PRB. Adding zero valent iron (ZVI) would abiotically destroy TCE and increase the adsorption space on the injection material without creating low adsorption daughter products. ZVI can also further reduce the environment, promoting the use of PHCs as electron donors for biotic dechlorination. Alternatively, the addition of dechlorinating bacteria, if not already naturally present, can assist in dechlorination and the increased availability of adsorption spaces on the PetroFix® PRB. The final option targets PHCs instead. Since the PHCs are present at significantly greater concentrations than chlorinated compounds, the addition of an electron acceptor, like oxygen, could rapidly increase the rate of PHC bioremediation. This would also increase the available adsorption space on the PetroFix®; however, this would stunt dechlorination.

POST-INJECTION MONITORING WELL INSTALLATION

An additional monitoring well will be installed after the completion of the injections in case the wells located near the PRB are impacted by the PetroFix® product entering the wells. The new well (GZM-05) will be installed in the downgradient area at the northern corner downgradient of the property as shown on **Figure 2**.

The GZM-05 will be set with a 10-foot screen interval at approximately 15 to 25 feet below grade, or at depths that straddle the observed water table, to sample the groundwater at the site. The well location will be predrilled with a DPT drill rig to confirm the depth to water and screen interval depths at the location and then over-drilled utilizing a hollow stem auger drill rig to install the monitoring well. The new monitoring well will be constructed of 2-inch diameter PVC with a 10-foot long 0.01-inch slotted well screen set in filter sand to two feet above the screen interval. Community Air Monitoring Plan (CAMP) will be conducted during intrusive work in accordance with the approved Remedial Action Workplan for the Site, dated May 2019.

The monitoring well will be developed prior to sampling by surging/bailing, using a centrifugal pump and dedicated polyethylene tubing, or by Waterra positive displacement pumps and dedicated polyethylene tubing, or other methods at the discretion of the Field Manager/Site Supervisor. The development water will be contained in a tank on site or in drums. The well will be developed until turbidity is less than 50 Nephelometric Turbidity Units (NTUs) for three (3) successive 5-minute reading or until at least three well volumes are purged. All monitoring well development will be overseen by a field geologist or engineer and the duration, method of development, and approximate volume of water removed will be recorded in the field book.

An updated Quality Assurance Project Plan (QAPP)/Field Sampling Plan (FSP) with activities included from this Work Plan describes specific protocols for field sampling, sample handling and storage, chain-of-custody, laboratory analysis, and data handling and management. The QAAP/FSP can be found in **Attachment B**.



GROUNDWATER SAMPLING

Two rounds of groundwater monitoring will be completed after the completion of the PRB injections. The first sampling event will occur eight weeks after the injections and the second round will occur 12 weeks after the injections before transitioning to quarterly sampling events under the Site Management Plan (SMP). The transition to quarterly sampling will be dependent on sample results and NYSDEC approval. During both sampling events, the 5 monitoring wells will be sampled in accordance with EPA's Low-Flow (minimal drawdown) Groundwater Sampling procedures. Groundwater sampling will be utilized to determine the effectiveness of the PRB by observing contaminant concentrations at wells within and downgradient of the PRB (GZM-1, GZM-2, and GZM-5).

The following materials, as required, shall be available during groundwater sampling:

- Sample pump (peristaltic)
- Sample tubing
- Power source (i.e., generator, battery)
- Dedicated or disposable bailers
- New disposable polypropylene rope
- Buckets to measure purge water
- Water-level interface probe
- Conductivity/temperature meter
- pH meter
- Turbidity meter
- Appropriate water sample containers
- Appropriate blanks (trip blank supplied by the laboratory)
- Appropriate transport containers (coolers) with ice and appropriate labeling, packing, and shipping materials
- Groundwater sampling logs
- Chain of Custody forms
- Indelible ink pens
- Site map with well locations

Prior to sampling, groundwater elevations will be measured at each monitoring well and the presence of LNAPL (if any) within the well will be evaluated. Depth to water, and depth to bottom measurements of each well will be collected using an interface probe and recorded on the sampling log sheet. The groundwater flow is expected to be to the north-northeast.

After groundwater elevations are measured and NAPLs are determined not to be present, groundwater will be purged from the wells. If NAPLs are determined present, then a NAPL sample and a representative groundwater sample, will be collected using a peristaltic pump or other method in consultation with the NYSDEC.

Tubing (for peristaltic pumps) will be lowered slowly into the well to a depth corresponding to the center of the saturated screen section of the well. Purging rates will not exceed 500 milliliters per minute. During well purging, monitor the field indicator parameters (turbidity, temperature, specific conductance, pH, dissolved oxygen [DO], and oxidation-reduction potential [ORP]) every three to five minutes (or as appropriate). The well is considered stabilized and ready for sample collection when the indicator parameters have stabilized for three consecutive readings at the following limits:

- **Turbidity** (10% for values greater than 5 NTU; if three Turbidity values are less than 5 NTU, consider the values as stabilized);
- **DO** (10% for values greater than 0.5 mg/L; if three dissolved oxygen values are less than 0.5 mg/L, consider the values as stabilized);
- **Specific Conductance** (3%);
- **Temperature** (3%);
- **pH** (± 0.1 unit);
- **ORP** (± 10 millivolts).



Readings will be recorded utilizing a Horiba multimeter with flow through cell (or equivalent).

Groundwater samples will be collected directly from the dedicated tubing into laboratory-issued bottle ware. The vials will be filled completely and checked to ensure that no air bubbles are present. Samples will be packaged in laboratory-issued sample containers by GZA personnel and stored on ice pending same day or overnight shipment to a New York State ELAP laboratory subcontracted by GZA. All samples will be uniquely identified, and all information associated with the samples will be recorded utilizing standard chain-of custody protocols. Sample containers will then be placed on ice until delivered to the laboratory.

Groundwater samples from each well, during the first round of sampling, will be analyzed for the full suite of parameters, including the following:

- Target Compound List (TCL) VOCs by EPA Method 8260;
- TCL SVOCs by EPA Method 8270;
- Dissolved Target Analyte List (TAL) metals by EPA Methods 6010/7470;
- Total TAL metals by EPA Methods 6010/7470;
- PCBs by EPA Method 8082A;
- Pesticides by EPA Method 8081;
- PFAS by EPA Method 1633;

Groundwater samples during subsequent sampling events will be analyzed for TCL VOCs, TCL SVOCs, and total and dissolved TAL metals. All analysis will be reported using NYSDEC ASP Category B deliverables.

During this round of sampling, the following samples will be collected for QA/QC purposes in accordance with the approved QAPP:

- 1 trip blank
- 1 field blank
- 1 duplicate sample

The groundwater laboratory data will be reviewed by a qualified Data Validator and a Data Usability Summary Report (DUSR) will be prepared. The laboratory analytical results of the samples will be compared to NYSDEC TOGS groundwater standards and guidance values. Preliminary laboratory results will be summarized in a data table and on a figure highlighting exceedances of NYSDEC TOGS standards and guidance values to be provided to NYSDEC within 14 days of receipt from the laboratory. Monitoring well installation logs will be generated and will be included as an Appendix in the revised submittal of the Final Engineering Report (FER). The logs will contain any local condition(s) that occurred during the sampling that may influence interpretation of the results (e.g., weather). Additionally, logs will include parameters recorded during low flow sampling, depth to water, depth to bottom, monitoring well screen information, and construction details. All purge water will be drummed and sampled for proper off-site disposal.

In the event of a dust or odor complaint, or any exceedance related to the CAMP, the NYSDEC and the NYSDOH project managers will be notified within 24 hours. Any CAMP exceedance will be reported to the NYSDEC and the NYSDOH on the same business day and as soon as possible. Notification of the exceedance will be sent via email long with the reason for the exceedance, the measure(s) taken to address the exceedance, and if the exceedance was resolved.



Daily work reports will be submitted to the NYSDEC and the NYSDOH PMs by the end of each day following the reporting period and will include:

- An update of progress made during each day;
- Locations of work being performed;
- Photographs of work completed each day;
- A summary of work planned for the following day;
- The names and titles of staff performing on-Site activities;
- A timeline of daily work activities, including start and end times;
- A summary of any and all complaints with relevant details (names, phone numbers);
- A summary of CAMP findings, including exceedances;
- A summary of any data collected in association with on-site activities;
- An explanation of notable site conditions; and,
- Any deviations from the approved work plan.

Daily reporting and CAMP will be conducted during ground intrusive activities and soil handling activities. NYSDEC and the NYSDOH will be notified of CAMP results that exceed action levels, including duration and actions taken in response to any such exceedance. Notifications will be provided to the NYSDEC and the NYSDOH as soon as possible, within a minimum 24 hours from the exceedance event.

Daily reports are not intended to be the mode of communication for notification to the NYSDEC or NYSDOH of emergencies (accidents, spills, etc.), requests for changes to the Monitoring Well Installation and Sampling Work Plan or other sensitive or time critical information. However, such conditions must also be included in the daily reports. Any and all site notifications, including emergency conditions and changes to the Monitoring Well Installation and Sampling Work Plan, will be addressed directly to the NYSDEC PM and the NYSDOH PM via personal communication.

If wells within the area of the PRB (GZM-1 and GZM-2) have evidence of the PetroFix® in the initial purge water, purging will cease, and the wells will not be sampled to prevent the treatment chemicals from being removed from the treatment target area. Regensis recommends delaying any monitoring if suspended PetroFix® concentrations exceed 100 ppm based on the test kit provided with the shipment. If groundwater samples cannot be delayed, there are multiple methods to navigate the situation. The first includes GZA will remobilizing to collect groundwater samples with Passive Diffusion Bags (PDBs). The PDBs consist of a sealed low-density polyethylene (LDPE) bag containing deionized water. The PDBs will be installed and left in place for two weeks to allow groundwater to equilibrate. The PDBs will be deployed at the screened interval of the wells. After two weeks, the PDBs will be retrieved from the wells and the samples will be decanted into laboratory supplied containers. PDBs will only be used to sample from wells within the PRB and will not be used to collect groundwater samples to be considered representative of downgradient conditions.

The sample vials will be filled completely and checked to ensure that no air bubbles are present. Samples will be packaged in laboratory-issued sample contained by GZA personnel and stored on ice pending same day or overnight shipment to a New York State ELAP laboratory subcontracted by GZA. All samples will be uniquely identified, and all information associated with the samples will be recorded utilizing standard chain-of custody protocols. Sample containers will then be placed on ice until delivered to the laboratory.

Groundwater samples from each well will be analyzed for Target Compound List (TCL) VOCs and TCL SVOCs. All analysis will be reported using NYSDEC ASP Category B deliverables. GZA also will collect samples for geochemical and biologic analyses to evaluate whether contamination is migrating offsite; evaluation of the



off-site groundwater will help determine that the migration of contamination was prevented, as the injection points are along the Site boundary. We will analyze for parameters that indicate the geochemical environment in the subsurface (i.e., DO, pH, ORP, conductivity); the geochemical parameters affecting contaminant degradation (i.e., nitrate and sulfate); and potential products of contaminant degradation (i.e., methane, and carbon dioxide).

HEALTH AND SAFETY

All remedial work performed under this work plan will be completed in accordance with the health and safety requirements established in the Construction Health and Safety Plan (CHASP) provided in the NYSDEC approved May 2019 RAWP. The CHASP was prepared to protect on-site personnel, visitors, and the public from physical harm and exposure to hazardous materials or waste during remedial activities. In accordance with the Occupational Safety and Health Administration (OSHA) 29 CFR Part 1910.120 Hazardous Waste Operations and Emergency Response Final rule, the CHASP addressed safety and health hazards related to excavation, loading, and other soil disturbance activities.

REPORTING

Following the groundwater sampling event, the results will be summarized in a groundwater monitoring letter report submitted to the NYSDEC. Data from the injections for the PRB and the groundwater monitoring events will be summarized and included in the revised FER submittal.

SCHEDULE

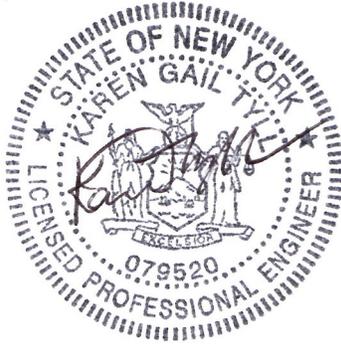
The table below presents a schedule for the permeable reactive barrier and reporting. If the schedule activities change, it will be updated and submitted to NYSDEC, as necessary.

Milestone	Anticipated Start Date	Anticipated End Date
Installation of Monitoring Wells	10/22/2025	10/27/2025
Pre-Injection Sampling Event	11/4/2025	11/5/2025
Pre-Injection Groundwater Sampling Letter	11/6/2025	12/11/2025
Permeable Reactive Barrier Injections	2/18/2026	2/27/2026
Post-Injection Sampling Event 1	4/24/2026	4/25/2026
Post-Injection Sampling Event 2	5/22/2026	5/23/2026
FER Resubmittal	6/1/2026	



CERTIFICATIONS

I, Karen G. Tyll, certify that I am currently a [NYS registered professional engineer or Qualified Environmental Professional as defined in 6 NYCRR Part 375] and that this Report [Remedial Design, Remedial Action Work Plan] was prepared in accordance with all applicable statutes and regulations and in substantial conformance with the DER Technical Guidance for Site Investigation and Remediation (DER-10).



12/31/2025

NYS Professional Engineer #

Date

Signature



Should you have any questions, please contact Victoria Whelan at Victoria.Whelan@gza.com.

Very truly yours,
GZA GEOENVIRONMENTAL OF NEW YORK


Zachary Landis, P.E.
Project Manager


Stephen M. Kline, P.E.
Consultant Reviewer


Victoria Whelan, P.G.
Vice President

Cc: Karen Tyll – TEC
Craig Livingston – Blondell Equities, LLC

Attachments:

Figure 1: Post Remediation Groundwater Analytical Results

Figure 2: Proposed Permeable Reactive Barrier Map

Table 1 – Sample Summary

Table 2 – VOCs in Groundwater

Table 3 – SVOCs in Groundwater

Table 4 – Total and Dissolved Metals in Groundwater

Table 5 – PCBs in Groundwater

Table 6 – PFAS in Groundwater

Table 7 – Pesticides in Groundwater

Attachment A - Regenesis PetroFix® Installation Data

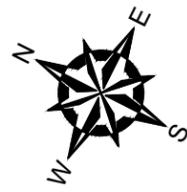
Attachment B – Quality Assurance Project Plan/Field Sampling Plan

Attachment C – November 2025 Post-Remediation Groundwater Purge Logs

Attachment D – November 2025 Post-Remediation Groundwater Laboratory Report



FIGURE 1
POST REMEDIATION GROUNDWATER ANALYTICAL RESULTS



GZM-2 - 11/4/25		GZM-DUP - 11/4/25	
SVOCs		SVOCs	
Benzo(a)anthracene	0.11	Benzo(a)anthracene	0.17
Dissolved Metals		Benzo(b)fluoranthene 0.04 J	
Iron, Dissolved	333	Dissolved Metals	
Manganese, Dissolved	1,855	Iron, Dissolved	626
Sodium, Dissolved	43,100	Manganese, Dissolved	1,942
Total Metals		Sodium, Dissolved 42,100	
Iron, Total	23,000	Total Metals	
Manganese, Total	1,814	Iron, Total	25,400
Sodium, Total	42,300	Manganese, Total	1,984
PFAS		Sodium, Total 42,800	
PFOS	0.0714	PFAS	
PFOA	0.025	PFOS	0.0688
		PFOA	0.0252

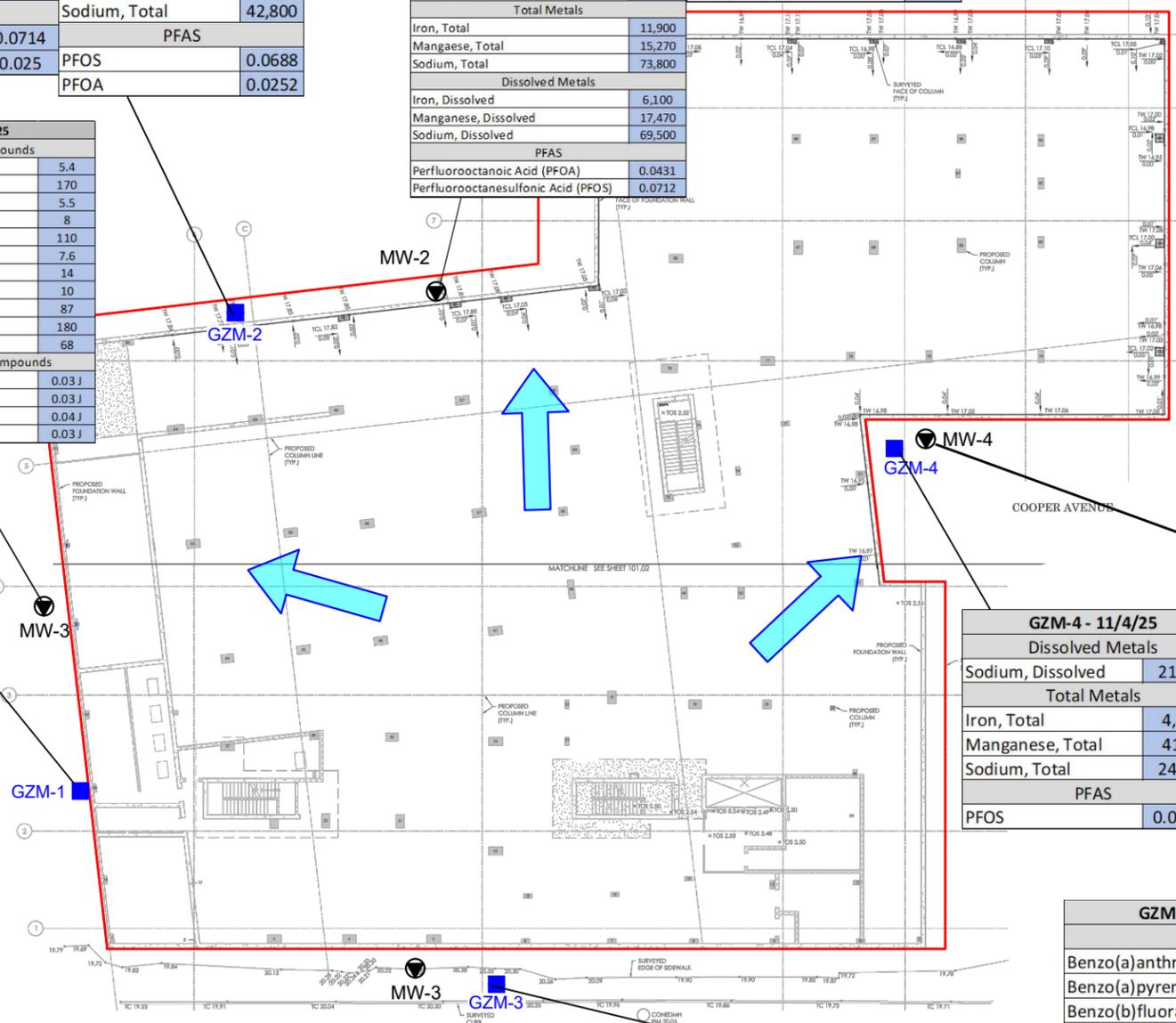
MW-2 - 11/13/2024		MW-2 - 1/14/2025	
Volatile Organic Compounds		Volatile Organic Compounds	
1,2,4,5-Tetramethylbenzene	10	1,2,4,5-Tetramethylbenzene	2.5
cis-1,2-Dichloroethene	13	cis-1,2-Dichloroethene	54
n-Propylbenzene	7.7	n-Propylbenzene	0.95
sec-Butylbenzene	5.5	sec-Butylbenzene	0.8
Trichloroethene	26	Trichloroethene	16
Vinyl chloride	0.22	Vinyl chloride	2
Semi-Volatile Organic Compounds		Semi-Volatile Organic Compounds	
Benzo(a)anthracene	0.08 J	Benzo(a)anthracene	0.08 J
Benzo(a)pyrene	0.05 J	Benzo(a)pyrene	0.07 J
Benzo(b)fluoranthene	0.07 J	Benzo(b)fluoranthene	0.09 J
Benzo(k)fluoranthene	0.1 U	Benzo(k)fluoranthene	0.04 J
Chrysene	0.05 J	Chrysene	0.08 J
Indeno(1,2,3-cd)pyrene	0.04 J	Indeno(1,2,3-cd)pyrene	0.07 J
Total Metals		Total Metals	
Iron, Total	11,900	Iron, Total	11,900
Manganese, Total	15,270	Manganese, Total	15,270
Sodium, Total	73,800	Sodium, Total	73,800
Dissolved Metals		Dissolved Metals	
Iron, Dissolved	6,100	Iron, Dissolved	6,100
Manganese, Dissolved	17,470	Manganese, Dissolved	17,470
Sodium, Dissolved	69,500	Sodium, Dissolved	69,500
PFAS		PFAS	
Perfluorooctanoic Acid (PFOA)	0.0431	Perfluorooctanoic Acid (PFOA)	0.0431
Perfluorooctanesulfonic Acid (PFOS)	0.0712	Perfluorooctanesulfonic Acid (PFOS)	0.0712

GZM-1 - 11/4/25	
VOCs	
1,2,4,5-Tetramethylbenzene	74
1,2,4-Trimethylbenzene	880
1,3,5-Trimethylbenzene	190
Benzene	85
Ethylbenzene	690
Isopropylbenzene	77
n-Butylbenzene	23 J
n-Propylbenzene	190
o-Xylene	280
p/m-Xylene	1,300
Toluene	25
SVOCs	
Naphthalene	34
Dissolved Metals	
Manganese, Dissolved	3,573
Sodium, Dissolved	244,000
Total Metals	
Iron, Total	21,200
Manganese, Total	3,825
Sodium, Total	229,000
PFAS	
PFOS	0.0347
PFOA	0.017

MW-1 - 1/14/2025	
Volatile Organic Compounds	
1,2,4,5-Tetramethylbenzene	5.4
1,2,4-Trimethylbenzene	170
1,3,5-Trimethylbenzene	5.5
Benzene	8
Ethylbenzene	110
Isopropylbenzene	7.6
n-Propylbenzene	14
Naphthalene	10
o-Xylene	87
p/m-Xylene	180
Toluene	68
Semi-Volatile Organic Compounds	
Benzo(a)anthracene	0.03 J
Benzo(a)pyrene	0.03 J
Benzo(b)fluoranthene	0.04 J
Indeno(1,2,3-cd)pyrene	0.03 J

NYSDEC TOGS 1.1.1 Ambient Water Quality Standards	
Analyte	Volatile Organic Compounds (VOCs) (µg/L)
1,2,4,5-Tetramethylbenzene	5
1,2,4-Trimethylbenzene	5
1,3,5-Trimethylbenzene	5
Benzene	1
Ethylbenzene	5
Isopropylbenzene	5
n-Butylbenzene	5
n-Propylbenzene	5
o-Xylene	5
p/m-Xylene	5
Toluene	5
Analyte	Semivolatile Organic Compounds - SIM (SVOCs) (µg/L)
Benzo(a)anthracene	0.002
Benzo(a)pyrene	0
Benzo(b)fluoranthene	0.002
Benzo(k)fluoranthene	0.002
Chrysene	0.002
Indeno(1,2,3-cd)pyrene	0.002
Naphthalene	10
Analyte	Dissolved Metals (µg/L)
Iron, Dissolved	300
Manganese, Dissolved	300
Sodium, Dissolved	20,000
Analyte	Total Metals (µg/L)
Iron, Total	300
Manganese, Total	300
Sodium, Total	20,000
Analyte	Per- and Polyfluoroalkyl Substances (PFAS) (µg/L)
Perfluorooctanesulfonic Acid (PFOS)	0.0027
Perfluorooctanoic Acid (PFOA)	0.0067

Value exceeds its New York TOGS 1.1.1. Ambient Water Quality Standard.



GZM-4 - 11/4/25	
Dissolved Metals	
Sodium, Dissolved	21,000
Total Metals	
Iron, Total	4,420
Manganese, Total	411.1
Sodium, Total	24,600
PFAS	
PFOS	0.00975

GZM-3 - 11/4/25	
SVOCs	
Benzo(a)anthracene	0.27
Benzo(a)pyrene	0.2
Benzo(b)fluoranthene	0.26
Benzo(k)fluoranthene	0.1
Chrysene	0.17
Indeno(1,2,3-cd)pyrene	0.16
Dissolved Metals	
Manganese, Dissolved	344.5
Sodium, Dissolved	265,000
Total Metals	
Iron, Total	1,620
Manganese, Total	489.3
Sodium, Total	248,000
PFAS	
PFOS	0.031
PFOA	0.0213

MW-4 - 11/13/2024		MW-4 - 1/14/2025	
Total Metals		Semi-Volatile Organic Compounds	
Iron, Total	3,520	Benzo(a)anthracene	0.03 J
Lead, Total	124.9	Benzo(a)pyrene	0.03 J
Manganese, Total	318.7	Benzo(b)fluoranthene	0.05 J
Sodium, Total	168,000	Indeno(1,2,3-cd)pyrene	0.05
Dissolved Metals		Dissolved Metals	
Sodium, Dissolved	165,000		
PFAS		PFAS	
Perfluorooctanoic Acid (PFOA)	0.012		
Perfluorooctanesulfonic Acid (PFOS)	0.0163		

GENERAL NOTES

1. BASE MAP DEVELOPED FROM DRAWING TITLED "CELLAR LEVEL WALLS AND COLUMNS AS-BUILT SURVEY", PREPARED BY "DOWNEY LAYOUT LLC", ORIGINAL SCALE 1"=20', DATED AUGUST 21, 2024.
2. GROUNDWATER MONITORING WELL LOCATIONS BASED ON SURVEY TITLED "MONITORING WELL LOCATION MAP" PREPARED BY "DPK LAND SURVEYING" ON NOVEMBER 7, 2025, ORIGINAL SCALE 1" = 50'.
3. SEE LABORATORY REPORTS FOR ADDITIONAL INFORMATION, INCLUDING QUALIFIER DESCRIPTIONS.
4. ONLY EXCEEDANCES OF THE APPLICABLE NYSDEC TOGS 1.1.1. AMBIENT WATER QUALITY STANDARDS ARE SHOWN. FOR FULL ANALYTICAL RESULTS, SEE TABLES LOCATED IN REPORT.

LEGEND

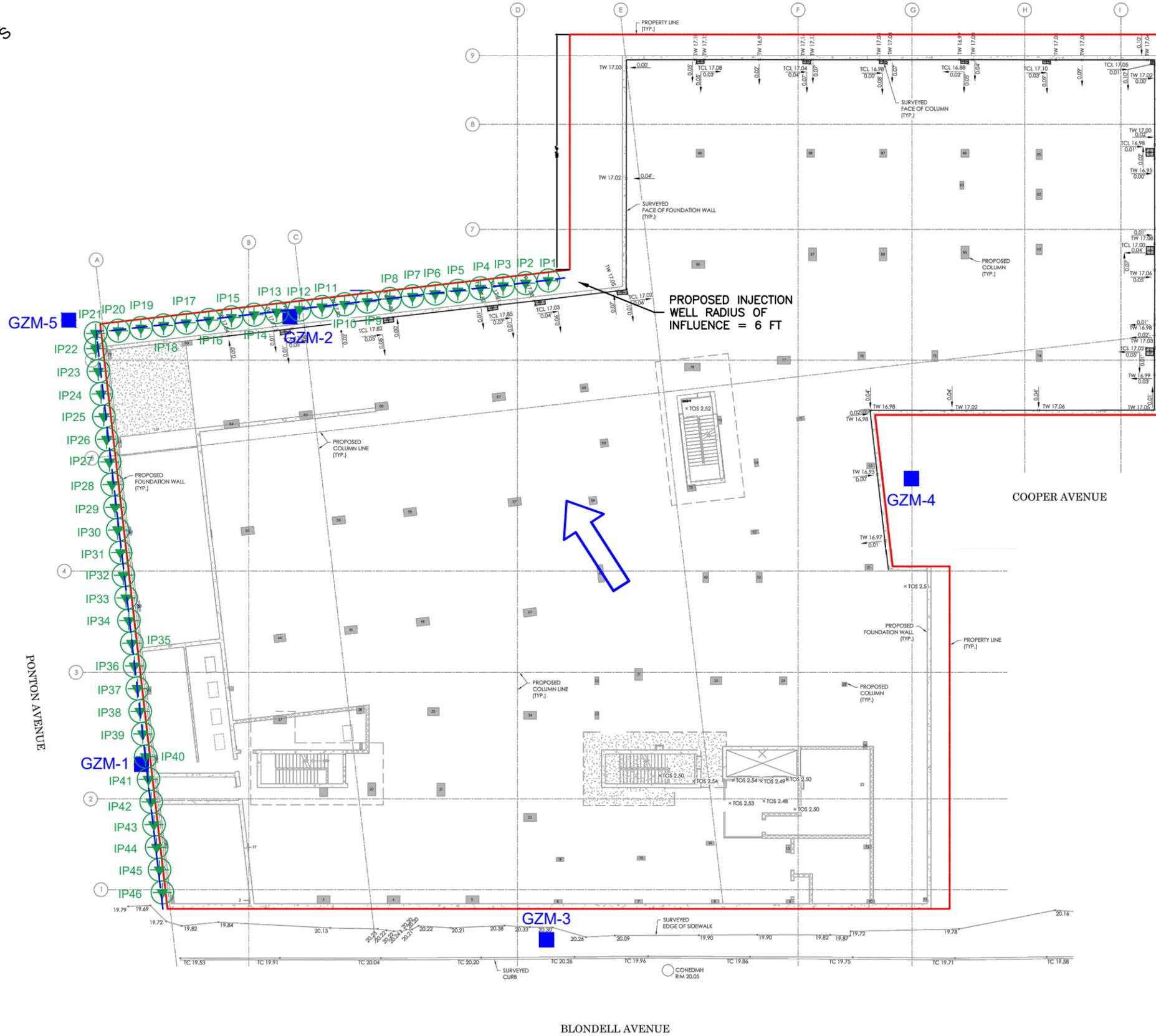
- BCP SITE BOUNDARY
- APPROXIMATE MONITORING WELL LOCATION
- GZM-1
- ➔ INFERRED GROUNDWATER FLOW DIRECTION



1346 BLONDELL AVENUE BRONX, NEW YORK	
POST-REMEDATION GROUNDWATER ANALYTICAL RESULTS	
PREPARED BY: GZA GeoEnvironmental of NY Engineers and Scientists www.gza.com	PREPARED FOR: BLONDELL EQUITIES LLC
PROJ MGR: ZL DESIGNED BY: SG DATE: DECEMBER 2025	REVIEWED BY: ZL DRAWN BY: SG PROJECT NO. 41.0163256.00
CHECKED BY: VW SCALE: 1" = 40' REVISION NO. -	FIGURE 1 SHEET NO.



FIGURE 2
PROPOSED PERMEABLE REACTIVE BARRIER MAP



GENERAL NOTES

1. BASE MAP DEVELOPED FROM DRAWING TITLED "CELLAR LEVEL WALLS AND COLUMNS AS-BUILT SURVEY", PREPARED BY "DOWNEY LAYOUT LLC", ORIGINAL SCALE 1"=20', DATED AUGUST 21, 2024.

LEGEND

- APPROXIMATE SITE BOUNDARY
- ➔ GROUNDWATER FLOW DIRECTION
- GZM-1 PROPOSED NEW MONITORING WELL LOCATIONS
- PROPOSED PERMEABLE REACTIVE BARRIER LOCATION
- ▼ IP PROPOSED INJECTION WELL LOCATION



NO.	ISSUE/DESCRIPTION	BY	DATE

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1346 BLONDELL AVENUE
 BRONX, NEW YORK

PROPOSED INJECTION MAP

PREPARED BY: GZA GeoEnvironmental of NY Engineers and Scientists www.gza.com		PREPARED FOR: BLONDELL EQUITIES LLC	
PROJ MGR: ZL DESIGNED BY: SG DATE: NOVEMBER 2025	REVIEWED BY: ZL DRAWN BY: SG PROJECT NO. 41.0163256.00	CHECKED BY: VW SCALE: 1" = 60' REVISION NO. -	FIGURE 2 SHEET NO.

SMH
 RIM 20.38

BLONDELL AVENUE

SMH
 RIM 20.01



TABLE 1
SAMPLE SUMMARY

Table 1 - Sample Summary

1346 Blondell Avenue
Bronx, New York

WELL INSTALLATION DETAILS						GROUNDWATER SAMPLING DETAILS			
Well ID	Lab Sample ID	Installation Date	Installation Area	Approximate TOC Elevation (NAVD 88)	Screen Interval (feet)	DTW (feet below TOC)	GWE (NAVD 88)	Sample Collection Date	Analysis
GZM-1	L2569921-02	10/23/25	West of Building	22.29	17-27	16.55	5.74	11/4/2025	VOCs, SVOCs, Metals (Total & Dissolved), PCBs, PFAS, Pesticides
GZM-2	L2569921-01	10/24/25	North of Building	12.21	5-15	6.59	5.62	11/4/2025	VOCs, SVOCs, Metals (Total & Dissolved), PCBs, PFAS, Pesticides
GZM-3	L2569921-03	10/23/25	South of Building	21.57	10-20	15.54	6.03	11/4/2025	VOCs, SVOCs, Metals (Total & Dissolved), PCBs, PFAS, Pesticides
GZM-4	L2569921-04	10/23/25	East of Building	12.22	5-10	6.71	5.51	11/4/2025	VOCs, SVOCs, Metals (Total & Dissolved), PCBs, PFAS, Pesticides

Notes:

TOC = Top of Casing

NAVD 88 - Elevation is feet above (+) mean sea level based on the North American Vertical Datum of 1988

DTW - Depth to water



TABLE 2
VOLATILE ORGANIC COMPOUNDS IN GROUNDWATER

Table 2 - Volatile Organic Compounds in Groundwater

1346 Blondell Avenue
Bronx, New York

LOCATION SAMPLING DATE LAB SAMPLE ID SAMPLE TYPE	NYSDEC TOGS 1.1.1 Ambient Water Quality Standards	GZM-1		GZM-2		GZM-3		GZM-4		GZM-DUP		FIELD BLANK		TRIP BLANK	
		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025	
		L2569921-02		L2569921-01		L2569921-03		L2569921-04		L2569921-05		L2569921-06		L2569921-07	
		WATER		WATER		WATER		WATER		WATER		WATER		WATER	
		Results	Qual												
Volatile Organics by GC/MS (µg/L)															
1,1,1,2-Tetrachloroethane	5	25	U	2.5	U										
1,1,1-Trichloroethane	5	25	U	2.5	U										
1,1,2,2-Tetrachloroethane	5	5	U	0.5	U										
1,1,2-Trichloroethane	1	15	U	1.5	U										
1,1-Dichloroethane	5	25	U	2.5	U										
1,1-Dichloroethene	5	5	U	0.5	U										
1,1-Dichloropropene	5	25	U	2.5	U										
1,2,3-Trichlorobenzene	5	25	U	2.5	U										
1,2,3-Trichloropropane	0.04	25	U	2.5	U										
1,2,4,5-Tetramethylbenzene	5	74		2	U	2	U	2	U	2	U	2	U	2	U
1,2,4-Trichlorobenzene	5	25	U	2.5	U										
1,2,4-Trimethylbenzene	5	880		2.5	U	1.3	J	0.73	J	2.5	U	2.5	U	2.5	U
1,2-Dibromo-3-chloropropane	0.04	25	U	2.5	U										
1,2-Dibromoethane	0.0006	20	U	2	U	2	U	2	U	2	U	2	U	2	U
1,2-Dichlorobenzene	3	25	U	2.5	U										
1,2-Dichloroethane	0.6	5	U	0.5	U										
1,2-Dichloroethene, Total	~	25	U	2.5	U										
1,2-Dichloropropane	1	10	U	1	U	1	U	1	U	1	U	1	U	1	U
1,3,5-Trimethylbenzene	5	190		2.5	U										
1,3-Dichlorobenzene	3	25	U	2.5	U										
1,3-Dichloropropane	5	25	U	2.5	U										
1,3-Dichloropropene, Total	~	5	U	0.5	U										
1,4-Dichlorobenzene	3	25	U	2.5	U										
1,4-Dioxane	0.35	2,500	U	250	U										
2,2-Dichloropropane	5	25	U	2.5	U										
2-Butanone	50	50	U	5	U	5	U	5	U	5	U	5	U	5	U
2-Hexanone	50	50	U	5	U	5	U	5	U	5	U	5	U	5	U
4-Methyl-2-pentanone	~	50	U	5	U	5	U	5	U	5	U	5	U	5	U
Acetone	50	50	U	5	U	7.3		3.3	J	5	U	5	U	5	U
Acrylonitrile	5	50	U	5	U	5	U	5	U	5	U	5	U	5	U
Benzene	1	85		0.85		0.35	J	0.5	U	0.78		0.5	U	0.5	U
Bromobenzene	5	25	U	2.5	U										
Bromochloromethane	5	25	U	2.5	U										
Bromodichloromethane	50	5	U	0.5	U										
Bromoform	50	20	U	2	U	2	U	2	U	2	U	2	U	2	U
Bromomethane	5	25	U	2.5	U										
Carbon disulfide	60	50	U	5	U	5	U	5	U	5	U	5	U	5	U
Carbon tetrachloride	5	5	U	0.5	U										
Chlorobenzene	5	25	U	2.5	U										
Chloroethane	5	25	U	2.5	U										
Chloroform	7	25	U	2.5	U										
Chloromethane	~	25	U	2.5	U										
cis-1,2-Dichloroethene	5	25	U	2.5	U										
cis-1,3-Dichloropropene	0.4	5	U	0.5	U										
Dibromochloromethane	50	5	U	0.5	U										
Dibromomethane	5	50	U	5	U	5	U	5	U	5	U	5	U	5	U
Dichlorodifluoromethane	5	50	U	5	U	5	U	5	U	5	U	5	U	5	U
Ethyl ether	~	25	U	2.5	U										
Ethylbenzene	5	690		2.5	U	1.1	J	2.5	U	2.5	U	2.5	U	2.5	U
Hexachlorobutadiene	0.5	25	U	2.5	U										
Isopropylbenzene	5	77		2.5	U										
Methyl tert butyl ether	10	25	U	0.51	J	2.5	U	2.5	U	0.52	J	2.5	U	2.5	U
Methylene chloride	5	25	U	2.5	U										
Naphthalene	10	25	U	2.5	U										
n-Butylbenzene	5	23	J	2.5	U										
n-Propylbenzene	5	190		2.5	U										
o-Chlorotoluene	5	25	U	2.5	U										
o-Xylene	5	280		2.5	U	0.97	J	2.5	U	2.5	U	2.5	U	2.5	U
p/m-Xylene	5	1,300		2.5	U	2.6		2.5	U	2.5	U	2.5	U	2.5	U
p-Chlorotoluene	5	25	U	2.5	U										
p-Diethylbenzene	~	93		2	U	2	U	2	U	2	U	2	U	2	U
p-Ethyltoluene	~	660		2	U	1.2	J	0.96	J	2	U	2	U	2	U
p-Isopropyltoluene	5	25	U	2.5	U										
sec-Butylbenzene	5	25	U	2.5	U										
Styrene	5	25	U	2.5	U										
tert-Butylbenzene	5	25	U	2.5	U										
Tetrachloroethene	5	5	U	0.5	U										
Toluene	5	25		2.5	U										
trans-1,2-Dichloroethene	5	25	U	2.5	U										
trans-1,3-Dichloropropene	0.4	5	U	0.5	U										
trans-1,4-Dichloro-2-butene	5	25	U	2.5	U										
Trichloroethene	5	5	U	0.5	U										
Trichlorofluoromethane	5	25	U	2.5	U										
Vinyl acetate	~	50	U	5	U	5	U	5	U	5	U	5	U	5	U
Vinyl chloride	2	10	U	0.12	J	1	U	1	U	0.13	J	1	U	1	U
Xylenes, Total	~	1,600		2.5	U	3.6	J	2.5	U	2.5	U	2.5	U	2.5	U

Table Notes:

- ~: No guidance value.
- µg/L: Micrograms per Liter.
- U: Not detected at the reported detection limit for the sample.
- J: Indicates an estimate value.
- Value exceeds its New York TOGS 1.1.1. Ambient Water Quality Standard.



TABLE 3
SEMIVOLATILE ORGANIC COMPOUNDS IN GROUNDWATER

Table 3 - Semivolatile Organic Compounds in Groundwater

1346 Blondell Avenue
Bronx, New York

LOCATION SAMPLING DATE LAB SAMPLE ID SAMPLE TYPE	NYSDEC TOGS 1.1.1 Ambient Water Quality Standards	GZM-1		GZM-2		GZM-3		GZM-4		GZM-DUP		FIELD BLANK	
		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025	
		L2569921-02		L2569921-01		L2569921-03		L2569921-04		L2569921-05		L2569921-06	
		WATER		WATER		WATER		WATER		WATER		WATER	
		Results	Qual										
Semivolatile Organics by GC/MS (µg/L)													
1,2,4,5-Tetrachlorobenzene	5	10	U										
1,2,4-Trichlorobenzene	5	5	U	5	U	5	U	5	U	5	U	5	U
1,2-Dichlorobenzene	3	2	U	2	U	2	U	2	U	2	U	2	U
1,3-Dichlorobenzene	3	2	U	2	U	2	U	2	U	2	U	2	U
1,4-Dichlorobenzene	3	2	U	2	U	2	U	2	U	2	U	2	U
2,4,5-Trichlorophenol	~	5	U	5	U	5	U	5	U	5	U	5	U
2,4,6-Trichlorophenol	~	5	U	5	U	5	U	5	U	5	U	5	U
2,4-Dichlorophenol	1	5	U	5	U	5	U	5	U	5	U	5	U
2,4-Dimethylphenol	50	3.5	J	5	U	5	U	5	U	5	U	5	U
2,4-Dinitrophenol	10	20	U										
2,4-Dinitrotoluene	5	5	U	5	U	5	U	5	U	5	U	5	U
2,6-Dinitrotoluene	5	5	U	5	U	5	U	5	U	5	U	5	U
2-Chlorophenol	~	2	U	2	U	2	U	2	U	2	U	2	U
2-Methylphenol	~	5	U	5	U	5	U	5	U	5	U	5	U
2-Nitroaniline	5	5	U	5	U	5	U	5	U	5	U	5	U
2-Nitrophenol	~	10	U										
3,3'-Dichlorobenzidine	5	5	U	5	U	5	U	5	U	5	U	5	U
3-Methylphenol/4-Methylphenol	~	3.9	J	5	U	5	U	5	U	5	U	5	U
3-Nitroaniline	5	5	U	5	U	5	U	5	U	5	U	5	U
4,6-Dinitro-o-cresol	~	10	U										
4-Bromophenyl phenyl ether	~	2	U	2	U	2	U	2	U	2	U	2	U
4-Chloroaniline	5	5	U	5	U	5	U	5	U	5	U	5	U
4-Chlorophenyl phenyl ether	~	2	U	2	U	2	U	2	U	2	U	2	U
4-Nitroaniline	5	5	U	5	U	5	U	5	U	5	U	5	U
4-Nitrophenol	~	10	U										
Acetophenone	~	5	U	5	U	5	U	5	U	5	U	5	U
Benzoic Acid	~	50	U	50	U	12	J	50	U	50	U	50	U
Benzyl Alcohol	~	2	U	2	U	2	U	2	U	2	U	2	U
Biphenyl	~	2	U	2	U	2	U	2	U	2	U	2	U
Bis(2-chloroethoxy)methane	5	5	U	5	U	5	U	5	U	5	U	5	U
Bis(2-chloroethyl)ether	1	2	U	2	U	2	U	2	U	2	U	2	U
Bis(2-chloroisopropyl)ether	5	2	U	2	U	2	U	2	U	2	U	2	U
Bis(2-ethylhexyl)phthalate	5	3	U	3	U	3	U	3	U	3	U	3	U
Butyl benzyl phthalate	50	5	U	5	U	5	U	5	U	5	U	5	U
Carbazole	~	2	U	2	U	2	U	2	U	2	U	2	U
Dibenzofuran	~	2	U	2	U	2	U	2	U	2	U	2	U
Diethyl phthalate	50	5	U	5	U	5	U	5	U	5	U	5	U
Dimethyl phthalate	50	5	U	5	U	5	U	5	U	5	U	5	U
Di-n-butylphthalate	50	1	J	5	U	5	U	5	U	1	J	5	U
Di-n-octylphthalate	50	5	U	5	U	5	U	5	U	5	U	5	U
Hexachlorocyclopentadiene	5	20	U										
Isophorone	50	5	U	5	U	5	U	5	U	5	U	5	U
NDPA/DPA	50	2	U	2	U	2	U	2	U	2	U	2	U
Nitrobenzene	0.4	2	U	2	U	2	U	2	U	2	U	2	U
n-Nitrosodi-n-propylamine	~	5	U	5	U	5	U	5	U	5	U	5	U
p-Chloro-m-cresol	~	2	U	2	U	2	U	2	U	2	U	2	U
Phenol	1	5	U	5	U	5	U	5	U	5	U	5	U
Semivolatile Organics by GC/MS-SIM (µg/L)													
2-Chloronaphthalene	10	0.2	U										
2-Methylnaphthalene	~	12		0.05	J	0.14		0.08	J	0.05	J	0.1	U
Acenaphthene	20	0.24		0.1	U	0.11		0.12		0.1	U	0.1	U
Acenaphthylene	~	0.07	J	0.1	U								
Anthracene	50	0.05	J	0.1	U	0.03	J	0.09	J	0.06	J	0.1	U
Benzo(a)anthracene	0.002	0.1	U	0.11		0.1	U	0.27		0.17		0.1	U
Benzo(a)pyrene	0	0.1	U	0.1	U	0.1	U	0.2		0.1	U	0.1	U
Benzo(b)fluoranthene	0.002	0.1	U	0.1	U	0.1	U	0.26		0.04	J	0.1	U
Benzo(ghi)perylene	~	0.1	U	0.1	U	0.1	U	0.15		0.1	U	0.1	U
Benzo(k)fluoranthene	0.002	0.1	U	0.1	U	0.1	U	0.1		0.1	U	0.1	U
Chrysene	0.002	0.1	U	0.1	U	0.1	U	0.17		0.1	U	0.1	U
Dibenzo(a,h)anthracene	~	0.1	U	0.1	U	0.1	U	0.04	J	0.1	U	0.1	U
Fluoranthene	50	0.11		0.1	U	0.05	J	0.44		0.06	J	0.1	U
Fluorene	50	0.26		0.1	U	0.1	U	0.07	J	0.1	U	0.1	U
Hexachlorobenzene	0.04	0.8	U										
Hexachlorobutadiene	0.5	0.5	U										
Hexachloroethane	5	0.8	U										
Indeno(1,2,3-cd)pyrene	0.002	0.1	U	0.1	U	0.1	U	0.16		0.1	U	0.1	U
Naphthalene	10	34		0.4		0.99		0.43		0.45		0.1	U
Pentachlorophenol	1	0.8	U	0.8	U	0.8	U	0.8	U	0.07	J	0.8	U
Phenanthrene	50	0.21		0.1	U	0.06	J	0.27		0.05	J	0.1	U
Pyrene	50	0.09	J	0.1	U	0.05	J	0.39		0.07	J	0.1	U

Table Notes:

- ~: No guidance value.
- µg/L: Micrograms per Liter.
- U: Not detected at the reported detection limit for the sample.
- J: Indicates an estimate value.
- Value exceeds its New York TOGS 1.1.1. Ambient Water Quality Standard.



TABLE 4
TOTAL AND DISSOLVED METALS IN GROUNDWATER

Table 4 - Total and Dissolved Metals in Groundwater

1346 Blondell Avenue
Bronx, New York

LOCATION SAMPLING DATE LAB SAMPLE ID SAMPLE TYPE	NYSDEC TOGS 1.1.1 Ambient Water Quality Standards	GZM-1		GZM-2		GZM-3		GZM-4		GZM-DUP		FIELD BLANK	
		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025	
		L2569921-02		L2569921-01		L2569921-03		L2569921-04		L2569921-05		L2569921-06	
		WATER		WATER		WATER		WATER		WATER		WATER	
		Results	Qual										
Dissolved Metals (µg/L)													
Aluminum, Dissolved	~	10	U	4.31	J	55.1		5.51	J	10.8		10	U
Antimony, Dissolved	3	4	U	4	U	0.8	J	2.01	J	4	U	4	U
Arsenic, Dissolved	25	0.6		0.67		2.12		2.72		0.75		0.5	U
Barium, Dissolved	1,000	314.9		139		110.9		38.89		145		0.5	U
Beryllium, Dissolved	3	0.5	U										
Cadmium, Dissolved	5	0.2	U										
Calcium, Dissolved	~	121,000		192,000		70,600		121,000		193,000		61.4	J
Chromium, Dissolved	50	1	U	0.22	J	0.38	J	1	U	0.62	J	1	U
Cobalt, Dissolved	~	0.29	J	1.2		0.37	J	0.85		1.28		0.5	U
Copper, Dissolved	200	1	U	1	U	0.89	J	0.66	J	1	U	3.5	
Iron, Dissolved	300	35.8	J	333		19.7	J	50	U	626		50	U
Lead, Dissolved	25	0.6	J	1	U	1	U	0.41	J	1	U	1	U
Magnesium, Dissolved	35,000	25,100		22,600		9,560		22,200		22,000		70	U
Manganese, Dissolved	300	3,573		1,855		344.5		202.6		1,942		1	U
Mercury, Dissolved	0.7	0.2	U										
Nickel, Dissolved	100	0.71	J	2.27		2.23		3.73		2.2		3.05	
Potassium, Dissolved	~	18,800		22,200		28,200		12,000		21,500		100	U
Selenium, Dissolved	10	5	U	5	U	5	U	5	U	5	U	5	U
Silver, Dissolved	50	0.4	U										
Sodium, Dissolved	20,000	244,000		43,100		265,000		21,000		42,100		184	J
Thallium, Dissolved	0.5	1	U	1	U	1	U	1	U	1	U	1	U
Vanadium, Dissolved	~	5	U	5	U	2.72	J	5	U	5	U	5	U
Zinc, Dissolved	2,000	10	U	4.33	J	10	U	21.83		10	U	10	U
Total Metals (µg/L)													
Aluminum, Total	~	1,890		247		1,170		2,600		370		10	U
Antimony, Total	3	4	U	4	U	0.55	J	2.12	J	4	U	4	U
Arsenic, Total	25	3.11		3.57		2.39		5.57		3.99		0.5	U
Barium, Total	1,000	436.3		178.1		132.4		106.4		187.3		0.18	JB
Beryllium, Total	3	0.5	U	0.5	U	0.5	U	0.18	J	0.5	U	0.5	U
Cadmium, Total	5	0.2	U	0.07	J	0.2	U	0.35		0.09	J	0.2	U
Calcium, Total	~	127,000		182,000		69,100		177,000		192,000		100	U
Chromium, Total	50	4.75		1.28		2.66		4.86		1.03		1	U
Cobalt, Total	~	1.95		1.48		1.42		2.82		1.61		0.5	U
Copper, Total	200	18.84		3.94		16.3		13.67		4.34		1	U
Iron, Total	300	21,200		23,000		1,620		4,420		25,400		50	U
Lead, Total	25	3.89		4.12		2.94		83.69		5.14		1	U
Magnesium, Total	35,000	26,600		21,600		9,610		29,600		22,300		70	U
Manganese, Total	300	3,825		1,814		489.3		411.1		1,984		1	U
Mercury, Total	0.7	0.2	U	0.2	U	0.2	U	0.09	J	0.2	U	0.2	U
Nickel, Total	100	4.61		3.58		4.41		9.07		3.57		2	U
Potassium, Total	~	19,600		19,500		27,500		12,800		20,000		100	U
Selenium, Total	10	5	U	5	U	5	U	5	U	5	U	5	U
Silver, Total	50	0.4	U										
Sodium, Total	20,000	229,000		42,300		248,000		24,600		42,800		180	J
Thallium, Total	0.5	1	U	1	U	1	U	1	U	1	U	1	U
Vanadium, Total	~	5.49		5	U	5.27		8.75		1.64	J	5	U
Zinc, Total	2,000	10.22		17.49		6.2	J	97.18		19.88		10	U

Table Notes:

- ~: No guidance value.
- µg/L: Micrograms per Liter.
- U: Not detected at the reported detection limit for the sample.
- J: Indicates an estimate value.
- B: Indicates analyte found in the analysis batch blank.
- Value exceeds its New York TOGS 1.1.1. Ambient Water Quality Standard.





TABLE 5
POLYCHLORINATED BIPHENYLS IN GROUNDWATER



TABLE 6
PER- AND POLYFLUOROALKYL SUBSTANCES IN GROUNDWATER

Table 6 - Per- and Polyfluoroalkyl Substances in Groundwater

1346 Blondell Avenue
Bronx, New York

LOCATION SAMPLING DATE LAB SAMPLE ID SAMPLE TYPE	NYSDEC TOGS 1.1.1 Ambient Water Quality Standards	GZM-1		GZM-2		GZM-3		GZM-4		GZM-DUP		FIELD BLANK	
		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025	
		L2569921-02		L2569921-01		L2569921-03		L2569921-04		L2569921-05		L2569921-06	
		WATER		WATER		WATER		WATER		WATER		WATER	
		Results	Qual										
Perfluorinated Alkyl Acids by EPA 1633 (µg/L)													
11-Chloroeicosfluoro-3-Oxaundecane-1-Sulfonic Acid (11Cl-PF3OUdS)	~	0.00626	U	0.00634	U	0.00574	U	0.00579	U	0.00633	U	0.00563	U
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2FTS)	~	0.00626	U	0.00634	U	0.00574	U	0.00579	U	0.00633	U	0.00563	U
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2FTS)	~	0.00626	U	0.00634	U	0.00574	U	0.00579	U	0.00633	U	0.00563	U
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2FTS)	~	0.0225	J	0.00634	U	0.00327	J	0.00579	U	0.00438	J	0.00563	U
2H,2H,3H,3H-Perfluorooctanoic Acid (5:3FTCA)	~	0.0391	U	0.0396	U	0.0359	U	0.0362	U	0.0396	U	0.0352	U
3-Perfluoroheptyl Propanoic Acid (7:3FTCA)	~	0.0391	U	0.0396	U	0.0359	U	0.0362	U	0.0396	U	0.0352	U
3-Perfluoropropyl Propanoic Acid (3:3FTCA)	~	0.00783	U	0.00792	U	0.00717	U	0.00724	U	0.00791	U	0.00704	U
4,8-Dioxo-3h-Perfluorononanoic Acid (ADONA)	~	0.00626	U	0.00634	U	0.00574	U	0.00579	U	0.00633	U	0.00563	U
9-Chlorohexadecafluoro-3-Oxanone-1-Sulfonic Acid (9Cl-PF3ONS)	~	0.00626	U	0.00634	U	0.00574	U	0.00579	U	0.00633	U	0.00563	U
Hexafluoropropylene Oxide Dimer Acid (HFPO-DA)	~	0.00626	U	0.00634	U	0.00574	U	0.00579	U	0.00633	U	0.00563	U
N-Ethyl Perfluorooctane Sulfonamide (NEtFOSA)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
N-Ethyl Perfluorooctanesulfonamido Ethanol (NEtFOSE)	~	0.0156	U	0.0158	U	0.0143	U	0.0145	U	0.0158	U	0.0141	U
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (NEtFOSAA)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
N-Methyl Perfluorooctane Sulfonamide (NMeFOSA)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
N-Methyl Perfluorooctanesulfonamido Ethanol (NMeFOSE)	~	0.0156	U	0.0158	U	0.0143	U	0.0145	U	0.0158	U	0.0141	U
N-Methyl Perfluorooctanesulfonamidoacetic Acid (NMeFOSAA)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)	~	0.00313	U	0.00317	U	0.00287	U	0.00289	U	0.00316	U	0.00281	U
Perfluoro(2-Ethoxyethane)Sulfonic Acid (PFEEESA)	~	0.00313	U	0.00317	U	0.00287	U	0.00289	U	0.00316	U	0.00281	U
Perfluoro-3-Methoxypropanoic Acid (PFMPA)	~	0.000376	J	0.0158	U	0.00287	U	0.00289	U	0.0158	U	0.00281	U
Perfluoro-4-Methoxybutanoic Acid (PFMBA)	~	0.00313	U	0.00317	U	0.00287	U	0.00289	U	0.00316	U	0.00281	U
Perfluorobutanesulfonic Acid (PFBS)	~	0.00501	U	0.00448	U	0.00877	U	0.00342	U	0.00429	U	0.00141	U
Perfluorobutanoic Acid (PFBA)	~	0.0103	U	0.0317	U	0.0122	U	0.0086	U	0.0196	J	0.00563	U
Perfluorodecanesulfonic Acid (PFDS)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
Perfluorodecanoic Acid (PFDA)	~	0.00147	J	0.00114	J	0.00171	U	0.00097	J	0.00117	J	0.00141	U
Perfluorododecanesulfonic Acid (PFDoS)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
Perfluorododecanoic Acid (PFDoA)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
Perfluoroheptanesulfonic Acid (PFHpS)	~	0.00156	U	0.000729	J	0.00143	U	0.00145	U	0.00108	J	0.00141	U
Perfluoroheptanoic Acid (PFHpA)	~	0.00531	U	0.008	U	0.00644	U	0.00256	U	0.00707	U	0.00141	U
Perfluorohexanesulfonic Acid (PFHxS)	~	0.00298	U	0.00284	U	0.00235	U	0.000738	J	0.00293	U	0.00141	U
Perfluorohexanoic Acid (PFHxA)	~	0.0195	U	0.0111	U	0.0129	U	0.00824	U	0.0129	U	0.00141	U
Perfluorononanesulfonic Acid (PFNS)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
Perfluorononanoic Acid (PFNA)	~	0.0114	U	0.0043	U	0.00373	U	0.00132	J	0.00478	U	0.00141	U
Perfluorooctanesulfonamide (PFOSA)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
Perfluorooctanesulfonic Acid (PFOS)	0.0027	0.0347	U	0.0714	U	0.031	U	0.00975	U	0.0688	U	0.000577	J
Perfluorooctanoic Acid (PFOA)	0.0067	0.017	U	0.025	U	0.0213	U	0.00424	U	0.0252	U	0.00141	U
Perfluoropentanesulfonic Acid (PFPeS)	~	0.00156	U	0.000792	J	0.000344	JF	0.00145	U	0.000807	J	0.00141	U
Perfluoropentanoic Acid (PFPeA)	~	0.0218	U	0.0122	U	0.0157	U	0.00615	U	0.0115	U	0.00281	U
Perfluorotetradecanoic Acid (PFTeDA)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
Perfluorotridecanoic Acid (PFTrDA)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U
Perfluoroundecanoic Acid (PFUnA)	~	0.00156	U	0.00158	U	0.00143	U	0.00145	U	0.00158	U	0.00141	U

Table Notes:

- ~: No guidance value.
- µg/L: Micrograms per Liter.
- U: Not detected at the reported detection limit for the sample.
- J: Indicates an estimate value.
- F: The ratio of quantifier ion response to qualifier ion response falls outside of the laboratory criteria. Results are considered to be an estimated maximum concentration.

Value exceeds its New York TOGS 1.1.1. Ambient Water Quality Standard.



TABLE 7
PESTICIDES IN GROUNDWATER

Table 7 - Pesticides in Groundwater

1346 Blondell Avenue
Bronx, New York

LOCATION SAMPLING DATE LAB SAMPLE ID SAMPLE TYPE	NYSDEC TOGS 1.1.1 Ambient Water Quality Standards	GZM-1		GZM-2		GZM-3		GZM-4		GZM-DUP		FIELD BLANK	
		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025		11/4/2025	
		L2569921-02		L2569921-01		L2569921-03		L2569921-04		L2569921-05		L2569921-06	
		WATER		WATER		WATER		WATER		WATER		WATER	
		Results	Qual										
Organochlorine Pesticides by GC (µg/L)													
Delta-BHC	0.04	0.014	U										
Lindane	0.05	0.014	U										
Alpha-BHC	0.01	0.014	U										
Beta-BHC	0.04	0.02	U										
Heptachlor	0.04	0.014	U										
Aldrin	0	0.014	U										
Heptachlor epoxide	0.03	0.014	U										
Endrin	0	0.029	U										
Endrin aldehyde	5	0.03	U										
Endrin ketone	5	0.029	U										
Dieldrin	0.004	0.029	U										
4,4'-DDE	0.2	0.029	U										
4,4'-DDD	0.3	0.029	U										
4,4'-DDT	0.2	0.029	U	0.029	U	0.024	J	0.029	U	0.029	U	0.025	J
Endosulfan I	~	0.014	U										
Endosulfan II	~	0.029	U										
Endosulfan sulfate	~	0.029	U										
Methoxychlor	35	0.143	U										
Toxaphene	0.06	0.2	U										
cis-Chlordane	~	0.02	U										
trans-Chlordane	~	0.02	U										
Chlordane	0.05	0.143	U										

Table Notes:

~: No guidance value.

µg/L: Micrograms per Liter.

U: Not detected at the reported detection limit for the sample.

J: Indicates an estimate value.





ATTACHMENT A
REGENESIS PETROFIX® INSTALLATION DATA

	0.22	Vinyl chloride	2
Volatile Organic Compounds		Semi-Volatile Organic Compounds	
	0.08 J	Benzo(a)anthracene	0.08 J
	0.05 J	Benzo(a)pyrene	0.07 J
	0.07 J	Benzo(b)fluoranthene	0.09 J
	0.1 U	Benzo(k)fluoranthene	0.04 J
	0.05 J	Chrysene	0.08 J
	0.04 J	Indeno(1,2,3-cd)pyrene	0.07 J
Total Metals			
	11,900		
	15,270		
	73,800		
Dissolved Metals			
	6,100		
	17,470		
	69,500		
PFAS			
	0.0431		
	0.0712		



Blondell Ave
 GZA GeoEnvironmental
 September 24, 2025

Figure 1-Treatment Area Map





Injection Grid Application Summary



*Blondell
PRB*

PetroFix Amount	8,800 lb
Electron Acceptor	440 lb
Treatment Surface Area	3,240.0 ft ²
Delivery Points	46
Point Spacing	8.4 ft
Top of Treatment Interval	16.0 ft bgs
Bottom of Treatment Interval	26.0 ft bgs
Vertical Treatment Interval Thickness	10.0 ft
Treatment Volume	1,200 yd ³
PetroFix Dose	7.33 lb/yd ³

Total Volume	24,238 gal
Product Volume	901 gal
Water Volume	23,338 gal
Injection Volume/Point	527 gal
Inject Volume/Vertical ft	53 gal
Product/Point	19.6 gal
Water/Point	507.3 gal
Soil Type	Mix of coarse and fine
Effective Pore Volume Fill %	50%

Mix Tank Volume*	275.0 gal
Dilution Factor*	26.91
PetroFix per Mix Tank	10 gal
Water per Mix Tank	265 gal
Electron Acceptor per Mix Tank	5 lb
Total Batches Required	88.14

Specific Area Notes
Native Soil Type: Mix of coarse and fine

Reported Ground Water Concentrations (µg/L)

Benzene	85	Naphthalenes	34
Toluene	25	MTBE	0
Ethylbenzene	690	TPH-GRO	5,948
Xylenes	1,580	TPH-DRO	0
Trimethylbenzenes	1,000	Sum of Dissolved Concentrations:	6,982

PetroFix[™] Specification Sheet

PetroFix Technical Description

PetroFix is a new remedial technology designed to treat petroleum fuel spills in soil and groundwater. A simple-to-use fluid that can be applied under low pressure into the subsurface or simply poured into open excavations, PetroFix offers a cost-effective solution for environmental practitioners and responsible parties to address petroleum hydrocarbon contaminants quickly and effectively.

PetroFix has a dual function; quickly removing hydrocarbons from the dissolved phase, by absorbing them onto the activated carbon particles, while added electron acceptors stimulate hydrocarbon biodegradation in-place. PetroFix does not require high pressure “fracking” for application and can be applied with ease using readily available equipment associated with direct push technology.

The remedial fluid is a highly concentrated water-based suspension consisting of micron-scale activated carbon and biostimulating electron acceptors. PetroFix has a viscosity higher than water and is black in appearance. Its environmentally-compatible formulation of micron-scale activated carbon (1-2 microns) is combined with both slow and quick-release inorganic electron acceptors. A blend of additional electron acceptors is included along with the PetroFix fluid. Practitioners can select between a sulfate and nitrate combination blend (recommended), or sulfate only for the additional electron acceptors required.



PetroFix Design Assistant



REGENESIS has developed a proprietary web-based design assistant called PetroFix Design Assistant[™] that provides environmental professionals the ability to input their site parameters, determine the required product amount, and order the product through REGENESIS' customer service. The PetroFix Design Assistant includes defaults and warnings throughout the process to guide users toward effective designs that will offer best results.

To access the PetroFix Design Assistant, create an account and login at www.PetroFix.com

PetroFix Fluid Chemical Composition	Properties
Activated Carbon - CAS 7440-44-0 > 30% Calcium Sulfate Dihydrate - CAS 10101-41-4 < 10%	Appearance: Black Fluid Viscosity: 1500-3500 cP (corn syrup-like) pH: 8-10

PetroFix Electron Acceptor Powder Chemical Composition	Properties
OPTION 1 - EA Blend (preferred) Sodium Nitrate - CAS 7631-99-4, 50% Ammonium Sulfate - CAS 7783-20-2, 50% OPTION 2 - EA Blend NF Potassium Sulfate - CAS 7778-80-5, 50% Ammonium Sulfate - CAS 7783-20-2, 50%	Appearance: White Powder

Storage and Handling Guidelines	
Storage: <ul style="list-style-type: none"> • Store away from incompatible materials • Store in original closed container • Store at temperatures between 40°F and 95°F • Do not allow material to freeze or store in direct sunlight. • Freezing and hot weather technical memo can be accessed at www.petrofix.com/resources or at this link here. • Dispose of waste and residues in accordance with local authority requirements 	Handling: <ul style="list-style-type: none"> • Never add additives to solution prior to mixing with water • Wear appropriate personal protective equipment • Do not taste or ingest • Observe good industrial hygiene practices • Wash hands after handling

Applications

PetroFix is mixed with water on-site and easily applied onto the sub-surface using low pressure injections, or mixed in excavations. PetroFix is compatible with and can be used with ORC Advanced® to expedite rates of biodegradation. For more information about co-application with ORC Advanced, contact REGENESIS.



ATTACHMENT B
QUALITY ASSURANCE PROJECT PLAN/FIELD SAMPLING PLAN



Known for excellence.
Built on trust.

QUALITY ASSURANCE PROJECT PLAN (QAPP) / FIELD SAMPLING PLAN (FSP)

**1346 Blondell Avenue
Bronx, New York 10461
Block 4134, Lot 1
NYSDEC BCP Site No. C203089**

December 2025

PREPARED FOR:

Blondell Equities LLC

150 East 52nd Street – 14th Floor

New York, NY 10022

PREPARED BY:

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169 Commack Road – Suite H173 | Commack, NY 11725

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File No. 41.0163256.01



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1.0 INTRODUCTION

This Quality Assurance Project Plan (QAPP) and Field Sampling Plan (FSP) presents the organization, objectives, planned activities, and specific quality assurance/quality control (QA/QC) procedures associated with the Site Management Plan (SMP) at 1346 Blondell Avenue, Bronx, New York (Site). **Figure 1** presents a Site location map.

This QAPP/FSP describes specific protocols for field sampling, sample handling and storage, chain-of-custody, laboratory analysis, and data handling and management. Preparation of the Plan was based on EPA Quality Assurance Project Plan guidance documents, including:

- *EPA Requirements for Quality Assurance Project Plans* (EPA QA/R-5, March 2001); and
- *Guidance for Quality Assurance Project Plans* (EPA QA/G-5, December 2002).

Potential parameters to be analyzed, including their respective quantitation limits (QLs), and data quality levels (DQLs), are provided in **Tables 1A through 1C**.

2.0 PROJECT ORGANIZATION AND RESPONSIBILITY

A qualified person will coordinate and manage the Site sampling and analysis program, data reduction, QA/QC, data validation, analysis, and reporting. A Victoria Whalen, P.G. is a qualified environmental professional (QEP), as defined by the New York State Department of Environmental Conservation (NYSDEC) and will direct the sampling activities and coordinate with the laboratory. The intent of this QAPP/FSP is to be performed in accordance with the technical guidance applicable to Technical Guidance for Site Investigation and Remediation (DER-10), and Sampling, Analysis and Assessment of Per- and Polyfluoroalkyl Substances (PFAS) under NYSDEC's Part 375 Remedial Programs dated April 2023.

A qualified person will ensure that the QA/QC plan is implemented. GZA's Senior Technical Specialist, Dr. Chunhua Liu will provide oversight and technical support for the sampling and analytical procedures followed acting as the project QA Officer. This individual has the broad authority to approve or disapprove project plans, specific analyses, and final reports. The QEP is independent from the data generation activities. In general, the QA officer will be responsible for reviewing and advising on all QA/QC aspects of this program.

Laboratories used will be New York State Department of Health Environmental (NYSDOH) Laboratory Approval Program (ELAP) certified laboratories. The laboratories will communicate directly with the sampler regarding the analytical results and reporting and will be responsible for providing all labels, sample containers, field blank water, trip blanks, shipping coolers, and laboratory documentation. Qualifications of the QA officer are provided in **Attachment A**.

3.0 QA OBJECTIVES FOR DATA MANAGEMENT

The analytical data will be provided by the laboratory using the NYSDEC Category B deliverable format. Analytical data collected for disposal characteristics that may be requested by off-site wastewater disposal facilities will be provided in the format that the facility requests.



All analytical measurements will be made so that the results are representative of the media sampled and the conditions measured. Data will be reported in micrograms per liter ($\mu\text{g/L}$) or milligrams per liter (mg/L) for aqueous samples. **Table 2** presents the proposed samples, sampling and analytical parameters, analytical methods, sample preservation requirements and containers.

Quantitation Limits (QLs) are laboratory-specific and reflect those values achievable by the laboratory performing the analyses. Data Quality Levels (DQLs) are those reporting limits required to meet the objectives of the program (i.e., program action levels, cleanup standards, etc.). Data Quality Objectives (DQOs) define the quality of data and documentation required to support decisions made in the various phases of the data collection activities. The DQOs are dependent on the end uses of the data to be collected and are also expressed in terms of objectives for precision, accuracy, representativeness, completeness, and comparability.

The analytical methods to be used at this Site provide the highest level of data quality and can be used for purposes of risk assessment, evaluation of remedial alternatives and verification that cleanup standards have been met. However, in order to ensure that the analytical methodologies are capable of achieving the DQOs, measurement performance criteria have been set for the analytical measurements in terms of accuracy, precision, and completeness.

The overall QA objective is to develop and implement procedures for field sampling, chain-of-custody, laboratory analysis, and reporting which will provide results that are scientifically valid, and the levels of which are sufficient to meet DQOs. Specific procedures for sampling, chain of custody, laboratory instrument calibration, laboratory analysis, reporting of data, internal quality control, and corrective action are described in other sections of this QAPP/FSP.

Tables 3, 4, and 5 present the precision and accuracy requirements for each parameter to be analyzed. For quantitation limits for parameters associated with groundwater samples, the laboratory will be required to attempt to meet or surpass the parameter-specific limits for groundwater from the Division of Water Technical and Operational Guidance Series (TOGS 1.1.1) Ambient Water Quality Standards and Guidance Values. In certain instances, if the TOGS criteria are not achievable due to analytical limitations, the laboratory will report the lowest possible quantitation limit.

The QA objectives are defined as follows:

Accuracy is the closeness of agreement between an observed value and an accepted reference value. The difference between the observed value and the reference value includes components of both systematic error (bias) and random error.

Accuracy in the field is assessed through the adherence to all field instrument calibration procedures, sample handling, preservation, and holding time requirements, and through the collection of equipment blanks prior to the collection of samples for each type of equipment being used (e.g., groundwater sampling pumps).

The laboratory will assess the overall accuracy of their instruments and analytical methods (independent of sample or matrix effects) through the measurement of “standards,” materials of accepted reference value. Accuracy will vary from analysis to analysis because of individual sample and matrix effects. In an individual analysis, accuracy will be measured in terms of blank results, the percent recovery (%R) of surrogate compounds in organic analyses, or %R of spiked compounds



in matrix spikes (MSs), matrix spike duplicates (MSDs) and/or laboratory control samples (LCSs). This gives an indication of expected recovery for analytes tending to behave chemically like the spiked or surrogate compounds. **Tables 3, 4, and 5** summarize the laboratory accuracy requirements.

Precision is the agreement among a set of replicate measurements without consideration of the “true” or accurate value: i.e., variability between measurements of the same material for the same analyte. Precision is measured in a variety of ways including statistically, such as calculating variance or standard deviation.

Precision in the field is assessed through the collection and measurement of field duplicates (one extra sample in addition to the original field sample). Field duplicates will be collected at a frequency of one per twenty investigative samples per matrix per analytical parameter. Precision will be measured through the calculation of relative percent differences (RPDs). The resulting information will be used to assess sampling and analytical variability. Field duplicate RPDs must be ≤ 50 for soil samples and ≤ 30 for aqueous samples. These criteria apply only if the sample and/or duplicate results are $>5x$ the quantitation limit; if both results are $\leq 5x$ the quantitation limit, the criterion will be doubled. Due to the uncertainty of available representative soil gas volume, field duplicates will not be collected for this matrix.

Precision in the laboratory is assessed through the calculation of RPD for duplicate samples. For water analyses, laboratory precision will be assessed through the analysis of MS/MSD samples and field duplicates. For the inorganic analyses, laboratory precision will be assessed through the analysis of matrix duplicates and field duplicates. MS/MSD samples or matrix duplicates will be performed at a frequency of one per twenty investigative samples per matrix per parameter. **Tables 3, 4, and 5** summarize the laboratory precision requirements.

Completeness is a measure of the amount of valid data obtained from a measurement system compared to the amount that was expected to be obtained under normal conditions. “Normal conditions” are defined as the conditions expected if the sampling plan was implemented as planned.

Field completeness is a measure of the amount of (1) valid measurements obtained from all the measurements taken in the project and (2) valid samples collected. The field completeness objective is greater than 90 percent.

Laboratory completeness is a measure of the amount of valid measurements obtained from all valid samples submitted to the laboratory. The laboratory completeness objective is greater than 95 percent.

Representativeness is a qualitative parameter that expresses the degree to which data accurately and precisely represent either a characteristic of a population, parameter variations at a sampling point, a process condition, or an environmental condition within a defined spatial and/or temporal boundary. To ensure representativeness, the sampling locations have been selected to provide coverage over a wide area and to highlight potential trends in the data. In addition, field duplicate samples will provide an additional measure of representativeness at a given location.



Representativeness is dependent upon the proper design of the sampling program and will be satisfied by ensuring that the Work Plans and QAPP are followed, and that proper sampling, sample handling, and sample preservation techniques are used.

Representativeness in the laboratory is ensured by using proper analytical procedures, appropriate methods, and meeting sample holding times.

Comparability expresses the confidence with which one data set can be compared to another. Comparability is dependent upon the proper design of the sampling program and will be satisfied by ensuring that the Work Plans and QAPP are followed and that proper sampling techniques are used. Maximization of comparability with previous data sets is expected because the sampling design and field protocols are consistent with those previously used.

Comparability is dependent on the use of recognized EPA or equivalent analytical methods and the reporting of data in standardized units. Laboratory procedures are consistent with those used for previous sampling efforts.

4.0 PERMENANT WELL INSTALLATION AND DEVELOPMENT

4.1. Well Installation

To collect representative groundwater samples, soil borings drilled with the sonic drilling method will be converted into permanent two-inch diameter monitoring wells. Groundwater monitoring wells will be constructed of threaded two-inch diameter PVC well casing and 0.01-inch slotted well screen. The 10-foot screen will be set straddling the measured water table. Clean silica sand, Morie No. 1 or equivalent, will be placed in the annular space around the well to a minimum of two feet above the top of the well screen. Solid PVC riser, attached to the well screen, will extend to grade or above if the well is a stick-up. For a two-inch diameter well, the annular space for the filter pack should be 4 inches thick. A two-foot thick bentonite seal will then be placed above the sand pack and moistened with potable water for a minimum of 15 minutes before backfilling the remaining space with a cement-bentonite grout or just cement. If warranted by depth, filling will be completed using a tremie pipe placed below the surface of the grout. A stick-up or flush-mount protective casing with a locking well cap will then be installed, and a measuring point marked on each PVC well riser. Well construction diagrams will be prepared for each well.

4.2. Well Development

Following installation, the groundwater monitoring wells will be developed using a submersible pump(s) (or equivalent) until the water is reasonably free of turbidity and field readings (pH, conductivity, temperature, and dissolved oxygen) sufficiently stabilize. Fifty nephelometric turbidity units (NTUs) or less will be the turbidity goal but not an absolute value. The wells will be developed aggressively to remove fines from the formation and sand pack. The wells will be allowed to equilibrate for a minimum seven days prior to sampling. The volume of water removed, the well development time, and field instrument readings will be recorded in the logbook.



5.0 SAMPLING PLAN

Environmental sampling will include groundwater sampling. Additionally, purge water generated during groundwater monitoring events will be sampled and tested for characterization for disposal. Groundwater samples will be collected using bailers or peristaltic, bladder or submersible pumps.

5.1. Groundwater Sampling

Groundwater sampling of permanent monitoring wells is described according to the following distinct phases of this work: well installation/construction, well development, well purging, and well sampling.

5.1.1. *Well Purging*

The objective is to purge monitoring wells until turbidity stabilizes to a level as low as possible and this parameter will be given the greatest weight in determining when groundwater sampling may begin. With this objective in mind, a low-flow pump will be used to avoid entrainment of particulates within the well or from the formation. Groundwater from each well will be purged until parameters have stabilized. A turbidity level of fifty NTUs or less is the well purging goal, but not an absolute value before sampling. Other field parameters including temperature, conductivity, pH, and dissolved oxygen (DO) will also be monitored. As practical, all field measurements will be taken from the flow cell and will be recorded during and after purging, and before sampling. Field parameters should generally be within ± 10 percent for three consecutive readings, one minute apart, prior to sampling.

Upon opening each monitoring well and point, the concentration of VOCs in the headspace will be measured using a PID and water level measurements will be recorded using an electronic interface probe. The depth to product (if present), depth to water, and the total depth will be measured from the top of the marked PVC casings. Water level and free product measurements will first be made and the volume of water in the well determined. The volume of water in the well will be calculated so that the number of well volumes purged and an estimate of the time required to purge the well can be made. Before sampling, the wells will be purged utilizing a low-flow submersible stainless steel pump using dedicated Teflon[®] or Teflon[®]-lined polyethylene tubing connected to a flow cell. Very low purging rates are proposed, on the order of 100 ml/minute to 500 ml/minute, to minimize suspension of particulate matter in the well.

Purging will be done with the pump intake placed at the midpoint of the well screen or the midpoint of the water column (to be determined based on the depth and length of the screen interval) to ensure that all stagnant water in the well is removed, while not stirring up sediment that may have accumulated on the bottom of the well. Equipment will be lowered into the well very carefully to prevent suspension of bottom sediment and subsequent entrainment onto sampling equipment. Surging will be avoided. Tubing will be replaced between each well. Pumps must be carefully cleaned between wells according to the procedures specified in **Section 4.15**, below. It is anticipated that no more than three well volumes



will be purged in order for turbidity to reach a minimum and the other parameters to stabilize. Ideally, pumping rates will be at a rate so that no drawdown of the groundwater level occurs (i.e., pumping rate is less than recharge rate). During purging, the sampler will actively monitor and track the volume of water purged and the field parameter readings. Data will be recorded in the field logbook. For example, the sampler will record the running total volume purged from each well and note the readings for the corresponding field parameters.

5.1.2. *Well Sampling*

Once groundwater conditions have stabilized and groundwater levels have recovered, samples will be collected from the flow cell outlet (connected to the low-flow submersible pump). All non-disposable/non-dedicated (re-usable) sampling equipment will be cleaned according to the procedures specified in **Section 4.15**.

Sampling will be performed with the pump intake at the same location used for purging. Pumping rates for withdrawing the samples will be similar to those followed for well purging.

The samples will be collected in sample bottles (pre-preserved, if appropriate), placed in iced coolers and removed from light immediately after collection. In addition, all sample bottles must be filled to the top so that no aeration of the samples occurs during transport. All bottles will be filled to avoid cascading and aeration of the samples, the goal being to minimize any precipitation of colloidal matter. Samples will be transported to a NYSDOH ELAP certified laboratory under proper chain of custody procedures for analysis. Samples for dissolved metals will be collected in unpreserved containers and will be filtered and preserved at the laboratory within 24 hours of sampling.

5.2. **Monitoring Well Abandonment**

There may be occasions when monitoring wells will require abandonment. For temporary monitoring wells, the approach will be to pull the PVC well materials from the borehole and backfill the remaining open portion of the borehole with cement/bentonite grout to approximately 0.5 feet below the ground surface. The ground surface will be restored to a similar condition as the surrounding grade (e.g., topsoil, asphalt, or concrete). For permanent overburden and bedrock monitoring wells, depending on the site-specific subsurface geologic conditions and nature of contamination, the abandonment approach will be in accordance with NYSDEC Policy CP-43 – Groundwater Monitoring Well Decommissioning Policy.

5.3. **Waste Characterization Sampling**

Waste classification sampling may be conducted to characterize soil, liquids and/or groundwater for the purpose of proper off-site waste disposal. Specific methods for sampling liquid wastes are briefly discussed below.

5.3.1. *Liquid Waste*



Liquid sampling methods include utilizing dedicated dippers, glass tube samplers, pump and tubing, kemmerer bottles, and Bacon Bomb samplers. Dippers are used to collect samples from the surface of the liquid and are appropriate for wastes that are homogeneous. Glass tube samplers consist of glass tubes of varying length and diameter used to collect a full-depth liquid sample from a drum or similar container. Pump and tubing (e.g., bladder pump or peristaltic pump) are used to collect liquid samples from a depth (up to approximately 20 feet below grade), and are typically relied upon for sampling subsurface structures, such as underground storage tanks. To minimize the loss of volatile organic components in the liquid, the lowest achievable flow rate is utilized for collecting the sample by this method. Kemmerer bottles and Bacon Bomb samplers are discrete-depth samplers. These samplers are lowered into the liquid and opened to collect a sample at a desired depth.

5.3.2. *Grab versus Composite Sampling*

Waste characterization of a liquid can involve grab or composite sampling depending upon the homogeneity and the volume of the waste. Grab sampling consists of collecting a discrete sample or samples of a material and submitting each sample for separate analysis. Grab sampling is appropriate for characterizing small quantities of waste as well as waste streams of varying content (e.g., drums of different contents). Composite sampling consists of taking discrete grab samples of a material and combining them into a smaller number of samples for analysis. Composite sampling generally is appropriate for large volumes of a homogenous waste material. The specific number of composite and grab samples largely will depend upon the size, nature of the waste, as well as the analysis required for characterization of the waste.

5.4. **QC Sample Collection**

QC samples will include equipment blanks, trip blanks, field duplicates and MS/MSDs.

Equipment blanks will consist of distilled water and will be used to check for potential contamination of the equipment that may cause sample contamination. Equipment blanks will be collected by routing the distilled water through the sampling equipment prior to sample collection. Equipment blanks will be submitted to the laboratory at a frequency of one per day per matrix per type of equipment being used per parameter. Equipment blanks will not be collected with samples for analysis for parameters associated with wastewater samples and samples collected for disposal purposes.

Trip blanks will consist of distilled water (supplied by the laboratory) and will be used to assess the potential for volatile organic compound contamination of groundwater samples due to contaminant migration during sample shipment and storage. Trip blanks will be transported to the site unopened, stored with the investigative samples, and kept closed until analyzed by the laboratory. Trip blanks will be submitted to the laboratory at a frequency of one per cooler that contains groundwater samples for analysis for VOCs.



Field duplicates are an additional aliquot of the same sample submitted for the same parameters as the original sample. Field duplicates will be used to assess the sampling and analytical reproducibility. Field duplicates will be collected by alternately filling sample bottles from the source being sampled. Field duplicates will be submitted at a frequency of one per 20 samples for all matrices and all parameters with the exception of TCLP parameters, parameters associated with wastewater samples, and samples collected for waste characterization purposes.

MSs and MSDs are two additional aliquots of the same sample submitted for the same parameters as the original sample. However, the additional aliquots are spiked with the compounds of concern. Matrix spikes provide information about the effect of the sample matrix on the measurement methodology. MS/MSDs will be submitted at a frequency of one per 20 investigative samples per matrix for organic parameters for soil, sediment, and groundwater. MSs will be submitted at a frequency of one per 20 investigative samples per matrix for inorganic parameters.

5.5. Sample Preservation and Containerization

The analytical laboratory will supply the sample containers for the chemical samples. These containers will be cleaned by the manufacturer to meet or exceed all analyte specifications established in the latest U.S. EPA's *Specifications and Guidance for Contaminant-Free Sample Containers*. Certificates of analysis are provided with each bottle lot and maintained on file to document conformance to EPA specifications. The containers will be pre-preserved, where appropriate (see **Table 2**).

Table 6 presents a summary of QC sample preservation and container requirements.

5.6. Equipment Decontamination

Re-usable Teflon[®], stainless steel, and aluminum sampling equipment shall be cleaned between each use in the following manner:

- Tap water rinse
- Wash and scrub with Alconox and water mixture
- Tap water rinse
- Distilled/deionized water rinse
- Air dry

Cleaned equipment shall be wrapped in aluminum foil if not used immediately after air-drying.

Groundwater sampling pumps will be cleaned by washing and scrubbing with an Alconox/water mixture, rinsing with tap water and irrigating with distilled/deionized water.



6.0 DOCUMENTATION AND CHAIN-OF-CUSTODY

6.1. Sample Collection Documentation

6.1.1. *Field Notes*

Field team members will keep a field logbook to document all field activities. Field logbooks will provide the means of recording the chronology of data collection activities performed during the remediation. As such, entries will be described in as much detail as possible so that a particular situation could be reconstructed without reliance on memory.

The logbook will be a bound notebook with water-resistant pages. Logbook entries will be dated, legible, and contain accurate and inclusive documentation of the activity. The title page of each logbook should contain the following:

- Person to whom the logbook is assigned
- The logbook number
- Project name and number
- Site name and location
- Project start date
- End date

Entries into the logbook will contain a variety of information. At the beginning of each entry, the date, start time, weather, and names of sampling team members present will be entered. Each page of the logbook will be signed and dated by the person making the entry. All entries will be made in permanent ink, signed, and dated and no erasures or obliterations will be made. If an incorrect entry is made, the information will be crossed out with a single strike mark that is signed and dated by the sampler. The correction shall be written adjacent to the error.

Field activities will be fully documented. Information included in the logbook should include, but may not be limited to, the following:

- Chronology of activities, including entry and exit times
- Names of all people involved in sampling activities
- Level of personal protection used
- Any changes made to planned protocol
- Names of visitors to the site during sampling and reason for their visit
- Sample location and identification
- Changes in weather conditions
- Dates (month/day/year) and times (military) of sample collection
- Measurement equipment identification (model/manufacturer) and calibration information
- Sample collection methods and equipment



- Sample depths
- Whether grab or composite sample collected
- How sample composited, if applicable
- Sample description (color, odor, texture, etc.)
- Sample identification code
- Tests or analyses to be performed
- Sample preservation and storage conditions
- Equipment decontamination procedures
- QC sample collection
- Unusual observations
- Record of photographs
- Sketches or diagrams
- Signature of person recording the information

Field logbooks will be reviewed on a daily basis by the Field Team Leader. Logbooks will be supported by standardized forms.

6.1.2. Chain-of-Custody Records

On a regular basis (daily or on such a basis that all holding times will be met), samples will be transferred to the custody of the respective laboratories, via third-party commercial carriers or via laboratory courier service.

Chain-of-custody records are initiated by the samplers in the field. The field portion of the custody documentation should include: (1) the project name; (2) signatures of samplers; (3) the sample number, date and time of collection, and whether the sample is grab or composite; (4) signatures of individuals involved in sampling; and (5) if applicable, air bill or other shipping number. Sample receipt and log-in procedures at the laboratory are described in **Section 5.2.2** of this Plan.

6.1.3. Sample Labeling

Immediately upon collection, each sample will be labeled with a pre-printed adhesive label, which includes the date and time of collection, sampler's initials, tests to be performed, preservative (if applicable), and a unique identifier.

A. The following identification scheme will be used:

Groundwater wells will be assigned sequential numbers. Groundwater samples will be identified by the well that the sample was collected from.

Examples:

GMW-01 = groundwater sample collected from permanent well point #1



Duplicate samples will be labeled as blind duplicates by giving them sample numbers indistinguishable from a normal sample.

Blanks should be spelled out and identify the associated matrix, e.g., Equipment Blank, Soil

MS/MSDs will be noted in the Comments column of the COC.

- B. The analysis required will be indicated for each sample.
Example: SVOC
- C. Date taken will be the date the sample was collected, using the format: MM-DD-YY.
Example: 04-22-25
- D. Time will be the time the sample was collected, using military time.
Example: 14:30
- E. The sampler’s name will be printed in the “Sampled By” section.
- F. Other information relevant to the sample.
Example: Equipment Blank

An example sample label is presented below:

Job No. _____
 Client: _____
 Sample Number _____
 Date _____ Sample Time _____
 Sample Matrix _____
 Grab or Composite (explain) _____
 Preservatives _____
 Analyses _____
 Sampler Signature _____

This sample label contains the authoritative information for the sample. Inconsistencies with other documents will be settled in favor of the vial or container label unless otherwise corrected in writing from the field personnel collecting samples or the QEP.

6.2. Sample Custody

Custody is one of several factors that are necessary for the admissibility of environmental data as evidence in a court of law. Custody procedures help to satisfy the two major requirements for



admissibility: relevance and authenticity. Sample custody is addressed in three parts: field sample collection, laboratory analysis, and final evidence files.

A sample or evidence file is considered to be under a person's custody if

- the item is in the actual possession of a person
- the item is in the view of the person after being in actual possession of the person
- the item was in the actual physical possession of the person but is locked up to prevent tampering
- the item is in a designated and identified secure area

6.2.1. *Field Custody Procedures*

Samples will be collected following the sampling procedures documented in **Section 4.0** of this Plan. Documentation of sample collection is described in **Section 5.1** of this Plan. Sample chain-of-custody and packaging procedures are summarized below. These procedures are intended to ensure that the samples will arrive at the laboratory with the chain-of-custody intact.

- The field sampler is personally responsible for the care and custody of the samples until they are transferred or dispatched properly. Field procedures have been designed such that as few people as possible will handle the samples.
- All bottles will be identified by the use of sample labels with sample numbers, sampling locations, date/time of collection, and type of analysis.
- Sample labels will be completed for each sample using waterproof ink unless prohibited by weather conditions. For example, a logbook notation would explain that a pencil was used to fill out the sample label because the pen would not function in wet weather.
- Samples will be accompanied by a properly completed chain-of-custody form. The sample numbers and locations will be listed on the chain-of-custody form. When transferring the possession of samples, the individuals relinquishing and receiving will sign, date, and note the time on the record. This record documents the transfer of custody of samples from the sampler to another person, to a mobile laboratory, to the permanent laboratory, or to/from a secure storage location.
- All shipments will be accompanied by the chain-of-custody record identifying the contents. The original record will accompany the shipment, and copies will be retained by the sampler and placed in the project files.
- Samples will be properly packaged for shipment and dispatched to the appropriate laboratory for analysis, with a separate signed custody record enclosed in and secured to the inside top of each sample box or cooler. If third party commercial carriers are used for transfer to the laboratory, shipping containers will be secured with strapping tape and custody seals prior to shipment. The



custody seals will be attached to the front right and back left of the cooler and covered with clear plastic tape after being signed by field personnel. The cooler will be strapped shut with strapping tape in at least two locations.

- If the samples are sent by third party commercial carrier, the air bill will be used. Air bills will be retained as part of the permanent documentation. Commercial carriers are not required to sign off on the custody forms since the custody forms will be sealed inside the sample cooler and the custody seals will remain intact.
- Samples remain in the custody of the sampler until transfer of custody is completed. This consists of delivery of samples to the laboratory courier or sample custodian, and signature of the laboratory courier or sample custodian on chain-of-custody document as receiving the samples and signature of sampler as relinquishing samples.

6.2.2. *Laboratory Custody Procedures*

Samples will be received and logged in by a designated sample custodian or his/her designee. Upon sample receipt, the sample custodian will

- Examine the shipping containers to verify that the custody tape is intact,
- Examine all sample containers for damage,
- Determine if the temperature required for the requested testing program has been maintained during shipment and document the temperature on the chain-of-custody records,
- Compare samples received against those listed on the chain-of-custody,
- Verify that sample holding times have not been exceeded,
- Examine all shipping records for accuracy and completeness,
- Determine sample pH (if applicable) and record on chain-of-custody forms,
- Sign and date the chain-of-custody immediately (if shipment is accepted) and attach the air bill,
- Note any problems associated with the coolers and/or samples on the cooler receipt form and notify the Laboratory Project Manager, who will be responsible for contacting the QEP,
- Attach laboratory sample container labels with unique laboratory identification and test, and
- Place the samples in the proper laboratory storage.

Following receipt, samples will be logged in according to the following procedure:

- The samples will be entered into the laboratory tracking system. At a minimum, the following information will be entered: project name or identification, unique sample numbers (both client and internal laboratory), type of sample, required tests, date and time of laboratory receipt of samples, and field ID provided by field personnel.
- The Laboratory Project Manager will be notified of sample arrival.
- The completed chain-of-custody, air bills, and any additional documentation will be placed in the final evidence file.



7.0 CALIBRATION PROCEDURES

7.1. Field Instruments

Field instruments will be calibrated according to the manufacturer's specifications. Calibration procedures performed will be documented in the field logbook and will include the date/time of calibration, name of person performing the calibration, reference standard used, temperature at which the readings were taken, and the readings.

7.2. Laboratory Instruments

Calibration procedures for a specific laboratory instrument will consist of initial calibrations, initial calibration verifications, and/or continuing calibration verification. Detailed descriptions of the calibration procedures for a specific laboratory instrument are included in the laboratory's standard operating procedures (SOPs), which describe the calibration procedures, their frequency, acceptance criteria, and the conditions that will require recalibration. These procedures are as required in the respective analytical methodologies (summarized in **Table 2** of this Plan). The initial calibration associated with all analyses must contain a low-level calibration standard which is less than or equal to the quantitation limit.

8.0 SAMPLE PREPARATION AND ANALYTICAL PROCEDURES

No field analyses are anticipated for this program. If site conditions were to warrant field analysis, the responsible contractor will prepare an addendum establishing the field analytical procedures. Analyses of all samples will be performed by NYSDOH ELAP certified laboratories. **Table 2** summarizes the analytical methods to be used during the groundwater monitoring events.

9.0 DATA REDUCTION, VALIDATION, AND REPORTING

Appropriate QC measures will be used to ensure the generation of reliable data from sampling and analysis activities. Proper collection and organization of accurate information followed by clear and concise reporting of the data is a primary goal in this project. Complete data packages suitable for data validation will be provided by the analytical laboratory.

For all analyses, the laboratory will report results that are below the laboratory's reporting limit; these results will be qualified as estimated (J) by the laboratory. The laboratory may be required to report tentatively identified compounds (TICs) for the VOC and SVOC analyses; this will be requested by the sampler on an as-needed basis. A Data Usability Summary Report (DUSR) will be prepared and will be included in the Periodic Review Report (PRR). The DUSR for this project is Ms. Christin Camardella. Qualifications of the DUSR preparer can be found in **Attachment A**.

9.1. Data Evaluation/Validation



9.1.1. Field Data Evaluation

Measurements and sample collection information will be transcribed directly into the field logbook or onto standardized forms. If errors are made, results will be legibly crossed out, initialed and dated by the person recording the data, and corrected in a space adjacent to the original (erroneous) entry. Daily reviews of the field records by the Field Team Leader will ensure that:

- Logbooks and standardized forms have been filled out completely and that the information recorded accurately reflects the activities that were performed.
- Records are legible and in accordance with good record keeping procedures, i.e., entries are signed and dated, data are not obliterated, changes are initialed, dated, and explained.
- Sample collection, handling, preservation, and storage procedures were conducted in accordance with the protocols described in the Plan, and that any deviations were documented and approved by the appropriate personnel.

9.1.2. Data Usability

A Data Usability Summary Report (DUSR) will be prepared in accordance with the DER Technical Guidance for Site Investigation and Remediation (DER-10).

The data usability evaluation will include reviewing the quality assurance/quality control (QA/QC) information including: (1) chain-of-custody; (2) the summary QA/QC information provided by the laboratory; and (3) the project narrative.

For each data package the following questions will be evaluated:

- Is the data package complete as defined under the requirements for the NYSDEC ASP Category B, USEPA CLP deliverables or other standards/guidance?
- Have all holding times and preservation requirements been met?
- Do the quality control (QC) data fall within the laboratory and project established limits and specifications?

9.2. Identification and Treatment of Outliers

Any data point which deviates markedly from others in its set of measurements will be investigated; however, the suspected outlier will be recorded and retained in the data set. One or both of the following tests will be used to identify outliers.

Dixon's test for extreme observations is an easily computed procedure for determining whether a single very large or very small value is consistent with the remaining data. The one-tailed t-test for difference may also be used in this case. It should be noted that these tests are designed for testing a single value.



If more than one outlier is suspected in the same data set, other statistical sources may be consulted and the most appropriate test of hypothesis will be used and documented, if warranted.

Since an outlier may result from unique circumstances at the time of sample analysis or data collection, those persons involved in the analysis and data reduction will be consulted. This may provide an experimental reason for the outlier. Further statistical analysis may be performed with and without the outlier to determine its effect on the conclusions. In many cases, two data sets may be reported, one including, and one excluding the outlier.

In summary, every effort will be made to include the outlying values in the reported data. If the value is rejected, it will be identified as an outlier, reported with its data set and its omission noted.

10.0 INTERNAL QUALITY CONTROL

The subcontracting laboratories' Quality Assurance Project Plans will identify the supplemental internal analytical quality control procedures to be used. At a minimum, this will include:

- Matrix spike and/or matrix spike duplicate samples
- Matrix duplicate analyses
- Laboratory control samples
- Instrument calibrations
- Instrument tunes for SW-846 8260B and 8270C analyses
- Method and/or instrument blanks
- Surrogate spikes for organic analyses
- Internal standard spikes for SW-846 8260B and 8270C analyses
- Quantitation limit determination and confirmation by analysis of low-level calibration standard

As outline on **Table 5** and summarized in **Section 4.13**, field quality control samples will include:

- Equipment blanks
- Field duplicate samples
- Trip blanks
- MS/MSDs

11.0 CORRECTIVE ACTION

The entire sampling program will be under the direction of the QEP. The emphasis in this program is on preventing problems by identifying potential errors, discrepancies, and gaps in the data-collection-laboratory-analysis-interpretation process. Any problems identified will be promptly resolved. Likewise, follow-up corrective action is always an option in the event that preventative corrective actions are not totally effective.



The acceptance limits for the sampling and analyses to be conducted in this program will be those stated in the method or defined by other means in the Plan. Corrective actions are likely to be immediate in nature and most often will be implemented by the contracted laboratory analyst or the Program Manager. The corrective action will usually involve recalculation, reanalysis, or resampling.

11.1. Immediate Corrective Action

Corrective action in the field may be needed when the sample network is changed (i.e., more/less samples, sampling locations other than those specified in the Plan), or when sampling procedures and/or field analytical procedures require modification, etc. due to unexpected conditions. The field team may identify the need for corrective action. The Field Team Leader will approve the corrective action and notify the Program Manager. The Program Manager will approve the corrective measure. The Field Team Leader will ensure that the corrective measure is implemented by the field team.

Corrective actions will be implemented and documented in the field logbook. Documentation will include:

- A description of the circumstances that initiated the corrective action,
- The action taken in response,
- The final resolution, and
- Any necessary approvals

No staff member will initiate corrective action without prior communication of findings through the proper channels.

Corrective action in the laboratory may occur prior to, during, and after initial analyses. A number of conditions such as broken sample containers, omissions or discrepancies with chain-of-custody documentation, low/high pH readings, and potentially high concentration samples may be identified during sample log-in or just prior to analysis. Following consultation with laboratory analysts and Laboratory Section Leaders, it may be necessary for the Laboratory QA Manager to approve the implementation of corrective action. The laboratory SOPs specify some conditions during or after analysis that may automatically trigger corrective action or optional procedures. These conditions may include dilution of samples, additional sample extract cleanup, automatic reinjection/reanalysis when certain QC criteria are not met, loss of sample through breakage or spillage, etc.

The analyst may identify the need for corrective action. The Laboratory Section Leader, in consultation with the staff, will approve the required corrective action to be implemented by the laboratory staff. The Laboratory QA Manager will ensure implementation and documentation of the corrective action. If the nonconformance causes project objectives not to be achieved, the QEP will be notified. The QEP will notify the Program Manager, who in turn will contact all levels of project management for concurrence with the proposed corrective action.

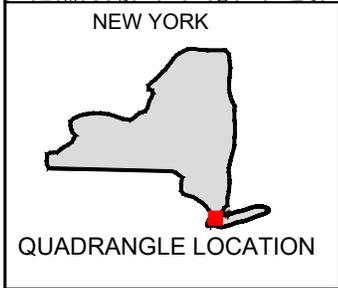
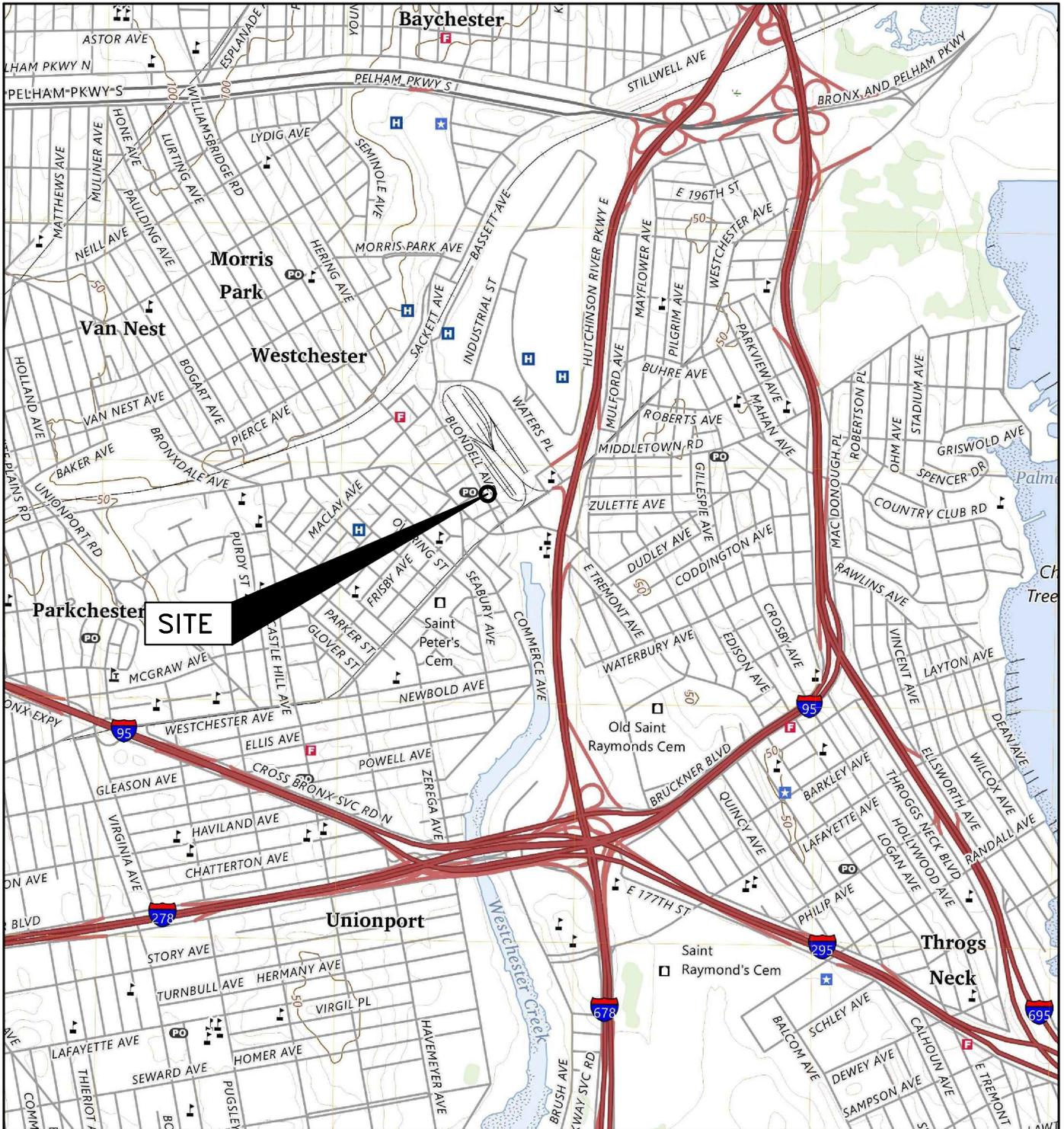


These corrective actions are performed prior to release of the data from the laboratory. The corrective action will be documented in both the laboratory's corrective action files, and the narrative data report sent from the laboratory to the Program Manager. If the corrective action does not rectify the situation, the laboratory will contact the Program Manager, who will determine the action to be taken and inform the appropriate personnel.

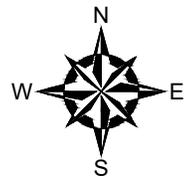
If potential problems are not solved as an immediate corrective action, the contractor will apply formalized long-term corrective action, if necessary.



FIGURES



SOURCE:
 USGS TOPOGRAPHIC MAPS: FLUSHING, NY (2023). CONTOUR INTERVAL 10FT., NAVD-1988, ORIGINAL SCALE 1:24,000 (1IN.=2,000FT.).



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1346 BLONDELL AVE, BRONX, NY

PREPARED BY:
 **GZA** GeoEnvironmental of NY
 Engineers and Scientists
 www.gza.com

PREPARED FOR:
 BLONDELL EQUITIES LLC

SITE LOCATION MAP

PROJ MGR: RM	REVIEWED BY: RM	CHECKED BY: VW
DESIGNED BY: SG	DRAWN BY: SG	SCALE: 1"=2000'
DATE: FEBRUARY 2025	PROJECT NO. 41.0163256.00	REVISION NO. -

FIGURE 1
 SHEET NO. 1 OF 12



TABLES

Table 1 A
Soil Criteria Table

1346 Blondell Avenue, Bronx, NY
BCP Site No. C203089
QAPP/FSP

Contaminant	Protection of Public Health					Protection of Ecological Resources ⁿ	Protection of Groundwater
	Unrestricted Use	Residential	Restricted-Residential	Commercial	Industrial		
All soil cleanup objectives (SCOs) are in parts per million (ppm); approximately equivalent to mg/kg.							
Metals							
Arsenic	13 ^m	16 ^f	17 ^f	18 ^f	19 ^f	13 ^f	16 ^f
Barium	350 ^m	350 ^f	400	400	10,000 ^d	433	820
Beryllium	7.2	14	72	590	2,700	10	47
Cadmium	2.5 ^m	2.5 ^f	4.3	9.3	60	4	7.5
Chromium, hexavalent ^h	1 ⁱ	22	110	400	800	1 ^e	19
Chromium, trivalent ^h	30 ^m	36	180	1,500	6,800	41	NS
Copper	50	270	270	270	10,000 ^d	50	1,720
Total Cyanide ^h	27	27	27	27	10,000 ^d	NS	40
Lead	63 ^m	400	400	1,000	3,900	63 ^f	450
Manganese	1600 ^m	2,000 ^f	2,000 ^f	10,000 ^d	10,000 ^d	1600 ^f	2,000 ^f
Total Mercury	0.18 ^m	0.81 ^j	0.81 ^j	2.8 ^j	5.7 ^j	0.18 ^f	0.73
Nickel	30	140	310	310	10,000 ^d	30	130
Selenium	3.9 ^m	36	180	1,500	6,800	3.9 ^f	4 ^f
Silver	2	36	180	1,500	6,800	2	8.3
Zinc	109 ^m	2200	10,000 ^d	10,000 ^d	10,000 ^d	109 ^f	2,480
PCBs/Pesticides							
2,4,5-TP Acid (Silvex)	3.8	58	100 ^a	500 ^b	1,000 ^c	NS	3.8
4,4'-DDE	0.0033 ^l	1.8	8.9	62	120	0.0033 ^e	17
4,4'-DDT	0.0033 ^l	1.7	7.9	47	94	0.0033 ^e	136
4,4'-DDD	0.0033 ^l	2.6	13	92	180	0.0033 ^e	14
Aldrin	0.005 ^m	0.019	0.097	0.68	1.4	0.14	0.19
alpha-BHC	0.02	0.097	0.48	3.4	6.8	0.04 ^g	0.02
beta-BHC	0.036	0.072	0.36	3	14	0.6	0.09
Chlordane (alpha)	0.094	0.91	4.2	24	47	1.3	2.9
delta-BHC	0.04	100 ^a	100 ^a	500 ^b	1,000 ^c	0.04 ^e	0.25
Dibenzofuran	7	14	59	350	1,000 ^c	NS	210
Dieldrin	0.005 ^m	0.039	0.2	1.4	2.8	0.006	0.1
Endosulfan I	2.4	4.8 ⁱ	24 ⁱ	200 ⁱ	920 ⁱ	NS	102
Endosulfan II	2.4	4.8 ⁱ	24 ⁱ	200 ⁱ	920 ⁱ	NS	102
Endosulfan sulfate	2.4	4.8 ⁱ	24 ⁱ	200 ⁱ	920 ⁱ	NS	1,000 ^c
Endrin	0.014	2.2	11	89	410	0.014	0.06
Heptachlor	0.042	0.42	2.1	15	29	0.14	0.38
Lindane	0.1	0.28	1.3	9.2	23	6	0.1
Polychlorinated biphenyls	0.1	1	1	1	25	1	3.2
Semivolatile							
Acenaphthene	20	100 ^a	100 ^a	500 ^b	1,000 ^c	20	98
Acenaphthylene	100 ^k	100 ^a	100 ^a	501 ^b	1,000 ^c	NS	107
Anthracene	100 ^k	100 ^a	100 ^a	502 ^b	1,000 ^c	NS	1,000 ^c
Benz(a)anthracene	1 ^m	1 ^f	1 ^f	5.6	11	NS	1 ^f
Benzo(a)pyrene	1 ^m	1 ^f	1 ^f	1 ^f	1.1	2.6	22
Benzo(b)fluoranthene	1 ^m	1 ^f	1 ^f	5.6	11	NS	1.7
Benzo(g,h,i)perylene	100	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	1,000 ^c
Benzo(k)fluoranthene	0.8 ^m	1	3.9	56	110	NS	1.7
Chrysene	1 ^m	1 ^f	3.9	56	110	NS	1 ^f
Dibenz(a,h)anthracene	0.33 ⁱ	0.33 ^e	0.33 ^e	0.56	1.1	NS	1,000 ^c
Fluoranthene	100 ^k	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	1,000 ^c
Fluorene	30	100 ^a	100 ^a	500 ^b	1,000 ^c	30	386
Indeno(1,2,3-cd)pyrene	0.5 ^m	0.5 ^f	0.5 ^f	5.6	11	NS	8.2
m-Cresol	0.33 ⁱ	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	0.33 ^e
Naphthalene	12	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	12
o-Cresol	0.33 ⁱ	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	0.33 ^e
p-Cresol	0.33 ⁱ	34	100 ^a	500 ^b	1,000 ^c	NS	0.33 ^e
Pentachlorophenol	0.8 ⁱ	2.4	6.7	6.7	55	0.8 ^e	0.8 ^e
Phenanthrene	100	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	1,000 ^c
Phenol	0.33 ⁱ	100 ^a	100 ^a	500 ^b	1,000 ^c	30	0.33 ^e
Pyrene	100	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	1,000 ^c

**Table 1 A
Soil Criteria Table**

**1346 Blondell Avenue, Bronx, NY
BCP Site No. C203089
QAPP/FSP**

Contaminant	Protection of Public Health					Protection of Ecological Resources ⁿ	Protection of Groundwater
	Unrestricted Use	Residential	Restricted-Residential	Commercial	Industrial		
All soil cleanup objectives (SCOs) are in parts per million (ppm); approximately equivalent to mg/kg.							
Volatiles							
1,1,1-Trichloroethane	0.68	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	0.68
1,1-Dichloroethane	0.27	19	26	240	480	NS	0.27
1,1-Dichloroethene	0.33	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	0.33
1,2-Dichlorobenzene	1.1	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	1.1
1,2-Dichloroethane	0.02 ^m	2.3	3.1	30	60	10	0.02 ^f
cis-1,2-Dichloroethene	0.25	59	100 ^a	500 ^b	1,000 ^c	NS	0.25
trans-1,2-Dichloroethene	0.19	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	0.19
1,3-Dichlorobenzene	2.4	17	49	280	560	NS	2.4
1,4-Dichlorobenzene	1.8	9.8	13	130	250	20	1.8
1,4-Dioxane	0.1 ^l	9.8	13	130	250	0.1 ^e	0.1 ^e
Acetone	0.05	100 ^a	100 ^b	500 ^b	1,000 ^c	2.2	0.05
Benzene	0.06	2.9	4.8	44	89	70	0.06
Butylbenzene	12	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	12
Carbon tetrachloride	0.76	1.4	2.4	22	44	NS	0.76
Chlorobenzene	1.1	100 ^a	100 ^a	500 ^b	1,000 ^c	40	1.1
Chloroform	0.37	10	49	350	700	12	0.37
Ethylbenzene	1	30	41	390	780	NS	1
Hexachlorobenzene	0.33 ^l	0.33 ^e	1.2	6	12	NS	3.2
Methyl ethyl ketone	0.12	100 ^a	100 ^a	500 ^b	1,000 ^c	100 ^a	0.12
Methyl tert-butyl ether	0.93	62	100 ^a	500 ^b	1,000 ^c	NS	0.93
Methylene chloride	0.05	51	100 ^a	500 ^b	1,000 ^c	12	0.05
n-Propylbenzene	3.9	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	3.9
sec-Butylbenzene	11	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	11
tert-Butylbenzene	5.9	100 ^a	100 ^a	500 ^b	1,000 ^c	NS	5.9
Tetrachloroethene	1.3	5.5	19	150	300	2	1.3
Toluene	0.7	100 ^a	100 ^a	500 ^b	1,000 ^c	36	0.7
Trichloroethene	0.47	10	21	200	400	2	0.47
1,2,4-Trimethylbenzene	3.6	47	52	190	380	NS	3.6
1,3,5-Trimethylbenzene	8.4	47	52	190	380	NS	8.4
Vinyl chloride	0.02	0.21	0.9	13	27	NS	0.02
Xylene (mixed)	0.26	100 ^a	100 ^a	500 ^b	1,000 ^c	0.26	1.6
Per- and Polyfluoroalkyl Substances (PFAs)^o							
PFOA	0.00066	0.0066	0.033	0.5	0.6	NS	0.0011
PFOS	0.00088	0.0088	0.044	0.44	0.44	NS	0.0037

Notes:

- ^a The SCOs for residential, restricted-residential and ecological resources use were capped at a maximum value of 100 ppm
- ^b The SCOs for commercial use were capped at a maximum value of 500 ppm
- ^c The SCOs for industrial use and the protection of groundwater were capped at a maximum value of 1000 ppm
- ^d The SCOs for metals were capped at a maximum value of 10,000 ppm.
- ^e For constituents where the calculated SCO was lower than the contract required quantitation limit (CRQL), the CRQL is used as the SCO value
- ^f For constituents where the calculated SCO was lower than the rural soil background concentration as determined by the Department and Department of Health rural soil survey, the rural soil background concentration is used as the Track 2 SCO value for this use of the site.
- ^g This SCO is derived from data on mixed isomers of BHC.
- ^h The SCO for this specific compound (or family of compounds) is considered to be met if the analysis for the total species of this contaminant is below the specific SCO. This SCO is for the sum of endosulfan I, endosulfan II, and endosulfan sulfate
- ⁱ This SCO is the lower of the values for mercury (elemental) or mercury (inorganic salts)
- ^k The SCOs for unrestricted use were capped at a maximum value of 100 ppm.
- ^l For constituents where the calculated SCO was lower than the contract required quantitation limit (CRQL), the CRQL is used as the Track 1 SCO value
- ^m For constituents where the calculated SCO was lower than the rural soil background concentration, as determined by the Department and Department of Health rural soil survey, the rural soil background concentration is used as the Track 1 SCO value for this use of the site.
- ⁿ Protection of ecological resources SCOs were not developed for contaminants identified in Table 375-6.8(b) with "NS". Where such contaminants appear in Table 375-6.8(a), the applicant may be required by the Department to calculate a protection of ecological resources SCO according to the TSD.
- ^o SCOs for PFAs are taken from the NYSDEC Sampling, Analysis, and Assessment of Per- and Polyfluoroalkyl Substances (PFAS) under NYSDEC's Part 375 Remedial Programs, dated April 2023.

Table 1B
Groundwater Criteria Table

1346 Blondell Avenue, Bronx, NY
BCP Site No. C203089

Contaminant	Aqueous Water Quality Standards ¹ , ug/L
Metals	
Antimony	3
Arsenic	---
Arsenic	25
Barium	1,000
Beryllium	3
Cadmium	5
Chromium, hexavalent	---
Chromium, trivalent	50
Copper	200
Cyanide	---
Iron	300
Lead	25
Magnesium	35,000
Manganese	300
Mercury	0.7
Nickel	100
Selenium	10
Silver	50
Sodium	20,000
Thallium	0.5
Zinc	2000
PCBs/Pesticides	
alpha-BHC	0.01
2,4,5-TP Acid (Silvex)	---
4,4'-DDD	0.3
4,4'-DDE	0.2
4,4'-DDT	0.2
Aldrin	---
beta-BHC	0.04
Chlordane (alpha)	---
Dibenzofuran	---
Dieldrin	0.004
Endosulfan I	0.12
Endosulfan II	0.12
Endosulfan sulfate	0.12
Endrin	---
Endrin aldehyde	5
Endrin ketone	5
gamma-BHC (Lindane)	0.05

Table 1B
Groundwater Criteria Table

1346 Blondell Avenue, Bronx, NY
BCP Site No. C203089

Contaminant	Aqueous Water Quality Standards ¹ , ug/L
PCBs/Pesticides, Con't.	
gamma-Chlordane	0.12
Heptachlor	0.04
Heptachlor epoxide	0.03
Lindane	---
Methoxychlor	35
Polychlorinated biphenyls	---
Toxaphene	0.06
Semivolatiles	
1,1'-Biphenyl	5
2,2'-oxybis(1-Chloropropane)	5
2,4,5-Trichlorophenol	1
2,4-Dichlorophenol	1
2,4-Dimethylphenol	50
2,4-Dinitrophenol	10
2,4-Dinitrotoluene	5
2,6-Dinitrotoluene	5
2-Chloronaphthalene	10
2-Chlorophenol	1
2-Methylnaphthalene	502
2-Methylphenol	1
2-Nitroaniline	5
2-Nitrophenol	1
3,3'-Dichlorobenzidine	5
3-Nitroaniline	5
4-Chloro-3-methylphenol	1
4-Chloroaniline	5
4-Methylphenol	1
4-Nitroaniline	5
4-Nitrophenol	1
Acenaphthene	20
Acenaphthylene	202
Anthracene	50
Atrazine	7.5
Benz(a)anthracene	0.002
Benzo(a)pyrene	---
Benzo(b)fluoranthene	0.002
Benzo(g,h,i)perylene	52
Benzo(k)fluoranthene	0.002
bis(2-Chloroethoxy)methane	5

Table 1B
Groundwater Criteria Table

1346 Blondell Avenue, Bronx, NY
BCP Site No. C203089

Contaminant	Aqueous Water Quality Standards ¹ , ug/L
Semivolatiles, Con't.	
Bis(2-Chloroethyl)ether	1
bis(2-Ethylhexyl)phthalate	5
Butylbenzylphthalate	50
Chrysene	0.002
Dibenz(a,h)anthracene	502
Dibenzofuran	52
Diethylphthalate	50
Dimethylphthalate	50
Di-n-butylphthalate	50
Di-n-octylphthalate	50
Fluoranthene	50
Fluorene	50
Hexachlorobenzene	0.04
Hexachlorobutadiene	0.5
Hexachlorocyclopentadiene	5
Hexachloroethane	5
Indeno(1,2,3-cd)pyrene	0.002
Isophorone	50
m-Cresol	---
Naphthalene	10
Nitrobenzene	0.4
N-Nitrosodiphenylamine	50
o-Cresol	---
p-Cresol	---
Pentachlorophenol	1
Phenanthrene	50
Phenol	1
Pyrene	50
Volatiles	
1,1,1-Trichloroethane	5
1,1,2,2-Tetrachloroethane	5
1,1,2-Trichloro-1,2,2-trifluoroethane	5
1,1,2-Trichloroethane	1
1,1-Dichloroethane	5
1,1-Dichloroethene	5
1,1-Dichloroethylene	---
1,2,4-Trichlorobenzene	---

Table 1B
Groundwater Criteria Table

1346 Blondell Avenue, Bronx, NY
BCP Site No. C203089

Contaminant	Aqueous Water Quality Standards ¹ , ug/L
Volatiles, Con't.	
1,2,4-Trimethylbenzene	5
1,2-Dibromo-3-chloropropane	0.04
1,2-Dibromoethane	0.0006
1,2-Dichlorobenzene	3
1,2-Dichloroethane	0.6
1,2-Dichloropropane	1
1,3,5- Trimethylbenzene	---
1,3-Butadiene	---
1,3-Dichlorobenzene	3
1,3-Dichlorobenzene	---
1,4-Dichlorobenzene	3
1,4-Dichlorobenzene	---
1,4-Dioxane	1 ²
2-Butanone	50
2-Hexanone	50
4-Methyl-2-pentanone	502
Acetone	50
Benzene	1
Bromodichloromethane	50
Bromoform	50
Bromomethane	5
Butylbenzene	---
Carbon Disulfide	60
Carbon tetrachloride	5
Chlorobenzene	5
Chloroethane	5
Chloroform	7
Chloromethane	5
Cis- 1,3-Dichloropropene	0.4
cis-1,2-Dichloroethene	5
cis-1,2-Dichloroethylene	---
Cyclohexane	---
Dibromochloromethane	50
Dichlorodifluoromethane	5
Ethyl Acetate	---
Ethylbenzene	5
Freon 113	---
Hexachlorobenzene	---

Table 1B
Groundwater Criteria Table

1346 Blondell Avenue, Bronx, NY
BCP Site No. C203089

Contaminant	Aqueous Water Quality Standards ¹ , ug/L
Volatiles, Con't.	
Hexachlorobutadiene	---
Hexane	---
Isopropylbenzene	5
m,p-Xylene	---
m-Dichlorobenzene	---
Methyl Acetate	NS
Methyl ethyl ketone	---
Methyl Isobutyl Ketone	---
Methyl tert-butyl ether	10
Methylcyclohexane	---
Methylene chloride	5
n-Propylbenzene	---
o-Dichlorobenzene	---
o-Xylene	---
p-Dichlorobenzene	---
sec-Butylbenzene	---
Styrene	5
tert-Butylbenzene	---
Tertiary Butyl Alcohol	---
Tetrachloroethene	5
Toluene	5
trans-1,2-Dichloroethene	5
trans-1,3-Dichloropropene	0.4
Trichloroethene	5
Trichlorofluoromethane	5
Vinyl Acetate	---
Vinyl Chloride	2
Xylene (mixed)	5
Per- and Polyfluoroalkyl Substances (PFAS)	
PFOA	0.01 ²
PFOS	0.01 ²
<p>Notes:</p> <p>¹ - Division of Water Technical and Operational Guidance Values (TOGS) Ambient Water Quality Standards and Guidance Values (AWQS), ug/L</p> <p>² - Guidance value for 1,4-Dioxane, PFOA, and PFOS is from the NYSDEC Guidance to Regulate PFOA, PFOS, and 1,4-Dioxane in State Waters, dated October 5, 2021</p> <p>ug/L - micro gram per liter</p>	

Table 1C
Soil Vapor Criteria Table

1346 Blondell Avenue, Bronx, NY
NYSDEC BCP No. C203089

Volatile Organics in Air	CAS No.	NYSDOH Soil Vapor Intrusion Guidance Criteria				Toxicity	Decision Matrix
		1	2	3	4		
1,1,1-Trichloroethane	71556	2.5	20.6	-	-	L	B
1,1,2,2-Tetrachloroethane	79345	0.4	-	-	-	M	TD
1,1,2-Trichloroethane	79005	0.4	<1.5	-	-	H	TD
1,1-Dichloroethane	75343	0.4	<0.7	-	-	L	TD
1,1-Dichloroethene	75354	0.4	<1.4	-	-	M	B
1,2,4-Trichlorobenzene	120821	0.5	<6.8	-	-	NA	TD
1,2,4-Trimethylbenzene	95636	9.8	9.5	-	-	NA	D
1,2-Dibromoethane	106934	0.4	<1.5	-	-	H	TD
1,2-Dichlorobenzene	95501	0.5	<1.2	-	-	M	TD
1,2-Dichloroethane	107062	0.4	<0.9	-	-	H	TD
1,2-Dichloropropane	78875	0.4	<1.6	-	-	M	TD
1,3,5-Trimethylbenzene	108678	3.9	3.7	-	-	M	D
1,3-Butadiene	106990	-	<3.0	-	-	H	TD
1,3-Dichlorobenzene	541731	0.5	<2.4	-	-	M	TD
1,4-Dichlorobenzene	106467	1.2	5.5	344	-	M	TD
1,4-Dioxane	123911	-	-	-	-	M	TD
2,2,4-Trimethylpentane	540841	5	-	-	-	M	D
2-Butanone	78933	16	12	-	-	M	TD
2-Hexanone	591786	-	-	-	-	NA	TD
3-Chloropropene	107051	-	-	-	-	M	TD
4-Ethyltoluene	622968	-	3.6	-	-	NA	TD
4-Methyl-2-pentanone	108101	1.9	6	-	-	M	TD
Acetone	67641	115	98.9	45.8	-	L	TD
Benzene	71432	13	9.4	10	-	H	D
Benzyl chloride	100447	-	<6.8	-	-	H	TD
Bromodichloromethane	75274	-	-	-	-	M	TD
Bromoform	75252	-	-	-	-	M	TD
Bromomethane	74839	0.5	<1.7	-	-	M	TD
Carbon disulfide	75150	-	4.2	-	-	M	TD
Carbon tetrachloride	56235	1.3	<1.3	1.1	-	H	A
Chlorobenzene	108907	0.4	<0.9	-	-	M	TD
Chloroethane	75003	0.4	<1.1	-	-	L	TD
Chloroform	67663	1.2	1.1	6.34	-	H	TD
Chloromethane	74873	4.2	3.7	-	-	M	TD
cis-1,2-Dichloroethene	156592	0.4	<1.9	-	-	M	B
cis-1,3-Dichloropropene	10061015	0.4	<2.3	-	-	NA	TD
Cyclohexane	110827	6.3	-	-	-	L	D

Table 2
Typical Analytical Parameters, Methods, Preservation, Holding Time and Container Requirements

1346 Blondell Avenue, Bronx, NY
NYSDEC BCP Site No. C203089

Sample Matrix	Analytical Parameter	Number of Samples ¹	EPA Analytical Method	Sample Preservation	Holding Time ²	Sample Container ³
Soil	VOCs	45	SW-846 Method 8260C/5035	1 - Methanol, 2 - Water; Cool to 4° C;	14 days to analysis	(3) Vial
	(TCL)			no headspace		
Soil	PCBs	45	SW-846 Method 8082A	Cool to 4° C	365 days to analysis	(1) 250 mL amber glass jar
Soil	Pesticides	45	SW-846 Method 8081A	Cool to 4° C	14 days to extraction	(1) 250 mL amber glass jar
	(TCL)					
Soil	SVOCs	45	SW-846 Method 8270D	Cool to 4° C	14 days to extraction	(1) 250 mL amber glass jar
	(TCL)					
Soil	1,4-Dioxane	45	SW-846 Method 8270D	Cool to 4° C	7 days to extraction	(2) 250 mL amber glass jars
Soil	Metals	45	SW-846 Method 6010D Series	Cool to 4° C	180 days to analysis	(1) 60 mL glass jar
Soil	Mercury	45	SW-846 Method 7471B	Cool to 4° C	28 days to analysis	(1) 60 mL glass jar
Soil	Cyanide	45	SW-846 Method 9010C/9012B	Cool to 4° C	14 days to analysis	(1) 250 mL amber glass jar
Soil	Herbicides	45	SW-846 Method 8151A	Cool to 4° C	14 days to extraction	(1) 250 mL amber glass jar
Soil	PFAs	45	EPA Method 1633	Cool to 4° C	14 Days	(1) 250 mL plastic container
Groundwater	VOCs	10	SW-846 Method 8260C	HCl; Cool to 4° C; no headspace	14 days to analysis	(3) Vial
	(TCL)					
Groundwater	VOCs with TICs, including 1,4-Dioxane	10	SW-846 Method 8260C	HCl; Cool to 4° C; no headspace	14 days to analysis	(3) Vial
	(TCL)					
Groundwater	1,4-Dioxane	10	SW-846 Method 8270D	Cool to 4° C	7 days to analysis	(2) 250 mL amber glass jar
Groundwater	SVOCs	10	SW-846 Method 8270D	Cool to 4° C	7 days to extraction	(2) 250 mL amber glass jar
	(TCL)					
Groundwater	SVOCs with TICs	10	SW-846 Method 8270D	Cool to 4° C	7 days to extraction	(2) 250 mL amber glass jar
	(TCL)					
Groundwater	Metals- total	10	SW-846 Method 6020B/7470A Series	HNO ₃ ; Cool to 4° C	28 days to analysis for Hg; 180 days to analysis for other	(1) 500 mL plastic container
	(TAL)					
Groundwater	Metals-dissolved	10	SW-846 Method 6020B/7470A Series	HNO ₃ ; Cool to 4° C	28 days to analysis for Hg; 180 days to analysis for other metals	(1) 500 mL plastic container
	(TAL)					
Groundwater	Pesticides (TCL)	10	SW-846 Method 8081B	Cool to 4° C	7 days to extraction	(2) 120 mL amber glass jar
Groundwater	Herbicides (TCL)	10	SW-846 Method 8151A	Cool to 4° C	7 days to extraction	(2) 1000 mL amber glass jar
Groundwater	PCBs	10	SW-846 Method 8082A	Cool to 4° C	365 days to analysis	(1) 250 mL amber glass jar
Groundwater	Cyanide	10	SW-846 Method 9012A	Cool to 4° C	14 days to analysis	(1) 250 mL amber glass jar
Groundwater	Mercury	10	SW-846 Method 7470 A	HNO ₃ ; Cool to 4° C	28 days to analysis	(1) 250 mL plastic container
Groundwater	PFAs	10	EPA Method 1633	Cool to 4° C	14 Days	(1) 250 mL plastic container
Soil Gas	VOCs	10	EPA Method TO-15	None	14 days to analysis	(1) Evacuated 6-Liter SUMMA® canister

Notes:

¹ Actual number of samples may vary depending on field conditions, sample material availability, and field observations. See RIWP for estimates.

² Holding times listed are method holding time calculated from time of collection and not NYSDEC ASP holding times.

³ MS/MSDs require duplicate volume for all parameters for solid matrices; MS/MSDs require triplicate volume for organic parameters for aqueous matrices and duplicate volume for inorganic parameters for aqueous matrices

Table 3
Typical Laboratory Data Quality Objectives
Soil Samples

1346 Blondell Avenue, Bronx, NY
NYCDEC BCP Site No. C203089

Parameter	Method	Matrix	Accuracy Control Limits	Accuracy Frequency Requirements	Precision (RPD) Control Limits	Precision Frequency Requirements
VOCs (TCL)	SW-846 Methods 8260B/5035	Soil	<u>Surrogates</u> % Rec. 1,2-Dichloroethane-d4 70-130 4-Bromofluorobenzene 70-130 Dibromofluoromethane 70-130 Toluene-d8 70-130 2-Chloroethoxyethane 70-130 Matrix Spikes 30-151% recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 30 per matrix type	<u>Field Duplicates</u> RPD <30 MS/MSDs (RPD) RPD <30	<u>Field Duplicates:</u> One per 20 per soils MS/MSDs: One per 30 per matrix type
VOCs with Tentatively Identified Compounds (TICs)	SW-846 Method 8260C	Soil	<u>Surrogates</u> % Rec. 1,2-Dichloroethane-d4 70-130 4-Bromofluorobenzene 70-130 Dibromofluoromethane 70-130 Toluene-d8 70-130 Matrix Spikes 36-162 % recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20	<u>Field Duplicates</u> RPD <30 MS/MSDs RPD RPD<30	<u>Field Duplicates:</u> One per 20 MS/MSDs: One per 20
PCBs	SW-846 Method 8082A	Soil	<u>Surrogates</u> % Rec. 2,4,5,6-Tetrachloro-m-xylene 30-150 Decachlorobiphenyl 30-150 Matrix Spikes 40-140% recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20 per matrix type	<u>Field Duplicates</u> RPD <50 MS/MSDs (RPD) RPD<50	<u>Field Duplicates:</u> One per 20 per soils MS/MSDs: One per 20 per matrix type
SVOCs	SW-846 Method 8270D	Soil	<u>Surrogates</u> % Rec. Phenol-d6 10-120 2-Fluorophenol 25-120 2,4,6-Tribromophenol 10-136 Nitrobenzene-d5 23-120 2-Fluorobiphenyl 30-120 4-Terphenyl-d14 18-120 Matrix Spikes 14-144% recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 50 per matrix type	<u>Field Duplicates</u> RPD <50 MS/MSDs (RPD)	<u>Field Duplicates:</u> One per 20 per soils MS/MSDs: One per 20 per matrix type
SVOCs with TICs	SW-846 Method 8270D	Soil	<u>Surrogates</u> % Rec. Phenol-d5 10-120 2-Fluorophenol 21-120 2,4,6-Tribromophenol 10-120 Nitrobenzene-d5 23-120 2-Fluorobiphenyl 15-120 4-Terphenyl-d14 41-149 Matrix Spikes 14-144%	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20	<u>Field Duplicates</u> RPD <50 MS/MSDs RPD RPD<50	<u>Field Duplicates:</u> One per 20 MS/MSDs: One per 20
1,4-Dioxane	SW-846 Method 8270D	Soil	<u>Surrogates</u> % Rec. 1,4-Dioxane-d8 15-110 Matrix Spikes 40-140% recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20	<u>Field Duplicates</u> RPD <30 MS/MSDs (RPD) RPD<30	<u>Field Duplicates:</u> One per 20 per soils MS/MSDs: One per 20
Pesticides (TCL)	SW-846 Method 8081A	Soil	<u>Surrogates</u> % Rec. Decachlorobiphenyl 30-150 Tetrachloro-m-xylene 30-150 Matrix Spikes 30-150% Recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20 per matrix type	<u>Field Duplicates</u> RPD <50 MS/MSDs (RPD) RPD<50	<u>Field Duplicates:</u> One per 20 per soils MS/MSDs: One per 20 per matrix type
Total Petroleum Hydrocarbons	SW-846 Method 8015B	Soil	<u>Surrogates</u> % Rec. o-Terphenyl 27-153 Tetracosane-d50 28-148 5 α -androstane 27-148 TPH-DRO 10-149	<u>Surrogates:</u> All samples, standards, QC samples One per 20 per matrix type	<u>Field Duplicates</u> RPD <50 TPH-DRO 44	<u>Field Duplicates:</u> One per 20 per soils One per 20 per matrix type
Herbicides	SW-846 Method 8151A	Soil	<u>Surrogates</u> % Rec. 2,4-DCAA 30-150 Matrix Spikes 30-150% Recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20 per matrix type	<u>Field Duplicates</u> RPD <50 MS/MSDs (RPD) RPD<50	<u>Field Duplicates:</u> One per 20 per soils MS/MSDs: One per 20 per matrix type
Metals (TAL)	SW-846 Method 6010D	Soil	<u>Surrogates</u> % Rec. Matrix Spikes 75-125% recovery	<u>Surrogates:</u> <u>Matrix Spikes:</u> One per 20 per matrix type	<u>Field Duplicates</u> RPD <20 MS/MSDs (RPD) RPD <20	<u>Field Duplicates:</u> One per 20 per soils MS/MSDs: One per 20 per matrix type

Table 3
Typical Laboratory Data Quality Objectives
Soil Samples

1346 Blondell Avenue, Bronx, NY
NYCDEC BCP Site No. C203089

Parameter	Method	Matrix	Accuracy Control Limits	Accuracy Frequency Requirements	Precision (RPD) Control Limits	Precision Frequency Requirements
PFAs	LCMSMS- Isotope Dilution	Soil	<u>Surrogates</u> % Rec. Perfluoro[13C4]Butanoic Acid (MPFBA) 61-135 Perfluoro[13C4]Butanoic Acid (MPFBA) 58-132 Perfluoro[13C5]Pentanoic Acid (MSPFPEA) 62-163 Perfluoro[13C5]Pentanoic Acid (MSPFPEA) 58-150 Perfluoro[2,3,4-13C3]Butanesulfonic Acid (M3PFBS) 70-131 Perfluoro[2,3,4-13C3]Butanesulfonic Acid (M3PFBS) 74-139 Perfluoro[1,2,3,4,6-13C5]Hexanoic Acid (M5PFHxA) 57-129 Perfluoro[1,2,3,4,6-13C5]Hexanoic Acid (M5PFHxA) 66-128 Perfluoro[1,2,3,4-13C4]Heptanoic Acid (M4PFHpA) 60-129 Perfluoro[1,2,3,4-13C4]Heptanoic Acid (M4PFHpA) 71-129 Perfluoro[1,2,3-13C3]Hexanesulfonic Acid (M3PFHxS) 71-134 Perfluoro[1,2,3-13C3]Hexanesulfonic Acid (M3PFHxS) 78-139 Perfluoro[13C8]Octanoic Acid (M8PFOA) 62-129 Perfluoro[13C8]Octanoic Acid (M8PFOA) 75-130 1H,1H,2H,2H-Perfluoro[1,2-13C2]Octanesulfonic Acid (M2-6:2FtS) 14-147 1H,1H,2H,2H-Perfluoro[1,2-13C2]Octanesulfonic Acid (M2-6:2FtS) 20-154 Perfluoro[13C9]Nonanoic Acid (M9PFNA) 59-139 Perfluoro[13C9]Nonanoic Acid (M9PFNA) 72-140 Perfluoro[13C8]Octanesulfonic Acid (M8PFOS) 79-136 Perfluoro[13C8]Octanesulfonic Acid (M8PFOS) 69-131 Perfluoro[1,2,3,4,5,6-13C6]Decanoic Acid (M6PFDA) 75-130 Perfluoro[1,2,3,4,5,6-13C6]Decanoic Acid (M6PFDA) 62-124 1H,1H,2H,2H-Perfluoro[1,2-13C2]Decanesulfonic Acid (M2-8:2FtS) 19-175 1H,1H,2H,2H-Perfluoro[1,2-13C2]Decanesulfonic Acid (M2-8:2FtS) 10-162 N-Deuteriomethylperfluoro-1- octanesulfonamidoacetic Acid (d3-NMeFOSAA) 24-116 N-Deuteriomethylperfluoro-1- octanesulfonamidoacetic Acid (d3-NMeFOSAA) 31-134 Perfluoro[1,2,3,4,5,6,7-13C7]Undecanoic Acid (M7- PFUDA) 61-155 Perfluoro[1,2,3,4,5,6,7-13C7]Undecanoic Acid (M7- PFUDA) 55-137 Perfluoro[13C8]Octanesulfonamide (M8FOSA) 10-112 Perfluoro[13C8]Octanesulfonamide (M8FOSA) 10-117 N-Deuterioethylperfluoro-1- octanesulfonamidoacetic Acid (d5-NEtFOSAA) 34-137 N-Deuterioethylperfluoro-1- octanesulfonamidoacetic Acid (d5-NEtFOSAA) 27-126 Perfluoro[1,2-13C2]Dodecanoic Acid (MPFDOA) 48-131 Perfluoro[1,2-13C2]Dodecanoic Acid (MPFDOA) 54-150 Perfluoro[1,2-13C2]Tetradecanoic Acid (M2PFTEDA) 22-136 Perfluoro[1,2-13C2]Tetradecanoic Acid (M2PFTEDA) 24-159 <u>Matrix Spikes</u> 46-182% recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20 per matrix type	<u>Field Duplicates</u> RPD <30 <u>MS/MSDs</u> (RPD) RPD <30	<u>Field Duplicates:</u> One per 20 per soils <u>MS/MSDs:</u> One per 20 per matrix type
			<u>Surrogates</u> % Rec. <u>Matrix Spikes</u> 80-125% recovery	<u>Surrogates:</u> <u>Matrix Spikes:</u> One per 20 per matrix type	<u>Field Duplicates</u> RPD <20 <u>MS/MSDs</u> (RPD) RPD <20	<u>Field Duplicates:</u> One per 20 per soils <u>MS/MSDs:</u> One per 20 per matrix type
Cyanide	SW-846 Method 9012A	Soil	<u>Surrogates</u> % Rec. <u>Matrix Spikes</u> 75-125% Recovery	<u>Surrogates:</u> One per 35 per matrix type	<u>Field Duplicates</u> RPD <35 <u>MS/MSDs</u> (RPD) RPD <35	<u>Field Duplicates:</u> One per 20 per soils <u>MS/MSDs:</u> One per 20 per matrix type
			<u>Surrogates</u> % Rec. <u>Matrix Spikes</u> 80-125% recovery	<u>Surrogates:</u> One per 20 per matrix type	<u>Field Duplicates</u> RPD <20 <u>MS/MSDs</u> (RPD) RPD <20	<u>Field Duplicates:</u> One per 20 per soils <u>MS/MSDs:</u> One per 20 per matrix type

Recovery criteria for laboratory control samples must be at least as stringent as MS/MSD criteria.
Laboratory control limits are periodically updated. The latest control limits will be utilized at the time of sample analysis.

Table 4
 Typical Laboratory Data Quality Objectives
 Groundwater Samples

1346 Blondell Avenue, Bronx, NY
 NYSDEC BCP Site No. C203089

Parameter	Method	Matrix	Accuracy Control Limits	Accuracy Frequency Requirements	Precision (RPD) Control Limits	Precision Frequency Requirements
VOCs (TCL)	SW-846 Method 8260C	Groundwater	<u>Surrogates</u> % Rec. 1,2-Dichloroethane-d4 70-130 4-Bromofluorobenzene 70-130 Dibromofluoromethane 70-130 Toluene-d8 70-130 <u>Matrix Spikes</u> 36-162 % recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20	<u>Field Duplicates</u> RPD <20 <u>MS/MSDs</u> <u>RPD</u> RPD <20	<u>Field Duplicates:</u> One per 20 <u>MS/MSDs:</u> One per 20
VOCs with Tentatively Identified Compounds (TICs)	SW-846 Method 8260C	Groundwater	<u>Surrogates</u> % Rec. 1,2-Dichloroethane-d4 70-130 4-Bromofluorobenzene 70-130 Dibromofluoromethane 70-130 Toluene-d8 70-130 <u>Matrix Spikes</u> 36-162 % recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20	<u>Field Duplicates</u> RPD <20 <u>MS/MSDs</u> <u>RPD</u> RPD <20	<u>Field Duplicates:</u> One per 20 <u>MS/MSDs:</u> One per 20
SVOCs (TCL)	SW-846 Method 8270D	Groundwater	<u>Surrogates</u> % Rec. Phenol-d5 10-120 2-Fluorophenol 21-120 2,4,6-Tribromophenol 10-120 Nitrobenzene-d5 23-120 2-Fluorobiphenyl 15-120 4-Terphenyl-d14 41-149 <u>Matrix Spikes</u> 14-144%	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20	<u>Field Duplicates</u> RPD <50 <u>MS/MSDs</u> <u>RPD</u> RPD <50	<u>Field Duplicates:</u> One per 20 <u>MS/MSDs:</u> One per 20
SVOCs with TICs	SW-846 Method 8270D	Groundwater	<u>Surrogates</u> % Rec. Phenol-d5 10-120 2-Fluorophenol 21-120 2,4,6-Tribromophenol 10-120 Nitrobenzene-d5 23-120 2-Fluorobiphenyl 15-120 4-Terphenyl-d14 41-149 <u>Matrix Spikes</u> 14-144%	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20	<u>Field Duplicates:</u> RPD <50 <u>MS/MSDs</u> <u>RPD</u> RPD <50	<u>Field Duplicates:</u> One per 20 <u>MS/MSDs:</u> One per 20
1,4-Dioxane	SW-846 Method 8270D	Groundwater	<u>Surrogates</u> % Rec. 1,4-Dioxane-d8 15-110 <u>Matrix Spikes</u> 40-140% recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20	<u>Field Duplicates</u> RPD <30 <u>Matrix Duplicates</u> RPD <30	<u>Field Duplicates:</u> One per 20 per soils <u>MS/MSDs:</u> One per 20
Metals (Total and Dissolved)	SW-846 Methods 6020B	Groundwater	<u>Matrix Spikes</u> 75-125% recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20	<u>Field Duplicates</u> RPD <20 <u>Matrix Duplicates</u> RPD <20	<u>Field Duplicates:</u> One per 20 <u>MS/MSDs:</u> One per 20
Mercury (Total and Dissolved)	SW-846 Methods 7470A	Groundwater	<u>Matrix Spikes</u> 75-125% recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20	<u>Field Duplicates</u> RPD <35 (dissolved) RPD <20 (Total) <u>Matrix Duplicates</u> RPD <35 (dissolved) RPD <20 (Total)	<u>Field Duplicates:</u> One per 20 <u>MS/MSDs:</u> One per 20
PCBs	SW-846 Method 8082A	Groundwater	<u>Surrogates</u> % Rec. 2,4,5,6-Tetrachloro-m-xylene 30-150 Decachlorobiphenyl 30-150 <u>Matrix Spikes</u> 40-140% recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20 per matrix type	<u>Field Duplicates</u> RPD <50 <u>MS/MSDs</u> <u>(RPD)</u> RPD <50	<u>Field Duplicates:</u> One per 20 <u>MS/MSDs:</u> One per 20 per matrix type
Herbicides	SW-846 Method 8151A	Groundwater	<u>Surrogates</u> % Rec. 2,4-DCAA 30-150 <u>Matrix Spikes</u> 30-150% Recovery	<u>Surrogates:</u> All samples, standards, QC samples <u>Matrix Spikes:</u> One per 20 per matrix type	<u>Field Duplicates:</u> RPD <50 <u>MS/MSDs</u> <u>(RPD)</u> RPD <50	<u>Field Duplicates:</u> One per 20 <u>MS/MSDs:</u> One per 20 per matrix type

Table 5
 Typical Laboratory Data Quality Objectives
 Soil Vapor Samples

1346 Blondell Avenue, Bronx, NY
 NYSDEC BCP Site No, C203089

Parameter	Method	Matrix	Accuracy Control Limits	Accuracy Frequency Requirements	Precision (RPD) Control Limits	Precision Frequency Requirements
VOCs	EPA Method TO-15	Soil Gas	<u>Surrogates</u> <u>% Rec</u> 4-Bromofluorobenzene 78-124	<u>Surrogates:</u> All samples, standards, QC samples	<u>Matrix Duplicates</u> RPD £30	<u>Matrix Duplicates</u> One per 20

Table 6
QC Sample Preservation and Container Requirements

1346 Blondell Avenue, Bronx, NY
NYSDEC BCP Site No. C203089

Sample Matrix	Analytical Parameter	No. of Samples	EPA Analytical Method	Sample Preservation	Holding Time ¹	Sample Container
Soil	VOCs	2	SW-846 Method 8260C/5035	1 - Methanol, 2 - Water; Cool to 4° C;	14 days to analysis	(3) Vial Preserved
	(TCL)			no headspace		
Soil	PCBs	2	SW-846 Method 8082A	Cool to 4° C	365 days to analysis	(1) 250 mL amber glass jar
Soil	SVOCs	2	SW-846 Method 8270D	Cool to 4° C	14 days to extraction	(1) 250 mL amber glass jar
	(TCL)					
Soil	1,4-Dioxane	2	SW-846 Method 8270D	Cool to 4° C	7 days to extraction	(2) 250 mL amber glass jars
Soil	Metals (TAL)	2	SW-846 Method 6010D Series	Cool to 4° C	180 days to analysis	(1) 60 mL glass jar
Soil	Mercury	2	SW-846 Method 7471B	Cool to 4° C	28 days to analysis	(1) 60 mL glass jar
Soil	Cyanide	2	SW-846 Method 9010C/9012B	Cool to 4° C	14 days to analysis	(1) 250 mL amber glass jar
Soil	Herbicides	2	SW-846 Method 8151A	Cool to 4° C	14 days to extraction	(1) 250 mL amber glass jar
Soil	Pesticides	2	SW-846 Method 8141A ⁶	Cool to 4° C	14 days to extraction	(1) 300 mL amber glass jar
Soil	PFAs	2	EPA Method 1633	Cool to 4° C	14 Days	(1) 250 mL plastic container

Table 6
QC Sample Preservation and Container Requirements

1346 Blondell Avenue, Bronx, NY
NYSDEC BCP Site No. C203089

Groundwater	VOCs (TCL)	1	SW-846 Method 8260C	HCl; Cool to 4° C; no headspace	14 days to analysis	(3) Vial
Groundwater	1,4-Dioxane	1	SW-846 Method 8270D	Cool to 4° C	7 days to analysis	(2) 250 mL amber glass jar
Groundwater	SVOCs (TCL)	1	SW-846 Method 8270D	Cool to 4° C	7 days to extraction	(2) 250 mL amber glass jar
Groundwater	Metals- total (TAL)	1	SW-846 Method 6020B/7470A Series	HNO ₃ ; Cool to 4° C	28 days to analysis for Hg; 180 days to analysis for other metals	(1) 500 mL plastic container
Groundwater	Metals-dissolved (TAL)	1	SW-846 Method 6020B/7470A Series	HNO ₃ ; Cool to 4° C	28 days to analysis for Hg; 180 days to analysis for other metals	(1) 500 mL plastic container
Groundwater	Pesticides (TCL)	1	SW-846 Method 8081B	Cool to 4° C	7 days to extraction	(2) 120 mL amber glass jar
Groundwater	Herbicides (TCL)	1	SW-846 Method 8151A	Cool to 4° C	7 days to extraction	(2) 1000 mL amber glass jar
Groundwater	PCBs	1	SW-846 Method 8082A	Cool to 4° C	365 days to analysis	(1) 250 mL amber glass jar
Groundwater	Cyanide	1	SW-846 Method 9012A	Cool to 4° C	14 days to analysis	(1) 250 mL amber glass jar
Groundwater	Mercury	1	SW-846 Method 7470 A	HNO ₃ ; Cool to 4° C	28 days to analysis	(1) 250 mL plastic container
Groundwater	PFAs	1	EPA Method 1633	Cool to 4° C	14 Days	(1) 250 mL plastic container
Soil Gas	VOCs	1	EPA Method TO-15	None	14 days to analysis	(1) Evacuated 6-Liter SUMMA® canister

Notes:

1 Holding times listed are method holding time calculated from time of collection and not NYSDEC ASP holding times.



December 2025
File No. 41.0163256.01
QAPP/FSP – 1346 Blondell Avenue, Bronx, NY

**ATTACHMENT A
QUALIFICATIONS**



Chunhua Liu, PhD

Senior Technical Specialist

Summary of Experience

Dr. Liu is a senior chemist with more than 10 years of experience in analytical chemistry, data validation and management, and quality control and quality assurance for remedial investigations and remedial actions. Her experience includes laboratory chemical analysis, EPA Region I and Region II data validation and data usability evaluation, data usability evaluation for Massachusetts Contingency Plan (MCP), sampling and analysis plan development in accordance with the NYSDEC Analytical Service Protocol and Massachusetts Compendium of Quality Assurance and Quality Control Requirements (QA/QC) and Performance Standards for Selected Analytical Methods, and quality control and quality assurance for Superfund and MCP projects.

Dr. Liu majored in environmental chemistry and during her doctoral study at Harvard School of Public Health, she researched analytical methods for sediment and evaluated metal fate and transport in sediment. Dr. Liu worked at Parsons for over seven years and at Gradient for one year before joining GZA. At Parsons, Dr. Liu led the quality control and assurance and data management efforts from developing Quality Assurance Project Plan (QAPP) to assuring implementation of QA/QC requirements and from field sampling preparation and arrangement to chemical data management. Dr. Liu was responsible for the QA/QC and data validation and data usability evaluation for a 10,000-acre BRAC and Superfund NPL site in New York and assisted in the successful transfer of over 8,000 acres of land. Dr. Liu performed data usability evaluation for various Massachusetts Contingency Plan sites at Gradient and GZA.

Relevant Project Experience

Senior Technical Specialist. Leads GZA human health risk assessment efforts for federal and state level superfund and MCP projects. Dr. Liu is also responsible for data usability evaluation for various projects.

Technical Director. Directed preparation and submittal of the Site-Wide Sampling and Analysis Plan (SAP) and the Site-Wide Quality Assurance Project Plan (QAPP) for a 10,000-acre Superfund site in New York in accordance with the Department of Defense (DOD), NYSDEC ASP, EPA Region II and EPA guidance. Directed project field sampling and data management. Supervised data validation in accordance with EPA Region II SOPs and NYSDEC ASP based on the NYSDEC ASP Category B deliverables. Identified laboratories qualified for project chemical analyses and interfaced with various analytical laboratories to address analytical deficiencies. Submitted data summary report to EPA Region II on a quarterly basis.

Lead Chemist and Risk Assessor. Led data usability evaluation and supported the successful closure of a 125-acre Hingham Annex Guaranteed Fixed Price Remediation Project. Dr. Liu also led the risk assessment effort and the effort of evaluating pesticide fate and transport at the site and successfully demonstrated that the pesticide conditions at the site were related to the past normal use of pesticides and therefore were not associated with the release at the Site.

Technical Director. Directed preparation and submittal of the SAP and the QAPP for various Formerly Used Defense (FUD) Sites. Supervised field sampling and data

Education

B.E., 1992, Environmental Engineering,
Tsinghua University, Beijing, China
M.E., 1995, Environmental Engineering,
Tsinghua University, Beijing, China
M.S., 1998, Environmental Health, Harvard
School of Public Health
D.S., 2000, Environmental Chemistry,
Harvard School of Public Health

Areas of Specialization

- Human Health Risk Assessment
- Ecological Risk Assessment
- Data Usability Evaluation
- Project Quality Control and Assurance
- Fate and Transport Modeling



Chunhua Liu, PhD

Senior Technical Specialist

validation in accordance with guidance from various EPA regions. Reviewed data validation and data usability report.

Technical Director. Directed data validation for various Superfund sites in EPA Region I and Region II in accordance with the EPA regional and state SOPs and the EPA Functional Guidelines. Led data validation for numerous MCP sites for various analytical analyses including metal, VOC, SVOC, pesticide, PCB, EPH, VPH, and TPH analyses.

Project Chemist. Evaluated different analytical methods for hexavalent chromium analysis. Compared analytical methods developed by NJDEP and EPA and identified the appropriate method for a CERCLA site in New Jersey.

Project Chemist. Evaluated quantitatively potential impacts to metal data usability by interference caused by common metals in environmental samples for a CERCLA site in New York.

Project Chemist. Performed data validation for indoor air samples for various CERCLA and MCP Sites to assist evaluation of potential vapor intrusion pathway.

Project Chemist. Performed Level IV data validation for a Superfund site in New York for various analytical analyses including metal, VOC, SVOC, pesticide, and PCB analyses. Reviewed TIC identification and quantitation and assessed chromatograms and mass spectrums for VOCs and SVOCs.

Project Chemist. Provided technical support, prepared QAPPs, established proper data quality objectives (DQOs) for various projects, maintained project quality control, trained junior scientists, coordinated project field sampling and laboratory analyses, addressed non-conformance issues associated with the data produced by the laboratory, conducted statistical analysis, and prepared data validation reports on numerous RCRA/CERCLA and MCP projects.

ENVIRONMENTAL ASSESSMENT AND REMEDIATION PROJECTS

(Former Malden MGP) Senior Scientist - Lead Risk Assessor, Chemist, Site Assessment, Malden, Massachusetts. Leading ongoing risk assessment work at large, complex former MGP that encompasses more than 16 acres of land and more than 10 different properties. Work has included vapor intrusion pathway evaluation, imminent hazard evaluation, substantial hazard evaluation, data usability evaluation, and risk characterizations for evaluation of effectiveness of sub-slab depressurization systems and remediation, direction of Site remediation and investigations, and verification of the need for AULs. GZA was able to demonstrate that indoor air impacts in a residential area were not related to MGP residuals, allowing for closure of that portion of the Site. Risk characterizations also demonstrated that Site conditions did not pose substantial hazards, confirming the effectiveness of the Temporary Solution.

(Commercial Point LNG Facility) Senior Scientist - Lead Risk Assessor, Site Assessment and Closure, Dorchester Massachusetts. Directed risk characterization to support MCP closure of former MGP facility that was currently being used as a Liquefied Natural Gas (LNG) storage and distribution facility. Performed risk characterization to support supplemental Phase II – IV MCP investigations and Permanent Solution status while allowing for beneficial reuse of the facility for LNG operations and a solar power generating facility. Also performed focused risk characterizations in support of an AUL filing and potential reuse options for portions of the Site.

(Former Haverhill Holder Site) Senior Scientist - Lead Risk Assessor, Site Closure, Haverhill Massachusetts. Directed risk characterization to support MCP closure of former MGP gas holder facility where wastes had been disposed. Conducted risk characterization to facilitate development of cost-effective cleanup plan involving focused soil excavation and use restrictions that allowed for achievement of a Permanent Solution. Performed a Method 3 risk characterization to support the complex supplemental Phase II investigation. This complex site encompassed properties owned by seven different parties, including residential land and portions of a river/tributary system.

(Gloucester Former MGP) Senior Scientist - Lead Risk Assessor, Human Health Risk Assessment, North Shore, Massachusetts. Performed Method 3 Risk Characterization for multiple parcels to support the Supplemental Phase II Comprehensive Site Assessment initiated by other consultants under the Massachusetts Contingency Plan. This Site included MGP impacts to



Chunhua Liu, PhD

Senior Technical Specialist

approximately 45 acres of Gloucester Harbor sediment. GZA evaluated potential human health risks via exposure to soil, groundwater, sediment, surface water, homegrown produce, and consumption of fish. In addition, Risk Characterization was used in the early stages of the project to assist identification of data gaps and Site investigation.

(Salem Power Plant) Project Manager, Cost Recovery Negotiations, Salem, Massachusetts. Working through counsel, provided advice to a prior owner with respect to remedial obligations under the Massachusetts Contingency Plan (MCP) associated with impacts to soil at this power generating facility. Work included review of hot spot evaluation, risk characterization, and remediation performed at the Site. Given the Site use, we concluded that most of the claimed costs that had been incurred by the current owner were not necessary under the MCP and thus should not be subject to recovery from our client.

(Sawyer Passway) Senior Scientist - Lead Risk Assessor, Chemist, MGP Site Closure, Fitchburg, Massachusetts. Performed a Substantial Hazard Evaluation and a Method 3 Risk Characterization to support fast-track Massachusetts Contingency Plan (MCP) Phase II/III study of large, complex former MGP facility on the banks of a major New England river. The work had to be completed within two months to meet a key regulatory deadline. Work included a risk evaluation to support a streamlined supplemental field exploration program, a risk evaluation to direct the focused soil excavation, and a substantial hazard evaluation for the cost-effective temporary solution within the required regulatory deadlines. GZA's continuing work on this project has included technical support for an insurance cost-recovery claim, periodic evaluations of the temporary solution, completion of soil stabilization/solidification pilot studies, implementation of focused remedial programs during site building demolition work and development of remedial plans directed at achieving a Permanent Solution (PS). Based on updated evaluations, a cost-effective approach to a PS was developed in 2014 and a Method 3 Risk Characterization was performed to support a PSS in 2015.

(Former Army Depot Activity Site), Technical Director, Syracuse, New York. Directed preparation and submittal of the Site-Wide Sampling and Analysis Plan (SAP) and the Site-Wide QAPP for a 10,000-acre Superfund site in New York in accordance with the Department of Defense (DOD), NYSDEC ASP, USEPA Region II and USEPA guidance. Directed project field sampling and data management. Supervised data validation in accordance with USEPA Region II SOPs and NYSDEC ASP based on the NYSDEC ASP Category B deliverables. Identified laboratories qualified for project chemical analyses and interfaced with various analytical laboratories to address analytical deficiencies. Submitted data summary report to USEPA Region II on a quarterly basis.

(Waverley Oaks Road), Senior Scientist - Lead Risk Assessor, Human Health Risk Assessment, Waltham, Massachusetts. This property has been impacted by improper storage of large quantities of waste oil that was to be used to heat on-site green houses, or to be processed and resold. This Site is regulated under the Massachusetts Contingency Plan; Massachusetts Department of Environmental Protection reviewed and approved all work plans and reports for the site investigation and risk assessment; it was downgraded from a Tier 1A to a Tier 1B Site following completion of the Phase II investigations.

Waste oil releases have impacted nearly 10-acres of an on-Site pond, stream and wetland, including 3 acres that have visible oil presented within surficial wetland soil and sediment. GZA conducted a Method 3 Risk Characterization to support the permanent solution of the Site. For the Vapor Intrusion pathway, GZA identified constituents not related to Site release and verified the conclusion based on the evaluation of Site-specific attenuation factors.

Publications and Presentations

Liu, C., J. Jay, T. Ford. Evaluation of Environmental Effects on Metal Transport from Capped Contaminated Sediment under Conditions of Submarine Groundwater Discharge. *Env. Sci. Tech.* 2001 35: 4549-4555.

Liu, C., J. Jay, R. Ika, S. James, and T. Ford. Capping efficiency for metal-contaminated marine sediment under conditions of groundwater inflow. *Env. Sci. Tech.* 2001 35: 2334-2340.

Blanchet, R., Liu, C., Bowers, T. Summary of Available Freshwater and Marine Sediment Quality Guidelines and Their Use in North America. Abstract accepted at SEATEC Conference, November, 2001

Blanchet, R., Liu, C., Bowers, T. Estimation of Average Exposure Point Concentrations for Pesticides Assuming Accumulation and Degradation in the Environment. Abstract accepted at SEATEC Conference, November, 2001



Chunhua Liu, PhD

Senior Technical Specialist

Seeley, M.R., Schettler, S., Liu, C., Blanchet, R.J., Bowers, T.S. Assessing Cancer Risks Due to Use of Insecticides to Control the Mosquito-borne West Nile Virus: Use of the Margin of Exposure Approach. Abstract accepted at Society of Toxicology, 41st Annual Meeting, March 17-21, 2002.

Chunhua Liu, Jennifer Jay, Ravi Ika, Shine James, Timothy Ford. Capping Efficiency for Metal-Contaminated Marine Sediment under Conditions of Submarine Groundwater Discharge. Poster presentation at Conference on Dredged Material Management: Options and Environmental Considerations. December 3-6, 2000

Chunhua Liu, Jennifer Jay, Timothy Ford. Evaluation of Environmental Effects on Metal Transport from Capped Contaminated Sediment Under Conditions of Submarine Groundwater Discharge. Poster presentation at Conference on Dredged Material Management: Options and Environmental Considerations. December 3-6, 2000

Chunhua Liu, Jennifer Jay, Timothy Ford. Core analysis: Is it a good indicator of metal release and capping efficiency? Poster presentation at Conference on Dredged Material Management: Options and Environmental Considerations. December 3-6, 2000

Chunhua Liu. 2000. Capping Efficiency for Metal Contaminated Marine Sediment under Conditions of Submarine Groundwater Discharge. Doctoral Thesis. Harvard School of Public Health

Chunhua Liu, Ravi Ika, Tim Ford. 1998. Metal flux in near shore capping sites under conditions of submarine groundwater discharge. In: Fourth Marine & Estuarine Shallow Water Science & Management Conference. March 15-19, 1998

Wei Lin, Guowei Fu, Chunhua Liu. 1996. Study on allocating permissible pollutants discharge based on axioms system. Chin. J. Environ. Sci. 1996 17(3):35-37

Wei Lin, Chunhua Liu, Guowei Fu. 1995. Environmental conflict analysis and its application in environmental planning and management: siting of public facilities. Chin. J. Environ. Sci. 1995 16(6): 36-39

Chunhua Liu, Yongfeng Nie, Wei Lin. 1995. Application prospects of landfill gas utilization technique in China. Pollution Control Technology 1995 8(3): 143-145

Chunhua Liu. 1995. Evaluation of gas production from sanitary landfill. Master's thesis. Tsinghua University, Beijing, P.R.China

Wei Lin, Chunhua Liu. 1994. Rudimentary study on countermeasure to comprehensively control air pollution caused by motor vehicles in China. Pollution Control Technology 1994 7(4): 1-3

Xiurong Zhang, Chunhua Liu, Yanru Yang, Qingzhong Bai. 1993. Environmental impact report of wastewater treatment plant project in Xuanhua City, China.

Chunhua Liu, Yongfeng Nie. 1993. Water balance evaluation in Hongmei hazardous waste landfill. In: Environmental Impact Assessment of Hongmei Hazardous Waste Landfill: 25-33

Chunhua Liu. 1992. Modeling landfill leachate production and migration. Bachelor Thesis. Tsinghua University, Beijing, P.R.China

Chunhua Liu. 1991. A discussion with the author of "clean water extraction from ocean water". Technology of Water Purification 1991(1): 39-41

Affiliations/Memberships

- Member, LSP Association
- Member, Society for Risk Analysis
- Certified EIT in Massachusetts

Christine A. Camardella, P.G., PMP

54 Prentice Road

Levittown, New York 11756

chriscamardella@hotmail.com

516-946-1121

Data Validation/Data Review/ Environmental Consultant

Providing consultation, data validation and data review services. These services include evaluation of analytical data with the objective of determining whether the data presented meets the site/project specific criteria for data quality and data use. Validation and review will be performed in accordance with the NYSDEC DER10 and USEPA 600/8-91/003, EPA 540/R/94/093 & EPA 540/R/94/097, USEPA National Functional Guidelines for Data Validation and EPA Region 2 SOPs for Data Validation.

Over 25 years of experience in both the laboratory and the environmental consulting field, with expertise in chemistry and environmental technology. Working knowledge of state and federal laws, regulations, and guidelines for the environment, water quality, and health including NYSDEC, NYCDEP, OER and NYSDOH. Additional skills are in quality control and quality assurance, data validation, expert technical review, chemical analysis, client communication and service, staff supervision and mentoring, regulatory interface, compliance management, work plan and proposal preparation, soil and groundwater investigation oversight, project scheduling and tracking, and technical editing as well as extensive financial management experience including forecasting and budget management. Manage and train junior staff.

Experience:

Lead Project Manager, Complex Project Management – Gas

National Grid – April 2018 - Present

- Prioritize, plan and coordinate project development activities according to stakeholder requirements.
- Supervise project team on daily basis to execute assigned projects within deadlines and budget.
- Prepare project proposals and develop project plan, schedule, and budget.
- Determine resource requirements and identify resources with the right skills to successfully execute projects.
- Assess potential risks and technical challenges and develop appropriate mitigation plans.
- Perform cash flow analysis and process invoices in a timely fashion.
- Develop cost reduction initiatives while maintaining quality and productivity.
- Analyze and resolve project issues in a timely and accurate manner.
- Provide technical guidance for large-scale projects.

Senior Project Manager

Advanced Site Restoration, LLC – September 2016 - February 2017

- Provided management and support for several retail petroleum clients.
- Prepared Phase I and Phase II Environmental Site Assessments.
- Knowledge in state and federal laws, regulations, and guidelines for the environment, water quality, and health including NYSDEC, NYCDEP, NYC OER and NYSDOH.
- Prepared high-level technical reports including Remedial Action Plans.
- Provided high level technical review of reports.
- Acted as a liaison between client and regulatory agencies.
- Provided data validation and data summary usability reports for clients.
- Managed and train junior staff while providing mentoring and support.
- Prepared proposals for clients.
- Managed several complex projects involving chemistry data QA/QC, asbestos removal, shoring installation, soil excavation, UST removal, and remediation system installation, in-situ bioremediation and soil and groundwater remediation.

Project Manager

Impact Environmental Consulting, Inc. - January 2011 - September 2016

- Managed retail petroleum client portfolios for projects in Nassau, Suffolk, Brooklyn, Queens, Bronx and Westchester.
- Advised clients on most cost-effective remedial strategies to obtain cleanup objectives.
- Expertise in Phase I and Phase II Environmental Site Assessments, remedial systems, in-situ bioremediation, UST removal, excavation, Site redevelopment, and soil and groundwater chemistry.
- Worked closely with regulatory agencies as well as local municipalities.
- Prepared and reviewed technical reports for client and/or regulatory submittal.

Pro Developed budgets and cost estimates.

- Managed and trained junior staff.

Senior Project Manager

Sovereign Consulting, Inc.- February 2009 - August 2010

- Provided project management and support for a multi-million-dollar project for a major retail petroleum company.
- Served as technical support on project with client and other team members, which has resulted in additional assessment work to obtain soil and groundwater characteristics in area of major plume.
- Managed quarterly sampling activities for an extensive groundwater monitoring well network including staffing, equipment, regulatory coordination and permits.
- Provided additional project management support for several retail petroleum projects in Brooklyn, Queens, and Nassau & Suffolk Counties.
- Maintained excellent client relationship and adhere to client expectations.
- Prepared and review technical reports for the client and regulatory agency.
- Documented cost savings of over \$500,000 resulting in an award from the client.

Senior Project Manager

Kleinfelder, Inc. - January 2004 - February 2009

- Provided project management for a multi-million-dollar portfolio in Kings County for a major retail petroleum company.
- Supervised a staff of 5 professionals and provide mentorship and guidance.
- Communicated with regulatory agencies and maintain a working relationship with key regulators.
- Provided financial management and support.
- Assisted in providing technical guidance for large-scale projects, including reviewing and recommending new technologies.
- Provided management and support for several complex projects involving asbestos removal, shoring install, excavation, UST removal, and remediation system install and groundwater and soil remediation.
- Adhered to all company and client health and safety programs.

Project Environmental Scientist

Groundwater & Environmental Services, Inc.- December 2001 - December 2003

- Provided project management support activities including client involvement, work plan and proposal preparation, project scheduling and tracking, budget evaluation and analysis, and technical editing and report finalization.
- Developed and implemented Site work plans and proposals.
- Communicated with regulatory agencies.
- Performed oversight of soil and groundwater investigations.
- Conducted Phase I/II Site assessments for acquisitions and divestments of properties: including all associated fieldwork and reports.

Water Quality Specialist/Assistant Laboratory Director

Long Island Water Corporation - January 1992 – November 2001

- Responsible for maintaining laboratory certification for potable water via EPA and Nassau County Dept. of Health (NCDOH) approved testing methods.
- Assisted in evaluating laboratory programs, coordinated sampling/testing procedures to assure regulatory compliance.
- Established and directed quality assurance programs. Prepared reports and made recommendations as needed.

Chemist

Nassau County Department of Health, Division of Labs & Research - January 1990 - January 1992

- Analyzed potable and non-potable waters through EPA and NYSDOH requirements.
- Provided assistance with quality control systems and techniques.

Education:

New York Institute of Technology
Master Degree with Distinction, Environmental Technology, 2002

Adelphi University
Master Degree, Chemistry, 1994

St. Bonaventure University
Bachelor of Science Degree, Chemistry, 1990

Nassau Community College
Courses taken in Biology & Microbiology, 1998

Licenses:

Professional Geologist, License No. 1003

Certifications:

Project Management Professional (PMP)

Professional Memberships:

Project Management Institute (PMI)
Project Management Institute (PMI) Long Island Chapter
New York State Council of Professional Geologists (NYSCPG)
Long Island Association of Professional Geologists (LIAPG)



Victoria Whelan, PG, OEP

Associate Principal

Summary of Experience

Ms. Whelan is a Certified Professional Geologist and Qualified Environmental Professional with nearly 20 years of experience in environmental assessment. She has performed and managed field investigations and remedial activities at numerous sites on Long Island and throughout the Metro New York area. She has skillfully conducted all aspects of environmental investigations and remediation. Her primary focus is to accurately assess, investigate, remediate, and maintain environmental integrity for real estate transactions and the redevelopment of brownfield or similarly environmental impaired properties.

She manages all aspects of projects with the New York State Department of Environmental Conservation (NYSDEC) Brownfield (BCP) and Voluntary Cleanup Program (VCP), the New York City Office of Environmental Remediation (NYC OER), the New York City Department of Environmental Protection (NYCDEP) and the United States Environmental Protection Agency (USEPA).

Relevant Project Experience (Prior to GZA)

NYCOER PROJECTS

Project Manager, Chester Street Brooklyn Supportive Housing Project, Brooklyn, New York. Managed all aspects of environmental project from due diligence investigation services, Phase I Environmental Site Assessment, and Phase II Environmental Site Investigation services to assisting client through NYCOER Voluntary Cleanup Program (VCP). Submitted and received approval for remedial investigation work plan, remedial investigation report, remedial action work plan, and construction health and safety plan, including a community air monitoring program. Managed removal of 12 buried aboveground storage tanks (ASTs). Managed waste characterization study to evaluate various soil types for disposal. Cost effectively utilized the NYC Clean Soil Bank as a disposal site and backfill source. Secured grant funding after receiving Notice of Satisfaction (NOS) for a Track 1 Cleanup.

Environmental Project Manager, Manhattan Avenue, Affordable Housing Project, Brooklyn, New York. Member of team that helped Ownership develop a new seven-story residential building on former factory site. Proposed development covered nearly 8,000 square feet of the property, including affordable housing with amenities such as a rear yard, recreation space, and children's play place. Site's contaminants included heavy metals and semi-volatile organic compounds. Hazardous and non-hazardous waste and non-hazardous was removed from the property as part of remediation efforts to address source material. Goal Soil Cleanup Objectives (SCOs) could not be achieved after remediation due to shallow groundwater. A track 4 Cleanup was achieved on this site by installing a composite cover inclusive of a vapor barrier. The project was completed on-time and on budget for the client to receive a NOS.

Project Manager, Bronx Community Development Project, Bronx, New York. Provided environmental services as client purchased, investigated, and remediated site for 81-unit community development, parking area and recreational area. The project is enrolled in NYC OER's VCP. Completed a Phase I ESA, VEA, Phase II ESI, RAWP and RAR.

Education

B.S., Geology, State University of New York at Oswego, 2001-2005; James Cook University 2004-2005

Licenses & Registrations

Registered Professional Geologist – 2017, New York, # 000318

Certified Professional Geologist, New York State

Qualified Environmental Professional, Institute of Professional Environmental Practice

Areas of Specialization

- Geology
- NYCOER VCP
- NYSBCP
- Environmental Assessments
- Environmental Site Investigation and Remediation
- UST Closures/Assessments
- Regulatory Compliance Planning and Permitting



Victoria Whelan, PG, QEP

Associate Principal

During remediation perimeter air monitoring was performed as per the CAMP. Designed oversaw removal of contaminated soil and installation of chemical vapor barrier during redevelopment.

NYSDEC BROWNFIELD PROJECTS

Principal-in-Charge, Former Auto Wreckers Site, Bronx, New York. Project is in the NYSDEC BCP with a planned Track 1 Cleanup. The site was successfully rezoned, and the proposed project will include 212 affordable housing apartments, 22,000 square feet (sf) of retail space, and parking. As remedial excavation was conducted it was quickly determined that the initial remedial plan would not satisfy the requirements for the project. As PIC, worked with the ownership, architect, accountant and construction team to steer the project towards new remedial goals without impacting project schedule. Remedial elements include a large-scale groundwater treatment system to address petroleum impacted groundwater and excavation of all source material ranging from depths of 2 to 15 feet below grade.

Environmental Project Manager, Confidential Residential Development, Bronx, New York. The 1.5-acre property was enrolled in the NYSDEC Brownfield Cleanup Program. Remedial components included excavation of soil exceeding the Site-specific Track 4 SCOs ranging from 2-22 feet, construction and maintenance of a composite cover system, removal of multiple underground storage tanks (USTs) and injection of Regenox and ORC Advanced (ISCO treatment) into the groundwater. Remediation also involved implementation of a CAMP. The site building was equipped with a vapor barrier and an active sub-slab depressurization system (SSDS). Throughout the process, assisted with design, maintaining a schedule and development of a Site Management Plan (SMP) and Final Engineering Report (FER).

Principal-in-Charge, Clay Street, NYCOER to NYSBCP Site, Brooklyn New York. Project consists of three parcels that share a property boundary and is in an area known to have heavy contamination. As the Principal-in-Charge, guided a team including ownership, developer and architect from the NYCOER VCP to the NYSDEC BCP based on contamination identified during the initial Remedial Investigation. Strategically conducted additional investigation to get multiple parts of the project eligible for the program and to maximize the tax credits available. Development will include a much-needed community facility in the way of a medical center, an indoor children's play center, and residential house.

USEPA PROJECTS

Project Manager, Remediation System, Confidential Client, Hicksville, New York. Managed this USEPA Superfund site for nearly 15 years through the operations and maintenance phase including a long-term groundwater treatment program, off-site soil vapor intrusion evaluations, and a large-scale groundwater sampling program. Contaminants of concern included PCBs and volatile organic compounds (VOCs). The site was complicated by multiple overlapping plumes of groundwater contamination. Collaborated with multiple property owners and their consultants to successfully drive the remediation.

NYS SPILLS PROJECTS

Project Manager, Spill Investigation and Remediation Services, Hempstead, New York. Performed a Phase I Environmental Site Assessment (ESA) that identified a gas station on the Site from 1940 through 1962, until redevelopment in the 1970s as a current commercial building. A subsequent Phase II Environmental Site Investigation (ESI) identified petroleum impacted soils, groundwater, and the presence of light non-aqueous phase liquids (LNAPL). A NYSDEC Spill Case was opened, and a Spill Investigation Work Plan was approved. Managed the spill investigation activities which included a work plan of Vacuum Enhanced Fluid Recovery (VEFR) events to evaluate feasibility of collecting residual petroleum contamination from beneath the Site building using VEFR. As part of long-term remedial plan, Monitored Natural Attenuation (MNA) and biodegradation to assess MNA is viable remedial strategy for the Site after the remediation of the LNAPL.

Certifications/Training

- 40-Hour OSHA HAZWOPER Training and 8-Hour Refreshers
- 10-Hour OSHA Construction Safety Course



Victoria Whelan, PG, QEP

Associate Principal

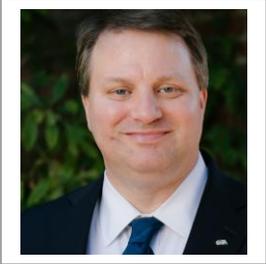
- First Aid/CPR Training
- LIRR Roadway Worker Training required by 49 CFR Part 214 Subpart C
- ARC Flash Training
- Confined Space Entry

Affiliations/Memberships

- Board Member - New York City Brownfield Partnership (NYCBP) 2022- present
- Committee Chair - Small Business Committee (NYCBP) 2022- present
- Member, New York State Council of Professional Geologists (NYSCPG)
- Member, American Council of Engineering Companies
- Member, Long Island Association of Professional Geologist

Honors & Awards

- Big Apple Brownfield Award - Hour Apartment House III
- Supportive Living Affordable Housing Award - Putnam Court
- Who's Who in Green Award - Atlantic Terrace



Stephen M. Kline, P.E.

Associate Principal/Client Manger

Summary of Experience

Mr. Kline is a registered Professional Engineer (P.E.) with over 27 years of environmental consulting and managerial experience. Mr. Kline possesses strong management, communication, and analytical skills, which make him an asset to any environmental investigation or design team. Mr. Kline has managed and has supervised operation, maintenance and monitoring programs, storage tank management programs, tank system compliance audits and engineering, and spill prevention/response planning projects throughout New York. His work has also included the installation of monitoring wells, recovery wells, dewatering wells, and remediation equipment; the design and implementation of remedial technologies; geohydrologic testing; and geophysical testing. He has managed a wide range of projects involving the construction of transit projects, power plants, schools, hospitals, residential buildings, and retail centers on urban and otherwise environmentally impacted properties in state brownfields cleanup programs and the New York City voluntary cleanup program. He is the principal author of project deliverables; including work plans and reports, site specific health and safety plans, quality assurance project plans, and contractor plans and specifications.

Relevant Project Experience

Program Manager, New York City School Construction Authority (NYC SCA), Environmental Consulting Term Contract, New York, New York. Mr. Kline has been the Program Manager for the Term-contract since 2017 and has been responsible for planning, coordination, and design of due diligence investigations at proposed New York City school sites in the five boroughs of New York. In addition, managed several the designs and installations of sub-slab depressurization systems (SSDS) and underground storage tank (UST) removals and spill closure projects. Prepared design specifications for SSDS/Vapor Barriers, UST removal and excavations, excavated material sampling, and transportation and disposal. Prior to becoming Program Manager, Mr. Kline has worked on the NYCSCA term contract since 2011.

Principal-In-Charge, West Side Federation of Senior and Supportive Housing, Mill Brook Terrace Development, Bronx, New York. Managed all aspects of the environmental project from due diligence investigation services, Phase I Environmental Site Assessment, and Phase II Environmental Site Investigation services to assisting the client through the New York City Office of Environmental Remediation (OER) Voluntary Cleanup Program (VCP). The investigation revealed that an area of hazardous lead levels in the subsurface soils. Managed a waste characterization study to evaluate various soil types for disposal. Submitted and received approval for a remedial investigation work plan, a remedial investigation report, remedial action work plan, and a construction health and safety plan with included a community air monitoring program. Managed the City/State Environmental Quality Review (CEQR), and National Environmental Policy Act (NEPA) environmental assessment services. The regulatory approval and development process required strict adherence to deadlines for deliverables, and close coordination with several city and state agencies.

Principal-In-Charge, BJs Wholesale Club, Inc, Brooklyn Bay Center Redevelopment, Brooklyn, New York. The remediation involved implementation Community Air

Education

M.B.A., Executive Management, St. John's University, Tobin College of Business, 2014

B.S., Environmental Systems Engineering, Cornell University, 1992

Licenses & Registrations

Professional Engineer:

Rhode Island, #6991 (Dormant)

Pennsylvania, #60796

New York, #80431

New Jersey, #24GE04468900

Certifications & Specialized Training

- OER Gold Turbo Training Certificate
- Transportation Worker Identification Credential
- 40-Hour OSHA Hazardous Waste Operations and Emergency Response
- 8-hour OSHA Supervisor
- 10-hour OSHA Construction Safety
- Track Safety Trained for Long Island Railroad and Metropolitan Transportation Authority (MTA)

Areas of Specialization

- Environmental Site Investigation and Remediation
- Environmental Construction Support and Dewatering
- Regulatory Compliance Planning and Permitting
- Hazardous Materials Assessment and Abatement
- Landfill Engineering and Monitoring



Stephen M. Kline, P.E.

Associate Principal

Monitoring Program (CAMP), and the management, handling, and off-site disposal of impacted material above site-specific soil cleanup objectives. The site building was equipped with a Liquid Boot™ spray applied vapor barrier and a passive sub-slab depressurization system (SSDS). The interior of the building was monitored for VOCs, methane and hydrogen sulfide using a gas detection system. Mr. Kline performed contractor submittal review and vapor barrier testing and PE inspections to document the installation of the engineering controls in accordance with the NYC OER RAP. Assisted in the development of a Site Management Plan (SMP) and Remedial Completion Report (RCR).

Principal-In-Charge, 512 West 143rd Street West 143rd Street, The Brotherhood / Sister Sol, New York, New York. Pursuant to Title 62, Chapter 5: Rules of for the City Environmental Quality Review (CEQR), amended January 26, 2014, GZA performed an Environmental Assessment Statement (EAS) Short Form in connection with the construction of the new Community Center. The project funding meant that the NYC Economic Development Corporation (NYCEDC) was the Lead Agency and The New York State grants are being administered by the Dormitory Authority of the State of New York (DASNY). GZA completed this EAS Short Form in general conformance with the CEQR Technical Manual dated March 2014, with revisions up to April 27, 2016. Additional studies included Shadows, Historical and Cultural Resources, Air Quality and Hazardous Materials. The full demolition of the existing building meant that in addition to the Phase I, Phase II Work Plan, Phase II Investigation, Remedial Action Work Plan (RAWP) and Construction Health and Safety Plan (CHASP), GZA also performed an ACM and Lead Paint Survey of the building. The project received a Negative Declaration.

Principal-In-Charge, 910 East 192nd Street, LLC, Spill Investigation and Remediation Services, Bronx, New York. Performed a Phase I Environmental Site Assessment (ESA) that identified a gas station on the Site from 1940 through 1962, until redevelopment in the 1970's as a current community center building. A subsequent Phase II Environmental Site Investigation (ESI) identified petroleum impacted soils, groundwater, and the presence of light non-aqueous phase liquids (LNAPL). A NYSDEC Spill Case was opened and a Spill Investigation Work Plan was approved. Managed the spill investigation activities and certified as a PE the Spill Investigation Report which included a work plan for an Interim Remedial Measure (IRM). The IRM consisted of Vacuum Enhanced Fluid Recovery (VEFR) events to evaluate the feasibility of collecting residual petroleum contamination from beneath the Site building using VEFR. Based on favorable IRM results, GZA negotiated conditional approved of a Remedial Action Work Plan to continue the VEFR events for eight quarters along with quarterly soil vapor and groundwater monitoring. As part of our remedial strategy, GZA collects Monitored Natural Attenuation (MNA) and biodegradation to assess MNA is a viable remedial strategy for the Site after the remediation of the LNAPL.

Project Manager, Columbia University, Consolidated Edison (Con Edison) Public Utility Regenerating Station (PURS), West Harlem New York. Managed the due diligence Site Investigation inform a property transaction between Columbia University and Con Edison. Plans for this Site include relocating the existing Con Edison PURS facility and redevelopment of the Site as part of a new campus construction, including complete excavation for subgrade basement levels up to 60 feet bgs. A comprehensive subsurface investigation included installation of five groundwater monitoring wells, collection of 22 soil samples collected and analyzed from 11 soil borings, collection of five soil vapor samples for analysis. Additionally, a utility clearance was conducted, Community Air Monitoring was completed; and hydraulic conductivity aquifer testing was completed. Vibration monitoring of sensitive Con Edison structures conducted during Sonic drilling. GZA's report documented the former Manufactured Gas Plant (MGP) Structures and subsurface soil and groundwater impacts requiring special handling during redevelopment of the Site.

Project Manager, Enercon Inc., Indian Point Energy Center, Buchanan, New York. Supervised a field team during the implementation of a comprehensive site investigation to delineate and determine the source of tritium, strontium and cesium detected in groundwater at the Indian Point Nuclear Plant. The investigation consisted of a thorough review of construction drawings, historic hydrogeologic data and historic groundwater chemistry data to prepare a CSM for the release. GZA advanced 36 bedrock and overburden borings at the Site to supplement the existing 15 groundwater monitoring wells at the Site. Rock cores were characterized for the presence of water bearing fractures as well as lithology. All bedrock borings were subject to downhole geophysical borehole logging consisting of acoustic televiewer, optical televiewer, temperature, conductivity, and heat pulse flow



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meter. Hydraulic conductivity was evaluated using a combination of extraction packer testing and sustain yield pump tests. Wells were completed using multilevel sampling systems resulting in 127 sample intervals. GZA then completed an organic dye tracer test to confirm contaminant flow paths and groundwater velocities. GZA was able to delineate the extent of horizontal and vertical groundwater contamination, determine the sources of the contamination and the post release flow paths. GZA then designed a pilot remediation system in order to allow for hydraulically contain contaminated groundwater.

Project Manager, TransCanada, Ravenswood Power Generating Station, Long Island City, New York. Managed the spill response investigation following the discovery of a 25,000-gallon kerosene release from an underground fuel oil line connecting a gas turbine generator with the 2-million-gallon aboveground storage tank (AST). GZA conducted a subsurface investigation to delineate the vertical and horizontal extent of kerosene associated with the release. Developed a Conceptual Site Model (CSM) that accounted for Site features (old foundation elements and utility conduits) and shallow bedrock which complicated kerosene migration patterns. Recommended IRM to begin collecting recoverable kerosene. The spill investigation (including off-site delineation and IRM) was completed within 3 months of mobilization to the Site. The spill investigation included: a vapor intrusion study, an inspection of the East River bulkhead, a tidal influence study, the installation of over 30 soil borings, installation of 14 monitoring wells, installation of 4 recovery sumps, and design of an IRM product recovery system. Within the first 4 months of the release 4,000 gallons of kerosene were recovered, and a report was submitted to the NYSDEC.

Project Manager, Dormitory Authority State of New York (DASNY), Environmental Term Contract, Various Locations, New York. Managed environmental sampling and analytical testing for DASNY-controlled building projects for the 6 years. Provided subcontractor selection and underground storage tank (UST) testing, repair and closure services for Manhattan Family Court building, Brooklyn College Heating Plant, and the Bronx Community College water supply system. In support of the Bellevue Hospital Office of the Chief Medical Examiner building construction, performed soil and groundwater investigation of an existing petroleum spill, and reported on the impacts of observed contamination on soil excavation, disposal, and construction dewatering. Brooklyn College Heating Plant remedial design included 48-hour constant rate pumping test, development of contract plans and specifications, New York City Department of Environmental Protection (NYCDEP) sewer discharge and New York State Department of Environmental Conservation (NYSDEC) Long Island Well Permits. Principle author of all deliverables including NYSDEC-approved: Site Investigation/Spill Investigation Work Plans; Remedial Action Work Plans; Spill Closure Reports; and Remedial Action/Corrective Action Reports. Performed extensive project administration, including coordination between subcontractors, DASNY, construction managers, on-site engineering staff, and the appropriate New York City and State regulatory agencies.

Environmental Project Manager, Metropolitan Transit Authority, Fulton Street Transit Center, New York. Developed and authored a work plan, a health and safety plan, and an emergency action plan for the subsurface investigation for a metropolitan New York City site investigation field program. During conceptual design and preliminary engineering phases performed to date, has managed the evaluating soil and groundwater conditions at the site as they relate to: soil excavation and the handling, transportation and disposal of potentially contaminated soil; and groundwater quality and groundwater disposal. In addition, has reviewed and supported consultant submissions for the Draft Environmental Impact Statement (DEIS), and authored various Design Criteria Manual and Constructability Report assessments, and contractor specifications.

Project Manager, Fort Washington Park EAS, Stantec, New York, New York. Managed the hazardous materials assessment portion of the Environmental Assessment Statement (EAS) for the 160-acre Fort Washington Park (FWP) as part of the New York City Environmental Quality Review (CEQR). The New York City Department of Parks and Recreation (DPR) parkland is located adjacent to the east bank of the Hudson River on the far Upper West Side of Manhattan from 135th Street in the south to Spuyten Duyvil in the north. GZA evaluated the potential for contaminated materials in the soil, groundwater or building materials of FWP to be disturbed during reconstruction and excavation activities. The preliminary contaminated materials assessment identified 73 potential sources of contamination and recommended additional investigations at 14 of these areas. Additional phases of work included: development and approval of Sampling and Analysis Plans; Preliminary Environmental Site Assessments; Construction Soil Management Plans; Construction Health and Safety Plans; and coordination with the NYSDEC and NYCDEP.



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Project Manager, Triangle Equities Development Company, The Junction Site Redevelopment, Brooklyn, New York. Under the City Environmental Quality Review (CEQR) procedures for redevelopment of New York City-owned properties, GZA performed a multi-phase site assessment of soil, groundwater and building conditions of three contiguous properties and an active Long Island Rail Road (LIRR) easement with a petroleum pipeline that bisects the planned development. Assessment techniques included surficial geophysical surveys, soil borings, monitoring well installations, test pits and soil vapor surveys of two former gasoline filling stations and auto repair facilities. Six abandoned USTs were located, registered, and closed by removal. All project deliverables were reviewed and approved by the NYCDEP Office of Environmental Planning and Assessment (OEPA). Performed all project administration, including coordination between subcontractors, MTA/LIRR, and New York City and State regulatory agencies.

Project Manager, Horrigan Development Partners, North 9th Street, Brooklyn, New York. Dr. Winslow managed CEQR “e” designated Site requirements associated with the redevelopment of a former industrial building for reuse as a residential building. GZA submitted a Site Investigation Work Plan to the NYCDEP for review. Following acceptance of the work plan, GZA conducted a Site Investigation and submitted a Site Investigation Report combined with a Remedial Action Plan to save time for the client. The Site was found to be contaminated with chlorinated solvents. GZA proposed a remedial action which coincided with the proposed redevelopment of the parcels. The remedial design consisted of excavation of soils to bedrock to accommodate the foundation design of the proposed building. Soil samples were collected in a grid across the Site at multiple depths to further characterize VOC contaminated soils, historic fill material and native soils for disposal purposes. This resulted identification of “cells” of soil that would be transported to different facilities for significant cost savings to the client. In addition, GZA recommended a soil vapor barrier and post excavation groundwater monitoring. Both the NYCDEP and the NYSDEC approved the work plans.

Project Manager, New York City Department of Corrections (DOC), Rikers Island, East Elmhurst, New York. Managed monthly groundwater elevation and Light Non-Aqueous Phase Liquid (LNAPL) monitoring services at Rikers Island Prison in the vicinity of the powerhouse complex. Scope of services has included sampling performed using the low-flow, pneumatic bladder pump equipment that the DOC installed in the monitoring wells, laboratory analysis, and NYSDEC reporting. Negotiated with the NYSDEC first to reduce from quarterly to annual sampling and subsequently to close Spill and cease plume monitoring. The DOC renewed the annual contract for five years prior to GZA receiving closure from the NYSDEC. Prepared a site-specific health and safety plan.

Project Manager, Columbia Presbyterian Hospital, Spill Investigation/Closure and Tank Repairs, New York. The 15,000-gallon emergency power generation UST was found to be leaking during the USEPA Region II compliance auditing. The soil and groundwater conditions were evaluated during the spill investigation. Recommended limited soil removal, tank repairs and tank upgrades over UST removal and replacement. Produced designs, plans and specifications to bring UST and associated piping into compliance with current standards. Provided both design and construction management services. Submitted spill investigation and closure reports to the NYSDEC. Closed Spill without removal of existing UST at considerable cost savings to the client.

Field Manager, NYSDEC Superfund Standby Contract (Contract No. D003060), Various Locations. Between 1998 and 2002, conducted project management, field oversight, historical review, and report preparation. This multi-year, multi-million-dollar term environmental contract covered the following work items related to sites within the New York State superfund program, included: Preliminary Site Assessments; Remedial Investigations; Feasibility Studies; Remedial Design; and Remedial Construction.

CONSTRUCTION SUPPORT AND DEWATERING

Project Manager, Westchester County Airport Proposed Baggage Screening Facility, TY LIN International, White Plains, New York. Managed an environmental site investigation of the airport prior to a new construction project. This Site Investigation Report (SIR) is based on our review of available historical and environmental records, information collected during field investigations, and laboratory analysis of collected soil and groundwater samples. Made recommendations and developed specifications for tanks closure services, soil handling, and above ground storage tank installation for the project.

Project Manager, NYCEDC, Willets Point Redevelopment Project, Stantec, Queens, New York. In charge of coordinating the dewatering specifications and pretreatment design for the dewatering of the infrastructure development of the 40-acre site. The



Stephen M. Kline, P.E.

Associate Principal

infrastructure development includes a sanitary sewer main (2,400 lf) through a private property (CitiField), two storm sewer outfalls (2,500 lf each) along 126th Street and 127th Street and an existing 72" diameter water main along Willets Point Boulevard. Included preliminary permitting of NYCDEP sewer discharge versus State Pollution Discharge Elimination System (SPDES) permit, and Long Island Well Permits for each alignment. Permitting included environmental justice zones, estuary discharge, and several contractor design changes.

Project Manager, PS-27K, - Dewatering Permits, Brooklyn, New York. As part of construction of a spill case closure and waterproofing work, performed a site investigation and designed a treatment system to obtain a NYCDEP wastewater quality control application, a temporary sewer discharge dewatering permit and a LI Well permit for dewatering discharges. Performed geohydraulic testing to determine pumping rates of up to 90 gallons per minute. Performed a treatability study to demonstrate adequate treatment of the petroleum-impacted groundwater prior to sewer discharged.

Project Manager, Romanoff Enterprises, 402 W 13th Street - Dewatering Permits, New York, New York. As part of construction of a commercial building, performed a site investigation and designed a treatment system to obtain a NYCDEP wastewater quality control application and a temporary sewer discharge dewatering permit for dewatering discharges. Performed geohydraulic testing to determine pumping rates of up to 200 gallons per minute. Performed a treatability study to demonstrate adequate treatment of the PCE impacted groundwater prior to sewer discharged. Also prepared an engineer's estimate of the volume of dewatering effluent discharged to the combined sewer for submittal to the NYCDEP. Reviewed dewatering contractor submittals.

HAZARDOUS MATERIALS ASSESSMENT / ABATEMENT

Project Manager, Wishcamper Group, Parkledge Apartments Hazardous Materials Assessment, Yonkers, New York. Managed the assessment of four Site buildings to be renovated for multi-family residential purposes. The asbestos containing materials (ACMs), lead paint, radon and hazardous material survey was conducted as part of "due diligence" activities. The objective was to facilitate the making of risk management decisions regarding the suitability of property for acquisition, and for general use in planning of abatement activities and for preparing operations and maintenance programs. Electrical/hydraulic equipment was inventoried and evaluated for special handling prior to proposed demolition and renovation activities at the Site. Subsequent phases of work have involved abatement of identified ACMs during renovations, and clearance sampling for re-occupancy of living areas after abatement.

Project Manager, DMJM+Harris, New York City Health and Hospitals Corporation, Various Locations, New York City, New York. As part of the HHC Emergency Power System Upgrades designed by DMJM+Harris, GZA conducted pre-demolition asbestos surveys of eight hospitals in the Five Boroughs. Managed the surveys, survey reports and the preparation of contractor abatement specifications. Abatement specifications were incorporated into bid documents produced by the DASNY.

Project Manager, DMJM+Harris, New York City Technical College, Brooklyn, New York. As part of the DASNY upgrade of the HVAC System Upgrades designed by DMJM+Harris, GZA conducted pre-demolition asbestos surveys of four campus buildings located on the Brooklyn campus. Managed the surveys, survey reports and the preparation of contractor abatement specifications. Abatement specifications were incorporated into bid documents produced by the DASNY.

Project Manager, Granary Associates, Memorial Sloan Kettering Cancer Center, New York, New York. Assisted in coordinating a pre-demolition laboratory and hazardous material survey and the preparation of the survey report, drawings, and data tabulation. The survey included an evaluation and inventory of all hazardous chemicals, waste and biohazards associated with each laboratory. Once the hazard assessment was completed, a survey was designed to determine impacts from use on building materials in light of planned demolition activities. Areas such as storage cabinets, duct work, fume hoods, bench tops and plumbing were evaluated for presence of hazardous substances. In addition, a lead-based paint survey and a universal waste survey were conducted. Laboratory decontamination and hazardous material specifications were created, and a contractor was selected. GZA personnel coordinated and over saw the decommissioning and abatement.



Stephen M. Kline, P.E.

Associate Principal

Project Manager, PANYNJ, Mold Assessment and Abatement, Central Terminal Building, LaGuardia Airport, Flushing, New York. Managed a pre- and post-abatement mold investigation of two specific locations within the Central Terminal Building of LaGuardia Airport. Investigation included collection of swab, bulk, air and carpet samples from areas exhibiting mold growth before, during and after the mold abatement activities. Worked with the PANYNJ's accelerated schedule, which included evening and weekend work. Based on the analytical results, assisted PANYNJ in notifying the employees working in the affected areas, and recommended specific mold abatement activities. The mold abatement included containing the two discrete, affected areas, removing the impacted materials for disposal, cleaning and HEPA vacuuming the permanent (non-disposable) items, and scrubbing the air in the containment areas for 72 hours following the active abatement. Managed periodic contractor oversight during abatement activities on behalf of PANYNJ.



Zachary Landis, P.E.

Project Manager

Summary of Experience

Mr. Landis is a Licensed Engineer with six years of environmental consulting, engineering, and research experience. His focus is on site investigations for contaminated soil, groundwater, and surface water and remedial construction. He has managed and implemented environmental activities in multiple states at residential, commercial, industrial, and publicly owned sites. Mr. Landis performs field work and supports the closure of sites utilizing source removal, in-situ and ex-situ remedial technologies, and waste removal and risk-based corrective action methods in accordance with state and federal regulations. He has managed complex multi-site projects requiring significant planning and coordination with subcontractors and vendors.

Prior to consulting, Mr. Landis worked as a researcher for the National Science Foundation and Virginia Tech. His projects were focused on water quality and erosion management in urban, rural, and suburban areas.

Relevant Project Experience

ENVIRONMENTAL CONSULTING

Site Supervisor, 645 Gates Avenue Redevelopment/The Astra, Brooklyn, New York. Oversaw remedial actions for construction of a six-story, mixed-income affordable housing building. The project involved excavation and removal of 11 underground storage tanks (USTs) and approximately 7,500 CY of impacted soils. During excavation and backfilling activities, completed confirmation sampling of each UST and completed excavation area. Conducted Community Air Monitoring Program (CAMP). Designed and oversaw installation of building's vapor barrier system.

Project Engineer, DLA Pipeline Spill, Atlantic Undersea Test and Evaluation Center, NAVFAC Southeast, Andros Island, Bahamas. Provided technical support for design and implementation of air sparging system at the US Navy's AUTEK Facility. Provided analysis of the effectiveness of varying air sparging system layouts and long-term monitoring of groundwater sampling at the facility after previous soil delineation and remedial actions for a pipeline spill and a tidal influence investigation had been completed. Coordinated complex field event logistics with subcontractors, field personnel, and clients at remote military installation.

Project Engineer, Bethany Hills Wells PFAS Response Action, USACE Omaha District, Off Base from Luke AFB, Arizona. Oversaw installation of a bypass pipeline at an existing municipal PFAS water treatment facility and designed and installed a treatment media evaluation study at the facility. The bypass pipeline was installed to divert an emergency source of previously treated municipal water supply around the PFAS treatments system to avoid chlorinated water from impacting treatment media. The treatment media evaluation study consisted of a four-column test skid that diverted untreated effluent from the PFAS treatment system to test four separate treatment medias (ion exchange resins, granular activated carbon, and an organoclay) and reintroduce treated water into the treatment system. The study was used to evaluate effectiveness of each media and design a full-scale treatment at this facility as well as additional Air Force operated facilities. Also conducted monthly analysis of

Education

B.S., Earth and Environmental Sciences,
Virginia Wesleyan University, 2017
M.E., Environmental Engineering, Old
Dominion University, 2018

Licenses & Registrations

Professional Engineer – Virginia,
#0402066593, New York #112679

Areas of Specialization

- Environmental Site Investigation and Remediation
- Construction Management
- Permitting & Compliance
- Soil, Groundwater, Air Sampling
- Air Quality and Surface Water Modeling
- Technical Specification Preparation
- Bid Evaluation
- Remediation Oversight



Zachary Landis, P.E.

Project Manager

PFAS breakthrough for the treatment system, prepared engineering evaluation and cost analysis of new PFAS extraction and treatment system at David-Monthan AFB.

Project Engineer and Quality Engineer, MSFIH Site 67, 4915, Navy Facilities Engineering Systems command, Naval Support Facility Indian Head, Maryland. Provided quality control and health and safety oversight for project involving excavation of seven soil removal areas with contaminants including heavy metals and VOCs. One area required removal of riverbed sediment from the shoreline to approximately 45 feet into a creek. Each removal area's excavated soils were stockpiled and evaluated separately until transportation and disposal to an offsite facility. Site restoration included backfilling, replanting of wetland/river areas that were disturbed, grass seeding, and repairing the stormwater drainage system. Reviewed executed work, provided client with progress updates and changes in conditions, and prepared all required submittals.

Project Engineer - Technical Lead, Fried Industries Superfund Site, East Brunswick, New Jersey. Provided technical support and design for excavation of approximately 8,000 CY of VOC impacted soils and installation and testing of a monitoring well and extraction well for the existing groundwater treatment facility. Excavation was completed within a wetland area and required a combination of slide rail shoring and open excavation with sloped sides. After backfilling and compaction, two wells were installed to go feet within the competent bedrock and tested for contaminant concentrations and yield. The more significantly impacted well was constructed as an extraction well; a below grade pipeline was installed to connect it to the existing groundwater treatment plant at the site. The other well was constructed as a monitoring well to be added to site's existing monitoring well network. After completion of well installation, wetland area was restored with native plantings and an impacted roadway was restored in kind.

STORMWATER MANAGEMENT

Project Engineer, Underserved Communities Stormwater Management, Virginia Department of Emergency Management, Multiple Locations, Virginia. Provided engineering support for FEMA applications for stormwater management funds, including engineering estimates and evaluations for project feasibility for stormwater management projects including home elevations, installation of subsurface drainage systems, wet ponds, breakwaters, and culverts. For several localities, provided onsite investigations and prepared full design packages and specifications for projects including a breakwater, a wet pond and subsurface drainage system, and a subsurface drainage system.

Research Aid, Virginia Tech, Biological Systems Engineering Department, Virginia Beach, Virginia. Aided research on stormwater management and water quality in the Hampton Roads, Virginia area, including rural, suburban, and urban, runoff assessments and modeling impacts to the Chesapeake Bay Watershed. Responsible for data collection and management, setting up sampling and monitoring equipment at various site locations, conducting particle size distribution analyses, and maintaining equipment in compliance with research targets.

International Researcher, National Science Foundation, Xilingol, China. He researched the aeolian and fluvial erosion conditions of the steppe grasslands of northern China within the Inner Mongolia Region. The research included the development of a mobile air tunnel and rainfall simulator for the testing of bare and vegetated portions of the grassland for the development of an erosion model for the region. He was responsible for the field testing and sampling of the air tunnel and rainfall simulator for data collection, analysis of the results, and the development of the model to account for both aeolian and fluvial erosion factors.

Certifications/Training

- Transportation Worker Identification Credential – Nationwide, 05860698
- NYC Site Safety Training, New York, 4M60011129
- First Aid and CPR
- Hazardous Materials Supervisor, Nationwide, UMD201285
- OSHA 10-hr Construction Safety & Health
- OSHA 30-hr Construction Safety & Health
- OSHA 40-hr Hazardous Waste Operations



Zachary Landis, P.E.

Project Manager

- OSHA 8-hr Fall Protection
- OSHA 2-hour Drug & Alcohol Awareness
- US Army Corps of Engineers Construction Quality Management for Contractors

Publications and Presentations

Zachary Landis, Xixi Wang, Nicholas Potter, Measuring and Modeling Bare Topsoil Erosion by Wind from a Steppe Grassland of China, AGU Fall Meeting, Washington D.C., 2018

Zachary Landis, International Research in Environmental Science and Engineering, Virginia Wesleyan University's Honors and Scholars Lecture Series, Virginia Beach, VA, 2018



December 2025
File No. 41.0163256.01
QAPP/FSP – 1346 Blondell Avenue, Bronx, NY

ATTACHMENT B
LABORATORY STANDARD OPERATING PROCEDURES

Method 1633 Analysis of Per- and Polyfluoroalkyl Substances (PFAS) in Aqueous, Solid, Biosolids, Oil and Tissue Samples by LC-MS/MS

References: Method 1633 - Analysis of Per- and Polyfluoroalkyl Substances (PFAS) in Aqueous, Solid, Biosolids, Oil and Tissue Samples by LC-MS/MS (2nd Draft -June 2022)

DOD QSM (US Department of Defense Quality Systems Manual for Environmental Laboratories, version 5.4, 20221)

1. Scope and Application

Matrices: Drinking water, Non-potable Water, Tissues, Oils, Biosolids and Solid Matrices

Definitions: Refer to Alpha Analytical Quality Manual.

- 1.1** Method 1633 is for use in the Clean Water Act (CWA) for the determination of the per- and polyfluoroalkyl substances (PFAS) in Table 1 in aqueous, solid (soil, biosolids, sediment) and tissue samples by liquid chromatography/mass spectrometry (LC-MS/MS).
- 1.2** The method calibrates and quantifies PFAS analytes using isotopically labeled standards. Where linear and branched isomers are present in the sample and either qualitative or quantitative standards containing branched and linear isomers are commercially available, the PFAS analyte is reported as a single analyte consisting of the sum of the linear and branched isomer concentrations
- 1.3** This is a liquid chromatography/tandem mass spectrometry (LC/MS/MS) method for the determination of selected perfluorinated alkyl substances (PFAS) in Non-Drinking Water, tissue soil and biosolid Matrices. Accuracy and precision data have been generated for the compounds listed in Table 1.
- 1.4** The data report packages present the documentation of any method modification related to the samples tested. Depending upon the nature of the modification and the extent of intended use, the laboratory may be required to demonstrate that the modifications will produce equivalent results for the matrix. Approval of all method modifications is by one or more of the following laboratory personnel before performing the modification: Area Supervisor, Department Supervisor, Laboratory Director, or Quality Assurance Officer.
- 1.5** This method is restricted to use by or under the supervision of analysts experienced in the operation of the LC/MS/MS and in the interpretation of LC/MS/MS data. Each analyst must demonstrate the ability to generate acceptable results with this method by performing an initial demonstration of capability.

2. Summary of Method

- 2.1** Environmental samples are prepared and extracted using method-specific procedures. Sample extracts are subjected to cleanup procedures designed to remove interferences. Analyses of the sample extracts are conducted by LC-MS/MS in the multiple reaction monitoring (MRM) mode. Sample concentrations are determined by isotope dilution or extracted internal standard quantification using isotopically labeled compounds added to the samples before extraction.

- 2.2** Aqueous samples are spiked with isotopically labeled standards, extracted using solid-phase extraction (SPE) cartridges and undergo cleanup using carbon before analysis.
- 2.3** Solid and Oil samples are spiked with isotopically labeled standards, extracted into basic methanol, and cleaned up by carbon and SPE cartridges before analysis
- 2.4** Tissue samples are spiked with isotopically labeled standards, extracted in potassium hydroxide and acetonitrile followed by basic methanol, and cleaned up by carbon and SPE cartridges before analysis.
- 2.5** A sample extract is injected into an LC equipped with a C18 column that is interfaced to an MS/MS). The analytes are separated and identified by comparing the acquired mass spectra and retention times to reference spectra and retention times for calibration standards acquired under identical LC/MS/MS conditions. The concentration of each analyte is determined by using the isotope dilution technique. Extracted Internal Standards (EIS) analytes are used to monitor the extraction efficiency of the method analytes.

2.6 Method Modifications from Reference

N/A

3. Reporting Limits

The reporting limit for PFAS's are listed in Table 8.

4. Interferences

- 4.1** PFAS standards, extracts and samples should not come in contact with any glass containers or pipettes as these analytes can potentially adsorb to glass surfaces. PFAS analyte and EIS standards commercially purchased in glass ampoules are acceptable; however, all subsequent transfers or dilutions performed by the analyst must be prepared and stored in polypropylene containers.
- 4.2** Method interferences may be caused by contaminants in solvents, reagents (including reagent water), sample bottles and caps, and other sample processing hardware that lead to discrete artifacts and/or elevated baselines in the chromatograms. The method analytes in this method can also be found in many common laboratory supplies and equipment, such as PTFE (polytetrafluoroethylene) products, LC solvent lines, methanol, aluminum foil, SPE sample transfer lines, etc. All items such as these must be routinely demonstrated to be free from interferences (less than 1/2 the RL for each method analyte) under the conditions of the analysis by analyzing laboratory reagent blanks as described in Section 9.1. Subtracting blank values from sample results is not permitted.
- 4.3** Matrix interferences may be caused by contaminants that are co-extracted from the sample. The extent of matrix interferences will vary considerably from source to source, depending upon the nature of the water. Humic and/or fulvic material can be co-extracted during SPE and high levels can cause enhancement and/or suppression in the electrospray ionization source or low recoveries on the SPE sorbent. Total organic carbon (TOC) is a good indicator of humic content of the sample.

- 4.4** SPE cartridges can be a source of interferences. The analysis of field and laboratory reagent blanks can provide important information regarding the presence or absence of such interferences. Brands and lots of SPE devices should be tested to ensure that contamination does not preclude analyte identification and quantitation.

5. Health and Safety

- 5.1** The toxicity or carcinogenicity of each reagent and standard used in this method is not fully established; however, each chemical compound should be treated as a potential health hazard. From this viewpoint, exposure to these chemicals must be reduced to the lowest possible level by whatever means available. A reference file of material safety data sheets is available to all personnel involved in the chemical analysis. Additional references to laboratory safety are available in the Chemical Hygiene Plan.
- 5.2** All personnel handling environmental samples known to contain or to have been in contact with municipal waste must follow safety practices for handling known disease causative agents.
- 5.3** PFOA has been described as “likely to be carcinogenic to humans.” Pure standard materials and stock standard solutions of these method analytes should be handled with suitable protection to skin and eyes, and care should be taken not to breathe the vapors or ingest the materials.

6. Sample Collection, Preservation, Shipping and Handling

6.1 Sample Collection for Aqueous Samples

- 6.1.1** Samples must be collected in two (2) 500-mL or 250-mL high density polyethylene (HDPE) container with an unlined plastic screw cap. All sample containers must have linerless HDPE or polypropylene caps.
- 6.1.2** The sample handler must wash their hands before sampling and wear nitrile gloves while filling and sealing the sample bottles. PFAS contamination during sampling can occur from a number of common sources, such as food packaging and certain foods and beverages. Proper hand washing and wearing nitrile gloves will aid in minimizing this type of accidental contamination of the samples.
- 6.1.3** Open the tap and allow the system to flush until the water temperature has stabilized (approximately 3 to 5 min). Collect samples from the flowing system.
- 6.1.4** Fill sample bottles. Samples do not need to be collected headspace free.
- 6.1.5** After collecting the sample and cap the bottle. Keep the sample sealed from time of collection until extraction.
- 6.1.6** Maintain all aqueous samples protected from light at 0 - 6 °C from the time of collection until shipped to the laboratory. Samples must be shipped as soon as practical with sufficient ice to maintain the sample temperature below 6 °C during transport and be received by the laboratory within 48 hours of collection. The laboratory must confirm that the sample temperature is 0 - 6 °C upon receipt. Once received by the laboratory, the samples must be stored at ≤ -20 °C until sample preparation.

6.2 Sample Collection for Solid and Oil samples.

- 6.2.1 Grab samples are collected in polypropylene containers. Sample containers and contact surfaces containing PTFE shall be avoided. Samples should fill no more than $\frac{3}{4}$ full.
- 6.2.2 Maintain solid samples protected from light (in HDPE containers) at 0 - 6 °C from the time of collection until receipt at the laboratory. The laboratory must confirm that the sample temperature is 0 - 6 °C upon receipt. Once received by the laboratory, the samples must be stored at ≤ -20 °C until sample preparation.

6.3 Sample Collection for fish and other tissue samples

- 6.3.1 If the time of collection to the time of receipt at the laboratory is expected to exceed 24 hours, the tissue samples must be frozen upon collection and shipped to the laboratory on dry ice.
- 6.3.2 Once received by the laboratory, the samples must be maintained protected from light at ≤ -20 °C until prepared. Store unused samples in HDPE containers or wrapped in aluminum foil at ≤ -20 °C.
- 6.3.3 The nature of the tissues of interest may vary by project. Field sampling plans and protocols should explicitly state the samples to be collected and if any processing will be conducted in the field (e.g., filleting of whole fish or removal of organs). All field procedures must involve materials and equipment that have been shown to be free of PFAS.

6.4 Sample Preservation

Not applicable.

6.5 Sample Shipping

Samples must be chilled during shipment and must not exceed 0 – 6 °C during the first 48 hours after collection. Sample temperature must be confirmed to be at or below 0 – 6 °C when the samples are received at the laboratory. Samples stored in the lab must be held at or below 6 °C until extraction but should not be frozen.

NOTE: Samples that are significantly above 0 – 6 °C, at the time of collection, may need to be iced or refrigerated for a period of time, in order to chill them prior to shipping. This will allow them to be shipped with sufficient ice to meet the above requirements.

6.6 Sample Handling

- 6.6.1 Aqueous samples (including leachates) should be analyzed as soon as possible; however, samples may be held in the laboratory for up to 90 days from collection, when stored at ≤ -20 °C and protected from the light. When stored at 0 - 6 °C and protected from the light, aqueous samples may be held for up to 28 days, with the caveat that issues were observed with certain perfluorooctane sulfonamide ethanols and perfluorooctane sulfonamidoacetic acids after 7 days. These issues are more likely to elevate the observed concentrations of other PFAS compounds via the transformation of these precursors if they are present in the sample.

- 6.6.2** Solid samples (soils and sediments), Oil and tissue samples may be held for up to 90 days, if stored by the laboratory in the dark at either 0 - 6 °C or ≤ -20 °C, with the caveat that samples may need to be extracted as soon as possible if NFDHA is an important analyte.
- 6.6.3** Biosolids samples may be held for up to 90 days, if stored by the laboratory in the dark at 0 - 6 °C or at -20 °C. Because microbiological activity in biosolids samples at 0 - 6 °C may lead to production of gases which may cause the sample to be expelled from the container when it is opened, as well as producing noxious odors, EPA recommends that samples be frozen if they need to be stored for more than a few days before extraction. Store sample extracts in the dark at less than 0 - 4 °C until analyzed. If stored in the dark at less than 0 - 4 °C, sample extracts may be stored for up to 90 days, with the caveat that issues were observed for some ether sulfonates after 28 days. These issues may elevate the observed concentrations of the ether sulfonates in the extract over time. Samples may need to be extracted as soon as possible if NFDHA is an important analyte.

7. Equipment and Supplies

- 7.1** SAMPLE CONTAINERS – 500-mL or 250-mL high density polyethylene (HDPE) bottles fitted with unlined screw caps. Sample bottles must be discarded after use.
- 7.2** SAMPLE JARS – 8-ounce wide mouth high density polyethylene (HDPE) bottles fitted with unlined screw caps. Sample bottles must be discarded after use.
- 7.3** POLYPROPYLENE BOTTLES – 4-mL narrow-mouth polypropylene bottles.
- 7.4** CENTRIFUGE TUBES – 50-mL conical polypropylene tubes with polypropylene screw caps for storing standard solutions and for collection of the extracts.
- 7.5** AUTOSAMPLER VIALS – Polypropylene 0.7-mL autosampler vials with polypropylene caps.
- 7.5.1** NOTE: Polypropylene vials and caps are necessary to prevent contamination of the sample from PTFE coated septa. However, polypropylene caps do not reseal, so evaporation occurs after injection. Thus, multiple injections from the same vial are not possible.
- 7.6** POLYPROPYLENE GRADUATED CYLINDERS – Suggested sizes include 25, 50, 100 and 1000-mL cylinders.
- 7.7** Auto Pipets – Suggested sizes include 5, 10, 25, 50, 100, 250, 500, 1000, 5000 and 10,000-µls.
- 7.8** PLASTIC PIPETS – Polypropylene or polyethylene disposable pipets.
- 7.9** Silanized glass wool (Sigma-Aldrich, Cat # 20411 or equivalent) – store in a clean glass jar and rinsed with methanol (2 times) prior to use.
- 7.10** Disposable syringe filter, 25-mm, 0.2-µm Nylon membrane, PALL/Acrodisc or equivalent
- 7.11** Variable volume pipettes with disposable HDPE or polypropylene tips (10 µL to 5 mL) used for preparation of calibration standards and spiked samples.
- 7.12** ANALYTICAL BALANCE – Capable of weighing to the nearest 0.0001 g.
- 7.13** ANALYTICAL BALANCE – Capable of weighing to the nearest 0.1 g.

7.14 SOLID PHASE EXTRACTION (SPE) APPARATUS FOR USING CARTRIDGES

7.14.1 SPE CARTRIDGES – (Phenomenex WAX 150 or 250mg or equivalent). The SPE sorbent must have a pKa above 8 so that it remains positively charged during the extraction.

7.14.1.1 Note: SPE cartridges with different bed volume (e.g., 500 mg) may be used; however, the laboratory must demonstrate that the bed volume does not negatively affect analyte absorption and elution, by performing the initial demonstration of capability analyses described in Section 13.

7.14.2 VACUUM EXTRACTION MANIFOLD – A manual vacuum manifold with large volume sampler for cartridge extractions, or an automatic/robotic sample preparation system designed for use with SPE cartridges, may be used if all QC requirements discussed in Section 9 are met. Extraction and/or elution steps may not be changed or omitted to accommodate the use of an automated system. Care must be taken with automated SPE systems to ensure the PTFE commonly used in these systems does not contribute to unacceptable analyte concentrations in the MB.

7.14.3 SAMPLE DELIVERY SYSTEM – Use of a polypropylene transfer tube system, which transfers the sample directly from the sample container to the SPE cartridge, is recommended, but not mandatory. Standard extraction manifolds come equipped with PTFE transfer tube systems. These can be replaced with 1/8" O.D. x 1/16" I.D. polypropylene or polyethylene tubing cut to an appropriate length to ensure no sample contamination from the sample transfer lines. Other types of non-PTFE tubing may be used provided it meets the MB and LCS QC requirements.

7.15 EXTRACT CONCENTRATION SYSTEM – Extracts are concentrated by evaporation with nitrogen using a water bath set no higher than 55 °C.

7.16 LABORATORY OR ASPIRATOR VACUUM SYSTEM – Sufficient capacity to maintain a vacuum of approximately 10 to 15 inches of mercury for extraction cartridges.

7.17 LIQUID CHROMATOGRAPHY (LC)/TANDEM MASS SPECTROMETER (MS/MS) WITH DATA SYSTEM

7.17.1 LC SYSTEM – Instrument capable of reproducibly injecting up to 10- μ L aliquots and performing binary linear gradients at a constant flow rate near the flow rate used for development of this method (0.4 mL/min). The LC must be capable of pumping the water/methanol mobile phase without the use of a degasser which pulls vacuum on the mobile phase bottle (other types of degassers are acceptable). Degassers which pull vacuum on the mobile phase bottle will volatilize the ammonium acetate mobile phase causing the analyte peaks to shift to earlier retention times over the course of the analysis batch. The usage of a column heater is optional.

7.17.2 LC/TANDEM MASS SPECTROMETER – The LC/MS/MS must be capable of negative ion electrospray ionization (ESI) near the suggested LC flow rate of 0.4 mL/min. The system must be capable of performing MS/MS to produce unique product ions for the method analytes within specified retention time segments. A minimum of 10 scans across the chromatographic peak is required to ensure adequate precision.

7.17.3 DATA SYSTEM – An interfaced data system is required to acquire, store, reduce, and output mass spectral data. The computer software should have the

capability of processing stored LC/MS/MS data by recognizing an LC peak within any given retention time window. The software must allow integration of the ion abundance of any specific ion within specified time or scan number limits. The software must be able to calculate relative response factors, construct linear regressions or quadratic calibration curves, and calculate analyte concentrations.

7.17.4 INSTRUMENT COLUMNS

7.17.4.1 ANALYTICAL: C18 column, 1.7 μm , 50 x 2.1 mm (Waters Acquity UPLC® BEH or equivalent)

7.17.4.2 OPTIONAL GUARD COLUMN: (Phenomenex Kinetex® Evo C18 or equivalent)

8. Reagents and Standards

8.1 GASES, REAGENTS, AND SOLVENTS – Reagent grade or better chemicals must be used.

8.1.1 REAGENT WATER – Purified water which does not contain any measurable quantities of any method analytes or interfering compounds greater than 1/2 the RL for each method analyte of interest. Prior to daily use, at least 3 L of reagent water should be flushed from the purification system to rinse out any build-up of analytes in the system's tubing.

8.1.2 METHANOL (CH_3OH , CAS#: 67-56-1) – High purity, demonstrated to be free of analytes and interferences.

8.1.3 AMMONIUM ACETATE ($\text{NH}_4\text{C}_2\text{H}_3\text{O}_2$, CAS#: 631-61-8) – High purity, demonstrated to be free of analytes and interferences. Store at 2-8° and replace 2 years after opening date.

8.1.4 ACETIC ACID (H_3CCOOH , CAS#: 64-19-7) - High purity, demonstrated to be free of analytes and interferences and stored at room temperature.

8.1.4.1 Acetic Acid (0.1%) – Dissolve acetic acid (1 mL) in reagent water (1 L), store at room temperature, replace after 3 months.

8.1.5 1M AMMONIUM ACETATE/REAGENT WATER – High purity, demonstrated to be free of analytes and interferences.

8.1.6 2mM AMMONIUM ACETATE/METHANOL:WATER (5:95) – To prepare, mix 2 ml of 1M AMMONIUM ACETATE, 1 ml ACETIC ACID and 50 ml METHANOL into 1 Liter of REAGENT WATER.

8.1.7 ACETONITRILE – UPLC grade or equivalent, store at room temperature

8.1.8 TOLUENE – HPLC grade or equivalent.

8.1.9 ACETONE – pesticide grade or equivalent

8.1.10 AMMONIUM HYDROXIDE (NH_3 , CAS#: 1336-21-6) – High purity, demonstrated to be free of analytes and interferences, and stored at room temperature.

- 8.1.11 AQUEOUS AMMONIUM HYDROXIDE (3%) – Add ammonium hydroxide (10 mL, 30%) to reagent water (90 mL), store at room temperature, replace after 3 months.
- 8.1.12 METHANOLIC AMMONIUM HYDROXIDE (0.3%) - add ammonium hydroxide (1 mL, 30%) to methanol (99 mL), store at room temperature, replace after 1 month
- 8.1.13 METHANOLIC AMMONIUM HYDROXIDE (1%) - add ammonium hydroxide (3.3 mL, 30%) to methanol (97 mL), store at room temperature, replace after 1 month
- 8.1.14 METHANOLIC AMMONIUM HYDROXIDE (2%) - add ammonium hydroxide (6.6 mL, 30%) to methanol (93.4 mL), store at room temperature, replace after 1 month
- 8.1.15 METHANOLIC POTASSIUM HYDROXIDE (0.05 M) – add 3.3 g of potassium hydroxide to 1 L of methanol, store at room temperature, replace after 3 months
- 8.1.16 METHANOL WITH 4% WATER, 1% AMMONIUM HYDROXIDE AND 0.625% ACETIC ACID - add ammonium hydroxide (3.3 mL, 30%), reagent water (1.7 mL) and acetic acid (0.625 mL) to methanol (92 mL), store at room temperature, replace after 1 month. This solution is used to prepare the instrument blank and calibration standards (Section 8.3.2).
- 8.1.17 FORMIC ACID – (greater than 96% purity or equivalent). Store at room temperature and replace after 2 years.
- 8.1.18 FORMIC ACID (aqueous, 0.1 M) - dissolve formic acid (4.6 g) in reagent water (1 L), store at room temperature, replace after 2 years.
- 8.1.19 FORMIC ACID (aqueous, 0.3 M) - dissolve formic acid (13.8 g) in reagent water (1 L), store at room temperature, replace after 2 years.
- 8.1.20 FORMIC ACID (aqueous, 5% v/v) - mix 5 mL formic acid with 95 mL reagent water, store at room temperature, replace after 2 years.
- 8.1.21 FORMIC ACID (methanolic 1:1, 0.1 M formic acid/methanol) - mix equal volumes of methanol and 0.1 M formic acid, store at room temperature, replace after 2 years.
- 8.1.22 FORMIC ACID (aqueous, 50% v/v) - mix 50 mL formic acid with 50 mL reagent water, store at room temperature, replace after 2 years.
- 8.1.23 POTASSIUM HYDROXIDE – certified ACS or equivalent, store at room temperature, replace after 2 years.
- 8.1.24 CARBON - – EnviCarb® 1-M-USP or equivalent, verified by lot number before use, stored at room temperature. Loose carbon allows for better adsorption of interferent organics. Note: The single-laboratory validation laboratory achieved better performance with loose carbon than carbon cartridges. Loose carbon will be used for the multi-laboratory validation to set statistically based method criteria.

- 8.1.25** NITROGEN – Used for the following purposes: Nitrogen aids in aerosol generation of the ESI liquid spray and is used as collision gas in some MS/MS instruments. The nitrogen used should meet or exceed instrument manufacturer's specifications. In addition, Nitrogen is used to concentrate sample extracts (Ultra High Purity or equivalent).
- 8.1.26** ARGON – Used as collision gas in some MS/MS instruments. Argon should meet or exceed instrument manufacturer's specifications. Nitrogen gas may be used as the collision gas provided sufficient sensitivity (product ion formation) is achieved.
- 8.2** REFERENCE MATRICES - Matrices in which PFAS and interfering compounds are not detected by this method. These matrices are to be used to prepare the batch QC samples, LOQ/MDL, and IDOC samples.
- 8.2.1** Reagent water - purified water, Type I
- 8.2.2** Solid reference matrix Ottawa Sand or equivalent
- 8.2.3** Tissue Reference matrix – Cod loin or other animal tissue demonstrated to be PFAS free.
- 8.3** STANDARD SOLUTIONS – When a compound purity is assayed to be 96% or greater, the weight can be used without correction to calculate the concentration of the stock standard. PFAS analyte and IS standards commercially purchased in glass ampoules are acceptable; however, all subsequent transfers or dilutions performed by the analyst must be prepared and stored in polypropylene containers and are stored at ≤ 4 °C. Standards for sample fortification generally should be prepared in the smallest volume that can be accurately measured to minimize the addition of excess organic solvent to aqueous samples.
- 8.3.1** Stock standards and diluted stock standards are stored at ≤ 4 °C. Prepare a spiking solution, containing the method analytes listed in Table 1, in methanol from prime stocks. The solution is used to prepare the calibration standards and to spike the known reference QC samples that are analyzed with every batch. Quantitative standards containing a mixture of branched and linear isomers must be used for method analytes if they are commercially available. Currently, these include PFOS, PFHxS, NEtFOSAA, and NMeFOSAA.
- 8.3.2** Calibration standard solutions – A series of calibration solutions containing the target analytes and the Labeled extracted internal standards (EIS) and non-extracted internal standards (NIS) is used to establish the initial calibration of the analytical instrument. Table 4 represents the concentrations of the native, EIS and NIS analytes of the calibration curve. Calibration standard solutions are made using the solution described in section 8.1.16.
- 8.3.3** ISOTOPE DILUTION EXTRACTED INTERNAL STANDARD (EIS) – Isotopically labelled analogs of the target analytes to be used for the quantification of target analytes. EIS stock standard solutions are purchased in glass ampoules and are stored in accordance with the manufacturer's recommendations. The EIS stock solution to be used for the fortification of samples and QC in accordance with the isotope dilution procedure. Table 2 represents the EIS concentrations and nominal sample amounts added to each field sample and QC element.

- 8.3.4 ISOTOPE DILUTION NON-EXTRACTED INTERNAL STANDARDS (NIS) – Isotopically labelled analogs to be added post extraction for the measurement of EIS extraction efficiency and is added to the final volume of all extractions. Table 3 represents the EIS concentrations and nominal sample amounts added to each field sample and QC element.

9. Quality Control

9.1 Method Blank

- 9.1.1 A Method Blank (MB) is required with each extraction batch to confirm that potential background contaminants are not interfering with the identification or quantitation of method analytes. An aliquot of reagent water that is treated exactly as a sample including exposure to all glassware, equipment, solvents, reagents and standards. Prep and analyze a MB for every 20 samples. If the MB produces a peak within the retention time window of any analyte that would prevent the determination of that analyte, determine the source of contamination, and eliminate the interference before processing samples. Background contamination must be reduced to an acceptable level before proceeding. Background from method analytes or other contaminants that interfere with the measurement of method analytes must be below the RL. If the method analytes are detected in the MB at concentrations equal to or greater than this level, then all data for the problem analyte(s) must be considered invalid for all samples in the extraction batch.

9.2 Laboratory Control Sample (LCS)

- 9.2.1 Low Level LCS or OPR (Ongoing Precision Recovery) sample is required with each extraction batch. A LLCS or OPR samples is a method blank spiked with known quantities of analytes. The fortified concentration of the LCS is spiked at 2X the LOQ. Default limits of 70-130% of the true value may be used for analytes until sufficient replicates have been analyzed to generate proper control limits. Calculate the percent recovery (%R) for each analyte using the equation:
- 9.2.2 An LCS or OPR (Ongoing Precision Recovery) sample is required with each extraction batch. A LCS or OPR samples is a method blank spiked with known quantities of analytes. The fortified concentration of the LCS is spiked at the midpoint of the calibration curve. Default limits of 70-130% of the true value may be used for analytes until sufficient replicates have been analyzed to generate proper control limits. Calculate the percent recovery (%R) for each analyte using the equation:

$$\%R = \frac{A \times 100}{B}$$

Where:

A = measured concentration in the fortified sample

B = fortification concentration.

- 9.1.1 Where applicable, in the absence of additional sample volume required to perform matrix specific QC, LCSD's are to be extracted and analyzed. The concentration and analyte

recovery criteria for the LCSD must be the same as the batch LCS. The RSD's must fall within $\leq 30\%$ of the true value for medium and high-level replicates, and $\leq 50\%$ for low level replicates. Calculate the relative percent difference (RPD) for duplicate MSs (MS and MSD) using the equation:

$$RPD = \frac{|LCS - LCSD|}{(LCS + LCSD) / 2} \times 100$$

- 9.1.2 If the LCS and or LCSD results do not meet these criteria for method analytes, then all data for the problem analyte(s) must be considered invalid for all samples in the extraction batch.

9.3 Non-extracted Internal Standard Area (NIS)

Each time an initial calibration is performed, use the data from all the initial calibration standards used to meet the linearity test in Section 10.3.3.3 to calculate the mean area response for each of the NIS compounds, using the equation below.

$$\text{Mean Area}_{\text{NIS}_i} = \sum \text{Area}_{\text{NIS}_i} / n$$

where:

Area_{NIS_i} = Area counts for the *i*th NIS, where *i* ranges from 1 to 7, for the seven NIS compounds listed in Table 1

n = The number of ICAL standards (the default value is *n* = 6). If a different number of standards is used for the ICAL, for example, to increase the calibration range or by dropping a point at either end of the range to meet the linearity criterion, change 6 to match the actual number of standards used)

Record the mean areas for each NIS for use in evaluating results for sample analyses. There is no acceptance criterion associated with the mean NIS area data.

9.4 Extracted Internal Standards (EIS)

- 9.4.1 The EIS standard is fortified into all samples, CCVs, MBs, LCSs, MSs, MSDs, FD, and FRB prior to extraction. It is also added to the CAL standards. The EIS is a means of assessing method performance from extraction to final chromatographic measurement. Calculate the recovery (%R) for the EIS using the following equation:

$$\%R = (A / B) \times 100$$

Where:

A = calculated EIS concentration for the QC or Field Sample

B = fortified concentration of the EIS.

- 9.4.2 Default limits of 50-150% may be used for analytes until sufficient replicates have been analyzed to generate proper control limits. A low or high percent recovery for a sample, blank, or CCV does not require discarding the analytical data but it may indicate a potential problem with future analytical data. When EIS recovery from a sample, blank, or CCV are outside control limits, check 1) calculations to locate

possible errors, 2) standard solutions for degradation, 3) contamination, and 4) instrument performance. For CCVs and QC elements spiked with all target analytes, if the recovery of the corresponding target analytes meet the acceptance criteria for the EIS in question, the data can be used but all potential biases in the recovery of the EIS must be documented in the sample report. If the associated target analytes do not meet the acceptance criteria, the data must be reanalyzed.

9.5 Matrix Spike (MS/MSD)

- 9.5.1 Analysis of an MS is prepared one per preparation batch (if required).
- 9.5.2 Aliquots of field samples that have been fortified with a known concentration of target compounds, prior to sample preparation and extraction, and analyzed to measure the effect of matrix interferences. The use of MS/MSD samples is generally not required in isotope dilution methods because the labeled compounds added to every sample provide more performance data than spiking a single sample in each preparation batch. Aliquots of field samples
- 9.5.3 Analyte recoveries may exhibit matrix bias. For samples fortified at or above their native concentration, recoveries should range between 50-150%. If the accuracy of any analyte falls outside the designated range, and the laboratory performance for that analyte is shown to be in control in the LCS, the recovery is judged to be matrix biased. The result for that analyte in the unfortified sample is labeled suspect/matrix to inform the data user that the results are suspect due to matrix effects.

9.6 Laboratory Duplicate

- 9.6.1 FIELD DUPLICATE OR LABORATORY FORTIFIED SAMPLE MATRIX DUPLICATE (FD or MSD) – Within each extraction batch (not to exceed 20 Field Samples), a minimum of one FD or MSD must be analyzed. Duplicates check the precision associated with sample collection, preservation, storage, and laboratory procedures. If method analytes are not routinely observed in Field Samples, an MSD should be analyzed rather than an FD.
- 9.6.2 Calculate the relative percent difference (RPD) for duplicate measurements (FD1 and FD2) using the equation:

$$RPD = \frac{|FD1 - FD2|}{(FD1 + FD2) / 2} \times 100$$

- 9.6.3 RPDs for FDs should be ≤30%. Greater variability may be observed when FDs have analyte concentrations that are within a factor of 2 of the RL. At these concentrations, FDs should have RPDs that are ≤50%. If the RPD of any analyte falls outside the designated range, and the laboratory performance for that analyte is shown to be in control in the CCV, the recovery is judged to be matrix biased. The result for that analyte in the unfortified sample is labeled suspect/matrix to inform the data user that the results are suspect due to matrix effects.
- 9.6.4 If an MSD is analyzed instead of a FD, calculate the relative percent difference (RPD) for duplicate MSs (MS and MSD) using the equation:

$$RPD = \frac{|MS - MSD|}{(MS + MSD) / 2} \times 100$$

9.6.5 RPDs for duplicate MSs should be $\leq 30\%$ for samples fortified at or above their native concentration. Greater variability may be observed when MSs are fortified at analyte concentrations that are within a factor of 2 of the RL. MSs fortified at these concentrations should have RPDs that are $\leq 50\%$ for samples fortified at or above their native concentration. If the RPD of any analyte falls outside the designated range, and the laboratory performance for that analyte is shown to be in control in the LCSD where applicable, the result is judged to be matrix biased. If no LCSD is present, the associated MS and MSD are to be re-analyzed to determine if any analytical has occurred. If the resulting RPDs are still outside control limits, the result for that analyte in the unfortified sample is labeled suspect/matrix to inform the data user that the results are suspect due to matrix effects.

9.7 Bile Salt Interference Check

9.7.1 The laboratory must analyze a TDCA standard after the initial calibration, prior to the analysis of tissue samples, to check for interferences caused by bile salts. If an interference is present, the chromatographic conditions must be modified to eliminate the interference from TDCA (e.g., changing the retention time of TDCA such that it falls outside the

9.8 Initial Calibration Verification (ICV)

9.8.1 After each ICAL, analyze a QCS sample from a source different from the source of the CAL standards. If a second vendor is not available, then a different lot of the standard should be used. The QCS should be prepared and analyzed just like a CCV. Acceptance criteria for the QCS are identical to the CCVs; the calculated amount for each analyte must be $\pm 30\%$ of the expected value. If measured analyte concentrations are not of acceptable accuracy, check the entire analytical procedure to locate and correct the problem.

9.9 Instrument Sensitivity Check (ISC)

9.9.1 At the start of each 12-hour shift, analyze a standard at the LOQ. The signal-to-noise ratio of the ISC standard must be greater than or equal to 3:1. If the requirements cannot be met, the problem must be corrected before analyses can proceed.

9.10 Continuing Calibration Verification (CCV)

9.10.1 CCV Standards must be analyzed at the beginning of each analysis batch, after every 10 Field Samples, and at the end of the analysis batch.

9.10.2 The recovery of native and isotopically labeled compounds for the CVs must be within 70 - 130%

9.11 Method-specific Quality Control Samples

9.11.1 Instrument Blank – During the analysis of a batch of samples, a solvent blank is analyzed after samples containing high level of target compounds (e.g., calibration, CV) to monitor carryover from the previous injection. The injection blank consists of the solution in Section 8.1.16 fortified with the EIS and NIS for quantitation purposes.

9.12 Example Method Sequence

- INSTRUMENT BLANK
- INSTRUMENT SENSITIVITY CHECK
- CALIBRATION VERIFICATION STANDARD
- QUALITATIVE IDENTIFICATION STANDARDS
- TDCA STANDARD (only if analyzing tissues)
- INSTRUMENT BLANK
- METHOD BLANK
- LOW-LEVEL LCS/OPR
- OPR/LCS
- SAMPLE (10 or fewer)
- CALIBRATION VERIFICATION STANDARD
- INSTRUMENT BLANK
- SAMPLE (10 or fewer)
- CALIBRATION VERIFICATION STANDARD
- INSTRUMENT BLANK

10. Procedure

10.1 Equipment Set-up

- 10.1.1** This procedure may be performed manually or in an automated mode using a robotic or automatic sample preparation device. If an automated system is used to prepare samples, follow the manufacturer's operating instructions, but all extraction and elution steps must be the same as in the manual procedure. Extraction and/or elution steps may not be changed or omitted to accommodate the use of an automated system. If an automated system is used, the MBs should be rotated among the ports to ensure that all the valves and tubing meet the MB requirements.
- 10.1.2** Some of the PFAS's adsorb to surfaces, including polypropylene. Therefore, the aqueous sample bottles must be rinsed with the elution solvent whether extractions are performed manually or by automation. The bottle rinse is passed through the cartridge to elute the method analytes and is then collected.
- 10.1.3** The SPE cartridges and sample bottles described in this section are designed as single use items and should be discarded after use. They may not be refurbished for reuse in subsequent analyses.
- 10.1.4** All SPE apparatus, including manifolds, tubing and sample ports must be thoroughly rinsed following each use with 1% methanolic ammonium hydroxide, followed by Methanol and then DI water. Additionally, sample manifold ports and transfer tubing should be inspected regularly for signs of wear and/or

discoloration. When such observations are made, the associated components should be replaced.

10.1.5 Prior to the start of any extraction, sample site information must be evaluated for any potentially high level PFAS concentrations or sample matrix irregularities that may impact the extraction process. If such samples are identified, aqueous samples may be pre-screened via direct aqueous injection prior to analysis to estimate the potential PFAS concentrations present.

10.1.6 To perform a direct aqueous injection (DAI) screen, the sample should be inverted several times to try and evenly disperse any organic matter present. A 1 ml aliquot (or less depending on the matrix) is to be taken from the parent sample, volume adjusted to 1 ml with reagent water if less than 1ml, fortified with EIS and NIS spiking solutions to match the concentrations of an extracted sample (typically 5 µl per 1 ml DAI), and then analyzed under the same analytical conditions as field samples.

10.2 Sample Preparation of Aqueous Samples

10.2.1 Samples are preserved, collected, and stored as presented in Section 6.

10.2.2 Determine sample volume. Weigh all samples to the nearest 1g. If visible sediment is present, centrifuge and decant into a new HDPE bottle and record the weight of the new container.

NOTE: Some of the PFAS's adsorb to surfaces, thus the sample volume may not be transferred to a graduated cylinder for volume measurement.

10.2.3 The MB, LCS and FRB may be prepared by measuring reagent water with a polypropylene graduated cylinder or filling an HDPE sample bottle near the top.

10.2.4 Check that the pH is 6.5 ± 0.5 . If necessary, adjust pH with 50% formic acid or ammonium hydroxide and 3% aqueous ammonium hydroxide. The extract is now ready for solid-phase extraction (SPE) and cleanup.

10.2.5 Add 20 µL of the EIS to each sample and QC, cap and invert to mix.

10.2.6 If the sample is an LCS, LCSD, MS, or MSD, add the necessary amount of analyte PDS. Cap and invert each sample to mix.

10.3 Sample Prep and Extraction Protocol for Solids.

10.3.1 Homogenize and weigh 5 grams of sample (measured to the nearest hundredth of a gram) into a 50 ml polypropylene centrifuge tube. For laboratory control blanks and spikes, 5 grams of clean sand is used.

10.3.1.1 For Biosolids and other complex matrices, a small aliquot may be required due to co-extracted matrix interferences.

10.3.1.2 For batch QC samples using 5 g of reference solid, add 2.5 g of reagent water. The addition of reagent water to the sand provides a matrix closer in composition to real-world samples.

10.3.2 Add 20 µL of the EIS to each sample and QC.

10.3.3 If the sample is an LCS, LCSD, MS, or MSD, add the necessary amount of analyte PDS. Cap and invert each sample to mix.

- 10.3.4** Vortex the samples to evenly disperse the spiking solutions and allow to equilibrate for 30 minutes.
- 10.3.5** To all samples, add 10 ml of 0.3% methanolic ammonium hydroxide, cap, vortex for 25 seconds.
- 10.3.6** Following mixing, shake each sample for 30 minutes on a shaker table.
- 10.3.7** Centrifuge each sample at 2800RPM for 10 minutes.
- 10.3.8** Remove the supernatant and transfer to a clean 50 ml polypropylene centrifuge tube.
- 10.3.9** Repeat steps 10.3.4 to 10.3.7, with 15 ml of 0.3% methanolic ammonium hydroxide, combining the supernatants.
- 10.3.10** Add 5ml of 0.3% methanolic ammonium hydroxide to the sample, vortex for 25 seconds and centrifuge each sample at 2800RPM for 10 minutes.
- 10.3.11** Remove the supernatant and transfer to the same 50 ml polypropylene centrifuge tube containing eluates from the previous cycles.
- 10.3.12** Add 10 mg of carbon to the combined extract, mix by occasional hand shaking for no more than five minutes and then centrifuge at 2800 rpm for 10 minutes. Immediately decant the extract into a 50 ml polypropylene centrifuge tube.
- 10.3.13** Dilute to approximately 35 mL with reagent water. Samples containing more than 50% water may yield extracts that are greater than 35 mL in volume; therefore, do not add water to these. Determine the water content in the sample as follows (percent moisture is determined from the % solids):
- $$\text{Water Content in Sample} = (\text{Sample Weight} * \text{Percent moisture}) / 100$$
- 10.3.14** Concentrate each extract at approximately 55 °C with a gentle N2 flow to a final volume that is based on the water content of the sample (see table below). Allow extracts to concentrate for 10 minutes, then mix (by vortex if the volume is < 20. Continue concentrating and mixing every 5 minutes until the extract has been reduced to the required volume as specified in the table below. If the extract volume appears to stop dropping, the concentration must be stopped and the volume at which it was stopped recorded.

Water Content in Sample	Concentrated Final Volume
< 5 grams	15 ml
5-8 grams	15-20 ml
8-9 grams	20-22.5 ml
9-10 grams	22.5-25 ml

- 10.3.15** Add 40 - 50 mL of reagent water to the extract and vortex. Check that the pH is 6.5 ±0.5 and adjust as necessary with 50% formic acid or 30% ammonium hydroxide, or with 5% formic acid and 3% aqueous ammonium hydroxide. The extracts are ready for SPE and cleanup.

10.4 Sample Prep and Extraction Protocol for Oils.

- 10.4.1 Weigh 1-2 grams of sample (measured to the nearest hundredth of a gram) into a 50 ml polypropylene centrifuge tube. For laboratory control blanks and spikes, 1 grams of mineral oil is used.
 - 10.4.1.1 For batch QC samples use 1 g of reference oil.
- 10.4.2 Add 20 µL of the EIS to each sample and QC.
- 10.4.3 If the sample is an LCS, LCSD, MS, or MSD, add the necessary amount of analyte PDS. Cap and invert each sample to mix.
- 10.4.4 Vortex the samples to evenly disperse the spiking solutions and allow to equilibrate for 30 minutes.
- 10.4.5 To all samples, add 10 ml of 0.3% methanolic ammonium hydroxide, cap, vortex for 25 seconds.
- 10.4.6 Following mixing, shake each sample for 30 minutes on a shaker table.
- 10.4.7 Centrifuge each sample at 2800RPM for 10 minutes.
- 10.4.8 Remove the supernatant and transfer to a clean 50 ml polypropylene centrifuge tube.
- 10.4.9 Repeat steps 10.3.4 to 10.3.7, with 15 ml of 0.3% methanolic ammonium hydroxide, combining the supernatants.
- 10.4.10 Add 5ml of 0.3% methanolic ammonium hydroxide to the sample, vortex for 25 seconds and centrifuge each sample at 2800RPM for 10 minutes.
- 10.4.11 Remove the supernatant and transfer to the same 50 ml polypropylene centrifuge tube containing eluates from the previous cycles.
- 10.4.12 Add 10 mg of carbon to the combined extract, mix by occasional hand shaking for no more than five minutes and then centrifuge at 2800 rpm for 10 minutes. Immediately decant the extract into a 50 ml polypropylene centrifuge tube.

10.5 Sample Prep and Extraction Protocol for Tissues.

- 10.5.1 Homogenize and weigh 2 grams of sample (measured to the nearest hundredth of a gram) into a 50 ml polypropylene centrifuge tube. For laboratory control blanks and spikes, 2 grams of clean tissue is used.
- 10.5.2 Add 20 µL of the EIS PDS to each sample and QC.
- 10.5.3 If the sample is an LCS, LCSD, MS, or MSD, add the necessary amount of analyte PDS. Cap and invert each sample to mix.
- 10.5.4 Add 10 mL of 0.05M KOH in methanol to each sample. Vortex to disperse the tissue then place tubes on a mixing table to extract for at 16 hours. Centrifuge at 2800 rpm for 10 minutes and collect the supernatant in a 50-mL polypropylene centrifuge tube.
- 10.5.5 Add 10 mL of acetonitrile to remaining tissue in the 50-mL centrifuge tube, vortex to mix and disperse the tissue. Sonicate for 30 minutes. Centrifuge at 2800 rpm for 10 minutes and collect the supernatant, adding it to the 50-mL centrifuge tube containing the initial extract.
- 10.5.6 Add 5 mL of 0.05M KOH in methanol to the remaining sample in each centrifuge tube. Vortex to disperse the tissue and hand mix briefly. Centrifuge at 2800 rpm

for 10 minutes and collect the supernatant, adding it to the 50-mL centrifuge tube containing the first two extracts.

- 10.5.7 Add 10 mg of carbon to the combined extract, mix by occasional hand shaking over a period of no more than five minutes and then centrifuge at 2800 rpm for 10 minutes. Immediately decant the extract into a 50-mL centrifuge tube.
- 10.5.8 Add 1 mL of reagent water to each tube and concentrate each extract at approximately 55 °C with a gentle N₂ flow to a final volume of 2.5 mL.
- 10.5.9 Add reagent water to each evaporation/concentrator tube to dilute the extracts to 50 mL. Check that the pH = 6.5 ± 0.5 and adjust as needed with 50% formic acid, or ammonium hydroxide or with 5% formic acid and 3% aqueous ammonium hydroxide. The extracts are ready for SPE and cleanup.

10.6 SPE Extract: All matrices

- 10.6.1 Pack clean silanized glass wool to half the height of the WAX SPE cartridge barrel.
- 10.6.2 Pre-condition the cartridges by washing them with 3 X 5 mL of 1% methanolic ammonium hydroxide, discarding the wash volumes.
- 10.6.3 Rinse the cartridge with 5 mL of 0.3M formic acid, allowing the cartridge to drain using gravity only, discarding the rinse volume. Do not allow the cartridge to go dry
- 10.6.4 Adjust the vacuum so that the approximate flow rate is ~5 mL/min and load the sample across the cartridge. Do not allow the cartridge to go dry before all the sample has passed through.
- 10.6.5 Once all the sample has passed across the cartridge, rinse the walls of the reservoir with 2 X 5 mL reagent water, loading the rinse across the cartridge.
- 10.6.6 Rinse the walls of the reservoir with 5 mL of 1:1 0.1M formic acid/methanol and pass the rinse through the cartridge using vacuum. Dry the cartridge by pulling air through for 15 seconds.
- 10.6.7 Rinse the inside of the sample bottle with 5 mL of 1% methanolic ammonium hydroxide. Use vacuum to pull the elution solvent through the cartridge and into the collection tubes. When the cartridge bed and glass wool are submerged, stop the cartridge flow by closing the valve, keeping the sorbent bed and wool submerged.
- 10.6.8 Let the wetted sorbent bed and wool soak for 1 minute.
- 10.6.9 Open the cartridge valve and collect the eluate into a 15 mL polypropylene collection tube.
- 10.6.10 Add 25 µL of concentrated acetic acid to each sample eluted in the collection tubes and vortex to mix.
- 10.6.11 Add 10 mg of carbon to each sample and batch QC extract, using a 10-mg scoop. Handshake occasionally for no more than 5 minutes. It is important to minimize the time the sample extract is in contact with the carbon. Immediately vortex (30 seconds) and centrifuge at 2800 rpm for 10 minutes.
- 10.6.12 Add NIS solution to a clean collection tube. Place a syringe filter (25-mm filter, 0.2-µm nylon membrane) on a 5-mL polypropylene syringe. Take the plunger out and carefully decant the sample supernatant into the syringe barrel. Replace the

plunger and filter the entire extract into the new collection tube containing the NIS.

- 10.6.13** Vortex to mix and transfer a portion of the extract into a .7-mL polypropylene LC vial for LC-MS/MS analysis. Cap the collection tube containing the remaining extract and store at 4 °C

10.7 Sample Volume Determination

- 10.7.1** If using weight to determine volume, weigh the empty bottle to the nearest 1 g and determine the sample weight by subtraction of the empty bottle weight from the original sample weight. Assume a sample density of 1.0 g/mL. In either case, the sample volume will be used in the final calculations of the analyte concentration.

10.8 Initial Calibration - Demonstration and documentation of acceptable initial calibration is required before any samples are analyzed. After the initial calibration is successful, a CCV is required at the beginning and end of each period in which analyses are performed, and after every tenth Field Sample.

10.8.1 ESI-MS/MS TUNE

- 10.8.1.1** Calibrate the mass scale of the MS with the calibration compounds and procedures prescribed by the manufacturer.

- 10.8.1.2** Optimize the [M-H]⁻ or [M-CO₂]⁻ for each method analyte by infusing approximately 0.5-1.0 µg/mL of each analyte (prepared in the initial mobile phase conditions) directly into the MS at the chosen LC mobile phase flow rate (0.4 mL/min). This tune can be done on a mix of the method analytes. The MS parameters (voltages, temperatures, gas flows, etc.) are varied until optimal analyte responses are determined. The method analytes may have different optima requiring some compromise between the optima.

The Mass spec conditions found in Table 7 show the Sciex Triple Quad 5500+ operation conditions used in this method.

- 10.8.1.3** Optimize the product ion for each analyte by infusing approximately 0.5-1.0 µg/mL of each analyte (prepared in the initial mobile phase conditions) directly into the MS at the chosen LC mobile phase flow rate (approximately 0.4 mL/min). This tune can be done on a mix of the method analytes. The MS/MS parameters (collision gas pressure, collision energy, etc.) are varied until optimal analyte responses are determined. Typically, the carboxylic acids have very similar MS/MS conditions, and the sulfonic acids have similar MS/MS conditions.

The conditions found on table 5 are representative of expected tune optimizations for each analyte. If conditions other the ones close to the values provided in table 5 are achieved, the process should be re-performed and/or instrument maintenance performed to resolve the problem.

- 10.8.2** Establish LC operating parameters that optimize resolution and peak shape. Modifying the standard or extract composition to more aqueous content to prevent poor shape is not permitted.

Table 6 represents the operation conditions of a Sciex Exion LC system when running this method.

- 10.8.3** Inject 2 μ l of a mid-level CAL standard under LC/MS conditions to obtain the retention times of each method analyte. Divide the chromatogram into retention time windows each of which contains one or more chromatographic peaks. During MS/MS analysis, fragment a small number of selected precursor ions ([M-H]⁻) for the analytes in each window and choose the most abundant product ion. For maximum sensitivity, small mass windows of ± 0.5 daltons around the product ion mass were used for quantitation.
- 10.8.4** Inject a mid-level CAL standard under optimized LC/MS/MS conditions to ensure that each method analyte is observed in its MS/MS window and that there are at least 10 scans across the peak for optimum precision.

NOTE: PFHxS, PFOS, NMeFOSAA, and NEtFOSAA have multiple chromatographic peaks using the LC conditions in Table 7 due to chromatographic resolution of the linear and branched isomers of these compounds. Most PFAS's are produced by two different processes. One process gives rise to linear PFAS's only while the other process produces both linear and branched isomers. Thus, both branched and linear PFAS's can potentially be found in the environment. For the aforementioned compounds that give rise to more than one peak, all the chromatographic peaks observed in the standard must be integrated and the areas totaled. Chromatographic peaks in a sample must be integrated in the same way as the CAL standard.

- 10.8.5** Prepare a set of CAL standards as outlined in table 5. The lowest concentration CAL standard must be at or below the LOQ.
- 10.8.6** The LC/MS/MS system is calibrated using the isotope dilution technique. Target analytes are quantitated against their isotopically labeled analog (Extracted Internal Standard) where commercially available. If a labeled analog is not commercially available, the extracted internal standard with the closest retention time and /or closest chemical similarity is to be used. Use the LC/MS/MS data system software to generate a linear regression or quadratic calibration curve for each of the analytes. This curve must always be forced through zero and may be concentration weighted, if necessary. Forcing zero allows for a better estimate of the background levels of method analytes. A minimum of 6 calibration points are required for a linear or quadratic calibration model.
- 10.8.7 CALIBRATION ACCEPTANCE CRITERIA** – A linear fit is acceptable if the calculated RSD or RSE for each target analyte is $\leq 20\%$. If linear or Quadratic regressions are used, coefficient of determination (r^2) values must be greater than 0.99. When quantitated using the initial calibration curve, each calibration point at or above the LOQ for each analyte must calculate to be within 70-130% of its true value. The calculate value of each EIS analyte must be within 50-150% of its true value. If these criteria cannot be met, corrective action is taken to reanalyze the CAL standards, restrict the range of calibration.
- 10.8.8 Bile salts interference check** - The laboratory must analyze a TDCA standard after the initial calibration, prior to the analysis of tissue samples, to check for interferences caused by bile salts. If an interference is present, the chromatographic conditions must be modified to eliminate the interference from TDCA (e.g., changing the retention time of TDCA such that it falls outside the

retention window for PFOS by at least one minute), and the initial calibration repeated.

10.9 CONTINUING CALIBRATION CHECK (CCV) – Minimum daily calibration verification is as follows. Verify the initial calibration at the beginning and end of each group of analyses, and after every tenth sample during analyses. In this context, a “sample” is considered to be a Field Sample. MBs, CCVs, LCSs, MSs, FDs FRBs and MSDs are not counted as samples. The beginning CCV of each analysis batch must be at or below the RL in order to verify instrument sensitivity prior to any analyses. If standards have been prepared such that all low CAL points are not in the same CAL solution, it may be necessary to analyze two CAL standards to meet this requirement. Alternatively, the analyte concentrations in the analyte PDS may be customized to meet these criteria. Subsequent CCVs should alternate between a medium and Low concentration CAL standard.

10.9.1 Inject an aliquot of the appropriate concentration CAL standard and analyze with the same conditions used during the initial calibration.

10.9.2 Calculate the concentration of each analyte and EIS in the CCV. The calculated amount for each native and EIS analyte for medium level CCVs must be within $\pm 30\%$ of the true. If these conditions do not exist, then all data for the problem analyte must be considered invalid, and remedial action should be taken which may require recalibration. Any Field or QC Samples that have been analyzed since the last acceptable calibration verification should be reanalyzed after adequate calibration has been restored, with the following exception. If the CCV fails because the calculated concentration is greater than 130% for a particular method analyte, and Field Sample extracts show no detection for that method analyte, non-detects may be reported without re-analysis.

10.9.3 REMEDIAL ACTION – Failure to meet CCV QC performance criteria may require remedial action. Major maintenance, such as cleaning the electrospray probe, atmospheric pressure ionization source, cleaning the mass analyzer, replacing the LC column, etc., requires recalibration and verification of sensitivity by analyzing a CCV at or below the LOQ.

10.10 EXTRACT ANALYSIS

10.10.1 The same operating conditions used for the initial calibration and summarized in Tables 6 and 7 are to be used.

10.10.2 Prior to analysis of sample extracts, the Instrument mass calibration verification must be performed using standards whose mass range brackets the masses of interest and performed in the negative ion mode. The mass calibration is verified if the calculated mass is within $\pm .2$ daltons of the specified mass.

10.10.3 Establish an appropriate retention time window for each analyte. This should be based on measurements of actual retention time variation for each method analyte in CAL standard solutions analyzed on the LC over the course of time. A value of plus or minus three times the standard deviation of the retention time obtained for each method analyte while establishing the initial calibration can be used to calculate a suggested window size. However, the experience of the analyst should weigh heavily on the determination of the appropriate retention window size.

- 10.10.4** Calibrate the system by either the analysis of a calibration curve or by confirming the initial calibration is still valid by analyzing a CCV.
- 10.10.5** Begin analyzing Field Samples, including QC samples, at their appropriate frequency by injecting the same size aliquots under the same conditions used to analyze the CAL standards.
- 10.10.6** For concentrations at or above the method LOQ, the total (branched and linear isomer) quantification ion response to the total (branched and linear isomer) confirmation ion response ratio must fall within $\pm 50\%$ of the ratio observed in the midpoint initial calibration standard.
- 10.10.7** At the conclusion of data acquisition, use the same software that was used in the calibration procedure to identify peaks of interest in predetermined retention time windows. Use the data system software to examine the ion abundances of the peaks in the chromatogram. Identify an analyte by comparison of its retention time with that of the corresponding method analyte peak in a reference standard.
- 10.10.7.1** Method analyte, EIS analyte, and NIS analyte RTs must fall within 0.4 minutes of the predicted retention times from the midpoint standard of the ICAL or initial daily CV, whichever was used to establish the RT window position for the analytical batch. All branched isomer peaks identified in either the calibration standard or the qualitative (technical grade) standard must fall within in the retention time window for that analyte.
- 10.10.7.2** For all method analytes with exact corresponding isotopically labeled analogs, method analytes must elute within 0.1 minutes of the associated EIS.
- 10.10.8** The analyst must not extrapolate beyond the established calibration range. If an analyte peak area exceeds the range of the initial calibration curve, the sample should be re-extracted with a reduced sample volume in order to bring the out of range target analytes into the calibration range. If a smaller sample size would not be representative of the entire sample, the following options are recommended. Re-extract an additional aliquot of sufficient size to ensure that it is representative of the entire sample. Spike it with a higher concentration of internal standard. Prior to LC/MS analysis, dilute the sample so that it has a concentration of internal standard equivalent to that present in the calibration standard. Then, analyze the diluted extract.3
- 10.10.9** In instances where re-extraction is not an option, dilute a subsample of the sample extract with 0.1% acetic acid by a factor no greater than 10x adjust the amount of the NIS in the diluted extract, and analyze the diluted extract. If the responses for each EIS in the diluted extract meet the S/N and retention time, and the EIS recoveries from the analysis of the diluted extract are greater than 5%, then the compounds associated with those EISs may be quantified using isotope dilution. Use the EIS recoveries from the original analysis to select the dilution factor, with the objective of keeping the EIS recoveries in the dilution above that 5% lower limit. If the adjusted EIS recoveries are below 5%, the dilution is assumed invalid. If the adjusted EIS recoveries are greater than 5%, adjust the compound concentrations, detection limits, and minimum levels to account for the dilution.

11. Data Evaluation, Calculations and Reporting

11.1 Complete chromatographic resolution is not necessary for accurate and precise measurements of analyte concentrations using MS/MS. In validating this method, concentrations were calculated by measuring the product ions listed in Table 9.

11.2 Calculate analyte concentrations using the multipoint calibration established in Section 10.9. Do not use daily calibration verification data to quantitate analytes in samples. Adjust final analyte concentrations to reflect the actual sample volume determined in Section 10.8

$$C_{ex} = (\text{Area of target analyte} * \text{Concentration of Labeled analog}) / (\text{area of labeled analog} * \text{CF})$$

$$C_s = (C_{ex} / \text{sample volume in ml}) * 1000$$

C_{ex} = The concentration of the analyte in the extract
CF = calibration factor from calibration.

11.3 Prior to reporting the data, the chromatogram should be reviewed for any incorrect peak identification or poor integration.

11.4 PFHxS, PFOS, PFOA, NMeFOSAA, and NEtFOSAA have multiple chromatographic peaks using the LC conditions in Table 7 due to the linear and branch isomers of these compounds (Sect. 10.10.4.). The areas of all the linear and branched isomer peaks observed in the CAL standards for each of these analytes must be summed and the concentrations reported as a total for each of these analytes.

11.5 Calculations must utilize all available digits of precision, but final reported concentrations should be rounded to an appropriate number of significant figures (one digit of uncertainty), typically two, and not more than three significant figures.

12. Contingencies for Handling Out-of-Control Data or Unacceptable Data

12.1 Section 9.0 outlines sample batch QC acceptance criteria. If non-compliant organic compound results are to be reported, the Organic Section Head and/or the Laboratory Director, and the Operations Manager must approve the reporting of these results. The laboratory Project Manager shall be notified and may choose to relay the non-compliance to the client, for approval, or other corrective action, such as re-sampling and re-analysis. The analyst, Data Reviewer, or Department Supervisor performing the secondary review initiates the project narrative, and the narrative must clearly document the non-compliance and provide a reason for acceptance of these results.

12.2 All results for the organic compounds of interest are reportable without qualification if extraction and analytical holding times are met, preservation requirements (including cooler temperatures) are met, all QC criteria are met, and matrix interference is not suspected during extraction or analysis of the samples. If any of the below QC parameters are not met, all associated samples must be evaluated for re-extraction and/or re-analysis.

13. Method Performance

13.1 Detection Limit Study (DL) / Limit of Detection Study (LOD) / Limit of Quantitation (LOQ)

13.1.1 The laboratory follows the procedure to determine the DL, LOD, and/or LOQ as outlined in Alpha SOP ID 1732. These studies performed by the laboratory are maintained on file for review.

13.2 Demonstration of Capability Studies

13.2.1 Refer to Alpha SOP ID 1739 for further information regarding IDC/DOC Generation.

13.2.2 The analyst must make a continuing, annual, demonstration of the ability to generate acceptable accuracy and precision with this method.

14. Pollution Prevention and Waste Management

14.1 Refer to Alpha's Chemical Hygiene Plan and Hazardous Waste Management and Disposal SOP for further pollution prevention and waste management information.

14.2 This method utilizes SPE to extract analytes from water. It requires the use of very small volumes of organic solvent and very small quantities of pure analytes, thereby minimizing the potential hazards to both the analyst and the environment as compared to the use of large volumes of organic solvents in conventional liquid-liquid extractions.

14.3 The analytical procedures described in this method generate relatively small amounts of waste since only small amounts of reagents and solvents are used. The matrices of concern are finished drinking water or source water. However, laboratory waste management practices must be conducted consistent with all applicable rules and regulations, and that laboratories protect the air, water, and land by minimizing and controlling all releases from fume hoods and bench operations. Also, compliance is required with any sewage discharge permits and regulations, particularly the hazardous waste identification rules and land disposal restrictions.

15. Referenced Documents

Chemical Hygiene Plan – ID 2124

SOP ID 1732 Detection Limit (DL), Limit of Detection (LOD) & Limit of Quantitation (LOQ) SOP

SOP ID 1739 Demonstration of Capability (DOC) Generation SOP

SOP ID 1728 Hazardous Waste Management and Disposal SOP

16. Attachments

Table 1: Names, Abbreviations, and CAS Registry Numbers for Target PFAS, Extracted Internal Standards and Non-extracted Internal Standards

Parameter	Acronym	CAS
PER- and POLYFLUOROALKYLETHER CARBOXYLIC ACIDS (PFECAs)		
Tetrafluoro-2-(heptafluoropropoxy)propanoic acid	HFPO-DA	13252-13-6
4,8-dioxa-3H-perfluorononanoic acid	ADONA	919005-14-4
Perfluoro-3-methoxypropanoic acid	PFMPA	377-73-1
Perfluoro-4-methoxybutanoic acid	PFMBA	863090-89-5
Nonafluoro-3,6-dioxaheptanoic acid	NFDHA	151772-58-6
PERFLUOROALKYL CARBOXYLIC ACIDS (PFCAs)		
Perfluorobutanoic acid	PFBA	375-22-4
Perfluoropentanoic acid	PFPeA	2706-90-3
Perfluorohexanoic acid	PFHxA	307-24-4
Perfluoroheptanoic acid	PFHpA	375-85-9
Perfluorooctanoic acid	PFOA	335-67-1
Perfluorononanoic acid	PFNA	375-95-1
Perfluorodecanoic acid	PFDA	335-76-2
Perfluoroundecanoic acid	PFUnA	2058-94-8
Perfluorododecanoic acid	PFDoA	307-55-1
Perfluorotridecanoic acid	PFTrDA	72629-94-8
Perfluorotetradecanoic acid	PFTeDA	376-06-7
PERFLUOROALKYL SULFONIC ACIDS (PFASs)		
Perfluorobutanesulfonic acid	PFBS	375-73-5
Perfluoropentanesulfonic acid	PFPeS	2706-91-4
Perfluorohexanesulfonic acid	PFHxS	355-46-4
Perfluoroheptanesulfonic acid	PFHpS	375-92-8
Perfluorooctanesulfonic acid	PFOS	1763-23-1
Perfluorononanesulfonic acid	PFNS	68259-12-1
Perfluorodecanesulfonic acid	PFDS	335-77-3
Perfluorododecanesulfonic acid	PFDoS	79780-39-5
CHLORO-PERFLUOROALKYLSULFONATE		
11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid	11Cl-PF3OUdS	763051-92-9
Perfluoro(2-ethoxyethane)sulfonic acid	PFEESA	113507-82-7
9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid	9Cl-PF3ONS	756426-58-1
FLUOROTELOMER CARBOXYLIC ACIDS		
3-Perfluoropropyl propanoic acid	3:3FTCA	356-02-5
2H,2H,3H,3H-Perfluorooctanoic acid	5:3FTCA	914637-49-3
Perfluoroheptyl propanoic acid	7:3FTCA	812-70-4
PERFLUOROCTANESULFONAMIDES		

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Perfluorooctanesulfonamide	PFOSA	754-91-6
N-methylperfluoro-1-octanesulfonamide	NMeFOSA	31506-32-8
N-ethylperfluoro-1-octanesulfonamide	NEtFOSA	4151-50-2
PERFLUOROCTANE SULFONAMIDE ETHANOLS		
N-Methyl perfluorooctanesulfonamidoethanol	NMeFOSE	24448-09-7
N-ethyl perfluorooctanesulfonamidoethanol	NEtFOSE	1691-99-2
TELOMER SULFONIC ACIDS		
1H,1H,2H,2H-perfluorohexanesulfonic acid (4:2)	4:2FTS	757124-72-4
1H,1H,2H,2H-perfluorooctanesulfonic acid (6:2)	6:2FTS	27619-97-2
1H,1H,2H,2H-perfluorodecanesulfonic acid (8:2)	8:2FTS	39108-34-4
PERFLUOROCTANESULFONAMIDOACETIC ACIDS		
N-methyl perfluorooctanesulfonamidoacetic acid	NMeFOSAA	2355-31-9
N-ethyl perfluorooctanesulfonamidoacetic acid	NEtFOSAA	2991-50-6
PERFLUOROETHER AND POLYETHER CARBOXYLIC ACIDS		
Perfluoro-3-methoxypropanoic acid	PFMPA	377-73-1
Perfluoro-4-methoxybutanoic acid	PFMBA	863090-89-5
Perfluoro(2-ethoxyethane)sulfonic acid	PFEESA	113507-82-7
Nonafluoro-3,6-dioxaheptanoic acid	NFDHA	151772-58-6

Table 2: Stock and Nominal Extracted Internal Standard Concentrations

Isotope Labeled Standard	Conc. of EIS Stock (ng/mL)	Nominal amount of EIS added to extracts (ng)
M4PFBA	2000	40
M5PFPeA	1000	20
M5PFHxA	500	10
M4PFHpA	500	10
M8PFOA	500	10
M9PFNA	250	5
M6PFDA	250	5
M7PFUdA	250	5
MPFDoA	250	5
M2PFTeDA	250	5
M3PFBS	466	9.32
M3PFHxS	474	9.48
M8PFOS	479	9.58
M2-4:2FTS	938	18.8
M2-6:2FTS	951	19
M2-8:2FTS	960	19.2
M8FOSA	500	10
d3-N-MeFOSA	500	10
d5-N-EtFOSA	500	10
d3-N-MeFOSAA	1000	20
d5-N-EtFOSAA	1000	20
d7-N-MeFOSE	5000	100
d9-N-EtFOSE	5000	100
M3HFPO-DA	2000	40

Table 3: Stock and Nominal Non-Extracted Internal Standard Concentrations

Isotope Labeled Standard	Conc. of EIS Stock (ng/mL)	Nominal amount of EIS added to extracts (ng)
M3PFBA	1000	40
M2PFHxA	500	10
M4PFOA	500	10
M5PFNA	250	5
M2PFDA	250	5
18O2PFHxS	474	9.48
M4PFOS	479	9.58

Table 4: Initial Calibration levels and Concentrations

Analyte	Cal A	Cal B (LOQ)	CAL C	Cal D	Cal E (CCV)	Cal F	Cal G	Cal H	Cal I
PFBA	.4	.8	2	5	10	20	50	250	500
PFPeA	.2	.4	1	2.5	5	10	25	125	250
PFHxA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
PFHpA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
PFOA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
PFNA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
PFDA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
PFUnA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
PFDoA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
PFTrDA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
PFTA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
PFBS	0.089	0.177	0.444	1.11	2.22	4.44	11.1	55.4	111
PFPeS	0.094	0.188	0.471	1.18	2.35	4.71	11.8	58.8	118
PFHxS	0.091	0.183	0.457	1.14	2.29	4.57	11.4	57.1	114
PFHpS	0.095	0.191	0.477	1.19	2.38	4.77	11.9	59.6	119
PFOS	0.093	0.186	0.464	1.16	2.32	4.64	11.6	58	116
PFNS	0.096	0.192	0.481	1.20	2.41	4.81	12	60.1	120
PFDS	0.097	0.193	0.483	1.21	2.41	4.83	12.1	60.3	121
PFDOS	0.097	0.194	0.485	1.21	2.43	4.85	12.1	60.6	121.
4:2FTS	0.375	0.75	1.88	4.69	9.38	18.8	46.9	234	469
6:2FTS	0.38	0.76	1.9	4.75	9.5	19	47.5	238	475
8:2FTS	0.384	0.768	1.92	4.8	9.6	19.2	48	240	480
PFOSA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
NMeFOSA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
NEtFOSA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
NMeFOSAA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
NEtFOSAA	.1	.2	.5	1.25	2.5	5	12.5	62.5	125
NMeFOSE	1	2	5	12.5	25	50	125	625	1250
NEtFOSE	1	2	5	12.5	25	50	125	625	1250
HFPO-DA	.4	.8	2	5	10	20	50	250	500
ADONA	0.378	0.756	1.89	4.73	9.45	18.9	47.3	236	473
9Cl-PFONS	0.374	0.748	1.87	4.68	9.35	18.7	46.8	234	468
11Cl-PFOUdS	0.378	0.756	1.89	4.73	9.45	18.9	47.3	236	473
PFMPA	.2	.4	1	2.5	5	10	25	125	250
PFMBA	.2	.4	1	2.5	5	10	25	125	250
PFEESA	0.178	0.356	0.89	2.23	4.45	8.9	22.3	111	223
NFDHA	.2	.4	1	2.5	5	10	25	125	250

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3:3FTCA	.5	1	2.5	6.25	12.5	25	62.5	312	624
5:3FTCA	2.5	5	12.5	31.3	62.5	125	312	1560	3120
7:3FTCA	2.5	5	12.5	31.3	62.5	125	312	1560	3125
M4PFBA	10	10	10	10	10	10	10	10	10
M5PFPeA	5	5	5	5	5	5	5	5	5
M5PFHxA	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5
M4PFHpA	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5
M8PFOA	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5
M9PFNA	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25
M6PFDA	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25
M7PFUdA	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25
MPFDoA	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25
M2PFTeDA	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25
M3PFBS	2.33	2.33	2.33	2.33	2.33	2.33	2.33	2.33	2.33
M3PFHxS	2.37	2.37	2.37	2.37	2.37	2.37	2.37	2.37	2.37
M8PFOS	2.4	2.4	2.4	2.4	2.4	2.4	2.4	2.4	2.4
M2-4:2FTS	4.69	4.69	4.69	4.69	4.69	4.69	4.69	4.69	4.69
M2-6:2FTS	4.76	4.76	4.76	4.76	4.76	4.76	4.76	4.76	4.76
M2-8:2FTS	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8	4.8
M8FOSA	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5
d3-N-MeFOSA	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5
d5-N-EtFOSA	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5
d3-N-MeFOSAA	5	5	5	5	5	5	5	5	5
d5-N-EtFOSAA	5	5	5	5	5	5	5	5	5
d7-N-MeFOSE	25	25	25	25	25	25	25	25	25
d9-N-EtFOSE	25	25	25	25	25	25	25	25	25
M3HFPO-DA	10	10	10	10	10	10	10	10	10
M3PFBA	5	5	5	5	5	5	5	5	5
M2PFHxA	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5
M4PFOA	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5	2.5
M5PFNA	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25
M2PFDA	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25	1.25
18O2PFHxS	2.37	2.37	2.37	2.37	2.37	2.37	2.37	2.37	2.37
M4PFOS	2.4	2.4	2.4	2.4	2.4	2.4	2.4	2.4	2.4

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Table 5: Expected Mass Transitions and instrument conditions.

Q1	Q2	Analyte	DP Volts	CE Volts
213.032	169.022	PFBA	-50	-14
263.039	219.03	PFPeA	-55	-12
263.039	68.9	PFPeA_2	-55	-55
313.047	269.037	PFHxA	-45	-12
313.047	119	PFHxA_2	-45	-28
363.055	319.045	PFHpA	-60	-12
363.055	169.022	PFHpA_2	-60	-24
413.063	369.053	PFOA	-65	-14
413.063	169.022	PFOA_2	-65	-23
463.071	419.061	PFNA	-70	-14
463.071	219.03	PFNA_2	-70	-24
513.078	469.069	PFDA	-80	-16
513.078	219.03	PFDA_2	-80	-30
563.086	519.076	PFUnA	-85	-18
563.086	269.037	PFUnA_2	-85	-25
613.094	569.084	PFDoA	-85	-18
613.094	319.045	PFDoA_2	-85	-28
663.102	619.092	PFTTrDA	-85	-20
663.102	169.022	PFTTrDA_2	-85	-36
713.11	669.1	PFTA	-70	-22
713.11	169.022	PFTA_2	-70	-38
299.092	80.062	PFBS	-100	-65
299.092	99.061	PFBS_2	-100	-40
349.1	80.062	PFPeS	-100	-75
349.1	99.061	PFPeS_2	-100	-60
399.107	80.062	PFHxS	-120	-75
399.107	99.061	PFHxS_2	-120	-80
449.115	80.062	PFHpS	-140	-95
449.115	99.061	PFHpS_2	-140	-80
499.113	80.062	PFOS	-145	-108
499.113	99.061	PFOS_2	-145	-85
549.131	80.062	PFNS	-180	-100
549.131	99.061	PFNS_2	-180	-100
599.139	80.062	PFDS	-170	-110
599.138	99.061	PFDS_2	-170	-100
699.154	80.062	PFDoS	-160	-150
699.154	99.061	PFDoS_2	-160	-130

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327.146	307.139	4:2FTS	-100	-28
327.146	81.07	4:2FTS_2	-100	-50
427.161	407.155	6:2FTS	-120	-33
427.161	81.07	6:2FTS_2	-120	-65
527.177	507.17	8:2FTS	-140	-39
527.177	81.07	8:2FTS_2	-140	-85
498.146	78.07	FOSA	-150	-90
498.146	478	FOSA_2	-150	-35
512.163	219.03	NMeFOSA	-130	-35
512.163	169.022	NMeFOSA_2	-130	-40
526.192	219.03	NEtFOSA	-140	-35
526.192	169.022	NEtFOSA_2	-140	-35
570.202	419.061	NMeFOSAA	-100	-28
570.202	483	NMeFOSAA_2	-100	-22
584.229	419.061	NEtFOSAA	-100	-28
584.229	526.192	NEtFOSAA_2	-100	-38
616.1	58.9	NMeFOSE	-90	-70
630	58.9	NEtFOSE	-80	-75
285.035	169.022	HFPO-DA	-60	-12
285.035	184.9	HFPO-DA_2	-60	-18
377.06	251.028	ADONA	-65	-18
377.06	84.8	ADONA_2	-65	-48
530.8	351.05	9CI-PFONS	-130	-38
532.8	353	9CI-PFONS_2	-130	-38
630.9	451.031	11CI-PFOUdS	-145	-41
632.9	452.9	11CI-PFOUdS_2	-145	-41
241.085	177.069	3:3FTCA	-60	-12
241.085	117	3:3FTCA_2	-60	-50
341.101	237.072	5:3FTCA	-70	-20
341.101	217	5:3FTCA_2	-70	-35
441.117	316.9	7:3FTCA	-85	-30
441.117	337.088	7:3FTCA_2	-85	-20
315.093	135.013	PFEESA	-100	-35
315.093	82.9	PFEESA_2	-100	-25
229.032	85.006	PFMPA	-40	-25
279.042	85.006	PFMBA	-45	-25
295.032	201	NFDHA	-30	-15
295.032	84.9	NFDHA_2	-30	-40
217.001	171.999	MPFBA	-50	-14

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268.001	222.999	M5PFPeA	-55	-12
318.009	273.007	M5PFHxA	-45	-12
367.024	322.022	M4PFHpA	-60	-12
421.002	376	M8PFOA	-65	-14
472.002	427	M9PFNA	-70	-14
519.033	474.03	M6PFDA	-80	-16
570.033	525.031	M7-PFUdA	-85	-18
615.079	570.033	MPFDoA	-85	-18
715.094	670.092	M2PFTeDA	-70	-22
302.069	80.062	M3PFBS	-100	-65
402.084	80.062	M3PFHxS	-120	-74
507.062	80.062	M8PFOS	-145	-85
329.13	81.07	M2-4:2FTS	-100	-50
429.162	81.07	M2-6:2FTS	-120	-65
529.162	81.07	M2-8:2FTS	-140	-85
506.077	78.07	M8FOSA	-150	-90
515.183	219.03	d3-NMeFOSA	-130	-35
531.222	219.03	d5-NEtFOSA	-140	-35
573.22	419.061	d3-NMeFOSAA	-75	-28
589.259	419.061	d5-NEtFOSAA	-90	-28
623.2	58.9	d7-NMeFOSE	-100	-28
639.2	58.9	d9-NEtFOSE	-100	-28
287.02	169.022	M3HFPO-DA	-60	-12
216.009	171.999	M3PFBA	-50	-14
315.032	270.03	M2PFHxA	-45	-12
417.032	372.03	M4PFOA	-65	-14
468.032	423.03	M5PFNA	-70	-14
515.063	470.061	M2PFDA	-80	-16
403.107	84.062	18O2-PFHxS	-120	-74
503.093	80.062	M4PFOS	-145	-85

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Table 6: LC Method Conditions

Time (min)	2 mM Ammonium Acetate (5:95 CH/H ₂ O)	100% Acetonitrile	Gradient Curve
Initial	100.0	0.0	0
.2	100.0	0.0	2
4	70	30	7
7	45	55	8
9	25	80	8
10	5	95	6
10.4	98	2	10
11.8	100	0	7
12	100	0	1
Waters Aquity UPLC ® BEHC ₁₈ 2.1 x 50 mm packed with 1.7 µm BEH C ₁₈ stationary phase Flow rate of 0.4 mL/min 2 µL injection			

Table 7: ESI-MS Method Conditions

ESI Conditions	
Polarity	Negative ion
Curtain Gas	30
Collision gas	9
Ion Spray Voltage	-4500
Desolvation gas temp.	500 °C
Ion Source Gas 1	30
Ion Source Gas 2	50
Entrance Poitential	-10
Exic Cell Potential	-11

Table 8. Reporting Limits by Matrix

Compound	Aqueous (ng/L)	Solid (ng/g)	Tissue (ng/g)
PFBA	6.4	0.8	2
PFPeA	3.2	0.4	1
PFHxA	1.6	0.2	0.5
PFHpA	1.6	0.2	0.5
PFOA	1.6	0.2	0.5
PFNA	1.6	0.2	0.5
PFDA	1.6	0.2	0.5
PFUnA	1.6	0.2	0.5
PFDoA	1.6	0.2	0.5
PFTTrDA	1.6	0.2	0.5
PFTA	1.6	0.2	0.5
PFBS	1.6	0.2	0.5
PFPeS	1.6	0.2	0.5
PFHxS	1.6	0.2	0.5
PFHpS	1.6	0.2	0.5
PFOS	1.6	0.2	0.5
PFNS	1.6	0.2	0.5
PFDS	1.6	0.2	0.5
PFDoS	1.6	0.2	0.5
4:2FTS	6.4	0.8	2
6:2FTS	6.4	0.8	2
8:2FTS	6.4	0.8	2
FOSA	1.6	0.2	2
NMeFOSA	1.6	0.2	0.5
NEtFOSA	1.6	0.2	0.5
NMeFOSAA	1.6	0.2	0.5
NEtFOSAA	1.6	0.2	0.5
NMeFOSE	16	2	5
NEtFOSE	16	2	5
HFPO-DA	6.4	0.8	2
ADONA	6.4	0.8	2
9CI-PFONS	6.4	0.8	2
11CI-PFOUdS	6.4	0.8	2
3:3FTCA	8	1	2.5
5:3FTCA	40	5	12.5
7:3FTCA	40	5	12.5
PFEEA	3.2	0.4	1
PFMPA	3.2	0.4	1
PFMBA	3.2	0.4	1
NFDHA	3.2	0.4	1

17. TOP Assay SOP Addendum

The following modifications are applied to this SOP;

Section 7: Water Bath – Capable of monitoring recording to .1 C maintaining 85 C.

Section 8.1

1. Sodium Hydroxide (NaOH, CAS#: 1310-73-2) – High purity, demonstrated to be free of analytes and interferences.
2. Potassium Persulfate ($K_2S_2O_8$, CAS#: 7727-21-1) - High purity, demonstrated to be free of analytes and interferences.
3. Hydrochloric Acid (HCl, CAS#: 7647-01-0) - High purity, demonstrated to be free of analytes and interferences.

Table 1

Isotope Labeled Standard	Conc. Top Surr Stock (ng/mL)	Vol. of Top Surr Stock (μ l)	Final Vol. of Top Surr PDS (mL)	Final Conc. of Top Surr PDS (ng/mL)
$^{13}C_2$ -PFOA	50,000	40	4.0	500
$^{13}C_2D_4$ -4:2FTS	50,000	80	4.0	1000

Section 10

1. Pretreatment of Samples - For Aqueous samples, prior to Section 10.1, add to each 125 ml sample with Potassium Persulfate until 60 mM (about 2 grams). Add Sodium Hydroxide until the sample is 125 mM (about .625 grams). Sample should have a pH >12.
2. Fortify each sample with 20 μ l of TOP pre-assay surrogate containing 1 negative control surrogates and 1 positive control surrogates from table 1.
3. Place sample in a water bath at 85 C for 6 hours.
4. Remove sample from water bath and adjust pH 6-8 with Hydrochloric Acid.
5. Following pre-treatment, follow the aqueous extraction protocol from section 10 of SOP 45852 unaltered.

Section 11 - An additional calculation is added for the final reporting of the TOP assay results.

TOP Assay = Result of Post TOP assay extraction – Result of analysis from Pre-Top Assay extraction.



ATTACHMENT C

NOVEMBER 2025 POST REMEDIATION MONITORING WELL PURGE LOGS



ATTACHMENT D

NOVEMBER 2025 POST REMEDIATION GROUNDWATER LABORATORY REPORT



ANALYTICAL REPORT

Lab Number:	L2569921
Client:	GZA GeoEnvironmental, Inc. 104 West 29th Street, 10th Floor New York, NY 10001
ATTN:	Zach Landis
Phone:	(212) 594-8140
Project Name:	BLONDELL EQUITIES
Project Number:	41.0163256.00
Report Date:	11/11/25

The original project report/data package is held by Pace Analytical Services. This report/data package is paginated and should be reproduced only in its entirety. Pace Analytical Services holds no responsibility for results and/or data that are not consistent with the original.

Certifications & Approvals: MA (M-MA086), NH NELAP (2064), CT (PH-0826), IL (200077), IN (C-MA-03), KY (KY98045), ME (MA00086), MD (348), NJ (MA935), NY (11148), NC (25700/666), OR (MA-1316), PA (68-03671), RI (LAO00065), TX (T104704476), VT (VT-0935), VA (460195), USDA (Permit #525-23-122-91930A1).

Eight Walkup Drive, Westborough, MA 01581-1019
508-898-9220 (Fax) 508-898-9193 800-624-9220 - www.alphalab.com



Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Lab Sample ID	Client ID	Matrix	Sample Location	Collection Date/Time	Receive Date
L2569921-01	GZM-2	WATER	BRONX, NY	11/04/25 09:30	11/04/25
L2569921-02	GZM-1	WATER	BRONX, NY	11/04/25 10:30	11/04/25
L2569921-03	GZM-3	WATER	BRONX, NY	11/04/25 11:30	11/04/25
L2569921-04	GZM-4	WATER	BRONX, NY	11/04/25 12:30	11/04/25
L2569921-05	GZM-DUP	WATER	BRONX, NY	11/04/25 00:00	11/04/25
L2569921-06	FIELD BLANK	WATER	BRONX, NY	11/04/25 14:00	11/04/25
L2569921-07	TRIP BLANK	WATER	BRONX, NY	11/04/25 00:00	11/04/25

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Case Narrative

The samples were received in accordance with the Chain of Custody and no significant deviations were encountered during the preparation or analysis unless otherwise noted. Sample Receipt, Container Information, and the Chain of Custody are located at the back of the report.

Results contained within this report relate only to the samples submitted under this Pace Lab Number and meet NELAP requirements for all NELAP accredited parameters unless otherwise noted in the following narrative. The data presented in this report is organized by parameter (i.e. VOC, SVOC, etc.). Sample specific Quality Control data (i.e. Surrogate Spike Recovery) is reported at the end of the target analyte list for each individual sample, followed by the Laboratory Batch Quality Control at the end of each parameter. Tentatively Identified Compounds (TICs), if requested, are reported for compounds identified to be present and are not part of the method/program Target Compound List, even if only a subset of the TCL are being reported. If a sample was re-analyzed or re-extracted due to a required quality control corrective action and if both sets of data are reported, the Laboratory ID of the re-analysis or re-extraction is designated with an "R" or "RE", respectively.

When multiple Batch Quality Control elements are reported (e.g. more than one LCS), the associated samples for each element are noted in the grey shaded header line of each data table. Any Laboratory Batch, Sample Specific % recovery or RPD value that is outside the listed Acceptance Criteria is bolded in the report. In reference to questions H (CAM) or 4 (RCP) when "NO" is checked, the performance criteria for CAM and RCP methods allow for some quality control failures to occur and still be within method compliance. In these instances, the specific failure is not narrated but noted in the associated QC Outlier Summary Report, located directly after the Case Narrative. QC information is also incorporated in the Data Usability Assessment table (Format 11) of our Data Merger tool, where it can be reviewed in conjunction with the sample result, associated regulatory criteria and any associated data usability implications.

Soil/sediments and solids are reported on a dry weight basis unless otherwise noted. Tissues are reported "as received" or on a wet weight basis, unless otherwise noted. Definitions of all data qualifiers and acronyms used in this report are provided in the Glossary located at the back of the report.

HOLD POLICY - For samples submitted on hold, Pace's policy is to hold samples (with the exception of Air canisters) free of charge for 21 calendar days from the date the project is completed. After 21 calendar days, we will dispose of all samples submitted including those put on hold unless you have contacted your Pace Project Manager and made arrangements for Pace to continue to hold the samples. Air canisters will be disposed after 3 business days from the date the project is completed.

Please contact Project Management at 800-624-9220 with any questions.

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Case Narrative (continued)

Report Submission

All non-detect (ND) or estimated concentrations (J-qualified) have been quantitated to the limit noted in the MDL column.

Perfluorinated Alkyl Acids by 1633

L2569921-01: The Extracted Internal Standard recoveries were above the acceptance criteria for 1h,1h,2h,2h-perfluoro[1,2-13c2]hexanesulfonic acid (m2-4:2fts) (339%) and 1h,1h,2h,2h-perfluoro[1,2-13c2]octanesulfonic acid (m2-6:2fts) (374%). Since the sample was non-detect to the RL for all associated target analytes, re-analysis was not required.

L2569921-01 and -05: The Non-extracted Internal Standard (NIS) response was below the acceptance criteria for 13C3-PFBA; however, the criteria were achieved upon re-analysis on dilution. The results of the re-analysis are reported for the associated target analytes.

L2569921-02: The Extracted Internal Standard recoveries were above the acceptance criteria for 1h,1h,2h,2h-perfluoro[1,2-13c2]hexanesulfonic acid (m2-4:2fts) (253%) and 1h,1h,2h,2h-perfluoro[1,2-13c2]octanesulfonic acid (m2-6:2fts) (258%); however, any associated target analytes detected above the RL are not reported from this analysis.

L2569921-02D: The sample was re-analyzed on dilution due to EIS failure in the original analysis. The results of the re-analysis are reported for the associated target compounds.

L2569921-03R: The sample was re-analyzed due to QC failures in the original analysis. The results of the re-analysis are reported.

L2569921-04: The Extracted Internal Standard recoveries were above the acceptance criteria for 1h,1h,2h,2h-perfluoro[1,2-13c2]hexanesulfonic acid (m2-4:2fts) (248%) and 1h,1h,2h,2h-perfluoro[1,2-13c2]octanesulfonic acid (m2-6:2fts) (240%). Since the sample was non-detect to the RL for all associated target analytes, re-analysis was not required.

L2569921-05: The Extracted Internal Standard recoveries were above the acceptance criteria for 1h,1h,2h,2h-perfluoro[1,2-13c2]hexanesulfonic acid (m2-4:2fts) (360%) and 1h,1h,2h,2h-perfluoro[1,2-13c2]octanesulfonic acid (m2-6:2fts) (392%). Since the sample was non-detect to the RL for all associated

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Case Narrative (continued)

target analytes, re-analysis was not required.

L2569921-05D: The Extracted Internal Standard recovery was outside the acceptance criteria for perfluoro[13c8]octanesulfonamide (m8fosa) (36%); however, this recovery does not apply to the abbreviated list of target analytes reported.

Total Metals

The WG2138162-1 Method Blank associated with L2569921-01 through -06 has a concentration above the reporting limit for barium. Since the associated sample concentrations are either greater than 10x the blank concentration or non-detect to the RL for this target analyte, no corrective action is required. Any results detected below the reporting limit are qualified with a "B".

The WG2138162-3 MS recovery performed on L2569921-01 is outside the acceptance criteria for manganese (131%). A post digestion spike was performed and was within acceptance criteria.

The WG2138162-3 MS recoveries performed on L2569921-01 do not apply for calcium (200%), iron (240%) and sodium (127%) because the sample concentrations are greater than four times the spike amounts added.

Dissolved Metals

L2569921-06: The Field Blank has results for copper and nickel present above the reporting limits. The sample was verified as being labeled correctly by the laboratory and the previous analysis showed there was no potential for carry over.

I, the undersigned, attest under the pains and penalties of perjury that, to the best of my knowledge and belief and based upon my personal inquiry of those responsible for providing the information contained in this analytical report, such information is accurate and complete. This certificate of analysis is not complete unless this page accompanies any and all pages of this report.

Authorized Signature:  Cristin Walker

Title: Technical Director/Representative

Date: 11/11/25

ORGANICS

VOLATILES

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Analytical Method: 1,8260D

Analytical Date: 11/09/25 18:55

Analyst: MJV

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Methylene chloride	ND		ug/l	2.5	0.70	1
1,1-Dichloroethane	ND		ug/l	2.5	0.70	1
Chloroform	ND		ug/l	2.5	0.70	1
Carbon tetrachloride	ND		ug/l	0.50	0.13	1
1,2-Dichloropropane	ND		ug/l	1.0	0.14	1
Dibromochloromethane	ND		ug/l	0.50	0.15	1
1,1,2-Trichloroethane	ND		ug/l	1.5	0.50	1
Tetrachloroethene	ND		ug/l	0.50	0.18	1
Chlorobenzene	ND		ug/l	2.5	0.70	1
Trichlorofluoromethane	ND		ug/l	2.5	0.70	1
1,2-Dichloroethane	ND		ug/l	0.50	0.13	1
1,1,1-Trichloroethane	ND		ug/l	2.5	0.70	1
Bromodichloromethane	ND		ug/l	0.50	0.19	1
trans-1,3-Dichloropropene	ND		ug/l	0.50	0.16	1
cis-1,3-Dichloropropene	ND		ug/l	0.50	0.14	1
1,3-Dichloropropene, Total	ND		ug/l	0.50	0.14	1
1,1-Dichloropropene	ND		ug/l	2.5	0.70	1
Bromoform	ND		ug/l	2.0	0.65	1
1,1,1,2-Tetrachloroethane	ND		ug/l	0.50	0.17	1
Benzene	0.85		ug/l	0.50	0.16	1
Toluene	ND		ug/l	2.5	0.70	1
Ethylbenzene	ND		ug/l	2.5	0.70	1
Chloromethane	ND		ug/l	2.5	0.70	1
Bromomethane	ND		ug/l	2.5	0.70	1
Vinyl chloride	0.12	J	ug/l	1.0	0.07	1
Chloroethane	ND		ug/l	2.5	0.70	1
1,1-Dichloroethene	ND		ug/l	0.50	0.17	1
trans-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Trichloroethene	ND		ug/l	0.50	0.18	1
1,2-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,3-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,4-Dichlorobenzene	ND		ug/l	2.5	0.70	1
Methyl tert butyl ether	0.51	J	ug/l	2.5	0.17	1
p/m-Xylene	ND		ug/l	2.5	0.70	1
o-Xylene	ND		ug/l	2.5	0.70	1
Xylenes, Total	ND		ug/l	2.5	0.70	1
cis-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1
1,2-Dichloroethene, Total	ND		ug/l	2.5	0.70	1
Dibromomethane	ND		ug/l	5.0	1.0	1
1,2,3-Trichloropropane	ND		ug/l	2.5	0.70	1
Acrylonitrile	ND		ug/l	5.0	1.5	1
Styrene	ND		ug/l	2.5	0.70	1
Dichlorodifluoromethane	ND		ug/l	5.0	1.0	1
Acetone	ND		ug/l	5.0	1.5	1
Carbon disulfide	ND		ug/l	5.0	1.0	1
2-Butanone	ND		ug/l	5.0	1.9	1
Vinyl acetate	ND		ug/l	5.0	1.0	1
4-Methyl-2-pentanone	ND		ug/l	5.0	1.0	1
2-Hexanone	ND		ug/l	5.0	1.0	1
Bromochloromethane	ND		ug/l	2.5	0.70	1
2,2-Dichloropropane	ND		ug/l	2.5	0.70	1
1,2-Dibromoethane	ND		ug/l	2.0	0.65	1
1,3-Dichloropropane	ND		ug/l	2.5	0.70	1
1,1,1,2-Tetrachloroethane	ND		ug/l	2.5	0.70	1
Bromobenzene	ND		ug/l	2.5	0.70	1
n-Butylbenzene	ND		ug/l	2.5	0.70	1
sec-Butylbenzene	ND		ug/l	2.5	0.70	1
tert-Butylbenzene	ND		ug/l	2.5	0.70	1
o-Chlorotoluene	ND		ug/l	2.5	0.70	1
p-Chlorotoluene	ND		ug/l	2.5	0.70	1
1,2-Dibromo-3-chloropropane	ND		ug/l	2.5	0.70	1
Hexachlorobutadiene	ND		ug/l	2.5	0.70	1
Isopropylbenzene	ND		ug/l	2.5	0.70	1
p-Isopropyltoluene	ND		ug/l	2.5	0.70	1
Naphthalene	ND		ug/l	2.5	0.70	1



Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
n-Propylbenzene	ND		ug/l	2.5	0.70	1
1,2,3-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,3,5-Trimethylbenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trimethylbenzene	ND		ug/l	2.5	0.70	1
1,4-Dioxane	ND		ug/l	250	61.	1
p-Diethylbenzene	ND		ug/l	2.0	0.70	1
p-Ethyltoluene	ND		ug/l	2.0	0.70	1
1,2,4,5-Tetramethylbenzene	ND		ug/l	2.0	0.54	1
Ethyl ether	ND		ug/l	2.5	0.70	1
trans-1,4-Dichloro-2-butene	ND		ug/l	2.5	0.70	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
1,2-Dichloroethane-d4	88		70-130
Toluene-d8	103		70-130
4-Bromofluorobenzene	95		70-130
Dibromofluoromethane	100		70-130

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02 D

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Analytical Method: 1,8260D

Analytical Date: 11/09/25 20:41

Analyst: MJV

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Methylene chloride	ND		ug/l	25	7.0	10
1,1-Dichloroethane	ND		ug/l	25	7.0	10
Chloroform	ND		ug/l	25	7.0	10
Carbon tetrachloride	ND		ug/l	5.0	1.3	10
1,2-Dichloropropane	ND		ug/l	10	1.4	10
Dibromochloromethane	ND		ug/l	5.0	1.5	10
1,1,2-Trichloroethane	ND		ug/l	15	5.0	10
Tetrachloroethene	ND		ug/l	5.0	1.8	10
Chlorobenzene	ND		ug/l	25	7.0	10
Trichlorofluoromethane	ND		ug/l	25	7.0	10
1,2-Dichloroethane	ND		ug/l	5.0	1.3	10
1,1,1-Trichloroethane	ND		ug/l	25	7.0	10
Bromodichloromethane	ND		ug/l	5.0	1.9	10
trans-1,3-Dichloropropene	ND		ug/l	5.0	1.6	10
cis-1,3-Dichloropropene	ND		ug/l	5.0	1.4	10
1,3-Dichloropropene, Total	ND		ug/l	5.0	1.4	10
1,1-Dichloropropene	ND		ug/l	25	7.0	10
Bromoform	ND		ug/l	20	6.5	10
1,1,1,2-Tetrachloroethane	ND		ug/l	5.0	1.7	10
Benzene	85		ug/l	5.0	1.6	10
Toluene	25		ug/l	25	7.0	10
Ethylbenzene	690		ug/l	25	7.0	10
Chloromethane	ND		ug/l	25	7.0	10
Bromomethane	ND		ug/l	25	7.0	10
Vinyl chloride	ND		ug/l	10	0.71	10
Chloroethane	ND		ug/l	25	7.0	10
1,1-Dichloroethene	ND		ug/l	5.0	1.7	10
trans-1,2-Dichloroethene	ND		ug/l	25	7.0	10



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-02 D

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Trichloroethene	ND		ug/l	5.0	1.8	10
1,2-Dichlorobenzene	ND		ug/l	25	7.0	10
1,3-Dichlorobenzene	ND		ug/l	25	7.0	10
1,4-Dichlorobenzene	ND		ug/l	25	7.0	10
Methyl tert butyl ether	ND		ug/l	25	1.7	10
p/m-Xylene	1300		ug/l	25	7.0	10
o-Xylene	280		ug/l	25	7.0	10
Xylenes, Total	1600		ug/l	25	7.0	10
cis-1,2-Dichloroethene	ND		ug/l	25	7.0	10
1,2-Dichloroethene, Total	ND		ug/l	25	7.0	10
Dibromomethane	ND		ug/l	50	10.	10
1,2,3-Trichloropropane	ND		ug/l	25	7.0	10
Acrylonitrile	ND		ug/l	50	15.	10
Styrene	ND		ug/l	25	7.0	10
Dichlorodifluoromethane	ND		ug/l	50	10.	10
Acetone	ND		ug/l	50	15.	10
Carbon disulfide	ND		ug/l	50	10.	10
2-Butanone	ND		ug/l	50	19.	10
Vinyl acetate	ND		ug/l	50	10.	10
4-Methyl-2-pentanone	ND		ug/l	50	10.	10
2-Hexanone	ND		ug/l	50	10.	10
Bromochloromethane	ND		ug/l	25	7.0	10
2,2-Dichloropropane	ND		ug/l	25	7.0	10
1,2-Dibromoethane	ND		ug/l	20	6.5	10
1,3-Dichloropropane	ND		ug/l	25	7.0	10
1,1,1,2-Tetrachloroethane	ND		ug/l	25	7.0	10
Bromobenzene	ND		ug/l	25	7.0	10
n-Butylbenzene	23	J	ug/l	25	7.0	10
sec-Butylbenzene	ND		ug/l	25	7.0	10
tert-Butylbenzene	ND		ug/l	25	7.0	10
o-Chlorotoluene	ND		ug/l	25	7.0	10
p-Chlorotoluene	ND		ug/l	25	7.0	10
1,2-Dibromo-3-chloropropane	ND		ug/l	25	7.0	10
Hexachlorobutadiene	ND		ug/l	25	7.0	10
Isopropylbenzene	77		ug/l	25	7.0	10
p-Isopropyltoluene	ND		ug/l	25	7.0	10
Naphthalene	ND		ug/l	25	7.0	10



Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02 D

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
n-Propylbenzene	190		ug/l	25	7.0	10
1,2,3-Trichlorobenzene	ND		ug/l	25	7.0	10
1,2,4-Trichlorobenzene	ND		ug/l	25	7.0	10
1,3,5-Trimethylbenzene	190		ug/l	25	7.0	10
1,2,4-Trimethylbenzene	880		ug/l	25	7.0	10
1,4-Dioxane	ND		ug/l	2500	610	10
p-Diethylbenzene	93		ug/l	20	7.0	10
p-Ethyltoluene	660		ug/l	20	7.0	10
1,2,4,5-Tetramethylbenzene	74		ug/l	20	5.4	10
Ethyl ether	ND		ug/l	25	7.0	10
trans-1,4-Dichloro-2-butene	ND		ug/l	25	7.0	10

Surrogate	% Recovery	Qualifier	Acceptance Criteria
1,2-Dichloroethane-d4	85		70-130
Toluene-d8	101		70-130
4-Bromofluorobenzene	93		70-130
Dibromofluoromethane	95		70-130

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-03

Date Collected: 11/04/25 11:30

Client ID: GZM-3

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Analytical Method: 1,8260D

Analytical Date: 11/09/25 19:22

Analyst: MJV

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Methylene chloride	ND		ug/l	2.5	0.70	1
1,1-Dichloroethane	ND		ug/l	2.5	0.70	1
Chloroform	ND		ug/l	2.5	0.70	1
Carbon tetrachloride	ND		ug/l	0.50	0.13	1
1,2-Dichloropropane	ND		ug/l	1.0	0.14	1
Dibromochloromethane	ND		ug/l	0.50	0.15	1
1,1,2-Trichloroethane	ND		ug/l	1.5	0.50	1
Tetrachloroethene	ND		ug/l	0.50	0.18	1
Chlorobenzene	ND		ug/l	2.5	0.70	1
Trichlorofluoromethane	ND		ug/l	2.5	0.70	1
1,2-Dichloroethane	ND		ug/l	0.50	0.13	1
1,1,1-Trichloroethane	ND		ug/l	2.5	0.70	1
Bromodichloromethane	ND		ug/l	0.50	0.19	1
trans-1,3-Dichloropropene	ND		ug/l	0.50	0.16	1
cis-1,3-Dichloropropene	ND		ug/l	0.50	0.14	1
1,3-Dichloropropene, Total	ND		ug/l	0.50	0.14	1
1,1-Dichloropropene	ND		ug/l	2.5	0.70	1
Bromoform	ND		ug/l	2.0	0.65	1
1,1,2,2-Tetrachloroethane	ND		ug/l	0.50	0.17	1
Benzene	0.35	J	ug/l	0.50	0.16	1
Toluene	ND		ug/l	2.5	0.70	1
Ethylbenzene	1.1	J	ug/l	2.5	0.70	1
Chloromethane	ND		ug/l	2.5	0.70	1
Bromomethane	ND		ug/l	2.5	0.70	1
Vinyl chloride	ND		ug/l	1.0	0.07	1
Chloroethane	ND		ug/l	2.5	0.70	1
1,1-Dichloroethene	ND		ug/l	0.50	0.17	1
trans-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-03

Date Collected: 11/04/25 11:30

Client ID: GZM-3

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Trichloroethene	ND		ug/l	0.50	0.18	1
1,2-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,3-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,4-Dichlorobenzene	ND		ug/l	2.5	0.70	1
Methyl tert butyl ether	ND		ug/l	2.5	0.17	1
p/m-Xylene	2.6		ug/l	2.5	0.70	1
o-Xylene	0.97	J	ug/l	2.5	0.70	1
Xylenes, Total	3.6	J	ug/l	2.5	0.70	1
cis-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1
1,2-Dichloroethene, Total	ND		ug/l	2.5	0.70	1
Dibromomethane	ND		ug/l	5.0	1.0	1
1,2,3-Trichloropropane	ND		ug/l	2.5	0.70	1
Acrylonitrile	ND		ug/l	5.0	1.5	1
Styrene	ND		ug/l	2.5	0.70	1
Dichlorodifluoromethane	ND		ug/l	5.0	1.0	1
Acetone	7.3		ug/l	5.0	1.5	1
Carbon disulfide	ND		ug/l	5.0	1.0	1
2-Butanone	ND		ug/l	5.0	1.9	1
Vinyl acetate	ND		ug/l	5.0	1.0	1
4-Methyl-2-pentanone	ND		ug/l	5.0	1.0	1
2-Hexanone	ND		ug/l	5.0	1.0	1
Bromochloromethane	ND		ug/l	2.5	0.70	1
2,2-Dichloropropane	ND		ug/l	2.5	0.70	1
1,2-Dibromoethane	ND		ug/l	2.0	0.65	1
1,3-Dichloropropane	ND		ug/l	2.5	0.70	1
1,1,1,2-Tetrachloroethane	ND		ug/l	2.5	0.70	1
Bromobenzene	ND		ug/l	2.5	0.70	1
n-Butylbenzene	ND		ug/l	2.5	0.70	1
sec-Butylbenzene	ND		ug/l	2.5	0.70	1
tert-Butylbenzene	ND		ug/l	2.5	0.70	1
o-Chlorotoluene	ND		ug/l	2.5	0.70	1
p-Chlorotoluene	ND		ug/l	2.5	0.70	1
1,2-Dibromo-3-chloropropane	ND		ug/l	2.5	0.70	1
Hexachlorobutadiene	ND		ug/l	2.5	0.70	1
Isopropylbenzene	ND		ug/l	2.5	0.70	1
p-Isopropyltoluene	ND		ug/l	2.5	0.70	1
Naphthalene	ND		ug/l	2.5	0.70	1



Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-03
 Client ID: GZM-3
 Sample Location: BRONX, NY

Date Collected: 11/04/25 11:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
n-Propylbenzene	ND		ug/l	2.5	0.70	1
1,2,3-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,3,5-Trimethylbenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trimethylbenzene	1.3	J	ug/l	2.5	0.70	1
1,4-Dioxane	ND		ug/l	250	61.	1
p-Diethylbenzene	ND		ug/l	2.0	0.70	1
p-Ethyltoluene	1.2	J	ug/l	2.0	0.70	1
1,2,4,5-Tetramethylbenzene	ND		ug/l	2.0	0.54	1
Ethyl ether	ND		ug/l	2.5	0.70	1
trans-1,4-Dichloro-2-butene	ND		ug/l	2.5	0.70	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
1,2-Dichloroethane-d4	88		70-130
Toluene-d8	101		70-130
4-Bromofluorobenzene	92		70-130
Dibromofluoromethane	101		70-130

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-04
 Client ID: GZM-4
 Sample Location: BRONX, NY

Date Collected: 11/04/25 12:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8260D
 Analytical Date: 11/09/25 19:48
 Analyst: MJV

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Methylene chloride	ND		ug/l	2.5	0.70	1
1,1-Dichloroethane	ND		ug/l	2.5	0.70	1
Chloroform	ND		ug/l	2.5	0.70	1
Carbon tetrachloride	ND		ug/l	0.50	0.13	1
1,2-Dichloropropane	ND		ug/l	1.0	0.14	1
Dibromochloromethane	ND		ug/l	0.50	0.15	1
1,1,2-Trichloroethane	ND		ug/l	1.5	0.50	1
Tetrachloroethene	ND		ug/l	0.50	0.18	1
Chlorobenzene	ND		ug/l	2.5	0.70	1
Trichlorofluoromethane	ND		ug/l	2.5	0.70	1
1,2-Dichloroethane	ND		ug/l	0.50	0.13	1
1,1,1-Trichloroethane	ND		ug/l	2.5	0.70	1
Bromodichloromethane	ND		ug/l	0.50	0.19	1
trans-1,3-Dichloropropene	ND		ug/l	0.50	0.16	1
cis-1,3-Dichloropropene	ND		ug/l	0.50	0.14	1
1,3-Dichloropropene, Total	ND		ug/l	0.50	0.14	1
1,1-Dichloropropene	ND		ug/l	2.5	0.70	1
Bromoform	ND		ug/l	2.0	0.65	1
1,1,1,2-Tetrachloroethane	ND		ug/l	0.50	0.17	1
Benzene	ND		ug/l	0.50	0.16	1
Toluene	ND		ug/l	2.5	0.70	1
Ethylbenzene	ND		ug/l	2.5	0.70	1
Chloromethane	ND		ug/l	2.5	0.70	1
Bromomethane	ND		ug/l	2.5	0.70	1
Vinyl chloride	ND		ug/l	1.0	0.07	1
Chloroethane	ND		ug/l	2.5	0.70	1
1,1-Dichloroethene	ND		ug/l	0.50	0.17	1
trans-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-04

Date Collected: 11/04/25 12:30

Client ID: GZM-4

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Trichloroethene	ND		ug/l	0.50	0.18	1
1,2-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,3-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,4-Dichlorobenzene	ND		ug/l	2.5	0.70	1
Methyl tert butyl ether	ND		ug/l	2.5	0.17	1
p/m-Xylene	ND		ug/l	2.5	0.70	1
o-Xylene	ND		ug/l	2.5	0.70	1
Xylenes, Total	ND		ug/l	2.5	0.70	1
cis-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1
1,2-Dichloroethene, Total	ND		ug/l	2.5	0.70	1
Dibromomethane	ND		ug/l	5.0	1.0	1
1,2,3-Trichloropropane	ND		ug/l	2.5	0.70	1
Acrylonitrile	ND		ug/l	5.0	1.5	1
Styrene	ND		ug/l	2.5	0.70	1
Dichlorodifluoromethane	ND		ug/l	5.0	1.0	1
Acetone	3.3	J	ug/l	5.0	1.5	1
Carbon disulfide	ND		ug/l	5.0	1.0	1
2-Butanone	ND		ug/l	5.0	1.9	1
Vinyl acetate	ND		ug/l	5.0	1.0	1
4-Methyl-2-pentanone	ND		ug/l	5.0	1.0	1
2-Hexanone	ND		ug/l	5.0	1.0	1
Bromochloromethane	ND		ug/l	2.5	0.70	1
2,2-Dichloropropane	ND		ug/l	2.5	0.70	1
1,2-Dibromoethane	ND		ug/l	2.0	0.65	1
1,3-Dichloropropane	ND		ug/l	2.5	0.70	1
1,1,1,2-Tetrachloroethane	ND		ug/l	2.5	0.70	1
Bromobenzene	ND		ug/l	2.5	0.70	1
n-Butylbenzene	ND		ug/l	2.5	0.70	1
sec-Butylbenzene	ND		ug/l	2.5	0.70	1
tert-Butylbenzene	ND		ug/l	2.5	0.70	1
o-Chlorotoluene	ND		ug/l	2.5	0.70	1
p-Chlorotoluene	ND		ug/l	2.5	0.70	1
1,2-Dibromo-3-chloropropane	ND		ug/l	2.5	0.70	1
Hexachlorobutadiene	ND		ug/l	2.5	0.70	1
Isopropylbenzene	ND		ug/l	2.5	0.70	1
p-Isopropyltoluene	ND		ug/l	2.5	0.70	1
Naphthalene	ND		ug/l	2.5	0.70	1



Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-04
 Client ID: GZM-4
 Sample Location: BRONX, NY

Date Collected: 11/04/25 12:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
n-Propylbenzene	ND		ug/l	2.5	0.70	1
1,2,3-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,3,5-Trimethylbenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trimethylbenzene	0.73	J	ug/l	2.5	0.70	1
1,4-Dioxane	ND		ug/l	250	61.	1
p-Diethylbenzene	ND		ug/l	2.0	0.70	1
p-Ethyltoluene	0.96	J	ug/l	2.0	0.70	1
1,2,4,5-Tetramethylbenzene	ND		ug/l	2.0	0.54	1
Ethyl ether	ND		ug/l	2.5	0.70	1
trans-1,4-Dichloro-2-butene	ND		ug/l	2.5	0.70	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
1,2-Dichloroethane-d4	87		70-130
Toluene-d8	102		70-130
4-Bromofluorobenzene	95		70-130
Dibromofluoromethane	100		70-130

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-05

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Analytical Method: 1,8260D

Analytical Date: 11/09/25 20:15

Analyst: MJV

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Methylene chloride	ND		ug/l	2.5	0.70	1
1,1-Dichloroethane	ND		ug/l	2.5	0.70	1
Chloroform	ND		ug/l	2.5	0.70	1
Carbon tetrachloride	ND		ug/l	0.50	0.13	1
1,2-Dichloropropane	ND		ug/l	1.0	0.14	1
Dibromochloromethane	ND		ug/l	0.50	0.15	1
1,1,2-Trichloroethane	ND		ug/l	1.5	0.50	1
Tetrachloroethene	ND		ug/l	0.50	0.18	1
Chlorobenzene	ND		ug/l	2.5	0.70	1
Trichlorofluoromethane	ND		ug/l	2.5	0.70	1
1,2-Dichloroethane	ND		ug/l	0.50	0.13	1
1,1,1-Trichloroethane	ND		ug/l	2.5	0.70	1
Bromodichloromethane	ND		ug/l	0.50	0.19	1
trans-1,3-Dichloropropene	ND		ug/l	0.50	0.16	1
cis-1,3-Dichloropropene	ND		ug/l	0.50	0.14	1
1,3-Dichloropropene, Total	ND		ug/l	0.50	0.14	1
1,1-Dichloropropene	ND		ug/l	2.5	0.70	1
Bromoform	ND		ug/l	2.0	0.65	1
1,1,2,2-Tetrachloroethane	ND		ug/l	0.50	0.17	1
Benzene	0.78		ug/l	0.50	0.16	1
Toluene	ND		ug/l	2.5	0.70	1
Ethylbenzene	ND		ug/l	2.5	0.70	1
Chloromethane	ND		ug/l	2.5	0.70	1
Bromomethane	ND		ug/l	2.5	0.70	1
Vinyl chloride	0.13	J	ug/l	1.0	0.07	1
Chloroethane	ND		ug/l	2.5	0.70	1
1,1-Dichloroethene	ND		ug/l	0.50	0.17	1
trans-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-05

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Trichloroethene	ND		ug/l	0.50	0.18	1
1,2-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,3-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,4-Dichlorobenzene	ND		ug/l	2.5	0.70	1
Methyl tert butyl ether	0.52	J	ug/l	2.5	0.17	1
p/m-Xylene	ND		ug/l	2.5	0.70	1
o-Xylene	ND		ug/l	2.5	0.70	1
Xylenes, Total	ND		ug/l	2.5	0.70	1
cis-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1
1,2-Dichloroethene, Total	ND		ug/l	2.5	0.70	1
Dibromomethane	ND		ug/l	5.0	1.0	1
1,2,3-Trichloropropane	ND		ug/l	2.5	0.70	1
Acrylonitrile	ND		ug/l	5.0	1.5	1
Styrene	ND		ug/l	2.5	0.70	1
Dichlorodifluoromethane	ND		ug/l	5.0	1.0	1
Acetone	ND		ug/l	5.0	1.5	1
Carbon disulfide	ND		ug/l	5.0	1.0	1
2-Butanone	ND		ug/l	5.0	1.9	1
Vinyl acetate	ND		ug/l	5.0	1.0	1
4-Methyl-2-pentanone	ND		ug/l	5.0	1.0	1
2-Hexanone	ND		ug/l	5.0	1.0	1
Bromochloromethane	ND		ug/l	2.5	0.70	1
2,2-Dichloropropane	ND		ug/l	2.5	0.70	1
1,2-Dibromoethane	ND		ug/l	2.0	0.65	1
1,3-Dichloropropane	ND		ug/l	2.5	0.70	1
1,1,1,2-Tetrachloroethane	ND		ug/l	2.5	0.70	1
Bromobenzene	ND		ug/l	2.5	0.70	1
n-Butylbenzene	ND		ug/l	2.5	0.70	1
sec-Butylbenzene	ND		ug/l	2.5	0.70	1
tert-Butylbenzene	ND		ug/l	2.5	0.70	1
o-Chlorotoluene	ND		ug/l	2.5	0.70	1
p-Chlorotoluene	ND		ug/l	2.5	0.70	1
1,2-Dibromo-3-chloropropane	ND		ug/l	2.5	0.70	1
Hexachlorobutadiene	ND		ug/l	2.5	0.70	1
Isopropylbenzene	ND		ug/l	2.5	0.70	1
p-Isopropyltoluene	ND		ug/l	2.5	0.70	1
Naphthalene	ND		ug/l	2.5	0.70	1



Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-05

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
n-Propylbenzene	ND		ug/l	2.5	0.70	1
1,2,3-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,3,5-Trimethylbenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trimethylbenzene	ND		ug/l	2.5	0.70	1
1,4-Dioxane	ND		ug/l	250	61.	1
p-Diethylbenzene	ND		ug/l	2.0	0.70	1
p-Ethyltoluene	ND		ug/l	2.0	0.70	1
1,2,4,5-Tetramethylbenzene	ND		ug/l	2.0	0.54	1
Ethyl ether	ND		ug/l	2.5	0.70	1
trans-1,4-Dichloro-2-butene	ND		ug/l	2.5	0.70	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
1,2-Dichloroethane-d4	87		70-130
Toluene-d8	103		70-130
4-Bromofluorobenzene	97		70-130
Dibromofluoromethane	100		70-130

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-06
 Client ID: FIELD BLANK
 Sample Location: BRONX, NY

Date Collected: 11/04/25 14:00
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8260D
 Analytical Date: 11/09/25 13:48
 Analyst: MJV

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Methylene chloride	ND		ug/l	2.5	0.70	1
1,1-Dichloroethane	ND		ug/l	2.5	0.70	1
Chloroform	ND		ug/l	2.5	0.70	1
Carbon tetrachloride	ND		ug/l	0.50	0.13	1
1,2-Dichloropropane	ND		ug/l	1.0	0.14	1
Dibromochloromethane	ND		ug/l	0.50	0.15	1
1,1,2-Trichloroethane	ND		ug/l	1.5	0.50	1
Tetrachloroethene	ND		ug/l	0.50	0.18	1
Chlorobenzene	ND		ug/l	2.5	0.70	1
Trichlorofluoromethane	ND		ug/l	2.5	0.70	1
1,2-Dichloroethane	ND		ug/l	0.50	0.13	1
1,1,1-Trichloroethane	ND		ug/l	2.5	0.70	1
Bromodichloromethane	ND		ug/l	0.50	0.19	1
trans-1,3-Dichloropropene	ND		ug/l	0.50	0.16	1
cis-1,3-Dichloropropene	ND		ug/l	0.50	0.14	1
1,3-Dichloropropene, Total	ND		ug/l	0.50	0.14	1
1,1-Dichloropropene	ND		ug/l	2.5	0.70	1
Bromoform	ND		ug/l	2.0	0.65	1
1,1,1,2-Tetrachloroethane	ND		ug/l	0.50	0.17	1
Benzene	ND		ug/l	0.50	0.16	1
Toluene	ND		ug/l	2.5	0.70	1
Ethylbenzene	ND		ug/l	2.5	0.70	1
Chloromethane	ND		ug/l	2.5	0.70	1
Bromomethane	ND		ug/l	2.5	0.70	1
Vinyl chloride	ND		ug/l	1.0	0.07	1
Chloroethane	ND		ug/l	2.5	0.70	1
1,1-Dichloroethene	ND		ug/l	0.50	0.17	1
trans-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-06

Date Collected: 11/04/25 14:00

Client ID: FIELD BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Trichloroethene	ND		ug/l	0.50	0.18	1
1,2-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,3-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,4-Dichlorobenzene	ND		ug/l	2.5	0.70	1
Methyl tert butyl ether	ND		ug/l	2.5	0.17	1
p/m-Xylene	ND		ug/l	2.5	0.70	1
o-Xylene	ND		ug/l	2.5	0.70	1
Xylenes, Total	ND		ug/l	2.5	0.70	1
cis-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1
1,2-Dichloroethene, Total	ND		ug/l	2.5	0.70	1
Dibromomethane	ND		ug/l	5.0	1.0	1
1,2,3-Trichloropropane	ND		ug/l	2.5	0.70	1
Acrylonitrile	ND		ug/l	5.0	1.5	1
Styrene	ND		ug/l	2.5	0.70	1
Dichlorodifluoromethane	ND		ug/l	5.0	1.0	1
Acetone	ND		ug/l	5.0	1.5	1
Carbon disulfide	ND		ug/l	5.0	1.0	1
2-Butanone	ND		ug/l	5.0	1.9	1
Vinyl acetate	ND		ug/l	5.0	1.0	1
4-Methyl-2-pentanone	ND		ug/l	5.0	1.0	1
2-Hexanone	ND		ug/l	5.0	1.0	1
Bromochloromethane	ND		ug/l	2.5	0.70	1
2,2-Dichloropropane	ND		ug/l	2.5	0.70	1
1,2-Dibromoethane	ND		ug/l	2.0	0.65	1
1,3-Dichloropropane	ND		ug/l	2.5	0.70	1
1,1,1,2-Tetrachloroethane	ND		ug/l	2.5	0.70	1
Bromobenzene	ND		ug/l	2.5	0.70	1
n-Butylbenzene	ND		ug/l	2.5	0.70	1
sec-Butylbenzene	ND		ug/l	2.5	0.70	1
tert-Butylbenzene	ND		ug/l	2.5	0.70	1
o-Chlorotoluene	ND		ug/l	2.5	0.70	1
p-Chlorotoluene	ND		ug/l	2.5	0.70	1
1,2-Dibromo-3-chloropropane	ND		ug/l	2.5	0.70	1
Hexachlorobutadiene	ND		ug/l	2.5	0.70	1
Isopropylbenzene	ND		ug/l	2.5	0.70	1
p-Isopropyltoluene	ND		ug/l	2.5	0.70	1
Naphthalene	ND		ug/l	2.5	0.70	1



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-06

Date Collected: 11/04/25 14:00

Client ID: FIELD BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
n-Propylbenzene	ND		ug/l	2.5	0.70	1
1,2,3-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,3,5-Trimethylbenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trimethylbenzene	ND		ug/l	2.5	0.70	1
1,4-Dioxane	ND		ug/l	250	61.	1
p-Diethylbenzene	ND		ug/l	2.0	0.70	1
p-Ethyltoluene	ND		ug/l	2.0	0.70	1
1,2,4,5-Tetramethylbenzene	ND		ug/l	2.0	0.54	1
Ethyl ether	ND		ug/l	2.5	0.70	1
trans-1,4-Dichloro-2-butene	ND		ug/l	2.5	0.70	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
1,2-Dichloroethane-d4	106		70-130
Toluene-d8	100		70-130
4-Bromofluorobenzene	95		70-130
Dibromofluoromethane	108		70-130

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-07

Date Collected: 11/04/25 00:00

Client ID: TRIP BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Analytical Method: 1,8260D

Analytical Date: 11/09/25 14:11

Analyst: MJV

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Methylene chloride	ND		ug/l	2.5	0.70	1
1,1-Dichloroethane	ND		ug/l	2.5	0.70	1
Chloroform	ND		ug/l	2.5	0.70	1
Carbon tetrachloride	ND		ug/l	0.50	0.13	1
1,2-Dichloropropane	ND		ug/l	1.0	0.14	1
Dibromochloromethane	ND		ug/l	0.50	0.15	1
1,1,2-Trichloroethane	ND		ug/l	1.5	0.50	1
Tetrachloroethene	ND		ug/l	0.50	0.18	1
Chlorobenzene	ND		ug/l	2.5	0.70	1
Trichlorofluoromethane	ND		ug/l	2.5	0.70	1
1,2-Dichloroethane	ND		ug/l	0.50	0.13	1
1,1,1-Trichloroethane	ND		ug/l	2.5	0.70	1
Bromodichloromethane	ND		ug/l	0.50	0.19	1
trans-1,3-Dichloropropene	ND		ug/l	0.50	0.16	1
cis-1,3-Dichloropropene	ND		ug/l	0.50	0.14	1
1,3-Dichloropropene, Total	ND		ug/l	0.50	0.14	1
1,1-Dichloropropene	ND		ug/l	2.5	0.70	1
Bromoform	ND		ug/l	2.0	0.65	1
1,1,1,2-Tetrachloroethane	ND		ug/l	0.50	0.17	1
Benzene	ND		ug/l	0.50	0.16	1
Toluene	ND		ug/l	2.5	0.70	1
Ethylbenzene	ND		ug/l	2.5	0.70	1
Chloromethane	ND		ug/l	2.5	0.70	1
Bromomethane	ND		ug/l	2.5	0.70	1
Vinyl chloride	ND		ug/l	1.0	0.07	1
Chloroethane	ND		ug/l	2.5	0.70	1
1,1-Dichloroethene	ND		ug/l	0.50	0.17	1
trans-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-07

Date Collected: 11/04/25 00:00

Client ID: TRIP BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
Trichloroethene	ND		ug/l	0.50	0.18	1
1,2-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,3-Dichlorobenzene	ND		ug/l	2.5	0.70	1
1,4-Dichlorobenzene	ND		ug/l	2.5	0.70	1
Methyl tert butyl ether	ND		ug/l	2.5	0.17	1
p/m-Xylene	ND		ug/l	2.5	0.70	1
o-Xylene	ND		ug/l	2.5	0.70	1
Xylenes, Total	ND		ug/l	2.5	0.70	1
cis-1,2-Dichloroethene	ND		ug/l	2.5	0.70	1
1,2-Dichloroethene, Total	ND		ug/l	2.5	0.70	1
Dibromomethane	ND		ug/l	5.0	1.0	1
1,2,3-Trichloropropane	ND		ug/l	2.5	0.70	1
Acrylonitrile	ND		ug/l	5.0	1.5	1
Styrene	ND		ug/l	2.5	0.70	1
Dichlorodifluoromethane	ND		ug/l	5.0	1.0	1
Acetone	ND		ug/l	5.0	1.5	1
Carbon disulfide	ND		ug/l	5.0	1.0	1
2-Butanone	ND		ug/l	5.0	1.9	1
Vinyl acetate	ND		ug/l	5.0	1.0	1
4-Methyl-2-pentanone	ND		ug/l	5.0	1.0	1
2-Hexanone	ND		ug/l	5.0	1.0	1
Bromochloromethane	ND		ug/l	2.5	0.70	1
2,2-Dichloropropane	ND		ug/l	2.5	0.70	1
1,2-Dibromoethane	ND		ug/l	2.0	0.65	1
1,3-Dichloropropane	ND		ug/l	2.5	0.70	1
1,1,1,2-Tetrachloroethane	ND		ug/l	2.5	0.70	1
Bromobenzene	ND		ug/l	2.5	0.70	1
n-Butylbenzene	ND		ug/l	2.5	0.70	1
sec-Butylbenzene	ND		ug/l	2.5	0.70	1
tert-Butylbenzene	ND		ug/l	2.5	0.70	1
o-Chlorotoluene	ND		ug/l	2.5	0.70	1
p-Chlorotoluene	ND		ug/l	2.5	0.70	1
1,2-Dibromo-3-chloropropane	ND		ug/l	2.5	0.70	1
Hexachlorobutadiene	ND		ug/l	2.5	0.70	1
Isopropylbenzene	ND		ug/l	2.5	0.70	1
p-Isopropyltoluene	ND		ug/l	2.5	0.70	1
Naphthalene	ND		ug/l	2.5	0.70	1



Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-07
 Client ID: TRIP BLANK
 Sample Location: BRONX, NY

Date Collected: 11/04/25 00:00
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Volatile Organics by GC/MS - Westborough Lab						
n-Propylbenzene	ND		ug/l	2.5	0.70	1
1,2,3-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trichlorobenzene	ND		ug/l	2.5	0.70	1
1,3,5-Trimethylbenzene	ND		ug/l	2.5	0.70	1
1,2,4-Trimethylbenzene	ND		ug/l	2.5	0.70	1
1,4-Dioxane	ND		ug/l	250	61.	1
p-Diethylbenzene	ND		ug/l	2.0	0.70	1
p-Ethyltoluene	ND		ug/l	2.0	0.70	1
1,2,4,5-Tetramethylbenzene	ND		ug/l	2.0	0.54	1
Ethyl ether	ND		ug/l	2.5	0.70	1
trans-1,4-Dichloro-2-butene	ND		ug/l	2.5	0.70	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
1,2-Dichloroethane-d4	108		70-130
Toluene-d8	98		70-130
4-Bromofluorobenzene	94		70-130
Dibromofluoromethane	107		70-130

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8260D
Analytical Date: 11/09/25 13:04
Analyst: PID

Parameter	Result	Qualifier	Units	RL	MDL
Volatile Organics by GC/MS - Westborough Lab for sample(s): 06-07 Batch: WG2139532-5					
Methylene chloride	ND		ug/l	2.5	0.70
1,1-Dichloroethane	ND		ug/l	2.5	0.70
Chloroform	ND		ug/l	2.5	0.70
Carbon tetrachloride	ND		ug/l	0.50	0.13
1,2-Dichloropropane	ND		ug/l	1.0	0.14
Dibromochloromethane	ND		ug/l	0.50	0.15
1,1,2-Trichloroethane	ND		ug/l	1.5	0.50
Tetrachloroethene	ND		ug/l	0.50	0.18
Chlorobenzene	ND		ug/l	2.5	0.70
Trichlorofluoromethane	ND		ug/l	2.5	0.70
1,2-Dichloroethane	ND		ug/l	0.50	0.13
1,1,1-Trichloroethane	ND		ug/l	2.5	0.70
Bromodichloromethane	ND		ug/l	0.50	0.19
trans-1,3-Dichloropropene	ND		ug/l	0.50	0.16
cis-1,3-Dichloropropene	ND		ug/l	0.50	0.14
1,3-Dichloropropene, Total	ND		ug/l	0.50	0.14
1,1-Dichloropropene	ND		ug/l	2.5	0.70
Bromoform	ND		ug/l	2.0	0.65
1,1,2,2-Tetrachloroethane	ND		ug/l	0.50	0.17
Benzene	ND		ug/l	0.50	0.16
Toluene	ND		ug/l	2.5	0.70
Ethylbenzene	ND		ug/l	2.5	0.70
Chloromethane	ND		ug/l	2.5	0.70
Bromomethane	ND		ug/l	2.5	0.70

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8260D
Analytical Date: 11/09/25 13:04
Analyst: PID

Parameter	Result	Qualifier	Units	RL	MDL
Volatile Organics by GC/MS - Westborough Lab for sample(s): 06-07 Batch: WG2139532-5					
Vinyl chloride	ND		ug/l	1.0	0.07
Chloroethane	ND		ug/l	2.5	0.70
1,1-Dichloroethene	ND		ug/l	0.50	0.17
trans-1,2-Dichloroethene	ND		ug/l	2.5	0.70
Trichloroethene	ND		ug/l	0.50	0.18
1,2-Dichlorobenzene	ND		ug/l	2.5	0.70
1,3-Dichlorobenzene	ND		ug/l	2.5	0.70
1,4-Dichlorobenzene	ND		ug/l	2.5	0.70
Methyl tert butyl ether	ND		ug/l	2.5	0.17
p/m-Xylene	ND		ug/l	2.5	0.70
o-Xylene	ND		ug/l	2.5	0.70
Xylenes, Total	ND		ug/l	2.5	0.70
cis-1,2-Dichloroethene	ND		ug/l	2.5	0.70
1,2-Dichloroethene, Total	ND		ug/l	2.5	0.70
Dibromomethane	ND		ug/l	5.0	1.0
1,2,3-Trichloropropane	ND		ug/l	2.5	0.70
Acrylonitrile	ND		ug/l	5.0	1.5
Styrene	ND		ug/l	2.5	0.70
Dichlorodifluoromethane	ND		ug/l	5.0	1.0
Acetone	ND		ug/l	5.0	1.5
Carbon disulfide	ND		ug/l	5.0	1.0
2-Butanone	ND		ug/l	5.0	1.9
Vinyl acetate	ND		ug/l	5.0	1.0
4-Methyl-2-pentanone	ND		ug/l	5.0	1.0

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8260D
Analytical Date: 11/09/25 13:04
Analyst: PID

Parameter	Result	Qualifier	Units	RL	MDL
Volatile Organics by GC/MS - Westborough Lab for sample(s): 06-07 Batch: WG2139532-5					
2-Hexanone	ND		ug/l	5.0	1.0
Bromochloromethane	ND		ug/l	2.5	0.70
2,2-Dichloropropane	ND		ug/l	2.5	0.70
1,2-Dibromoethane	ND		ug/l	2.0	0.65
1,3-Dichloropropane	ND		ug/l	2.5	0.70
1,1,1,2-Tetrachloroethane	ND		ug/l	2.5	0.70
Bromobenzene	ND		ug/l	2.5	0.70
n-Butylbenzene	ND		ug/l	2.5	0.70
sec-Butylbenzene	ND		ug/l	2.5	0.70
tert-Butylbenzene	ND		ug/l	2.5	0.70
o-Chlorotoluene	ND		ug/l	2.5	0.70
p-Chlorotoluene	ND		ug/l	2.5	0.70
1,2-Dibromo-3-chloropropane	ND		ug/l	2.5	0.70
Hexachlorobutadiene	ND		ug/l	2.5	0.70
Isopropylbenzene	ND		ug/l	2.5	0.70
p-Isopropyltoluene	ND		ug/l	2.5	0.70
Naphthalene	ND		ug/l	2.5	0.70
n-Propylbenzene	ND		ug/l	2.5	0.70
1,2,3-Trichlorobenzene	ND		ug/l	2.5	0.70
1,2,4-Trichlorobenzene	ND		ug/l	2.5	0.70
1,3,5-Trimethylbenzene	ND		ug/l	2.5	0.70
1,2,4-Trimethylbenzene	ND		ug/l	2.5	0.70
1,4-Dioxane	ND		ug/l	250	61.
p-Diethylbenzene	ND		ug/l	2.0	0.70

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8260D
Analytical Date: 11/09/25 13:04
Analyst: PID

Parameter	Result	Qualifier	Units	RL	MDL
Volatile Organics by GC/MS - Westborough Lab for sample(s): 06-07 Batch: WG2139532-5					
p-Ethyltoluene	ND		ug/l	2.0	0.70
1,2,4,5-Tetramethylbenzene	ND		ug/l	2.0	0.54
Ethyl ether	ND		ug/l	2.5	0.70
trans-1,4-Dichloro-2-butene	ND		ug/l	2.5	0.70

Surrogate	%Recovery	Qualifier	Acceptance Criteria
1,2-Dichloroethane-d4	106		70-130
Toluene-d8	100		70-130
4-Bromofluorobenzene	97		70-130
Dibromofluoromethane	107		70-130

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8260D
Analytical Date: 11/09/25 12:16
Analyst: PID

Parameter	Result	Qualifier	Units	RL	MDL
Volatile Organics by GC/MS - Westborough Lab for sample(s): 01-05 Batch: WG2139879-5					
Methylene chloride	ND		ug/l	2.5	0.70
1,1-Dichloroethane	ND		ug/l	2.5	0.70
Chloroform	ND		ug/l	2.5	0.70
Carbon tetrachloride	ND		ug/l	0.50	0.13
1,2-Dichloropropane	ND		ug/l	1.0	0.14
Dibromochloromethane	ND		ug/l	0.50	0.15
1,1,2-Trichloroethane	ND		ug/l	1.5	0.50
Tetrachloroethene	ND		ug/l	0.50	0.18
Chlorobenzene	ND		ug/l	2.5	0.70
Trichlorofluoromethane	ND		ug/l	2.5	0.70
1,2-Dichloroethane	ND		ug/l	0.50	0.13
1,1,1-Trichloroethane	ND		ug/l	2.5	0.70
Bromodichloromethane	ND		ug/l	0.50	0.19
trans-1,3-Dichloropropene	ND		ug/l	0.50	0.16
cis-1,3-Dichloropropene	ND		ug/l	0.50	0.14
1,3-Dichloropropene, Total	ND		ug/l	0.50	0.14
1,1-Dichloropropene	ND		ug/l	2.5	0.70
Bromoform	ND		ug/l	2.0	0.65
1,1,2,2-Tetrachloroethane	ND		ug/l	0.50	0.17
Benzene	ND		ug/l	0.50	0.16
Toluene	ND		ug/l	2.5	0.70
Ethylbenzene	ND		ug/l	2.5	0.70
Chloromethane	ND		ug/l	2.5	0.70
Bromomethane	ND		ug/l	2.5	0.70

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8260D
Analytical Date: 11/09/25 12:16
Analyst: PID

Parameter	Result	Qualifier	Units	RL	MDL
Volatile Organics by GC/MS - Westborough Lab for sample(s): 01-05 Batch: WG2139879-5					
Vinyl chloride	ND		ug/l	1.0	0.07
Chloroethane	ND		ug/l	2.5	0.70
1,1-Dichloroethene	ND		ug/l	0.50	0.17
trans-1,2-Dichloroethene	ND		ug/l	2.5	0.70
Trichloroethene	ND		ug/l	0.50	0.18
1,2-Dichlorobenzene	ND		ug/l	2.5	0.70
1,3-Dichlorobenzene	ND		ug/l	2.5	0.70
1,4-Dichlorobenzene	ND		ug/l	2.5	0.70
Methyl tert butyl ether	ND		ug/l	2.5	0.17
p/m-Xylene	ND		ug/l	2.5	0.70
o-Xylene	ND		ug/l	2.5	0.70
Xylenes, Total	ND		ug/l	2.5	0.70
cis-1,2-Dichloroethene	ND		ug/l	2.5	0.70
1,2-Dichloroethene, Total	ND		ug/l	2.5	0.70
Dibromomethane	ND		ug/l	5.0	1.0
1,2,3-Trichloropropane	ND		ug/l	2.5	0.70
Acrylonitrile	ND		ug/l	5.0	1.5
Styrene	ND		ug/l	2.5	0.70
Dichlorodifluoromethane	ND		ug/l	5.0	1.0
Acetone	ND		ug/l	5.0	1.5
Carbon disulfide	ND		ug/l	5.0	1.0
2-Butanone	ND		ug/l	5.0	1.9
Vinyl acetate	ND		ug/l	5.0	1.0
4-Methyl-2-pentanone	ND		ug/l	5.0	1.0

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8260D
Analytical Date: 11/09/25 12:16
Analyst: PID

Parameter	Result	Qualifier	Units	RL	MDL
Volatile Organics by GC/MS - Westborough Lab for sample(s): 01-05 Batch: WG2139879-5					
2-Hexanone	ND		ug/l	5.0	1.0
Bromochloromethane	ND		ug/l	2.5	0.70
2,2-Dichloropropane	ND		ug/l	2.5	0.70
1,2-Dibromoethane	ND		ug/l	2.0	0.65
1,3-Dichloropropane	ND		ug/l	2.5	0.70
1,1,1,2-Tetrachloroethane	ND		ug/l	2.5	0.70
Bromobenzene	ND		ug/l	2.5	0.70
n-Butylbenzene	ND		ug/l	2.5	0.70
sec-Butylbenzene	ND		ug/l	2.5	0.70
tert-Butylbenzene	ND		ug/l	2.5	0.70
o-Chlorotoluene	ND		ug/l	2.5	0.70
p-Chlorotoluene	ND		ug/l	2.5	0.70
1,2-Dibromo-3-chloropropane	ND		ug/l	2.5	0.70
Hexachlorobutadiene	ND		ug/l	2.5	0.70
Isopropylbenzene	ND		ug/l	2.5	0.70
p-Isopropyltoluene	ND		ug/l	2.5	0.70
Naphthalene	ND		ug/l	2.5	0.70
n-Propylbenzene	ND		ug/l	2.5	0.70
1,2,3-Trichlorobenzene	ND		ug/l	2.5	0.70
1,2,4-Trichlorobenzene	ND		ug/l	2.5	0.70
1,3,5-Trimethylbenzene	ND		ug/l	2.5	0.70
1,2,4-Trimethylbenzene	ND		ug/l	2.5	0.70
1,4-Dioxane	ND		ug/l	250	61.
p-Diethylbenzene	ND		ug/l	2.0	0.70

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8260D
Analytical Date: 11/09/25 12:16
Analyst: PID

Parameter	Result	Qualifier	Units	RL	MDL
Volatile Organics by GC/MS - Westborough Lab for sample(s): 01-05 Batch: WG2139879-5					
p-Ethyltoluene	ND		ug/l	2.0	0.70
1,2,4,5-Tetramethylbenzene	ND		ug/l	2.0	0.54
Ethyl ether	ND		ug/l	2.5	0.70
trans-1,4-Dichloro-2-butene	ND		ug/l	2.5	0.70

Surrogate	%Recovery	Qualifier	Acceptance Criteria
1,2-Dichloroethane-d4	86		70-130
Toluene-d8	102		70-130
4-Bromofluorobenzene	96		70-130
Dibromofluoromethane	100		70-130

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Volatile Organics by GC/MS - Westborough Lab Associated sample(s): 06-07 Batch: WG2139532-3 WG2139532-4								
Methylene chloride	99		100		70-130	1		20
1,1-Dichloroethane	110		100		70-130	10		20
Chloroform	100		100		70-130	0		20
Carbon tetrachloride	100		97		63-132	3		20
1,2-Dichloropropane	100		100		70-130	0		20
Dibromochloromethane	88		94		63-130	7		20
1,1,2-Trichloroethane	100		100		70-130	0		20
Tetrachloroethene	96		100		70-130	4		20
Chlorobenzene	100		100		75-130	0		20
Trichlorofluoromethane	100		100		62-150	0		20
1,2-Dichloroethane	100		98		70-130	2		20
1,1,1-Trichloroethane	100		98		67-130	2		20
Bromodichloromethane	96		96		67-130	0		20
trans-1,3-Dichloropropene	91		95		70-130	4		20
cis-1,3-Dichloropropene	88		88		70-130	0		20
1,1-Dichloropropene	94		94		70-130	0		20
Bromoform	85		92		54-136	8		20
1,1,2,2-Tetrachloroethane	97		110		67-130	13		20
Benzene	100		100		70-130	0		20

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Volatile Organics by GC/MS - Westborough Lab Associated sample(s): 06-07 Batch: WG2139532-3 WG2139532-4								
Toluene	100		100		70-130	0		20
Ethylbenzene	100		100		70-130	0		20
Chloromethane	98		96		64-130	2		20
Bromomethane	71		67		39-139	6		20
Vinyl chloride	110		100		55-140	10		20
Chloroethane	110		110		55-138	0		20
1,1-Dichloroethene	91		89		61-145	2		20
trans-1,2-Dichloroethene	99		96		70-130	3		20
Trichloroethene	100		100		70-130	0		20
1,2-Dichlorobenzene	94		100		70-130	6		20
1,3-Dichlorobenzene	97		100		70-130	3		20
1,4-Dichlorobenzene	94		100		70-130	6		20
Methyl tert butyl ether	81		84		63-130	4		20
p/m-Xylene	100		105		70-130	5		20
o-Xylene	100		100		70-130	0		20
cis-1,2-Dichloroethene	95		96		70-130	1		20
Dibromomethane	98		100		70-130	2		20
1,2,3-Trichloropropane	94		100		64-130	6		20
Acrylonitrile	100		100		70-130	0		20

Lab Control Sample Analysis

Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS		LCSD		%Recovery Limits	RPD	Qual	RPD Limits
	%Recovery	Qual	%Recovery	Qual				
Volatile Organics by GC/MS - Westborough Lab Associated sample(s): 06-07 Batch: WG2139532-3 WG2139532-4								
Styrene	100		105		70-130	5		20
Dichlorodifluoromethane	100		99		36-147	1		20
Acetone	82		85		58-148	4		20
Carbon disulfide	99		99		51-130	0		20
2-Butanone	95		100		63-138	5		20
Vinyl acetate	120		120		70-130	0		20
4-Methyl-2-pentanone	73		78		59-130	7		20
2-Hexanone	77		79		57-130	3		20
Bromochloromethane	97		98		70-130	1		20
2,2-Dichloropropane	99		98		63-133	1		20
1,2-Dibromoethane	90		96		70-130	6		20
1,3-Dichloropropane	96		100		70-130	4		20
1,1,1,2-Tetrachloroethane	95		99		64-130	4		20
Bromobenzene	90		97		70-130	7		20
n-Butylbenzene	100		110		53-136	10		20
sec-Butylbenzene	98		100		70-130	2		20
tert-Butylbenzene	93		98		70-130	5		20
o-Chlorotoluene	100		110		70-130	10		20
p-Chlorotoluene	96		100		70-130	4		20

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS		LCSD		%Recovery Limits	RPD	Qual	RPD Limits
	%Recovery	Qual	%Recovery	Qual				
Volatile Organics by GC/MS - Westborough Lab Associated sample(s): 06-07 Batch: WG2139532-3 WG2139532-4								
1,2-Dibromo-3-chloropropane	75		82		41-144	9		20
Hexachlorobutadiene	87		88		63-130	1		20
Isopropylbenzene	91		98		70-130	7		20
p-Isopropyltoluene	93		98		70-130	5		20
Naphthalene	72		77		70-130	7		20
n-Propylbenzene	99		110		69-130	11		20
1,2,3-Trichlorobenzene	82		88		70-130	7		20
1,2,4-Trichlorobenzene	83		89		70-130	7		20
1,3,5-Trimethylbenzene	96		100		64-130	4		20
1,2,4-Trimethylbenzene	95		100		70-130	5		20
1,4-Dioxane	84		86		56-162	2		20
p-Diethylbenzene	92		98		70-130	6		20
p-Ethyltoluene	95		100		70-130	5		20
1,2,4,5-Tetramethylbenzene	85		92		70-130	8		20
Ethyl ether	100		100		59-134	0		20
trans-1,4-Dichloro-2-butene	87		93		70-130	7		20

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
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Volatile Organics by GC/MS - Westborough Lab Associated sample(s): 06-07 Batch: WG2139532-3 WG2139532-4

Surrogate	LCS %Recovery	Qual	LCSD %Recovery	Qual	Acceptance Criteria
1,2-Dichloroethane-d4	107		104		70-130
Toluene-d8	100		101		70-130
4-Bromofluorobenzene	90		93		70-130
Dibromofluoromethane	104		105		70-130

Lab Control Sample Analysis

Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS		LCSD		%Recovery Limits	RPD	Qual	RPD Limits
	%Recovery	Qual	%Recovery	Qual				
Volatile Organics by GC/MS - Westborough Lab Associated sample(s): 01-05 Batch: WG2139879-3 WG2139879-4								
Methylene chloride	89		89		70-130	0		20
1,1-Dichloroethane	88		88		70-130	0		20
Chloroform	88		91		70-130	3		20
Carbon tetrachloride	93		92		63-132	1		20
1,2-Dichloropropane	89		89		70-130	0		20
Dibromochloromethane	91		92		63-130	1		20
1,1,2-Trichloroethane	92		93		70-130	1		20
Tetrachloroethene	100		100		70-130	0		20
Chlorobenzene	96		97		75-130	1		20
Trichlorofluoromethane	82		82		62-150	0		20
1,2-Dichloroethane	78		80		70-130	3		20
1,1,1-Trichloroethane	90		90		67-130	0		20
Bromodichloromethane	83		86		67-130	4		20
trans-1,3-Dichloropropene	94		94		70-130	0		20
cis-1,3-Dichloropropene	89		89		70-130	0		20
1,1-Dichloropropene	90		90		70-130	0		20
Bromoform	92		92		54-136	0		20
1,1,2,2-Tetrachloroethane	94		95		67-130	1		20
Benzene	94		94		70-130	0		20

Lab Control Sample Analysis

Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Volatile Organics by GC/MS - Westborough Lab Associated sample(s): 01-05 Batch: WG2139879-3 WG2139879-4								
Toluene	94		94		70-130	0		20
Ethylbenzene	90		90		70-130	0		20
Chloromethane	87		88		64-130	1		20
Bromomethane	59		63		39-139	7		20
Vinyl chloride	69		69		55-140	0		20
Chloroethane	59		59		55-138	0		20
1,1-Dichloroethene	98		97		61-145	1		20
trans-1,2-Dichloroethene	96		98		70-130	2		20
Trichloroethene	91		92		70-130	1		20
1,2-Dichlorobenzene	93		94		70-130	1		20
1,3-Dichlorobenzene	93		93		70-130	0		20
1,4-Dichlorobenzene	92		94		70-130	2		20
Methyl tert butyl ether	85		86		63-130	1		20
p/m-Xylene	90		90		70-130	0		20
o-Xylene	90		90		70-130	0		20
cis-1,2-Dichloroethene	97		98		70-130	1		20
Dibromomethane	84		85		70-130	1		20
1,2,3-Trichloropropane	88		88		64-130	0		20
Acrylonitrile	81		84		70-130	4		20

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Volatile Organics by GC/MS - Westborough Lab Associated sample(s): 01-05 Batch: WG2139879-3 WG2139879-4								
Styrene	90		90		70-130	0		20
Dichlorodifluoromethane	88		88		36-147	0		20
Acetone	86		87		58-148	1		20
Carbon disulfide	95		95		51-130	0		20
2-Butanone	92		90		63-138	2		20
Vinyl acetate	110		110		70-130	0		20
4-Methyl-2-pentanone	74		80		59-130	8		20
2-Hexanone	86		83		57-130	4		20
Bromochloromethane	98		98		70-130	0		20
2,2-Dichloropropane	87		86		63-133	1		20
1,2-Dibromoethane	96		97		70-130	1		20
1,3-Dichloropropane	96		90		70-130	6		20
1,1,1,2-Tetrachloroethane	91		92		64-130	1		20
Bromobenzene	96		98		70-130	2		20
n-Butylbenzene	91		91		53-136	0		20
sec-Butylbenzene	89		89		70-130	0		20
tert-Butylbenzene	88		89		70-130	1		20
o-Chlorotoluene	90		90		70-130	0		20
p-Chlorotoluene	91		91		70-130	0		20

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Volatile Organics by GC/MS - Westborough Lab Associated sample(s): 01-05 Batch: WG2139879-3 WG2139879-4								
1,2-Dibromo-3-chloropropane	87		86		41-144	1		20
Hexachlorobutadiene	97		100		63-130	3		20
Isopropylbenzene	93		93		70-130	0		20
p-Isopropyltoluene	89		89		70-130	0		20
Naphthalene	88		89		70-130	1		20
n-Propylbenzene	90		90		69-130	0		20
1,2,3-Trichlorobenzene	92		93		70-130	1		20
1,2,4-Trichlorobenzene	95		95		70-130	0		20
1,3,5-Trimethylbenzene	89		89		64-130	0		20
1,2,4-Trimethylbenzene	88		88		70-130	0		20
1,4-Dioxane	80		82		56-162	2		20
p-Diethylbenzene	86		86		70-130	0		20
p-Ethyltoluene	90		91		70-130	1		20
1,2,4,5-Tetramethylbenzene	84		85		70-130	1		20
Ethyl ether	85		88		59-134	3		20
trans-1,4-Dichloro-2-butene	81		76		70-130	6		20

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
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Volatile Organics by GC/MS - Westborough Lab Associated sample(s): 01-05 Batch: WG2139879-3 WG2139879-4

Surrogate	LCS %Recovery	Qual	LCSD %Recovery	Qual	Acceptance Criteria
1,2-Dichloroethane-d4	83		83		70-130
Toluene-d8	102		101		70-130
4-Bromofluorobenzene	96		96		70-130
Dibromofluoromethane	96		97		70-130

SEMIVOLATILES

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 3510C

Analytical Method: 1,8270E

Extraction Date: 11/06/25 19:23

Analytical Date: 11/07/25 01:06

Analyst: SMZ

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
1,2,4-Trichlorobenzene	ND		ug/l	5.0	0.98	1
Bis(2-chloroethyl)ether	ND		ug/l	2.0	0.39	1
1,2-Dichlorobenzene	ND		ug/l	2.0	0.33	1
1,3-Dichlorobenzene	ND		ug/l	2.0	0.32	1
1,4-Dichlorobenzene	ND		ug/l	2.0	0.39	1
3,3'-Dichlorobenzidine	ND		ug/l	5.0	1.8	1
2,4-Dinitrotoluene	ND		ug/l	5.0	0.54	1
2,6-Dinitrotoluene	ND		ug/l	5.0	0.84	1
4-Chlorophenyl phenyl ether	ND		ug/l	2.0	0.39	1
4-Bromophenyl phenyl ether	ND		ug/l	2.0	0.24	1
Bis(2-chloroisopropyl)ether	ND		ug/l	2.0	0.40	1
Bis(2-chloroethoxy)methane	ND		ug/l	5.0	0.84	1
Hexachlorocyclopentadiene	ND		ug/l	20	1.2	1
Isophorone	ND		ug/l	5.0	0.86	1
Nitrobenzene	ND		ug/l	2.0	0.20	1
NDPA/DPA	ND		ug/l	2.0	0.92	1
n-Nitrosodi-n-propylamine	ND		ug/l	5.0	0.91	1
Bis(2-ethylhexyl)phthalate	ND		ug/l	3.0	1.4	1
Butyl benzyl phthalate	ND		ug/l	5.0	2.6	1
Di-n-butylphthalate	ND		ug/l	5.0	0.96	1
Di-n-octylphthalate	ND		ug/l	5.0	2.3	1
Diethyl phthalate	ND		ug/l	5.0	0.76	1
Dimethyl phthalate	ND		ug/l	5.0	0.92	1
Biphenyl	ND		ug/l	2.0	0.20	1
4-Chloroaniline	ND		ug/l	5.0	0.47	1
2-Nitroaniline	ND		ug/l	5.0	1.0	1
3-Nitroaniline	ND		ug/l	5.0	1.2	1
4-Nitroaniline	ND		ug/l	5.0	1.4	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
Dibenzofuran	ND		ug/l	2.0	0.40	1
1,2,4,5-Tetrachlorobenzene	ND		ug/l	10	0.24	1
Acetophenone	ND		ug/l	5.0	0.92	1
2,4,6-Trichlorophenol	ND		ug/l	5.0	2.1	1
p-Chloro-m-cresol	ND		ug/l	2.0	0.61	1
2-Chlorophenol	ND		ug/l	2.0	0.65	1
2,4-Dichlorophenol	ND		ug/l	5.0	1.7	1
2,4-Dimethylphenol	ND		ug/l	5.0	2.0	1
2-Nitrophenol	ND		ug/l	10	2.0	1
4-Nitrophenol	ND		ug/l	10	1.4	1
2,4-Dinitrophenol	ND		ug/l	20	5.4	1
4,6-Dinitro-o-cresol	ND		ug/l	10	2.3	1
Phenol	ND		ug/l	5.0	0.35	1
2-Methylphenol	ND		ug/l	5.0	2.3	1
3-Methylphenol/4-Methylphenol	ND		ug/l	5.0	1.4	1
2,4,5-Trichlorophenol	ND		ug/l	5.0	2.1	1
Benzoic Acid	ND		ug/l	50	2.6	1
Benzyl Alcohol	ND		ug/l	2.0	0.38	1
Carbazole	ND		ug/l	2.0	0.31	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	57		21-120
Phenol-d6	37		10-120
Nitrobenzene-d5	72		23-120
2-Fluorobiphenyl	64		15-120
2,4,6-Tribromophenol	82		10-120
4-Terphenyl-d14	68		41-149

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-01
 Client ID: GZM-2
 Sample Location: BRONX, NY

Date Collected: 11/04/25 09:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8270E-SIM
 Analytical Date: 11/07/25 10:57
 Analyst: JJW

Extraction Method: EPA 3510C
 Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS-SIM - Westborough Lab						
Acenaphthene	ND		ug/l	0.10	0.02	1
2-Chloronaphthalene	ND		ug/l	0.20	0.02	1
Fluoranthene	ND		ug/l	0.10	0.03	1
Hexachlorobutadiene	ND		ug/l	0.50	0.02	1
Naphthalene	0.40		ug/l	0.10	0.02	1
Benzo(a)anthracene	0.11		ug/l	0.10	0.03	1
Benzo(a)pyrene	ND		ug/l	0.10	0.02	1
Benzo(b)fluoranthene	ND		ug/l	0.10	0.03	1
Benzo(k)fluoranthene	ND		ug/l	0.10	0.03	1
Chrysene	ND		ug/l	0.10	0.03	1
Acenaphthylene	ND		ug/l	0.10	0.02	1
Anthracene	ND		ug/l	0.10	0.02	1
Benzo(ghi)perylene	ND		ug/l	0.10	0.02	1
Fluorene	ND		ug/l	0.10	0.03	1
Phenanthrene	ND		ug/l	0.10	0.04	1
Dibenzo(a,h)anthracene	ND		ug/l	0.10	0.02	1
Indeno(1,2,3-cd)pyrene	ND		ug/l	0.10	0.02	1
Pyrene	ND		ug/l	0.10	0.04	1
2-Methylnaphthalene	0.05	J	ug/l	0.10	0.03	1
Pentachlorophenol	ND		ug/l	0.80	0.06	1
Hexachlorobenzene	ND		ug/l	0.80	0.01	1
Hexachloroethane	ND		ug/l	0.80	0.02	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
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Semivolatile Organics by GC/MS-SIM - Westborough Lab

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	53		21-120
Phenol-d6	41		10-120
Nitrobenzene-d5	99		23-120
2-Fluorobiphenyl	74		15-120
2,4,6-Tribromophenol	131	Q	10-120
4-Terphenyl-d14	75		41-149

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 1633

Analytical Method: 168,1633A

Extraction Date: 11/10/25 01:35

Analytical Date: 11/10/25 12:37

Analyst: JW

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Perfluoropentanoic Acid (PFPeA)	12.2		ng/l	3.17	0.586	1
Perfluorobutanesulfonic Acid (PFBS)	4.48		ng/l	1.58	0.507	1
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2FTS)	ND		ng/l	6.34	1.47	1
Perfluorohexanoic Acid (PFHxA)	11.1		ng/l	1.58	0.380	1
Perfluoropentanesulfonic Acid (PFPeS)	0.792	J	ng/l	1.58	0.364	1
Perfluoroheptanoic Acid (PFHpA)	8.00		ng/l	1.58	0.491	1
Perfluorohexanesulfonic Acid (PFHxS)	2.84		ng/l	1.58	0.269	1
Perfluorooctanoic Acid (PFOA)	25.0		ng/l	1.58	0.396	1
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2FTS)	ND		ng/l	6.34	3.22	1
Perfluoroheptanesulfonic Acid (PFHpS)	0.729	J	ng/l	1.58	0.491	1
Perfluorononanoic Acid (PFNA)	4.30		ng/l	1.58	0.238	1
Perfluorooctanesulfonic Acid (PFOS)	71.4		ng/l	1.58	0.269	1
Perfluorodecanoic Acid (PFDA)	1.14	J	ng/l	1.58	0.412	1
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2FTS)	ND		ng/l	6.34	3.03	1
Perfluorononanesulfonic Acid (PFNS)	ND		ng/l	1.58	0.238	1
N-Methyl Perfluorooctanesulfonamidoacetic Acid (NMeFOSAA)	ND		ng/l	1.58	1.14	1
Perfluoroundecanoic Acid (PFUnA)	ND		ng/l	1.58	0.380	1
Perfluorodecanesulfonic Acid (PFDS)	ND		ng/l	1.58	0.269	1
Perfluorooctanesulfonamide (PFOSA)	ND		ng/l	1.58	0.317	1
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (NEFOSAA)	ND		ng/l	1.58	0.729	1
Perfluorododecanoic Acid (PFDoA)	ND		ng/l	1.58	0.444	1
Perfluorotridecanoic Acid (PFTTrDA)	ND		ng/l	1.58	0.269	1
Perfluorotetradecanoic Acid (PFTeDA)	ND		ng/l	1.58	0.190	1
Hexafluoropropylene Oxide Dimer Acid (HFPO-DA)	ND		ng/l	6.34	0.935	1
4,8-Dioxa-3h-Perfluorononanoic Acid (ADONA)	ND		ng/l	6.34	1.24	1
Perfluorododecanesulfonic Acid (PFDoS)	ND		ng/l	1.58	0.333	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
9-Chlorohexadecafluoro-3-Oxanone-1-Sulfonic Acid (9Cl-PF3ONS)	ND		ng/l	6.34	0.697	1
11-Chloroeicosafluoro-3-Oxaundecane-1-Sulfonic Acid (11Cl-PF3OUdS)	ND		ng/l	6.34	0.475	1
N-Methyl Perfluorooctane Sulfonamide (NMeFOSA)	ND		ng/l	1.58	0.618	1
N-Ethyl Perfluorooctane Sulfonamide (NEtFOSA)	ND		ng/l	1.58	0.460	1
N-Methyl Perfluorooctanesulfonamido Ethanol (NMeFOSE)	ND		ng/l	15.8	1.68	1
N-Ethyl Perfluorooctanesulfonamido Ethanol (NEtFOSE)	ND		ng/l	15.8	1.39	1
Perfluoro-4-Methoxybutanoic Acid (PFMBA)	ND		ng/l	3.17	0.824	1
Perfluoro(2-Ethoxyethane)Sulfonic Acid (PFEEESA)	ND		ng/l	3.17	0.396	1
Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)	ND		ng/l	3.17	0.935	1
3-Perfluoropropyl Propanoic Acid (3:3FTCA)	ND		ng/l	7.92	2.04	1
2H,2H,3H,3H-Perfluorooctanoic Acid (5:3FTCA)	ND		ng/l	39.6	11.4	1
3-Perfluoroheptyl Propanoic Acid (7:3FTCA)	ND		ng/l	39.6	12.0	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Surrogate			% Recovery	Qualifier		Acceptance Criteria
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)			95			5-130
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)			86			40-130
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)			81			40-135
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)			339	Q		40-200
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)			96			40-130
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)			100			40-130
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)			83			40-130
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)			88			40-130
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)			374	Q		40-200
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)			88			40-130
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)			72			40-130
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)			81			40-130
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)			241			40-300
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)			121			40-170
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)			84			30-130
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)			62			40-130
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)			106			25-135
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)			66			10-130
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)			64			10-130
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)			110			40-130
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)			43			10-130
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)			44			10-130
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)			50			10-130
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)			53			10-130

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01 D

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 1633

Analytical Method: 168,1633A

Extraction Date: 11/10/25 01:35

Analytical Date: 11/10/25 16:44

Analyst: ANH

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
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Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab

Perfluorobutanoic Acid (PFBA)	ND		ng/l	31.7	11.8	5
Perfluoro-3-Methoxypropanoic Acid (PFMPA)	ND		ng/l	15.8	0.713	5

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01 D

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Surrogate			% Recovery	Qualifier	Acceptance Criteria	
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)			61		5-130	
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)			54		40-130	
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)			51		40-135	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)			117		40-200	
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)			52		40-130	
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)			50		40-130	
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)			52		40-130	
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)			54		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)			111		40-200	
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)			52		40-130	
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)			50		40-130	
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)			52		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)			63		40-300	
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)			47		40-170	
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)			50		30-130	
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)			40		40-130	
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)			49		25-135	
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)			40		10-130	
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)			37		10-130	
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)			49		40-130	
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)			32		10-130	
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)			31		10-130	
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)			36		10-130	
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)			40		10-130	

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 3510C

Analytical Method: 1,8270E

Extraction Date: 11/06/25 19:23

Analytical Date: 11/07/25 01:29

Analyst: SMZ

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
1,2,4-Trichlorobenzene	ND		ug/l	5.0	0.98	1
Bis(2-chloroethyl)ether	ND		ug/l	2.0	0.39	1
1,2-Dichlorobenzene	ND		ug/l	2.0	0.33	1
1,3-Dichlorobenzene	ND		ug/l	2.0	0.32	1
1,4-Dichlorobenzene	ND		ug/l	2.0	0.39	1
3,3'-Dichlorobenzidine	ND		ug/l	5.0	1.8	1
2,4-Dinitrotoluene	ND		ug/l	5.0	0.54	1
2,6-Dinitrotoluene	ND		ug/l	5.0	0.84	1
4-Chlorophenyl phenyl ether	ND		ug/l	2.0	0.39	1
4-Bromophenyl phenyl ether	ND		ug/l	2.0	0.24	1
Bis(2-chloroisopropyl)ether	ND		ug/l	2.0	0.40	1
Bis(2-chloroethoxy)methane	ND		ug/l	5.0	0.84	1
Hexachlorocyclopentadiene	ND		ug/l	20	1.2	1
Isophorone	ND		ug/l	5.0	0.86	1
Nitrobenzene	ND		ug/l	2.0	0.20	1
NDPA/DPA	ND		ug/l	2.0	0.92	1
n-Nitrosodi-n-propylamine	ND		ug/l	5.0	0.91	1
Bis(2-ethylhexyl)phthalate	ND		ug/l	3.0	1.4	1
Butyl benzyl phthalate	ND		ug/l	5.0	2.6	1
Di-n-butylphthalate	1.0	J	ug/l	5.0	0.96	1
Di-n-octylphthalate	ND		ug/l	5.0	2.3	1
Diethyl phthalate	ND		ug/l	5.0	0.76	1
Dimethyl phthalate	ND		ug/l	5.0	0.92	1
Biphenyl	ND		ug/l	2.0	0.20	1
4-Chloroaniline	ND		ug/l	5.0	0.47	1
2-Nitroaniline	ND		ug/l	5.0	1.0	1
3-Nitroaniline	ND		ug/l	5.0	1.2	1
4-Nitroaniline	ND		ug/l	5.0	1.4	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
Dibenzofuran	ND		ug/l	2.0	0.40	1
1,2,4,5-Tetrachlorobenzene	ND		ug/l	10	0.24	1
Acetophenone	ND		ug/l	5.0	0.92	1
2,4,6-Trichlorophenol	ND		ug/l	5.0	2.1	1
p-Chloro-m-cresol	ND		ug/l	2.0	0.61	1
2-Chlorophenol	ND		ug/l	2.0	0.65	1
2,4-Dichlorophenol	ND		ug/l	5.0	1.7	1
2,4-Dimethylphenol	3.5	J	ug/l	5.0	2.0	1
2-Nitrophenol	ND		ug/l	10	2.0	1
4-Nitrophenol	ND		ug/l	10	1.4	1
2,4-Dinitrophenol	ND		ug/l	20	5.4	1
4,6-Dinitro-o-cresol	ND		ug/l	10	2.3	1
Phenol	ND		ug/l	5.0	0.35	1
2-Methylphenol	ND		ug/l	5.0	2.3	1
3-Methylphenol/4-Methylphenol	3.9	J	ug/l	5.0	1.4	1
2,4,5-Trichlorophenol	ND		ug/l	5.0	2.1	1
Benzoic Acid	ND		ug/l	50	2.6	1
Benzyl Alcohol	ND		ug/l	2.0	0.38	1
Carbazole	ND		ug/l	2.0	0.31	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	62		21-120
Phenol-d6	44		10-120
Nitrobenzene-d5	78		23-120
2-Fluorobiphenyl	69		15-120
2,4,6-Tribromophenol	84		10-120
4-Terphenyl-d14	72		41-149

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-02
 Client ID: GZM-1
 Sample Location: BRONX, NY

Date Collected: 11/04/25 10:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8270E-SIM
 Analytical Date: 11/07/25 11:15
 Analyst: JJW

Extraction Method: EPA 3510C
 Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS-SIM - Westborough Lab						
Acenaphthene	0.24		ug/l	0.10	0.02	1
2-Chloronaphthalene	ND		ug/l	0.20	0.02	1
Fluoranthene	0.11		ug/l	0.10	0.03	1
Hexachlorobutadiene	ND		ug/l	0.50	0.02	1
Naphthalene	34		ug/l	0.10	0.02	1
Benzo(a)anthracene	ND		ug/l	0.10	0.03	1
Benzo(a)pyrene	ND		ug/l	0.10	0.02	1
Benzo(b)fluoranthene	ND		ug/l	0.10	0.03	1
Benzo(k)fluoranthene	ND		ug/l	0.10	0.03	1
Chrysene	ND		ug/l	0.10	0.03	1
Acenaphthylene	0.07	J	ug/l	0.10	0.02	1
Anthracene	0.05	J	ug/l	0.10	0.02	1
Benzo(ghi)perylene	ND		ug/l	0.10	0.02	1
Fluorene	0.26		ug/l	0.10	0.03	1
Phenanthrene	0.21		ug/l	0.10	0.04	1
Dibenzo(a,h)anthracene	ND		ug/l	0.10	0.02	1
Indeno(1,2,3-cd)pyrene	ND		ug/l	0.10	0.02	1
Pyrene	0.09	J	ug/l	0.10	0.04	1
2-Methylnaphthalene	12		ug/l	0.10	0.03	1
Pentachlorophenol	ND		ug/l	0.80	0.06	1
Hexachlorobenzene	ND		ug/l	0.80	0.01	1
Hexachloroethane	ND		ug/l	0.80	0.02	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
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Semivolatile Organics by GC/MS-SIM - Westborough Lab

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	57		21-120
Phenol-d6	43		10-120
Nitrobenzene-d5	106		23-120
2-Fluorobiphenyl	80		15-120
2,4,6-Tribromophenol	134	Q	10-120
4-Terphenyl-d14	75		41-149

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 1633

Analytical Method: 168,1633A

Extraction Date: 11/10/25 01:35

Analytical Date: 11/10/25 12:46

Analyst: JW

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Perfluorobutanoic Acid (PFBA)	10.3		ng/l	6.26	2.33	1
Perfluoropentanoic Acid (PFPeA)	21.8		ng/l	3.13	0.579	1
Perfluorobutanesulfonic Acid (PFBS)	5.01		ng/l	1.56	0.501	1
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2FTS)	ND		ng/l	6.26	1.46	1
Perfluorohexanoic Acid (PFHxA)	19.5		ng/l	1.56	0.376	1
Perfluoropentanesulfonic Acid (PFPeS)	ND		ng/l	1.56	0.360	1
Perfluoroheptanoic Acid (PFHpA)	5.31		ng/l	1.56	0.485	1
Perfluorohexanesulfonic Acid (PFHxS)	2.98		ng/l	1.56	0.266	1
Perfluorooctanoic Acid (PFOA)	17.0		ng/l	1.56	0.391	1
Perfluoroheptanesulfonic Acid (PFHpS)	ND		ng/l	1.56	0.485	1
Perfluorononanoic Acid (PFNA)	11.4		ng/l	1.56	0.235	1
Perfluorooctanesulfonic Acid (PFOS)	34.7		ng/l	1.56	0.266	1
Perfluorodecanoic Acid (PFDA)	1.47	J	ng/l	1.56	0.407	1
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2FTS)	ND		ng/l	6.26	2.99	1
Perfluorononanesulfonic Acid (PFNS)	ND		ng/l	1.56	0.235	1
N-Methyl Perfluorooctanesulfonamidoacetic Acid (NMeFOSAA)	ND		ng/l	1.56	1.13	1
Perfluoroundecanoic Acid (PFUnA)	ND		ng/l	1.56	0.376	1
Perfluorodecanesulfonic Acid (PFDS)	ND		ng/l	1.56	0.266	1
Perfluorooctanesulfonamide (PFOSA)	ND		ng/l	1.56	0.313	1
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (NEtFOSAA)	ND		ng/l	1.56	0.720	1
Perfluorododecanoic Acid (PFDoA)	ND		ng/l	1.56	0.438	1
Perfluorotridecanoic Acid (PFTTrDA)	ND		ng/l	1.56	0.266	1
Perfluorotetradecanoic Acid (PFTeDA)	ND		ng/l	1.56	0.188	1
Hexafluoropropylene Oxide Dimer Acid (HFPO-DA)	ND		ng/l	6.26	0.924	1
4,8-Dioxa-3h-Perfluorononanoic Acid (ADONA)	ND		ng/l	6.26	1.22	1
Perfluorododecanesulfonic Acid (PFDoS)	ND		ng/l	1.56	0.329	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
9-Chlorohexadecafluoro-3-Oxanone-1-Sulfonic Acid (9Cl-PF3ONS)	ND		ng/l	6.26	0.689	1
11-Chloroeicosafluoro-3-Oxaundecane-1-Sulfonic Acid (11Cl-PF3OUdS)	ND		ng/l	6.26	0.470	1
N-Methyl Perfluorooctane Sulfonamide (NMeFOSA)	ND		ng/l	1.56	0.611	1
N-Ethyl Perfluorooctane Sulfonamide (NEtFOSA)	ND		ng/l	1.56	0.454	1
N-Methyl Perfluorooctanesulfonamido Ethanol (NMeFOSE)	ND		ng/l	15.6	1.66	1
N-Ethyl Perfluorooctanesulfonamido Ethanol (NEtFOSE)	ND		ng/l	15.6	1.38	1
Perfluoro-3-Methoxypropanoic Acid (PFMPA)	0.376	J	ng/l	3.13	0.141	1
Perfluoro-4-Methoxybutanoic Acid (PFMBA)	ND		ng/l	3.13	0.814	1
Perfluoro(2-Ethoxyethane)Sulfonic Acid (PFEEESA)	ND		ng/l	3.13	0.391	1
Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)	ND		ng/l	3.13	0.924	1
3-Perfluoropropyl Propanoic Acid (3:3FTCA)	ND		ng/l	7.83	2.02	1
2H,2H,3H,3H-Perfluorooctanoic Acid (5:3FTCA)	ND		ng/l	39.1	11.2	1
3-Perfluoroheptyl Propanoic Acid (7:3FTCA)	ND		ng/l	39.1	11.8	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Surrogate			% Recovery	Qualifier		Acceptance Criteria
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)			108			5-130
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)			99			40-130
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)			103			40-135
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)			253	Q		40-200
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)			109			40-130
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)			111			40-130
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)			103			40-130
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)			103			40-130
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)			258	Q		40-200
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)			83			40-130
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)			95			40-130
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)			86			40-130
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)			113			40-300
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)			96			40-170
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)			80			30-130
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)			68			40-130
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)			91			25-135
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)			59			10-130
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)			59			10-130
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)			97			40-130
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)			66			10-130
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)			67			10-130
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)			57			10-130
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)			63			10-130

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02 D

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 1633

Analytical Method: 168,1633A

Extraction Date: 11/10/25 01:35

Analytical Date: 11/10/25 16:53

Analyst: ANH

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
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Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab

1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2FTS)	22.5	J	ng/l	31.3	15.9	5
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Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02 D

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Surrogate			% Recovery	Qualifier	Acceptance Criteria	
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)			65		5-130	
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)			64		40-130	
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)			62		40-135	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)			65		40-200	
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)			69		40-130	
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)			70		40-130	
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)			62		40-130	
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)			63		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)			67		40-200	
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)			56		40-130	
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)			56		40-130	
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)			50		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)			54		40-300	
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)			48		40-170	
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)			50		30-130	
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)			46		40-130	
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)			42		25-135	
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)			41		10-130	
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)			41		10-130	
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)			61		40-130	
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)			44		10-130	
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)			40		10-130	
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)			39		10-130	
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)			42		10-130	

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-03
 Client ID: GZM-3
 Sample Location: BRONX, NY

Date Collected: 11/04/25 11:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8270E
 Analytical Date: 11/07/25 01:52
 Analyst: SMZ

Extraction Method: EPA 3510C
 Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
1,2,4-Trichlorobenzene	ND		ug/l	5.0	0.98	1
Bis(2-chloroethyl)ether	ND		ug/l	2.0	0.39	1
1,2-Dichlorobenzene	ND		ug/l	2.0	0.33	1
1,3-Dichlorobenzene	ND		ug/l	2.0	0.32	1
1,4-Dichlorobenzene	ND		ug/l	2.0	0.39	1
3,3'-Dichlorobenzidine	ND		ug/l	5.0	1.8	1
2,4-Dinitrotoluene	ND		ug/l	5.0	0.54	1
2,6-Dinitrotoluene	ND		ug/l	5.0	0.84	1
4-Chlorophenyl phenyl ether	ND		ug/l	2.0	0.39	1
4-Bromophenyl phenyl ether	ND		ug/l	2.0	0.24	1
Bis(2-chloroisopropyl)ether	ND		ug/l	2.0	0.40	1
Bis(2-chloroethoxy)methane	ND		ug/l	5.0	0.84	1
Hexachlorocyclopentadiene	ND		ug/l	20	1.2	1
Isophorone	ND		ug/l	5.0	0.86	1
Nitrobenzene	ND		ug/l	2.0	0.20	1
NDPA/DPA	ND		ug/l	2.0	0.92	1
n-Nitrosodi-n-propylamine	ND		ug/l	5.0	0.91	1
Bis(2-ethylhexyl)phthalate	ND		ug/l	3.0	1.4	1
Butyl benzyl phthalate	ND		ug/l	5.0	2.6	1
Di-n-butylphthalate	ND		ug/l	5.0	0.96	1
Di-n-octylphthalate	ND		ug/l	5.0	2.3	1
Diethyl phthalate	ND		ug/l	5.0	0.76	1
Dimethyl phthalate	ND		ug/l	5.0	0.92	1
Biphenyl	ND		ug/l	2.0	0.20	1
4-Chloroaniline	ND		ug/l	5.0	0.47	1
2-Nitroaniline	ND		ug/l	5.0	1.0	1
3-Nitroaniline	ND		ug/l	5.0	1.2	1
4-Nitroaniline	ND		ug/l	5.0	1.4	1

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-03
 Client ID: GZM-3
 Sample Location: BRONX, NY

Date Collected: 11/04/25 11:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
Dibenzofuran	ND		ug/l	2.0	0.40	1
1,2,4,5-Tetrachlorobenzene	ND		ug/l	10	0.24	1
Acetophenone	ND		ug/l	5.0	0.92	1
2,4,6-Trichlorophenol	ND		ug/l	5.0	2.1	1
p-Chloro-m-cresol	ND		ug/l	2.0	0.61	1
2-Chlorophenol	ND		ug/l	2.0	0.65	1
2,4-Dichlorophenol	ND		ug/l	5.0	1.7	1
2,4-Dimethylphenol	ND		ug/l	5.0	2.0	1
2-Nitrophenol	ND		ug/l	10	2.0	1
4-Nitrophenol	ND		ug/l	10	1.4	1
2,4-Dinitrophenol	ND		ug/l	20	5.4	1
4,6-Dinitro-o-cresol	ND		ug/l	10	2.3	1
Phenol	ND		ug/l	5.0	0.35	1
2-Methylphenol	ND		ug/l	5.0	2.3	1
3-Methylphenol/4-Methylphenol	ND		ug/l	5.0	1.4	1
2,4,5-Trichlorophenol	ND		ug/l	5.0	2.1	1
Benzoic Acid	12.	J	ug/l	50	2.6	1
Benzyl Alcohol	ND		ug/l	2.0	0.38	1
Carbazole	ND		ug/l	2.0	0.31	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	52		21-120
Phenol-d6	39		10-120
Nitrobenzene-d5	67		23-120
2-Fluorobiphenyl	66		15-120
2,4,6-Tribromophenol	78		10-120
4-Terphenyl-d14	66		41-149

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-03
 Client ID: GZM-3
 Sample Location: BRONX, NY

Date Collected: 11/04/25 11:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8270E-SIM
 Analytical Date: 11/07/25 11:32
 Analyst: JJW

Extraction Method: EPA 3510C
 Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS-SIM - Westborough Lab						
Acenaphthene	0.11		ug/l	0.10	0.02	1
2-Chloronaphthalene	ND		ug/l	0.20	0.02	1
Fluoranthene	0.05	J	ug/l	0.10	0.03	1
Hexachlorobutadiene	ND		ug/l	0.50	0.02	1
Naphthalene	0.99		ug/l	0.10	0.02	1
Benzo(a)anthracene	ND		ug/l	0.10	0.03	1
Benzo(a)pyrene	ND		ug/l	0.10	0.02	1
Benzo(b)fluoranthene	ND		ug/l	0.10	0.03	1
Benzo(k)fluoranthene	ND		ug/l	0.10	0.03	1
Chrysene	ND		ug/l	0.10	0.03	1
Acenaphthylene	ND		ug/l	0.10	0.02	1
Anthracene	0.03	J	ug/l	0.10	0.02	1
Benzo(ghi)perylene	ND		ug/l	0.10	0.02	1
Fluorene	ND		ug/l	0.10	0.03	1
Phenanthrene	0.06	J	ug/l	0.10	0.04	1
Dibenzo(a,h)anthracene	ND		ug/l	0.10	0.02	1
Indeno(1,2,3-cd)pyrene	ND		ug/l	0.10	0.02	1
Pyrene	0.05	J	ug/l	0.10	0.04	1
2-Methylnaphthalene	0.14		ug/l	0.10	0.03	1
Pentachlorophenol	ND		ug/l	0.80	0.06	1
Hexachlorobenzene	ND		ug/l	0.80	0.01	1
Hexachloroethane	ND		ug/l	0.80	0.02	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-03

Date Collected: 11/04/25 11:30

Client ID: GZM-3

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
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Semivolatile Organics by GC/MS-SIM - Westborough Lab

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	57		21-120
Phenol-d6	44		10-120
Nitrobenzene-d5	104		23-120
2-Fluorobiphenyl	79		15-120
2,4,6-Tribromophenol	134	Q	10-120
4-Terphenyl-d14	74		41-149

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-03 R

Date Collected: 11/04/25 11:30

Client ID: GZM-3

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 1633

Analytical Method: 168,1633A

Extraction Date: 11/10/25 01:35

Analytical Date: 11/10/25 15:56

Analyst: JW

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Perfluorobutanoic Acid (PFBA)	12.2		ng/l	5.74	2.14	1
Perfluoropentanoic Acid (PFPeA)	15.7		ng/l	2.87	0.531	1
Perfluorobutanesulfonic Acid (PFBS)	8.77		ng/l	1.43	0.459	1
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2FTS)	ND		ng/l	5.74	1.33	1
Perfluorohexanoic Acid (PFHxA)	12.9		ng/l	1.43	0.344	1
Perfluoropentanesulfonic Acid (PFPeS)	0.344	JF	ng/l	1.43	0.330	1
Perfluoroheptanoic Acid (PFHpA)	6.44		ng/l	1.43	0.445	1
Perfluorohexanesulfonic Acid (PFHxS)	2.35		ng/l	1.43	0.244	1
Perfluorooctanoic Acid (PFOA)	21.3		ng/l	1.43	0.359	1
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2FTS)	3.27	J	ng/l	5.74	2.91	1
Perfluoroheptanesulfonic Acid (PFHpS)	ND		ng/l	1.43	0.445	1
Perfluorononanoic Acid (PFNA)	3.73		ng/l	1.43	0.215	1
Perfluorooctanesulfonic Acid (PFOS)	31.0		ng/l	1.43	0.244	1
Perfluorodecanoic Acid (PFDA)	1.71		ng/l	1.43	0.373	1
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2FTS)	ND		ng/l	5.74	2.74	1
Perfluorononanesulfonic Acid (PFNS)	ND		ng/l	1.43	0.215	1
N-Methyl Perfluorooctanesulfonamidoacetic Acid (NMeFOSAA)	ND		ng/l	1.43	1.03	1
Perfluoroundecanoic Acid (PFUnA)	ND		ng/l	1.43	0.344	1
Perfluorodecanesulfonic Acid (PFDS)	ND		ng/l	1.43	0.244	1
Perfluorooctanesulfonamide (PFOSA)	ND		ng/l	1.43	0.287	1
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (NEtFOSAA)	ND		ng/l	1.43	0.660	1
Perfluorododecanoic Acid (PFDoA)	ND		ng/l	1.43	0.402	1
Perfluorotridecanoic Acid (PFTrDA)	ND		ng/l	1.43	0.244	1
Perfluorotetradecanoic Acid (PFTeDA)	ND		ng/l	1.43	0.172	1
Hexafluoropropylene Oxide Dimer Acid (HFPO-DA)	ND		ng/l	5.74	0.846	1
4,8-Dioxa-3h-Perfluorononanoic Acid (ADONA)	ND		ng/l	5.74	1.12	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-03 R

Date Collected: 11/04/25 11:30

Client ID: GZM-3

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Perfluorododecanesulfonic Acid (PFDoS)	ND		ng/l	1.43	0.301	1
9-Chlorohexadecafluoro-3-Oxanone-1-Sulfonic Acid (9Cl-PF3ONS)	ND		ng/l	5.74	0.631	1
11-Chloroeicosafluoro-3-Oxaundecane-1-Sulfonic Acid (11Cl-PF3OUdS)	ND		ng/l	5.74	0.430	1
N-Methyl Perfluorooctane Sulfonamide (NMeFOSA)	ND		ng/l	1.43	0.560	1
N-Ethyl Perfluorooctane Sulfonamide (NEtFOSA)	ND		ng/l	1.43	0.416	1
N-Methyl Perfluorooctanesulfonamido Ethanol (NMeFOSE)	ND		ng/l	14.3	1.52	1
N-Ethyl Perfluorooctanesulfonamido Ethanol (NEtFOSE)	ND		ng/l	14.3	1.26	1
Perfluoro-3-Methoxypropanoic Acid (PFMPA)	ND		ng/l	2.87	0.129	1
Perfluoro-4-Methoxybutanoic Acid (PFMBA)	ND		ng/l	2.87	0.746	1
Perfluoro(2-Ethoxyethane)Sulfonic Acid (PFEEESA)	ND		ng/l	2.87	0.359	1
Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)	ND		ng/l	2.87	0.846	1
3-Perfluoropropyl Propanoic Acid (3:3FTCA)	ND		ng/l	7.17	1.85	1
2H,2H,3H,3H-Perfluorooctanoic Acid (5:3FTCA)	ND		ng/l	35.9	10.3	1
3-Perfluoroheptyl Propanoic Acid (7:3FTCA)	ND		ng/l	35.9	10.8	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-03 R

Date Collected: 11/04/25 11:30

Client ID: GZM-3

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Surrogate			% Recovery	Qualifier	Acceptance Criteria	
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)			65		5-130	
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)			55		40-130	
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)			49		40-135	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)			118		40-200	
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)			59		40-130	
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)			57		40-130	
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)			52		40-130	
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)			47		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)			115		40-200	
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)			49		40-130	
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)			50		40-130	
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)			53		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)			49		40-300	
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)			42		40-170	
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)			52		30-130	
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)			40		40-130	
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)			43		25-135	
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)			33		10-130	
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)			32		10-130	
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)			54		40-130	
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)			42		10-130	
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)			38		10-130	
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)			33		10-130	
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)			34		10-130	

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-04
 Client ID: GZM-4
 Sample Location: BRONX, NY

Date Collected: 11/04/25 12:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8270E
 Analytical Date: 11/07/25 02:15
 Analyst: SMZ

Extraction Method: EPA 3510C
 Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
1,2,4-Trichlorobenzene	ND		ug/l	5.0	0.98	1
Bis(2-chloroethyl)ether	ND		ug/l	2.0	0.39	1
1,2-Dichlorobenzene	ND		ug/l	2.0	0.33	1
1,3-Dichlorobenzene	ND		ug/l	2.0	0.32	1
1,4-Dichlorobenzene	ND		ug/l	2.0	0.39	1
3,3'-Dichlorobenzidine	ND		ug/l	5.0	1.8	1
2,4-Dinitrotoluene	ND		ug/l	5.0	0.54	1
2,6-Dinitrotoluene	ND		ug/l	5.0	0.84	1
4-Chlorophenyl phenyl ether	ND		ug/l	2.0	0.39	1
4-Bromophenyl phenyl ether	ND		ug/l	2.0	0.24	1
Bis(2-chloroisopropyl)ether	ND		ug/l	2.0	0.40	1
Bis(2-chloroethoxy)methane	ND		ug/l	5.0	0.84	1
Hexachlorocyclopentadiene	ND		ug/l	20	1.2	1
Isophorone	ND		ug/l	5.0	0.86	1
Nitrobenzene	ND		ug/l	2.0	0.20	1
NDPA/DPA	ND		ug/l	2.0	0.92	1
n-Nitrosodi-n-propylamine	ND		ug/l	5.0	0.91	1
Bis(2-ethylhexyl)phthalate	ND		ug/l	3.0	1.4	1
Butyl benzyl phthalate	ND		ug/l	5.0	2.6	1
Di-n-butylphthalate	ND		ug/l	5.0	0.96	1
Di-n-octylphthalate	ND		ug/l	5.0	2.3	1
Diethyl phthalate	ND		ug/l	5.0	0.76	1
Dimethyl phthalate	ND		ug/l	5.0	0.92	1
Biphenyl	ND		ug/l	2.0	0.20	1
4-Chloroaniline	ND		ug/l	5.0	0.47	1
2-Nitroaniline	ND		ug/l	5.0	1.0	1
3-Nitroaniline	ND		ug/l	5.0	1.2	1
4-Nitroaniline	ND		ug/l	5.0	1.4	1

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-04
 Client ID: GZM-4
 Sample Location: BRONX, NY

Date Collected: 11/04/25 12:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
Dibenzofuran	ND		ug/l	2.0	0.40	1
1,2,4,5-Tetrachlorobenzene	ND		ug/l	10	0.24	1
Acetophenone	ND		ug/l	5.0	0.92	1
2,4,6-Trichlorophenol	ND		ug/l	5.0	2.1	1
p-Chloro-m-cresol	ND		ug/l	2.0	0.61	1
2-Chlorophenol	ND		ug/l	2.0	0.65	1
2,4-Dichlorophenol	ND		ug/l	5.0	1.7	1
2,4-Dimethylphenol	ND		ug/l	5.0	2.0	1
2-Nitrophenol	ND		ug/l	10	2.0	1
4-Nitrophenol	ND		ug/l	10	1.4	1
2,4-Dinitrophenol	ND		ug/l	20	5.4	1
4,6-Dinitro-o-cresol	ND		ug/l	10	2.3	1
Phenol	ND		ug/l	5.0	0.35	1
2-Methylphenol	ND		ug/l	5.0	2.3	1
3-Methylphenol/4-Methylphenol	ND		ug/l	5.0	1.4	1
2,4,5-Trichlorophenol	ND		ug/l	5.0	2.1	1
Benzoic Acid	ND		ug/l	50	2.6	1
Benzyl Alcohol	ND		ug/l	2.0	0.38	1
Carbazole	ND		ug/l	2.0	0.31	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	56		21-120
Phenol-d6	42		10-120
Nitrobenzene-d5	78		23-120
2-Fluorobiphenyl	61		15-120
2,4,6-Tribromophenol	82		10-120
4-Terphenyl-d14	77		41-149

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-04
 Client ID: GZM-4
 Sample Location: BRONX, NY

Date Collected: 11/04/25 12:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8270E-SIM
 Analytical Date: 11/07/25 11:50
 Analyst: JJW

Extraction Method: EPA 3510C
 Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS-SIM - Westborough Lab						
Acenaphthene	0.12		ug/l	0.10	0.02	1
2-Chloronaphthalene	ND		ug/l	0.20	0.02	1
Fluoranthene	0.44		ug/l	0.10	0.03	1
Hexachlorobutadiene	ND		ug/l	0.50	0.02	1
Naphthalene	0.43		ug/l	0.10	0.02	1
Benzo(a)anthracene	0.27		ug/l	0.10	0.03	1
Benzo(a)pyrene	0.20		ug/l	0.10	0.02	1
Benzo(b)fluoranthene	0.26		ug/l	0.10	0.03	1
Benzo(k)fluoranthene	0.10		ug/l	0.10	0.03	1
Chrysene	0.17		ug/l	0.10	0.03	1
Acenaphthylene	ND		ug/l	0.10	0.02	1
Anthracene	0.09	J	ug/l	0.10	0.02	1
Benzo(ghi)perylene	0.15		ug/l	0.10	0.02	1
Fluorene	0.07	J	ug/l	0.10	0.03	1
Phenanthrene	0.27		ug/l	0.10	0.04	1
Dibenzo(a,h)anthracene	0.04	J	ug/l	0.10	0.02	1
Indeno(1,2,3-cd)pyrene	0.16		ug/l	0.10	0.02	1
Pyrene	0.39		ug/l	0.10	0.04	1
2-Methylnaphthalene	0.08	J	ug/l	0.10	0.03	1
Pentachlorophenol	ND		ug/l	0.80	0.06	1
Hexachlorobenzene	ND		ug/l	0.80	0.01	1
Hexachloroethane	ND		ug/l	0.80	0.02	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-04

Date Collected: 11/04/25 12:30

Client ID: GZM-4

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
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Semivolatile Organics by GC/MS-SIM - Westborough Lab

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	53		21-120
Phenol-d6	40		10-120
Nitrobenzene-d5	99		23-120
2-Fluorobiphenyl	74		15-120
2,4,6-Tribromophenol	118		10-120
4-Terphenyl-d14	68		41-149

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-04

Date Collected: 11/04/25 12:30

Client ID: GZM-4

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 1633

Analytical Method: 168,1633A

Extraction Date: 11/10/25 01:35

Analytical Date: 11/10/25 13:04

Analyst: JW

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Perfluorobutanoic Acid (PFBA)	8.60		ng/l	5.79	2.16	1
Perfluoropentanoic Acid (PFPeA)	6.15		ng/l	2.89	0.535	1
Perfluorobutanesulfonic Acid (PFBS)	3.42		ng/l	1.45	0.463	1
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2FTS)	ND		ng/l	5.79	1.34	1
Perfluorohexanoic Acid (PFHxA)	8.24		ng/l	1.45	0.347	1
Perfluoropentanesulfonic Acid (PFPeS)	ND		ng/l	1.45	0.333	1
Perfluoroheptanoic Acid (PFHpA)	2.56		ng/l	1.45	0.449	1
Perfluorohexanesulfonic Acid (PFHxS)	0.738	J	ng/l	1.45	0.246	1
Perfluorooctanoic Acid (PFOA)	4.24		ng/l	1.45	0.362	1
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2FTS)	ND		ng/l	5.79	2.94	1
Perfluoroheptanesulfonic Acid (PFHpS)	ND		ng/l	1.45	0.449	1
Perfluorononanoic Acid (PFNA)	1.32	J	ng/l	1.45	0.217	1
Perfluorooctanesulfonic Acid (PFOS)	9.75		ng/l	1.45	0.246	1
Perfluorodecanoic Acid (PFDA)	0.970	J	ng/l	1.45	0.376	1
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2FTS)	ND		ng/l	5.79	2.76	1
Perfluorononanesulfonic Acid (PFNS)	ND		ng/l	1.45	0.217	1
N-Methyl Perfluorooctanesulfonamidoacetic Acid (NMeFOSAA)	ND		ng/l	1.45	1.04	1
Perfluoroundecanoic Acid (PFUnA)	ND		ng/l	1.45	0.347	1
Perfluorodecanesulfonic Acid (PFDS)	ND		ng/l	1.45	0.246	1
Perfluorooctanesulfonamide (PFOSA)	ND		ng/l	1.45	0.289	1
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (NEtFOSAA)	ND		ng/l	1.45	0.666	1
Perfluorododecanoic Acid (PFDoA)	ND		ng/l	1.45	0.405	1
Perfluorotridecanoic Acid (PFTrDA)	ND		ng/l	1.45	0.246	1
Perfluorotetradecanoic Acid (PFTeDA)	ND		ng/l	1.45	0.174	1
Hexafluoropropylene Oxide Dimer Acid (HFPO-DA)	ND		ng/l	5.79	0.854	1
4,8-Dioxa-3h-Perfluorononanoic Acid (ADONA)	ND		ng/l	5.79	1.13	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-04

Date Collected: 11/04/25 12:30

Client ID: GZM-4

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Perfluorododecanesulfonic Acid (PFDoS)	ND		ng/l	1.45	0.304	1
9-Chlorohexadecafluoro-3-Oxanone-1-Sulfonic Acid (9Cl-PF3ONS)	ND		ng/l	5.79	0.637	1
11-Chloroeicosafuoro-3-Oxaundecane-1-Sulfonic Acid (11Cl-PF3OUdS)	ND		ng/l	5.79	0.434	1
N-Methyl Perfluorooctane Sulfonamide (NMeFOSA)	ND		ng/l	1.45	0.564	1
N-Ethyl Perfluorooctane Sulfonamide (NEtFOSA)	ND		ng/l	1.45	0.420	1
N-Methyl Perfluorooctanesulfonamido Ethanol (NMeFOSE)	ND		ng/l	14.5	1.53	1
N-Ethyl Perfluorooctanesulfonamido Ethanol (NEtFOSE)	ND		ng/l	14.5	1.27	1
Perfluoro-3-Methoxypropanoic Acid (PFMPA)	ND		ng/l	2.89	0.130	1
Perfluoro-4-Methoxybutanoic Acid (PFMBA)	ND		ng/l	2.89	0.752	1
Perfluoro(2-Ethoxyethane)Sulfonic Acid (PFEEESA)	ND		ng/l	2.89	0.362	1
Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)	ND		ng/l	2.89	0.854	1
3-Perfluoropropyl Propanoic Acid (3:3FTCA)	ND		ng/l	7.24	1.87	1
2H,2H,3H,3H-Perfluorooctanoic Acid (5:3FTCA)	ND		ng/l	36.2	10.4	1
3-Perfluoroheptyl Propanoic Acid (7:3FTCA)	ND		ng/l	36.2	10.9	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-04

Date Collected: 11/04/25 12:30

Client ID: GZM-4

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Surrogate			% Recovery	Qualifier	Acceptance Criteria	
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)			105		5-130	
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)			100		40-130	
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)			97		40-135	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)			248	Q	40-200	
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)			105		40-130	
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)			103		40-130	
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)			104		40-130	
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)			103		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)			240	Q	40-200	
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)			84		40-130	
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)			80		40-130	
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)			82		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)			101		40-300	
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)			89		40-170	
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)			67		30-130	
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)			63		40-130	
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)			63		25-135	
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)			48		10-130	
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)			46		10-130	
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)			96		40-130	
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)			52		10-130	
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)			50		10-130	
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)			45		10-130	
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)			49		10-130	

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-05
Client ID: GZM-DUP
Sample Location: BRONX, NY

Date Collected: 11/04/25 00:00
Date Received: 11/04/25
Field Prep: Not Specified

Sample Depth:

Matrix: Water
Analytical Method: 1,8270E
Analytical Date: 11/07/25 02:37
Analyst: SMZ

Extraction Method: EPA 3510C
Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
1,2,4-Trichlorobenzene	ND		ug/l	5.0	0.98	1
Bis(2-chloroethyl)ether	ND		ug/l	2.0	0.39	1
1,2-Dichlorobenzene	ND		ug/l	2.0	0.33	1
1,3-Dichlorobenzene	ND		ug/l	2.0	0.32	1
1,4-Dichlorobenzene	ND		ug/l	2.0	0.39	1
3,3'-Dichlorobenzidine	ND		ug/l	5.0	1.8	1
2,4-Dinitrotoluene	ND		ug/l	5.0	0.54	1
2,6-Dinitrotoluene	ND		ug/l	5.0	0.84	1
4-Chlorophenyl phenyl ether	ND		ug/l	2.0	0.39	1
4-Bromophenyl phenyl ether	ND		ug/l	2.0	0.24	1
Bis(2-chloroisopropyl)ether	ND		ug/l	2.0	0.40	1
Bis(2-chloroethoxy)methane	ND		ug/l	5.0	0.84	1
Hexachlorocyclopentadiene	ND		ug/l	20	1.2	1
Isophorone	ND		ug/l	5.0	0.86	1
Nitrobenzene	ND		ug/l	2.0	0.20	1
NDPA/DPA	ND		ug/l	2.0	0.92	1
n-Nitrosodi-n-propylamine	ND		ug/l	5.0	0.91	1
Bis(2-ethylhexyl)phthalate	ND		ug/l	3.0	1.4	1
Butyl benzyl phthalate	ND		ug/l	5.0	2.6	1
Di-n-butylphthalate	1.0	J	ug/l	5.0	0.96	1
Di-n-octylphthalate	ND		ug/l	5.0	2.3	1
Diethyl phthalate	ND		ug/l	5.0	0.76	1
Dimethyl phthalate	ND		ug/l	5.0	0.92	1
Biphenyl	ND		ug/l	2.0	0.20	1
4-Chloroaniline	ND		ug/l	5.0	0.47	1
2-Nitroaniline	ND		ug/l	5.0	1.0	1
3-Nitroaniline	ND		ug/l	5.0	1.2	1
4-Nitroaniline	ND		ug/l	5.0	1.4	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-05

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
Dibenzofuran	ND		ug/l	2.0	0.40	1
1,2,4,5-Tetrachlorobenzene	ND		ug/l	10	0.24	1
Acetophenone	ND		ug/l	5.0	0.92	1
2,4,6-Trichlorophenol	ND		ug/l	5.0	2.1	1
p-Chloro-m-cresol	ND		ug/l	2.0	0.61	1
2-Chlorophenol	ND		ug/l	2.0	0.65	1
2,4-Dichlorophenol	ND		ug/l	5.0	1.7	1
2,4-Dimethylphenol	ND		ug/l	5.0	2.0	1
2-Nitrophenol	ND		ug/l	10	2.0	1
4-Nitrophenol	ND		ug/l	10	1.4	1
2,4-Dinitrophenol	ND		ug/l	20	5.4	1
4,6-Dinitro-o-cresol	ND		ug/l	10	2.3	1
Phenol	ND		ug/l	5.0	0.35	1
2-Methylphenol	ND		ug/l	5.0	2.3	1
3-Methylphenol/4-Methylphenol	ND		ug/l	5.0	1.4	1
2,4,5-Trichlorophenol	ND		ug/l	5.0	2.1	1
Benzoic Acid	ND		ug/l	50	2.6	1
Benzyl Alcohol	ND		ug/l	2.0	0.38	1
Carbazole	ND		ug/l	2.0	0.31	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	52		21-120
Phenol-d6	38		10-120
Nitrobenzene-d5	62		23-120
2-Fluorobiphenyl	59		15-120
2,4,6-Tribromophenol	85		10-120
4-Terphenyl-d14	64		41-149

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-05
 Client ID: GZM-DUP
 Sample Location: BRONX, NY

Date Collected: 11/04/25 00:00
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8270E-SIM
 Analytical Date: 11/07/25 12:08
 Analyst: JJW

Extraction Method: EPA 3510C
 Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS-SIM - Westborough Lab						
Acenaphthene	ND		ug/l	0.10	0.02	1
2-Chloronaphthalene	ND		ug/l	0.20	0.02	1
Fluoranthene	0.06	J	ug/l	0.10	0.03	1
Hexachlorobutadiene	ND		ug/l	0.50	0.02	1
Naphthalene	0.45		ug/l	0.10	0.02	1
Benzo(a)anthracene	0.17		ug/l	0.10	0.03	1
Benzo(a)pyrene	ND		ug/l	0.10	0.02	1
Benzo(b)fluoranthene	0.04	J	ug/l	0.10	0.03	1
Benzo(k)fluoranthene	ND		ug/l	0.10	0.03	1
Chrysene	ND		ug/l	0.10	0.03	1
Acenaphthylene	ND		ug/l	0.10	0.02	1
Anthracene	0.06	J	ug/l	0.10	0.02	1
Benzo(ghi)perylene	ND		ug/l	0.10	0.02	1
Fluorene	ND		ug/l	0.10	0.03	1
Phenanthrene	0.05	J	ug/l	0.10	0.04	1
Dibenzo(a,h)anthracene	ND		ug/l	0.10	0.02	1
Indeno(1,2,3-cd)pyrene	ND		ug/l	0.10	0.02	1
Pyrene	0.07	J	ug/l	0.10	0.04	1
2-Methylnaphthalene	0.05	J	ug/l	0.10	0.03	1
Pentachlorophenol	0.07	J	ug/l	0.80	0.06	1
Hexachlorobenzene	ND		ug/l	0.80	0.01	1
Hexachloroethane	ND		ug/l	0.80	0.02	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-05

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
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Semivolatile Organics by GC/MS-SIM - Westborough Lab

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	57		21-120
Phenol-d6	42		10-120
Nitrobenzene-d5	103		23-120
2-Fluorobiphenyl	77		15-120
2,4,6-Tribromophenol	137	Q	10-120
4-Terphenyl-d14	87		41-149

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-05
Client ID: GZM-DUP
Sample Location: BRONX, NY

Date Collected: 11/04/25 00:00
Date Received: 11/04/25
Field Prep: Not Specified

Sample Depth:

Matrix: Water
Analytical Method: 168,1633A
Analytical Date: 11/10/25 13:13
Analyst: JW

Extraction Method: EPA 1633
Extraction Date: 11/10/25 01:35

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Perfluoropentanoic Acid (PFPeA)	11.5		ng/l	3.16	0.585	1
Perfluorobutanesulfonic Acid (PFBS)	4.29		ng/l	1.58	0.506	1
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2FTS)	ND		ng/l	6.33	1.47	1
Perfluorohexanoic Acid (PFHxA)	12.9		ng/l	1.58	0.380	1
Perfluoropentanesulfonic Acid (PFPeS)	0.807	J	ng/l	1.58	0.364	1
Perfluoroheptanoic Acid (PFHpA)	7.07		ng/l	1.58	0.490	1
Perfluorohexanesulfonic Acid (PFHxS)	2.93		ng/l	1.58	0.269	1
Perfluorooctanoic Acid (PFOA)	25.2		ng/l	1.58	0.396	1
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2FTS)	4.38	J	ng/l	6.33	3.21	1
Perfluoroheptanesulfonic Acid (PFHpS)	1.08	J	ng/l	1.58	0.490	1
Perfluorononanoic Acid (PFNA)	4.78		ng/l	1.58	0.237	1
Perfluorooctanesulfonic Acid (PFOS)	68.8		ng/l	1.58	0.269	1
Perfluorodecanoic Acid (PFDA)	1.17	J	ng/l	1.58	0.411	1
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2FTS)	ND		ng/l	6.33	3.02	1
Perfluorononanesulfonic Acid (PFNS)	ND		ng/l	1.58	0.237	1
N-Methyl Perfluorooctanesulfonamidoacetic Acid (NMeFOSAA)	ND		ng/l	1.58	1.14	1
Perfluoroundecanoic Acid (PFUnA)	ND		ng/l	1.58	0.380	1
Perfluorodecanesulfonic Acid (PFDS)	ND		ng/l	1.58	0.269	1
Perfluorooctanesulfonamide (PFOSA)	ND		ng/l	1.58	0.316	1
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (NEFOSAA)	ND		ng/l	1.58	0.728	1
Perfluorododecanoic Acid (PFDoA)	ND		ng/l	1.58	0.443	1
Perfluorotridecanoic Acid (PFTTrDA)	ND		ng/l	1.58	0.269	1
Perfluorotetradecanoic Acid (PFTeDA)	ND		ng/l	1.58	0.190	1
Hexafluoropropylene Oxide Dimer Acid (HFPO-DA)	ND		ng/l	6.33	0.933	1
4,8-Dioxa-3h-Perfluorononanoic Acid (ADONA)	ND		ng/l	6.33	1.23	1
Perfluorododecanesulfonic Acid (PFDoS)	ND		ng/l	1.58	0.332	1

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-05
Client ID: GZM-DUP
Sample Location: BRONX, NY

Date Collected: 11/04/25 00:00
Date Received: 11/04/25
Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
9-Chlorohexadecafluoro-3-Oxanone-1-Sulfonic Acid (9Cl-PF3ONS)	ND		ng/l	6.33	0.696	1
11-Chloroeicosafluoro-3-Oxaundecane-1-Sulfonic Acid (11Cl-PF3OUdS)	ND		ng/l	6.33	0.475	1
N-Methyl Perfluorooctane Sulfonamide (NMeFOSA)	ND		ng/l	1.58	0.617	1
N-Ethyl Perfluorooctane Sulfonamide (NEtFOSA)	ND		ng/l	1.58	0.459	1
N-Methyl Perfluorooctanesulfonamido Ethanol (NMeFOSE)	ND		ng/l	15.8	1.68	1
N-Ethyl Perfluorooctanesulfonamido Ethanol (NEtFOSE)	ND		ng/l	15.8	1.39	1
Perfluoro-4-Methoxybutanoic Acid (PFMBA)	ND		ng/l	3.16	0.823	1
Perfluoro(2-Ethoxyethane)Sulfonic Acid (PFEESA)	ND		ng/l	3.16	0.396	1
Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)	ND		ng/l	3.16	0.933	1
3-Perfluoropropyl Propanoic Acid (3:3FTCA)	ND		ng/l	7.91	2.04	1
2H,2H,3H,3H-Perfluorooctanoic Acid (5:3FTCA)	ND		ng/l	39.6	11.4	1
3-Perfluoroheptyl Propanoic Acid (7:3FTCA)	ND		ng/l	39.6	11.9	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-05

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Surrogate			% Recovery	Qualifier	Acceptance Criteria	
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)			87		5-130	
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)			92		40-130	
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)			83		40-135	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)			360	Q	40-200	
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)			98		40-130	
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)			101		40-130	
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)			88		40-130	
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)			92		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)			392	Q	40-200	
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)			78		40-130	
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)			69		40-130	
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)			73		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)			258		40-300	
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)			105		40-170	
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)			67		30-130	
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)			55		40-130	
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)			93		25-135	
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)			52		10-130	
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)			47		10-130	
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)			112		40-130	
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)			42		10-130	
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)			44		10-130	
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)			40		10-130	
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)			44		10-130	

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-05 D

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 1633

Analytical Method: 168,1633A

Extraction Date: 11/10/25 01:35

Analytical Date: 11/10/25 17:02

Analyst: ANH

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
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Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab

Perfluorobutanoic Acid (PFBA)	19.6	J	ng/l	31.6	11.8	5
Perfluoro-3-Methoxypropanoic Acid (PFMPA)	ND		ng/l	15.8	0.712	5

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-05 D

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Surrogate			% Recovery	Qualifier	Acceptance Criteria	
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)			64		5-130	
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)			63		40-130	
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)			63		40-135	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)			136		40-200	
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)			70		40-130	
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)			69		40-130	
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)			64		40-130	
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)			61		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)			124		40-200	
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)			51		40-130	
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)			53		40-130	
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)			56		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)			65		40-300	
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)			47		40-170	
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)			42		30-130	
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)			36	Q	40-130	
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)			42		25-135	
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)			33		10-130	
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)			30		10-130	
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)			65		40-130	
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)			34		10-130	
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)			30		10-130	
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)			33		10-130	
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)			35		10-130	

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-06

Date Collected: 11/04/25 14:00

Client ID: FIELD BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 3510C

Analytical Method: 1,8270E

Extraction Date: 11/06/25 19:23

Analytical Date: 11/07/25 03:00

Analyst: SMZ

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
1,2,4-Trichlorobenzene	ND		ug/l	5.0	0.98	1
Bis(2-chloroethyl)ether	ND		ug/l	2.0	0.39	1
1,2-Dichlorobenzene	ND		ug/l	2.0	0.33	1
1,3-Dichlorobenzene	ND		ug/l	2.0	0.32	1
1,4-Dichlorobenzene	ND		ug/l	2.0	0.39	1
3,3'-Dichlorobenzidine	ND		ug/l	5.0	1.8	1
2,4-Dinitrotoluene	ND		ug/l	5.0	0.54	1
2,6-Dinitrotoluene	ND		ug/l	5.0	0.84	1
4-Chlorophenyl phenyl ether	ND		ug/l	2.0	0.39	1
4-Bromophenyl phenyl ether	ND		ug/l	2.0	0.24	1
Bis(2-chloroisopropyl)ether	ND		ug/l	2.0	0.40	1
Bis(2-chloroethoxy)methane	ND		ug/l	5.0	0.84	1
Hexachlorocyclopentadiene	ND		ug/l	20	1.2	1
Isophorone	ND		ug/l	5.0	0.86	1
Nitrobenzene	ND		ug/l	2.0	0.20	1
NDPA/DPA	ND		ug/l	2.0	0.92	1
n-Nitrosodi-n-propylamine	ND		ug/l	5.0	0.91	1
Bis(2-ethylhexyl)phthalate	ND		ug/l	3.0	1.4	1
Butyl benzyl phthalate	ND		ug/l	5.0	2.6	1
Di-n-butylphthalate	ND		ug/l	5.0	0.96	1
Di-n-octylphthalate	ND		ug/l	5.0	2.3	1
Diethyl phthalate	ND		ug/l	5.0	0.76	1
Dimethyl phthalate	ND		ug/l	5.0	0.92	1
Biphenyl	ND		ug/l	2.0	0.20	1
4-Chloroaniline	ND		ug/l	5.0	0.47	1
2-Nitroaniline	ND		ug/l	5.0	1.0	1
3-Nitroaniline	ND		ug/l	5.0	1.2	1
4-Nitroaniline	ND		ug/l	5.0	1.4	1

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-06
 Client ID: FIELD BLANK
 Sample Location: BRONX, NY

Date Collected: 11/04/25 14:00
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS - Westborough Lab						
Dibenzofuran	ND		ug/l	2.0	0.40	1
1,2,4,5-Tetrachlorobenzene	ND		ug/l	10	0.24	1
Acetophenone	ND		ug/l	5.0	0.92	1
2,4,6-Trichlorophenol	ND		ug/l	5.0	2.1	1
p-Chloro-m-cresol	ND		ug/l	2.0	0.61	1
2-Chlorophenol	ND		ug/l	2.0	0.65	1
2,4-Dichlorophenol	ND		ug/l	5.0	1.7	1
2,4-Dimethylphenol	ND		ug/l	5.0	2.0	1
2-Nitrophenol	ND		ug/l	10	2.0	1
4-Nitrophenol	ND		ug/l	10	1.4	1
2,4-Dinitrophenol	ND		ug/l	20	5.4	1
4,6-Dinitro-o-cresol	ND		ug/l	10	2.3	1
Phenol	ND		ug/l	5.0	0.35	1
2-Methylphenol	ND		ug/l	5.0	2.3	1
3-Methylphenol/4-Methylphenol	ND		ug/l	5.0	1.4	1
2,4,5-Trichlorophenol	ND		ug/l	5.0	2.1	1
Benzoic Acid	ND		ug/l	50	2.6	1
Benzyl Alcohol	ND		ug/l	2.0	0.38	1
Carbazole	ND		ug/l	2.0	0.31	1

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	62		21-120
Phenol-d6	43		10-120
Nitrobenzene-d5	83		23-120
2-Fluorobiphenyl	68		15-120
2,4,6-Tribromophenol	97		10-120
4-Terphenyl-d14	80		41-149

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-06
 Client ID: FIELD BLANK
 Sample Location: BRONX, NY

Date Collected: 11/04/25 14:00
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8270E-SIM
 Analytical Date: 11/07/25 12:25
 Analyst: JJW

Extraction Method: EPA 3510C
 Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Semivolatile Organics by GC/MS-SIM - Westborough Lab						
Acenaphthene	ND		ug/l	0.10	0.02	1
2-Chloronaphthalene	ND		ug/l	0.20	0.02	1
Fluoranthene	ND		ug/l	0.10	0.03	1
Hexachlorobutadiene	ND		ug/l	0.50	0.02	1
Naphthalene	ND		ug/l	0.10	0.02	1
Benzo(a)anthracene	ND		ug/l	0.10	0.03	1
Benzo(a)pyrene	ND		ug/l	0.10	0.02	1
Benzo(b)fluoranthene	ND		ug/l	0.10	0.03	1
Benzo(k)fluoranthene	ND		ug/l	0.10	0.03	1
Chrysene	ND		ug/l	0.10	0.03	1
Acenaphthylene	ND		ug/l	0.10	0.02	1
Anthracene	ND		ug/l	0.10	0.02	1
Benzo(ghi)perylene	ND		ug/l	0.10	0.02	1
Fluorene	ND		ug/l	0.10	0.03	1
Phenanthrene	ND		ug/l	0.10	0.04	1
Dibenzo(a,h)anthracene	ND		ug/l	0.10	0.02	1
Indeno(1,2,3-cd)pyrene	ND		ug/l	0.10	0.02	1
Pyrene	ND		ug/l	0.10	0.04	1
2-Methylnaphthalene	ND		ug/l	0.10	0.03	1
Pentachlorophenol	ND		ug/l	0.80	0.06	1
Hexachlorobenzene	ND		ug/l	0.80	0.01	1
Hexachloroethane	ND		ug/l	0.80	0.02	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-06

Date Collected: 11/04/25 14:00

Client ID: FIELD BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
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Semivolatile Organics by GC/MS-SIM - Westborough Lab

Surrogate	% Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	57		21-120
Phenol-d6	44		10-120
Nitrobenzene-d5	104		23-120
2-Fluorobiphenyl	79		15-120
2,4,6-Tribromophenol	122	Q	10-120
4-Terphenyl-d14	77		41-149

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-06
Client ID: FIELD BLANK
Sample Location: BRONX, NY

Date Collected: 11/04/25 14:00
Date Received: 11/04/25
Field Prep: Not Specified

Sample Depth:

Matrix: Water
Analytical Method: 168,1633A
Analytical Date: 11/10/25 13:22
Analyst: JW

Extraction Method: EPA 1633
Extraction Date: 11/10/25 01:35

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Perfluorobutanoic Acid (PFBA)	ND		ng/l	5.63	2.10	1
Perfluoropentanoic Acid (PFPeA)	ND		ng/l	2.81	0.520	1
Perfluorobutanesulfonic Acid (PFBS)	ND		ng/l	1.41	0.450	1
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2FTS)	ND		ng/l	5.63	1.31	1
Perfluorohexanoic Acid (PFHxA)	ND		ng/l	1.41	0.338	1
Perfluoropentanesulfonic Acid (PFPeS)	ND		ng/l	1.41	0.324	1
Perfluoroheptanoic Acid (PFHpA)	ND		ng/l	1.41	0.436	1
Perfluorohexanesulfonic Acid (PFHxS)	ND		ng/l	1.41	0.239	1
Perfluorooctanoic Acid (PFOA)	ND		ng/l	1.41	0.352	1
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2FTS)	ND		ng/l	5.63	2.86	1
Perfluoroheptanesulfonic Acid (PFHpS)	ND		ng/l	1.41	0.436	1
Perfluorononanoic Acid (PFNA)	ND		ng/l	1.41	0.211	1
Perfluorooctanesulfonic Acid (PFOS)	0.577	J	ng/l	1.41	0.239	1
Perfluorodecanoic Acid (PFDA)	ND		ng/l	1.41	0.366	1
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2FTS)	ND		ng/l	5.63	2.69	1
Perfluorononanesulfonic Acid (PFNS)	ND		ng/l	1.41	0.211	1
N-Methyl Perfluorooctanesulfonamidoacetic Acid (NMeFOSAA)	ND		ng/l	1.41	1.01	1
Perfluoroundecanoic Acid (PFUnA)	ND		ng/l	1.41	0.338	1
Perfluorodecanesulfonic Acid (PFDS)	ND		ng/l	1.41	0.239	1
Perfluorooctanesulfonamide (PFOSA)	ND		ng/l	1.41	0.281	1
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (NEtFOSAA)	ND		ng/l	1.41	0.647	1
Perfluorododecanoic Acid (PFDoA)	ND		ng/l	1.41	0.394	1
Perfluorotridecanoic Acid (PFTrDA)	ND		ng/l	1.41	0.239	1
Perfluorotetradecanoic Acid (PFTeDA)	ND		ng/l	1.41	0.169	1
Hexafluoropropylene Oxide Dimer Acid (HFPO-DA)	ND		ng/l	5.63	0.830	1
4,8-Dioxa-3h-Perfluorononanoic Acid (ADONA)	ND		ng/l	5.63	1.10	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-06

Date Collected: 11/04/25 14:00

Client ID: FIELD BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Perfluorododecanesulfonic Acid (PFDoS)	ND		ng/l	1.41	0.295	1
9-Chlorohexadecafluoro-3-Oxanone-1-Sulfonic Acid (9Cl-PF3ONS)	ND		ng/l	5.63	0.619	1
11-Chloroeicosafluoro-3-Oxaundecane-1-Sulfonic Acid (11Cl-PF3OUdS)	ND		ng/l	5.63	0.422	1
N-Methyl Perfluorooctane Sulfonamide (NMeFOSA)	ND		ng/l	1.41	0.549	1
N-Ethyl Perfluorooctane Sulfonamide (NEtFOSA)	ND		ng/l	1.41	0.408	1
N-Methyl Perfluorooctanesulfonamido Ethanol (NMeFOSE)	ND		ng/l	14.1	1.49	1
N-Ethyl Perfluorooctanesulfonamido Ethanol (NEtFOSE)	ND		ng/l	14.1	1.24	1
Perfluoro-3-Methoxypropanoic Acid (PFMPA)	ND		ng/l	2.81	0.127	1
Perfluoro-4-Methoxybutanoic Acid (PFMBA)	ND		ng/l	2.81	0.732	1
Perfluoro(2-Ethoxyethane)Sulfonic Acid (PFEEESA)	ND		ng/l	2.81	0.352	1
Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)	ND		ng/l	2.81	0.830	1
3-Perfluoropropyl Propanoic Acid (3:3FTCA)	ND		ng/l	7.04	1.82	1
2H,2H,3H,3H-Perfluorooctanoic Acid (5:3FTCA)	ND		ng/l	35.2	10.1	1
3-Perfluoroheptyl Propanoic Acid (7:3FTCA)	ND		ng/l	35.2	10.6	1

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-06

Date Collected: 11/04/25 14:00

Client ID: FIELD BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab						
Surrogate			% Recovery	Qualifier	Acceptance Criteria	
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)			70		5-130	
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)			68		40-130	
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)			75		40-135	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)			66		40-200	
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)			59		40-130	
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)			66		40-130	
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)			63		40-130	
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)			64		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)			64		40-200	
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)			69		40-130	
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)			57		40-130	
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)			66		40-130	
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)			63		40-300	
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)			67		40-170	
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)			71		30-130	
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)			60		40-130	
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)			59		25-135	
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)			54		10-130	
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)			60		10-130	
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)			66		40-130	
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)			53		10-130	
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)			55		10-130	
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)			59		10-130	
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)			67		10-130	

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8270E
Analytical Date: 11/07/25 00:44
Analyst: SMZ

Extraction Method: EPA 3510C
Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL
Semivolatile Organics by GC/MS - Westborough Lab for sample(s): 01-06 Batch: WG2138276-1					
Acenaphthene	ND		ug/l	2.0	0.40
1,2,4-Trichlorobenzene	ND		ug/l	5.0	0.98
Hexachlorobenzene	ND		ug/l	2.0	0.45
Bis(2-chloroethyl)ether	ND		ug/l	2.0	0.39
2-Chloronaphthalene	ND		ug/l	2.0	0.35
1,2-Dichlorobenzene	ND		ug/l	2.0	0.33
1,3-Dichlorobenzene	ND		ug/l	2.0	0.32
1,4-Dichlorobenzene	ND		ug/l	2.0	0.39
3,3'-Dichlorobenzidine	ND		ug/l	5.0	1.8
2,4-Dinitrotoluene	ND		ug/l	5.0	0.54
2,6-Dinitrotoluene	ND		ug/l	5.0	0.84
Fluoranthene	ND		ug/l	2.0	0.41
4-Chlorophenyl phenyl ether	ND		ug/l	2.0	0.39
4-Bromophenyl phenyl ether	ND		ug/l	2.0	0.24
Bis(2-chloroisopropyl)ether	ND		ug/l	2.0	0.40
Bis(2-chloroethoxy)methane	ND		ug/l	5.0	0.84
Hexachlorobutadiene	ND		ug/l	2.0	0.36
Hexachlorocyclopentadiene	ND		ug/l	20	1.2
Hexachloroethane	ND		ug/l	2.0	0.20
Isophorone	ND		ug/l	5.0	0.86
Naphthalene	ND		ug/l	2.0	0.54
Nitrobenzene	ND		ug/l	2.0	0.20
NDPA/DPA	ND		ug/l	2.0	0.92
n-Nitrosodi-n-propylamine	ND		ug/l	5.0	0.91

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8270E
Analytical Date: 11/07/25 00:44
Analyst: SMZ

Extraction Method: EPA 3510C
Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL
Semivolatile Organics by GC/MS - Westborough Lab for sample(s): 01-06 Batch: WG2138276-1					
Bis(2-ethylhexyl)phthalate	ND		ug/l	3.0	1.4
Butyl benzyl phthalate	ND		ug/l	5.0	2.6
Di-n-butylphthalate	ND		ug/l	5.0	0.96
Di-n-octylphthalate	ND		ug/l	5.0	2.3
Diethyl phthalate	ND		ug/l	5.0	0.76
Dimethyl phthalate	ND		ug/l	5.0	0.92
Benzo(a)anthracene	ND		ug/l	2.0	0.32
Benzo(a)pyrene	ND		ug/l	2.0	0.37
Benzo(b)fluoranthene	ND		ug/l	2.0	0.53
Benzo(k)fluoranthene	ND		ug/l	2.0	0.62
Chrysene	ND		ug/l	2.0	0.22
Acenaphthylene	ND		ug/l	2.0	0.32
Anthracene	ND		ug/l	2.0	0.47
Benzo(ghi)perylene	ND		ug/l	2.0	0.37
Fluorene	ND		ug/l	2.0	0.44
Phenanthrene	ND		ug/l	2.0	0.42
Dibenzo(a,h)anthracene	ND		ug/l	2.0	0.29
Indeno(1,2,3-cd)pyrene	ND		ug/l	2.0	0.48
Pyrene	ND		ug/l	2.0	0.41
Biphenyl	ND		ug/l	2.0	0.20
4-Chloroaniline	ND		ug/l	5.0	0.47
2-Nitroaniline	ND		ug/l	5.0	1.0
3-Nitroaniline	ND		ug/l	5.0	1.2
4-Nitroaniline	ND		ug/l	5.0	1.4

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8270E
Analytical Date: 11/07/25 00:44
Analyst: SMZ

Extraction Method: EPA 3510C
Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL
Semivolatile Organics by GC/MS - Westborough Lab for sample(s): 01-06 Batch: WG2138276-1					
Dibenzofuran	ND		ug/l	2.0	0.40
2-Methylnaphthalene	ND		ug/l	2.0	0.37
1,2,4,5-Tetrachlorobenzene	ND		ug/l	10	0.24
Acetophenone	ND		ug/l	5.0	0.92
2,4,6-Trichlorophenol	ND		ug/l	5.0	2.1
p-Chloro-m-cresol	ND		ug/l	2.0	0.61
2-Chlorophenol	ND		ug/l	2.0	0.65
2,4-Dichlorophenol	ND		ug/l	5.0	1.7
2,4-Dimethylphenol	ND		ug/l	5.0	2.0
2-Nitrophenol	ND		ug/l	10	2.0
4-Nitrophenol	ND		ug/l	10	1.4
2,4-Dinitrophenol	ND		ug/l	20	5.4
4,6-Dinitro-o-cresol	ND		ug/l	10	2.3
Pentachlorophenol	ND		ug/l	10	2.5
Phenol	ND		ug/l	5.0	0.35
2-Methylphenol	ND		ug/l	5.0	2.3
3-Methylphenol/4-Methylphenol	ND		ug/l	5.0	1.4
2,4,5-Trichlorophenol	ND		ug/l	5.0	2.1
Benzoic Acid	ND		ug/l	50	2.6
Benzyl Alcohol	ND		ug/l	2.0	0.38
Carbazole	ND		ug/l	2.0	0.31

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8270E
Analytical Date: 11/07/25 00:44
Analyst: SMZ

Extraction Method: EPA 3510C
Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL
Semivolatile Organics by GC/MS - Westborough Lab for sample(s): 01-06 Batch: WG2138276-1					

Surrogate	%Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	57		21-120
Phenol-d6	41		10-120
Nitrobenzene-d5	75		23-120
2-Fluorobiphenyl	59		15-120
2,4,6-Tribromophenol	82		10-120
4-Terphenyl-d14	66		41-149

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8270E-SIM
Analytical Date: 11/07/25 10:40
Analyst: JJW

Extraction Method: EPA 3510C
Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL
Semivolatile Organics by GC/MS-SIM - Westborough Lab for sample(s): 01-06 Batch: WG2138277-1					
Acenaphthene	ND		ug/l	0.10	0.02
2-Chloronaphthalene	ND		ug/l	0.20	0.02
Fluoranthene	ND		ug/l	0.10	0.03
Hexachlorobutadiene	ND		ug/l	0.50	0.02
Naphthalene	ND		ug/l	0.10	0.02
Benzo(a)anthracene	ND		ug/l	0.10	0.03
Benzo(a)pyrene	ND		ug/l	0.10	0.02
Benzo(b)fluoranthene	ND		ug/l	0.10	0.03
Benzo(k)fluoranthene	ND		ug/l	0.10	0.03
Chrysene	ND		ug/l	0.10	0.03
Acenaphthylene	ND		ug/l	0.10	0.02
Anthracene	ND		ug/l	0.10	0.02
Benzo(ghi)perylene	ND		ug/l	0.10	0.02
Fluorene	ND		ug/l	0.10	0.03
Phenanthrene	ND		ug/l	0.10	0.04
Dibenzo(a,h)anthracene	ND		ug/l	0.10	0.02
Indeno(1,2,3-cd)pyrene	ND		ug/l	0.10	0.02
Pyrene	ND		ug/l	0.10	0.04
2-Methylnaphthalene	ND		ug/l	0.10	0.03
Pentachlorophenol	ND		ug/l	0.80	0.06
Hexachlorobenzene	ND		ug/l	0.80	0.01
Hexachloroethane	ND		ug/l	0.80	0.02

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8270E-SIM
Analytical Date: 11/07/25 10:40
Analyst: JJW

Extraction Method: EPA 3510C
Extraction Date: 11/06/25 19:23

Parameter	Result	Qualifier	Units	RL	MDL
Semivolatile Organics by GC/MS-SIM - Westborough Lab for sample(s): 01-06 Batch: WG2138277-1					

Surrogate	%Recovery	Qualifier	Acceptance Criteria
2-Fluorophenol	56		21-120
Phenol-d6	45		10-120
Nitrobenzene-d5	100		23-120
2-Fluorobiphenyl	75		15-120
2,4,6-Tribromophenol	113		10-120
4-Terphenyl-d14	68		41-149

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 168,1633A
Analytical Date: 11/10/25 11:35
Analyst: JW

Extraction Method: EPA 1633
Extraction Date: 11/10/25 01:35

Parameter	Result	Qualifier	Units	RL	MDL
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab for sample(s): 01-06 Batch: WG2139125-1					
Perfluorobutanoic Acid (PFBA)	ND		ng/l	6.40	2.38
Perfluoropentanoic Acid (PFPeA)	ND		ng/l	3.20	0.592
Perfluorobutanesulfonic Acid (PFBS)	ND		ng/l	1.60	0.512
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2FTS)	ND		ng/l	6.40	1.49
Perfluorohexanoic Acid (PFHxA)	ND		ng/l	1.60	0.384
Perfluoropentanesulfonic Acid (PFPeS)	ND		ng/l	1.60	0.368
Perfluoroheptanoic Acid (PFHpA)	ND		ng/l	1.60	0.496
Perfluorohexanesulfonic Acid (PFHxS)	0.384	J	ng/l	1.60	0.272
Perfluorooctanoic Acid (PFOA)	ND		ng/l	1.60	0.400
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2FTS)	ND		ng/l	6.40	3.25
Perfluoroheptanesulfonic Acid (PFHpS)	ND		ng/l	1.60	0.496
Perfluorononanoic Acid (PFNA)	ND		ng/l	1.60	0.240
Perfluorooctanesulfonic Acid (PFOS)	0.448	J	ng/l	1.60	0.272
Perfluorodecanoic Acid (PFDA)	ND		ng/l	1.60	0.416
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2FTS)	ND		ng/l	6.40	3.06
Perfluorononanesulfonic Acid (PFNS)	ND		ng/l	1.60	0.240
N-Methyl Perfluorooctanesulfonamidoacetic Acid (NMeFOSAA)	ND		ng/l	1.60	1.15
Perfluoroundecanoic Acid (PFUnA)	ND		ng/l	1.60	0.384
Perfluorodecanesulfonic Acid (PFDS)	ND		ng/l	1.60	0.272
Perfluorooctanesulfonamide (PFOSA)	ND		ng/l	1.60	0.320
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (NEtFOSAA)	ND		ng/l	1.60	0.736
Perfluorododecanoic Acid (PFDoA)	ND		ng/l	1.60	0.448
Perfluorotridecanoic Acid (PFTrDA)	ND		ng/l	1.60	0.272



Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 168,1633A
Analytical Date: 11/10/25 11:35
Analyst: JW

Extraction Method: EPA 1633
Extraction Date: 11/10/25 01:35

Parameter	Result	Qualifier	Units	RL	MDL
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab for sample(s): 01-06 Batch: WG2139125-1					
Perfluorotetradecanoic Acid (PFTeDA)	ND		ng/l	1.60	0.192
Hexafluoropropylene Oxide Dimer Acid (HFPO-DA)	ND		ng/l	6.40	0.944
4,8-Dioxa-3h-Perfluorononanoic Acid (ADONA)	ND		ng/l	6.40	1.25
Perfluorododecanesulfonic Acid (PFDoS)	ND		ng/l	1.60	0.336
9-Chlorohexadecafluoro-3-Oxanone-1-Sulfonic Acid (9Cl-PF3ONS)	ND		ng/l	6.40	0.704
11-Chloroeicosfluoro-3-Oxaundecane-1-Sulfonic Acid (11Cl-PF3OUdS)	ND		ng/l	6.40	0.480
N-Methyl Perfluorooctane Sulfonamide (NMeFOSA)	ND		ng/l	1.60	0.624
N-Ethyl Perfluorooctane Sulfonamide (NEtFOSA)	ND		ng/l	1.60	0.464
N-Methyl Perfluorooctanesulfonamido Ethanol (NMeFOSE)	ND		ng/l	16.0	1.70
N-Ethyl Perfluorooctanesulfonamido Ethanol (NEtFOSE)	ND		ng/l	16.0	1.41
Perfluoro-3-Methoxypropanoic Acid (PFMPA)	ND		ng/l	3.20	0.144
Perfluoro-4-Methoxybutanoic Acid (PFMBA)	ND		ng/l	3.20	0.832
Perfluoro(2-Ethoxyethane)Sulfonic Acid (PFEESA)	ND		ng/l	3.20	0.400
Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)	ND		ng/l	3.20	0.944
3-Perfluoropropyl Propanoic Acid (3:3FTCA)	ND		ng/l	8.00	2.06
2H,2H,3H,3H-Perfluorooctanoic Acid (5:3FTCA)	ND		ng/l	40.0	11.5
3-Perfluoroheptyl Propanoic Acid (7:3FTCA)	ND		ng/l	40.0	12.1

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 168,1633A
Analytical Date: 11/10/25 11:35
Analyst: JW

Extraction Method: EPA 1633
Extraction Date: 11/10/25 01:35

Parameter	Result	Qualifier	Units	RL	MDL
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab for sample(s): 01-06 Batch: WG2139125-1					

Surrogate	%Recovery	Qualifier	Acceptance Criteria
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)	88		5-130
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)	95		40-130
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)	91		40-135
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)	88		40-200
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)	95		40-130
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)	86		40-130
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)	81		40-130
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)	91		40-130
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)	95		40-200
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)	88		40-130
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)	78		40-130
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)	79		40-130
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)	77		40-300
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)	68		40-170
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)	79		30-130
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)	75		40-130
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)	73		25-135
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)	67		10-130
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)	60		10-130
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)	87		40-130
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)	58		10-130
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)	63		10-130
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)	75		10-130
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)	76		10-130

Lab Control Sample Analysis

Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Semivolatile Organics by GC/MS - Westborough Lab Associated sample(s): 01-06 Batch: WG2138276-2 WG2138276-3								
Acenaphthene	65		69		37-111	6		30
1,2,4-Trichlorobenzene	59		61		39-98	3		30
Hexachlorobenzene	68		72		40-140	6		30
Bis(2-chloroethyl)ether	65		69		40-140	6		30
2-Chloronaphthalene	58		61		40-140	5		30
1,2-Dichlorobenzene	58		64		40-140	10		30
1,3-Dichlorobenzene	57		58		40-140	2		30
1,4-Dichlorobenzene	56		60		36-97	7		30
3,3'-Dichlorobenzidine	68		71		40-140	4		30
2,4-Dinitrotoluene	73		76		48-143	4		30
2,6-Dinitrotoluene	62		60		40-140	3		30
Fluoranthene	64		65		40-140	2		30
4-Chlorophenyl phenyl ether	62		66		40-140	6		30
4-Bromophenyl phenyl ether	62		67		40-140	8		30
Bis(2-chloroisopropyl)ether	56		58		40-140	4		30
Bis(2-chloroethoxy)methane	67		69		40-140	3		30
Hexachlorobutadiene	49		48		40-140	2		30
Hexachlorocyclopentadiene	53		52		40-140	2		30
Hexachloroethane	60		63		40-140	5		30

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Semivolatile Organics by GC/MS - Westborough Lab Associated sample(s): 01-06 Batch: WG2138276-2 WG2138276-3								
Isophorone	68		72		40-140	6		30
Naphthalene	59		63		40-140	7		30
Nitrobenzene	67		71		40-140	6		30
NDPA/DPA	65		68		40-140	5		30
n-Nitrosodi-n-propylamine	66		74		29-132	11		30
Bis(2-ethylhexyl)phthalate	86		90		40-140	5		30
Butyl benzyl phthalate	79		82		40-140	4		30
Di-n-butylphthalate	74		72		40-140	3		30
Di-n-octylphthalate	84		86		40-140	2		30
Diethyl phthalate	70		73		40-140	4		30
Dimethyl phthalate	61		61		40-140	0		30
Benzo(a)anthracene	66		66		40-140	0		30
Benzo(a)pyrene	71		74		40-140	4		30
Benzo(b)fluoranthene	72		73		40-140	1		30
Benzo(k)fluoranthene	67		69		40-140	3		30
Chrysene	65		64		40-140	2		30
Acenaphthylene	64		62		45-123	3		30
Anthracene	64		64		40-140	0		30
Benzo(ghi)perylene	59		62		40-140	5		30

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Semivolatile Organics by GC/MS - Westborough Lab Associated sample(s): 01-06 Batch: WG2138276-2 WG2138276-3								
Fluorene	67		68		40-140	1		30
Phenanthrene	63		64		40-140	2		30
Dibenzo(a,h)anthracene	63		66		40-140	5		30
Indeno(1,2,3-cd)pyrene	66		69		40-140	4		30
Pyrene	63		64		26-127	2		30
Biphenyl	57		57		40-140	0		30
4-Chloroaniline	60		56		40-140	7		30
2-Nitroaniline	72		72		52-143	0		30
3-Nitroaniline	76		75		25-145	1		30
4-Nitroaniline	75		79		51-143	5		30
Dibenzofuran	65		68		40-140	5		30
2-Methylnaphthalene	59		62		40-140	5		30
1,2,4,5-Tetrachlorobenzene	51		50		2-134	2		30
Acetophenone	64		66		39-129	3		30
2,4,6-Trichlorophenol	64		67		30-130	5		30
p-Chloro-m-cresol	72		75		23-97	4		30
2-Chlorophenol	66		71		27-123	7		30
2,4-Dichlorophenol	63		72		30-130	13		30
2,4-Dimethylphenol	70		75		30-130	7		30

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCS %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Semivolatile Organics by GC/MS - Westborough Lab Associated sample(s): 01-06 Batch: WG2138276-2 WG2138276-3								
2-Nitrophenol	75		77		30-130	3		30
4-Nitrophenol	70		68		10-80	3		30
2,4-Dinitrophenol	65		64		20-130	2		30
4,6-Dinitro-o-cresol	68		74		20-164	8		30
Pentachlorophenol	79		86		9-103	8		30
Phenol	40		44		12-110	10		30
2-Methylphenol	67		70		30-130	4		30
3-Methylphenol/4-Methylphenol	66		69		30-130	4		30
2,4,5-Trichlorophenol	68		69		30-130	1		30
Benzoic Acid	81		77		10-164	5		30
Benzyl Alcohol	70		76		26-116	8		30
Carbazole	65		66		55-144	2		30

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
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Semivolatile Organics by GC/MS - Westborough Lab Associated sample(s): 01-06 Batch: WG2138276-2 WG2138276-3

Surrogate	LCS %Recovery	Qual	LCSD %Recovery	Qual	Acceptance Criteria
2-Fluorophenol	52		61		21-120
Phenol-d6	44		46		10-120
Nitrobenzene-d5	76		80		23-120
2-Fluorobiphenyl	61		61		15-120
2,4,6-Tribromophenol	84		91		10-120
4-Terphenyl-d14	68		70		41-149

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS		LCSD		%Recovery Limits	RPD	Qual	RPD Limits
	%Recovery	Qual	%Recovery	Qual				
Semivolatile Organics by GC/MS-SIM - Westborough Lab Associated sample(s): 01-06 Batch: WG2138277-2 WG2138277-3								
Acenaphthene	67		69		40-140	3		40
2-Chloronaphthalene	67		69		40-140	3		40
Fluoranthene	72		68		40-140	6		40
Hexachlorobutadiene	59		64		40-140	8		40
Naphthalene	60		63		40-140	5		40
Benzo(a)anthracene	70		71		40-140	1		40
Benzo(a)pyrene	72		74		40-140	3		40
Benzo(b)fluoranthene	69		71		40-140	3		40
Benzo(k)fluoranthene	69		70		40-140	1		40
Chrysene	68		69		40-140	1		40
Acenaphthylene	71		73		40-140	3		40
Anthracene	68		71		40-140	4		40
Benzo(ghi)perylene	76		76		40-140	0		40
Fluorene	69		70		40-140	1		40
Phenanthrene	63		65		40-140	3		40
Dibenzo(a,h)anthracene	85		85		40-140	0		40
Indeno(1,2,3-cd)pyrene	82		84		40-140	2		40
Pyrene	73		68		40-140	7		40
2-Methylnaphthalene	65		67		40-140	3		40

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCS %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Semivolatile Organics by GC/MS-SIM - Westborough Lab Associated sample(s): 01-06 Batch: WG2138277-2 WG2138277-3								
Pentachlorophenol	99		99		40-140	0		40
Hexachlorobenzene	70		76		40-140	8		40
Hexachloroethane	60		64		40-140	6		40

Surrogate	LCS %Recovery	Qual	LCS %Recovery	Qual	Acceptance Criteria
2-Fluorophenol	53		56		21-120
Phenol-d6	42		44		10-120
Nitrobenzene-d5	92		98		23-120
2-Fluorobiphenyl	67		70		15-120
2,4,6-Tribromophenol	121	Q	123	Q	10-120
4-Terphenyl-d14	73		69		41-149

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	Low Level LCS %Recovery	Qual	Low Level LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab Associated sample(s): 01-06 Batch: WG2139125-2 LOW LEVEL								
Perfluorobutanoic Acid (PFBA)	96		-		70-140	-		
Perfluoropentanoic Acid (PFPeA)	98		-		65-135	-		
Perfluorobutanesulfonic Acid (PFBS)	89		-		60-145	-		
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2FTS)	100		-		70-145	-		
Perfluorohexanoic Acid (PFHxA)	98		-		70-145	-		
Perfluoropentanesulfonic Acid (PFPeS)	106		-		65-140	-		
Perfluoroheptanoic Acid (PFHpA)	104		-		70-150	-		
Perfluorohexanesulfonic Acid (PFHxS)	117		-		65-145	-		
Perfluorooctanoic Acid (PFOA)	118		-		70-150	-		
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2FTS)	100		-		65-155	-		
Perfluoroheptanesulfonic Acid (PFHpS)	109		-		70-150	-		
Perfluorononanoic Acid (PFNA)	112		-		70-150	-		
Perfluorooctanesulfonic Acid (PFOS)	105		-		55-150	-		
Perfluorodecanoic Acid (PFDA)	111		-		70-140	-		
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2FTS)	107		-		60-150	-		
Perfluorononanesulfonic Acid (PFNS)	80		-		65-145	-		
N-Methyl Perfluorooctanesulfonamidoacetic Acid (NMeFOSAA)	80		-		50-140	-		
Perfluoroundecanoic Acid (PFUnA)	96		-		70-145	-		

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	Low Level LCS %Recovery	Qual	Low Level LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab Associated sample(s): 01-06 Batch: WG2139125-2 LOW LEVEL								
Perfluorodecanesulfonic Acid (PFDS)	70		-		60-145	-		
Perfluorooctanesulfonamide (PFOSA)	88		-		70-145	-		
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (NEtFOSAA)	132		-		70-145	-		
Perfluorododecanoic Acid (PFDoA)	92		-		70-140	-		
Perfluorotridecanoic Acid (PFTrDA)	113		-		65-140	-		
Perfluorotetradecanoic Acid (PFTeDA)	114		-		60-140	-		
Hexafluoropropylene Oxide Dimer Acid (HFPO-DA)	106		-		70-140	-		
4,8-Dioxa-3h-Perfluorononanoic Acid (ADONA)	111		-		65-145	-		
Perfluorododecanesulfonic Acid (PFDoS)	69		-		50-145	-		
9-Chlorohexadecafluoro-3-Oxanone- 1-Sulfonic Acid (9Cl-PF3ONS)	101		-		70-155	-		
11-Chloroeicosafluoro-3- Oxaundecane-1-Sulfonic Acid (11Cl- PF3OUdS)	79		-		55-160	-		
N-Methyl Perfluorooctane Sulfonamide (NMeFOSA)	110		-		60-150	-		
N-Ethyl Perfluorooctane Sulfonamide (NEtFOSA)	102		-		65-145	-		
N-Methyl Perfluorooctanesulfonamido Ethanol (NMeFOSE)	106		-		70-145	-		
N-Ethyl Perfluorooctanesulfonamido Ethanol (NEtFOSE)	105		-		70-135	-		
Perfluoro-3-Methoxypropanoic Acid (PFMPA)	98		-		55-140	-		

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	Low Level LCS %Recovery	Qual	Low Level LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab Associated sample(s): 01-06 Batch: WG2139125-2 LOW LEVEL								
Perfluoro-4-Methoxybutanoic Acid (PFMBA)	108		-		60-150	-		
Perfluoro(2-Ethoxyethane)Sulfonic Acid (PFEEESA)	89		-		70-140	-		
Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)	101		-		50-150	-		
3-Perfluoropropyl Propanoic Acid (3:3FTCA)	101		-		65-130	-		
2H,2H,3H,3H-Perfluorooctanoic Acid (5:3FTCA)	82		-		70-135	-		
3-Perfluoroheptyl Propanoic Acid (7:3FTCA)	76		-		50-145	-		

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	Low Level LCS		Low Level LCSD		%Recovery Limits		RPD	
	%Recovery	Qual	%Recovery	Qual	RPD	Qual	RPD	
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab Associated sample(s): 01-06 Batch: WG2139125-2 LOW LEVEL								

Surrogate	LCS %Recovery	Qual	LCSD %Recovery	Qual	Acceptance Criteria
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)	93				5-130
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)	89				40-130
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)	92				40-135
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)	90				40-200
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)	93				40-130
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)	89				40-130
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)	82				40-130
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)	89				40-130
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)	93				40-200
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)	88				40-130
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)	77				40-130
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)	80				40-130
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)	76				40-300
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)	55				40-170
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)	72				30-130
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)	62				40-130
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)	52				25-135
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)	53				10-130
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)	50				10-130
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)	89				40-130
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)	54				10-130
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)	57				10-130
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)	61				10-130
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)	64				10-130

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab Associated sample(s): 01-06 Batch: WG2139125-3								
Perfluorobutanoic Acid (PFBA)	96		-		70-140	-		
Perfluoropentanoic Acid (PFPeA)	104		-		65-135	-		
Perfluorobutanesulfonic Acid (PFBS)	90		-		60-145	-		
1H,1H,2H,2H-Perfluorohexanesulfonic Acid (4:2FTS)	100		-		70-145	-		
Perfluorohexanoic Acid (PFHxA)	92		-		70-145	-		
Perfluoropentanesulfonic Acid (PFPeS)	99		-		65-140	-		
Perfluoroheptanoic Acid (PFHpA)	95		-		70-150	-		
Perfluorohexanesulfonic Acid (PFHxS)	94		-		65-145	-		
Perfluorooctanoic Acid (PFOA)	114		-		70-150	-		
1H,1H,2H,2H-Perfluorooctanesulfonic Acid (6:2FTS)	93		-		65-155	-		
Perfluoroheptanesulfonic Acid (PFHpS)	98		-		70-150	-		
Perfluorononanoic Acid (PFNA)	107		-		70-150	-		
Perfluorooctanesulfonic Acid (PFOS)	95		-		55-150	-		
Perfluorodecanoic Acid (PFDA)	93		-		70-140	-		
1H,1H,2H,2H-Perfluorodecanesulfonic Acid (8:2FTS)	101		-		60-150	-		
Perfluorononanesulfonic Acid (PFNS)	89		-		65-145	-		
N-Methyl Perfluorooctanesulfonamidoacetic Acid (NMeFOSAA)	104		-		50-140	-		
Perfluoroundecanoic Acid (PFUnA)	102		-		70-145	-		

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab Associated sample(s): 01-06 Batch: WG2139125-3								
Perfluorodecanesulfonic Acid (PFDS)	78		-		60-145	-		
Perfluorooctanesulfonamide (PFOSA)	95		-		70-145	-		
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (NEtFOSAA)	96		-		70-145	-		
Perfluorododecanoic Acid (PFDoA)	96		-		70-140	-		
Perfluorotridecanoic Acid (PFTrDA)	110		-		65-140	-		
Perfluorotetradecanoic Acid (PFTeDA)	106		-		60-140	-		
Hexafluoropropylene Oxide Dimer Acid (HFPO-DA)	98		-		70-140	-		
4,8-Dioxa-3h-Perfluorononanoic Acid (ADONA)	96		-		65-145	-		
Perfluorododecanesulfonic Acid (PFDoS)	77		-		50-145	-		
9-Chlorohexadecafluoro-3-Oxanone- 1-Sulfonic Acid (9Cl-PF3ONS)	104		-		70-155	-		
11-Chloroeicosafluoro-3- Oxaundecane-1-Sulfonic Acid (11Cl- PF3OUdS)	95		-		55-160	-		
N-Methyl Perfluorooctane Sulfonamide (NMeFOSA)	95		-		60-150	-		
N-Ethyl Perfluorooctane Sulfonamide (NEtFOSA)	98		-		65-145	-		
N-Methyl Perfluorooctanesulfonamido Ethanol (NMeFOSE)	101		-		70-145	-		
N-Ethyl Perfluorooctanesulfonamido Ethanol (NEtFOSE)	100		-		70-135	-		
Perfluoro-3-Methoxypropanoic Acid (PFMPA)	96		-		55-140	-		

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab Associated sample(s): 01-06 Batch: WG2139125-3								
Perfluoro-4-Methoxybutanoic Acid (PFMBA)	107		-		60-150	-		
Perfluoro(2-Ethoxyethane)Sulfonic Acid (PFEEESA)	85		-		70-140	-		
Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)	94		-		50-150	-		
3-Perfluoropropyl Propanoic Acid (3:3FTCA)	91		-		65-130	-		
2H,2H,3H,3H-Perfluorooctanoic Acid (5:3FTCA)	84		-		70-135	-		
3-Perfluoroheptyl Propanoic Acid (7:3FTCA)	78		-		50-145	-		

Lab Control Sample Analysis

Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCS %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
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Perfluorinated Alkyl Acids by EPA 1633 - Mansfield Lab Associated sample(s): 01-06 Batch: WG2139125-3

Surrogate	LCS %Recovery	Qual	LCS %Recovery	Qual	Acceptance Criteria
Perfluoro-n-[13C4]Butanoic Acid (13C4-PFBA)	95				5-130
Perfluoro-n-[13C5]Pentanoic Acid (13C5-PFPeA)	88				40-130
Perfluoro-1-[2,3,4-13C3]Butanesulfonic Acid (13C3-PFBS)	95				40-135
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Hexanesulfonic Acid (13C2-4:2FTS)	91				40-200
Perfluoro-n-[1,2,3,4,6-13C5]Hexanoic Acid (13C5-PFHxA)	98				40-130
Perfluoro-n-[1,2,3,4-13C4]Heptanoic Acid (13C4-PFHpA)	91				40-130
Perfluoro-1-[1,2,3-13C3]Hexanesulfonic Acid (13C3-PFHxS)	88				40-130
Perfluoro-n-[13C8]Octanoic Acid (13C8-PFOA)	79				40-130
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Octanesulfonic Acid (13C2-6:2FTS)	109				40-200
Perfluoro-n-[13C9]Nonanoic Acid (13C9-PFNA)	91				40-130
Perfluoro-1-[13C8]Octanesulfonic Acid (13C8-PFOS)	83				40-130
Perfluoro-n-[1,2,3,4,5,6-13C6]Decanoic Acid (13C6-PFDA)	93				40-130
1H,1H,2H,2H-Perfluoro-1-[1,2-13C2]Decanesulfonic Acid (13C2-8:2FTS)	91				40-300
N-Methyl-d3-perfluoro-1-octanesulfonamidoacetic Acid (D3-NMeFOSAA)	62				40-170
Perfluoro-n-[1,2,3,4,5,6,7-13C7]Undecanoic Acid (13C7-PFUnA)	80				30-130
Perfluoro-1-[13C8]Octanesulfonamide (13C8-PFOSA)	72				40-130
N-Ethyl-d5-perfluoro-1-octanesulfonamidoacetic Acid (D5-NEtFOSAA)	70				25-135
Perfluoro-n-[1,2-13C2]Dodecanoic Acid (13C2-PFDoA)	65				10-130
Perfluoro-n-[1,2-13C2]Tetradecanoic Acid (13C2-PFTeDA)	63				10-130
Tetrafluoro-2-heptafluoropropoxy-[13C3]-propanoic acid (13C3-HFPO-DA)	88				40-130
N-Methyl-d3-Perfluoro-1-Octanesulfonamide (D3-NMeFOSA)	69				10-130
N-Ethyl-d5-Perfluoro-1-Octanesulfonamide (D5-NEtFOSA)	68				10-130
N-Methyl-d7-Perfluorooctanesulfonamidoethanol (D7-NMeFOSE)	72				10-130
N-Ethyl-d9-Perfluorooctanesulfonamidoethanol (D9-NEtFOSE)	77				10-130

PCBS

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-01
 Client ID: GZM-2
 Sample Location: BRONX, NY

Date Collected: 11/04/25 09:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8082A
 Analytical Date: 11/07/25 14:05
 Analyst: SDC

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:39
 Cleanup Method: EPA 3665A
 Cleanup Date: 11/07/25
 Cleanup Method: EPA 3660B
 Cleanup Date: 11/07/25

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Polychlorinated Biphenyls by GC - Westborough Lab							
Aroclor 1016	ND		ug/l	0.071	0.021	1	A
Aroclor 1221	ND		ug/l	0.071	0.026	1	A
Aroclor 1232	ND		ug/l	0.071	0.026	1	A
Aroclor 1242	ND		ug/l	0.071	0.026	1	A
Aroclor 1248	ND		ug/l	0.071	0.026	1	A
Aroclor 1254	ND		ug/l	0.071	0.026	1	A
Aroclor 1260	ND		ug/l	0.071	0.026	1	A
Aroclor 1262	ND		ug/l	0.071	0.026	1	A
Aroclor 1268	ND		ug/l	0.071	0.026	1	A
PCBs, Total	ND		ug/l	0.071	0.021	1	A

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	74		30-150	A
Decachlorobiphenyl	60		30-150	A
2,4,5,6-Tetrachloro-m-xylene	81		30-150	B
Decachlorobiphenyl	56		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-02
 Client ID: GZM-1
 Sample Location: BRONX, NY

Date Collected: 11/04/25 10:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8082A
 Analytical Date: 11/07/25 14:13
 Analyst: SDC

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:39
 Cleanup Method: EPA 3665A
 Cleanup Date: 11/07/25
 Cleanup Method: EPA 3660B
 Cleanup Date: 11/07/25

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Polychlorinated Biphenyls by GC - Westborough Lab							
Aroclor 1016	ND		ug/l	0.071	0.021	1	A
Aroclor 1221	ND		ug/l	0.071	0.026	1	A
Aroclor 1232	ND		ug/l	0.071	0.026	1	A
Aroclor 1242	ND		ug/l	0.071	0.026	1	A
Aroclor 1248	ND		ug/l	0.071	0.026	1	A
Aroclor 1254	ND		ug/l	0.071	0.026	1	A
Aroclor 1260	ND		ug/l	0.071	0.026	1	A
Aroclor 1262	ND		ug/l	0.071	0.026	1	A
Aroclor 1268	ND		ug/l	0.071	0.026	1	A
PCBs, Total	ND		ug/l	0.071	0.021	1	A

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	83		30-150	A
Decachlorobiphenyl	66		30-150	A
2,4,5,6-Tetrachloro-m-xylene	90		30-150	B
Decachlorobiphenyl	67		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-03
 Client ID: GZM-3
 Sample Location: BRONX, NY

Date Collected: 11/04/25 11:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8082A
 Analytical Date: 11/07/25 14:21
 Analyst: SDC

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:39
 Cleanup Method: EPA 3665A
 Cleanup Date: 11/07/25
 Cleanup Method: EPA 3660B
 Cleanup Date: 11/07/25

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Polychlorinated Biphenyls by GC - Westborough Lab							
Aroclor 1016	ND		ug/l	0.071	0.021	1	A
Aroclor 1221	ND		ug/l	0.071	0.026	1	A
Aroclor 1232	ND		ug/l	0.071	0.026	1	A
Aroclor 1242	ND		ug/l	0.071	0.026	1	A
Aroclor 1248	ND		ug/l	0.071	0.026	1	A
Aroclor 1254	ND		ug/l	0.071	0.026	1	A
Aroclor 1260	ND		ug/l	0.071	0.026	1	A
Aroclor 1262	ND		ug/l	0.071	0.026	1	A
Aroclor 1268	ND		ug/l	0.071	0.026	1	A
PCBs, Total	ND		ug/l	0.071	0.021	1	A

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	72		30-150	A
Decachlorobiphenyl	59		30-150	A
2,4,5,6-Tetrachloro-m-xylene	77		30-150	B
Decachlorobiphenyl	57		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-04
 Client ID: GZM-4
 Sample Location: BRONX, NY

Date Collected: 11/04/25 12:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8082A
 Analytical Date: 11/07/25 14:30
 Analyst: SDC

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:39
 Cleanup Method: EPA 3665A
 Cleanup Date: 11/07/25
 Cleanup Method: EPA 3660B
 Cleanup Date: 11/07/25

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Polychlorinated Biphenyls by GC - Westborough Lab							
Aroclor 1016	ND		ug/l	0.071	0.021	1	A
Aroclor 1221	ND		ug/l	0.071	0.026	1	A
Aroclor 1232	ND		ug/l	0.071	0.026	1	A
Aroclor 1242	ND		ug/l	0.071	0.026	1	A
Aroclor 1248	ND		ug/l	0.071	0.026	1	A
Aroclor 1254	0.088		ug/l	0.071	0.026	1	B
Aroclor 1260	ND		ug/l	0.071	0.026	1	A
Aroclor 1262	ND		ug/l	0.071	0.026	1	A
Aroclor 1268	ND		ug/l	0.071	0.026	1	A
PCBs, Total	0.088		ug/l	0.071	0.021	1	B

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	68		30-150	A
Decachlorobiphenyl	55		30-150	A
2,4,5,6-Tetrachloro-m-xylene	71		30-150	B
Decachlorobiphenyl	54		30-150	B

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-05

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Extraction Method: EPA 3510C

Analytical Method: 1,8082A

Extraction Date: 11/07/25 05:39

Analytical Date: 11/07/25 14:38

Cleanup Method: EPA 3665A

Analyst: SDC

Cleanup Date: 11/07/25

Cleanup Method: EPA 3660B

Cleanup Date: 11/07/25

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Polychlorinated Biphenyls by GC - Westborough Lab							
Aroclor 1016	ND		ug/l	0.071	0.021	1	A
Aroclor 1221	ND		ug/l	0.071	0.026	1	A
Aroclor 1232	ND		ug/l	0.071	0.026	1	A
Aroclor 1242	ND		ug/l	0.071	0.026	1	A
Aroclor 1248	ND		ug/l	0.071	0.026	1	A
Aroclor 1254	ND		ug/l	0.071	0.026	1	A
Aroclor 1260	ND		ug/l	0.071	0.026	1	A
Aroclor 1262	ND		ug/l	0.071	0.026	1	A
Aroclor 1268	ND		ug/l	0.071	0.026	1	A
PCBs, Total	ND		ug/l	0.071	0.021	1	A

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	65		30-150	A
Decachlorobiphenyl	54		30-150	A
2,4,5,6-Tetrachloro-m-xylene	70		30-150	B
Decachlorobiphenyl	55		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-06
 Client ID: FIELD BLANK
 Sample Location: BRONX, NY

Date Collected: 11/04/25 14:00
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8082A
 Analytical Date: 11/07/25 14:46
 Analyst: SDC

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:39
 Cleanup Method: EPA 3665A
 Cleanup Date: 11/07/25
 Cleanup Method: EPA 3660B
 Cleanup Date: 11/07/25

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Polychlorinated Biphenyls by GC - Westborough Lab							
Aroclor 1016	ND		ug/l	0.071	0.021	1	A
Aroclor 1221	ND		ug/l	0.071	0.026	1	A
Aroclor 1232	ND		ug/l	0.071	0.026	1	A
Aroclor 1242	ND		ug/l	0.071	0.026	1	A
Aroclor 1248	ND		ug/l	0.071	0.026	1	A
Aroclor 1254	ND		ug/l	0.071	0.026	1	A
Aroclor 1260	ND		ug/l	0.071	0.026	1	A
Aroclor 1262	ND		ug/l	0.071	0.026	1	A
Aroclor 1268	ND		ug/l	0.071	0.026	1	A
PCBs, Total	ND		ug/l	0.071	0.021	1	A

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	78		30-150	A
Decachlorobiphenyl	36		30-150	A
2,4,5,6-Tetrachloro-m-xylene	85		30-150	B
Decachlorobiphenyl	37		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8082A
Analytical Date: 11/07/25 11:01
Analyst: SDC

Extraction Method: EPA 3510C
Extraction Date: 11/06/25 15:53
Cleanup Method: EPA 3665A
Cleanup Date: 11/07/25
Cleanup Method: EPA 3660B
Cleanup Date: 11/07/25

Parameter	Result	Qualifier	Units	RL	MDL	Column
Polychlorinated Biphenyls by GC - Westborough Lab for sample(s): 01-06 Batch: WG2138169-1						
Aroclor 1016	ND		ug/l	0.071	0.021	A
Aroclor 1221	ND		ug/l	0.071	0.026	A
Aroclor 1232	ND		ug/l	0.071	0.026	A
Aroclor 1242	ND		ug/l	0.071	0.026	A
Aroclor 1248	ND		ug/l	0.071	0.026	A
Aroclor 1254	ND		ug/l	0.071	0.026	A
Aroclor 1260	ND		ug/l	0.071	0.026	A
Aroclor 1262	ND		ug/l	0.071	0.026	A
Aroclor 1268	ND		ug/l	0.071	0.026	A
PCBs, Total	ND		ug/l	0.071	0.021	A

Surrogate	%Recovery	Qualifier	Acceptance	
			Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	66		30-150	A
Decachlorobiphenyl	57		30-150	A
2,4,5,6-Tetrachloro-m-xylene	71		30-150	B
Decachlorobiphenyl	63		30-150	B

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits	Column
Polychlorinated Biphenyls by GC - Westborough Lab Associated sample(s): 01-06 Batch: WG2138169-2 WG2138169-3									
Aroclor 1016	68		70		40-140	3		50	A
Aroclor 1260	69		76		40-140	9		50	A

Surrogate	LCS %Recovery	Qual	LCSD %Recovery	Qual	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	69		68		30-150	A
Decachlorobiphenyl	36		62		30-150	A
2,4,5,6-Tetrachloro-m-xylene	74		73		30-150	B
Decachlorobiphenyl	38		50		30-150	B

PESTICIDES

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-01
 Client ID: GZM-2
 Sample Location: BRONX, NY

Date Collected: 11/04/25 09:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8081B
 Analytical Date: 11/07/25 15:22
 Analyst: PEG

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:37

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							
Delta-BHC	ND		ug/l	0.014	0.006	1	A
Lindane	ND		ug/l	0.014	0.005	1	A
Alpha-BHC	ND		ug/l	0.014	0.006	1	A
Beta-BHC	ND		ug/l	0.020	0.014	1	A
Heptachlor	ND		ug/l	0.014	0.005	1	A
Aldrin	ND		ug/l	0.014	0.005	1	A
Heptachlor epoxide	ND		ug/l	0.014	0.005	1	A
Endrin	ND		ug/l	0.029	0.008	1	A
Endrin aldehyde	ND		ug/l	0.030	0.018	1	A
Endrin ketone	ND		ug/l	0.029	0.014	1	A
Dieldrin	ND		ug/l	0.029	0.004	1	A
4,4'-DDE	ND		ug/l	0.029	0.010	1	A
4,4'-DDD	ND		ug/l	0.029	0.010	1	A
4,4'-DDT	ND		ug/l	0.029	0.024	1	A
Endosulfan I	ND		ug/l	0.014	0.005	1	A
Endosulfan II	ND		ug/l	0.029	0.008	1	A
Endosulfan sulfate	ND		ug/l	0.029	0.007	1	A
Methoxychlor	ND		ug/l	0.143	0.014	1	A
Toxaphene	ND		ug/l	0.200	0.150	1	A
cis-Chlordane	ND		ug/l	0.020	0.007	1	A
trans-Chlordane	ND		ug/l	0.020	0.011	1	A
Chlordane	ND		ug/l	0.143	0.098	1	A

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	109		30-150	A
Decachlorobiphenyl	74		30-150	A
2,4,5,6-Tetrachloro-m-xylene	123		30-150	B
Decachlorobiphenyl	102		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-02
 Client ID: GZM-1
 Sample Location: BRONX, NY

Date Collected: 11/04/25 10:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8081B
 Analytical Date: 11/07/25 15:34
 Analyst: PEG

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:37

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							
Delta-BHC	ND		ug/l	0.014	0.006	1	A
Lindane	ND		ug/l	0.014	0.005	1	A
Alpha-BHC	ND		ug/l	0.014	0.006	1	A
Beta-BHC	ND		ug/l	0.020	0.014	1	A
Heptachlor	ND		ug/l	0.014	0.005	1	A
Aldrin	ND		ug/l	0.014	0.005	1	A
Heptachlor epoxide	ND		ug/l	0.014	0.005	1	A
Endrin	ND		ug/l	0.029	0.008	1	A
Endrin aldehyde	ND		ug/l	0.030	0.018	1	A
Endrin ketone	ND		ug/l	0.029	0.014	1	A
Dieldrin	ND		ug/l	0.029	0.004	1	A
4,4'-DDE	ND		ug/l	0.029	0.010	1	A
4,4'-DDD	ND		ug/l	0.029	0.010	1	A
4,4'-DDT	ND		ug/l	0.029	0.024	1	B
Endosulfan I	ND		ug/l	0.014	0.005	1	A
Endosulfan II	ND		ug/l	0.029	0.008	1	A
Endosulfan sulfate	ND		ug/l	0.029	0.007	1	A
Methoxychlor	ND		ug/l	0.143	0.014	1	A
Toxaphene	ND		ug/l	0.200	0.150	1	A
cis-Chlordane	ND		ug/l	0.020	0.007	1	A
trans-Chlordane	ND		ug/l	0.020	0.011	1	A
Chlordane	ND		ug/l	0.143	0.098	1	A

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	110		30-150	A
Decachlorobiphenyl	70		30-150	A
2,4,5,6-Tetrachloro-m-xylene	120		30-150	B
Decachlorobiphenyl	98		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-03
 Client ID: GZM-3
 Sample Location: BRONX, NY

Date Collected: 11/04/25 11:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8081B
 Analytical Date: 11/07/25 15:47
 Analyst: PEG

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:37

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							
Delta-BHC	ND		ug/l	0.014	0.006	1	A
Lindane	ND		ug/l	0.014	0.005	1	A
Alpha-BHC	ND		ug/l	0.014	0.006	1	A
Beta-BHC	ND		ug/l	0.020	0.014	1	A
Heptachlor	ND		ug/l	0.014	0.005	1	A
Aldrin	ND		ug/l	0.014	0.005	1	A
Heptachlor epoxide	ND		ug/l	0.014	0.005	1	A
Endrin	ND		ug/l	0.029	0.008	1	A
Endrin aldehyde	ND		ug/l	0.030	0.018	1	A
Endrin ketone	ND		ug/l	0.029	0.014	1	A
Dieldrin	ND		ug/l	0.029	0.004	1	A
4,4'-DDE	ND		ug/l	0.029	0.010	1	A
4,4'-DDD	ND		ug/l	0.029	0.010	1	A
4,4'-DDT	0.024	J	ug/l	0.029	0.024	1	A
Endosulfan I	ND		ug/l	0.014	0.005	1	A
Endosulfan II	ND		ug/l	0.029	0.008	1	A
Endosulfan sulfate	ND		ug/l	0.029	0.007	1	A
Methoxychlor	ND		ug/l	0.143	0.014	1	A
Toxaphene	ND		ug/l	0.200	0.150	1	A
cis-Chlordane	ND		ug/l	0.020	0.007	1	A
trans-Chlordane	ND		ug/l	0.020	0.011	1	A
Chlordane	ND		ug/l	0.143	0.098	1	A

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-03

Date Collected: 11/04/25 11:30

Client ID: GZM-3

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	108		30-150	A
Decachlorobiphenyl	68		30-150	A
2,4,5,6-Tetrachloro-m-xylene	109		30-150	B
Decachlorobiphenyl	86		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-04
 Client ID: GZM-4
 Sample Location: BRONX, NY

Date Collected: 11/04/25 12:30
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8081B
 Analytical Date: 11/07/25 15:59
 Analyst: PEG

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:37

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							
Delta-BHC	ND		ug/l	0.014	0.006	1	A
Lindane	ND		ug/l	0.014	0.005	1	A
Alpha-BHC	ND		ug/l	0.014	0.006	1	A
Beta-BHC	ND		ug/l	0.020	0.014	1	A
Heptachlor	ND		ug/l	0.014	0.005	1	A
Aldrin	ND		ug/l	0.014	0.005	1	A
Heptachlor epoxide	ND		ug/l	0.014	0.005	1	A
Endrin	ND		ug/l	0.029	0.008	1	A
Endrin aldehyde	ND		ug/l	0.030	0.018	1	A
Endrin ketone	ND		ug/l	0.029	0.014	1	A
Dieldrin	ND		ug/l	0.029	0.004	1	A
4,4'-DDE	ND		ug/l	0.029	0.010	1	A
4,4'-DDD	ND		ug/l	0.029	0.010	1	A
4,4'-DDT	ND		ug/l	0.029	0.024	1	A
Endosulfan I	ND		ug/l	0.014	0.005	1	A
Endosulfan II	ND		ug/l	0.029	0.008	1	A
Endosulfan sulfate	ND		ug/l	0.029	0.007	1	A
Methoxychlor	ND		ug/l	0.143	0.014	1	A
Toxaphene	ND		ug/l	0.200	0.150	1	A
cis-Chlordane	ND		ug/l	0.020	0.007	1	A
trans-Chlordane	ND		ug/l	0.020	0.011	1	A
Chlordane	ND		ug/l	0.143	0.098	1	A

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-04

Date Collected: 11/04/25 12:30

Client ID: GZM-4

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	120		30-150	A
Decachlorobiphenyl	90		30-150	A
2,4,5,6-Tetrachloro-m-xylene	114		30-150	B
Decachlorobiphenyl	99		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-05
 Client ID: GZM-DUP
 Sample Location: BRONX, NY

Date Collected: 11/04/25 00:00
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8081B
 Analytical Date: 11/07/25 16:11
 Analyst: PEG

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:37

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							
Delta-BHC	ND		ug/l	0.014	0.006	1	A
Lindane	ND		ug/l	0.014	0.005	1	A
Alpha-BHC	ND		ug/l	0.014	0.006	1	A
Beta-BHC	ND		ug/l	0.020	0.014	1	A
Heptachlor	ND		ug/l	0.014	0.005	1	A
Aldrin	ND		ug/l	0.014	0.005	1	A
Heptachlor epoxide	ND		ug/l	0.014	0.005	1	A
Endrin	ND		ug/l	0.029	0.008	1	A
Endrin aldehyde	ND		ug/l	0.030	0.018	1	A
Endrin ketone	ND		ug/l	0.029	0.014	1	A
Dieldrin	ND		ug/l	0.029	0.004	1	A
4,4'-DDE	ND		ug/l	0.029	0.010	1	A
4,4'-DDD	ND		ug/l	0.029	0.010	1	A
4,4'-DDT	ND		ug/l	0.029	0.024	1	A
Endosulfan I	ND		ug/l	0.014	0.005	1	A
Endosulfan II	ND		ug/l	0.029	0.008	1	A
Endosulfan sulfate	ND		ug/l	0.029	0.007	1	A
Methoxychlor	ND		ug/l	0.143	0.014	1	A
Toxaphene	ND		ug/l	0.200	0.150	1	A
cis-Chlordane	ND		ug/l	0.020	0.007	1	A
trans-Chlordane	ND		ug/l	0.020	0.011	1	A
Chlordane	ND		ug/l	0.143	0.098	1	A

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-05

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	102		30-150	A
Decachlorobiphenyl	74		30-150	A
2,4,5,6-Tetrachloro-m-xylene	110		30-150	B
Decachlorobiphenyl	100		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-06
 Client ID: FIELD BLANK
 Sample Location: BRONX, NY

Date Collected: 11/04/25 14:00
 Date Received: 11/04/25
 Field Prep: Not Specified

Sample Depth:

Matrix: Water
 Analytical Method: 1,8081B
 Analytical Date: 11/07/25 16:24
 Analyst: PEG

Extraction Method: EPA 3510C
 Extraction Date: 11/07/25 05:37

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							
Delta-BHC	ND		ug/l	0.014	0.006	1	A
Lindane	ND		ug/l	0.014	0.005	1	A
Alpha-BHC	ND		ug/l	0.014	0.006	1	A
Beta-BHC	ND		ug/l	0.020	0.014	1	A
Heptachlor	ND		ug/l	0.014	0.005	1	A
Aldrin	ND		ug/l	0.014	0.005	1	A
Heptachlor epoxide	ND		ug/l	0.014	0.005	1	A
Endrin	ND		ug/l	0.029	0.008	1	A
Endrin aldehyde	ND		ug/l	0.030	0.018	1	A
Endrin ketone	ND		ug/l	0.029	0.014	1	A
Dieldrin	ND		ug/l	0.029	0.004	1	A
4,4'-DDE	ND		ug/l	0.029	0.010	1	A
4,4'-DDD	ND		ug/l	0.029	0.010	1	A
4,4'-DDT	0.025	J	ug/l	0.029	0.024	1	A
Endosulfan I	ND		ug/l	0.014	0.005	1	A
Endosulfan II	ND		ug/l	0.029	0.008	1	A
Endosulfan sulfate	ND		ug/l	0.029	0.007	1	A
Methoxychlor	ND		ug/l	0.143	0.014	1	A
Toxaphene	ND		ug/l	0.200	0.150	1	A
cis-Chlordane	ND		ug/l	0.020	0.007	1	A
trans-Chlordane	ND		ug/l	0.020	0.011	1	A
Chlordane	ND		ug/l	0.143	0.098	1	A

Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-06

Date Collected: 11/04/25 14:00

Client ID: FIELD BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Column
Organochlorine Pesticides by GC - Westborough Lab							

Surrogate	% Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	114		30-150	A
Decachlorobiphenyl	44		30-150	A
2,4,5,6-Tetrachloro-m-xylene	113		30-150	B
Decachlorobiphenyl	53		30-150	B

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8081B
Analytical Date: 11/07/25 12:41
Analyst: JAG

Extraction Method: EPA 3510C
Extraction Date: 11/06/25 15:53

Parameter	Result	Qualifier	Units	RL	MDL	Column
Organochlorine Pesticides by GC - Westborough Lab for sample(s): 01-06 Batch: WG2138168-1						
Delta-BHC	ND		ug/l	0.014	0.006	A
Lindane	ND		ug/l	0.014	0.005	A
Alpha-BHC	ND		ug/l	0.014	0.006	A
Beta-BHC	ND		ug/l	0.020	0.014	A
Heptachlor	ND		ug/l	0.014	0.005	A
Aldrin	ND		ug/l	0.014	0.005	A
Heptachlor epoxide	ND		ug/l	0.014	0.005	A
Endrin	ND		ug/l	0.029	0.008	A
Endrin aldehyde	ND		ug/l	0.030	0.018	A
Endrin ketone	ND		ug/l	0.029	0.014	A
Dieldrin	ND		ug/l	0.029	0.004	A
4,4'-DDE	ND		ug/l	0.029	0.010	A
4,4'-DDD	ND		ug/l	0.029	0.010	A
4,4'-DDT	ND		ug/l	0.029	0.024	A
Endosulfan I	ND		ug/l	0.014	0.005	A
Endosulfan II	ND		ug/l	0.029	0.008	A
Endosulfan sulfate	ND		ug/l	0.029	0.007	A
Methoxychlor	ND		ug/l	0.143	0.014	A
Toxaphene	ND		ug/l	0.200	0.150	A
cis-Chlordane	ND		ug/l	0.020	0.007	A
trans-Chlordane	ND		ug/l	0.020	0.011	A
Chlordane	ND		ug/l	0.143	0.098	A

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis
Batch Quality Control

Analytical Method: 1,8081B
Analytical Date: 11/07/25 12:41
Analyst: JAG

Extraction Method: EPA 3510C
Extraction Date: 11/06/25 15:53

Parameter	Result	Qualifier	Units	RL	MDL	Column
Organochlorine Pesticides by GC - Westborough Lab for sample(s): 01-06 Batch: WG2138168-1						

Surrogate	%Recovery	Qualifier	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	95		30-150	A
Decachlorobiphenyl	104		30-150	A
2,4,5,6-Tetrachloro-m-xylene	89		30-150	B
Decachlorobiphenyl	111		30-150	B

Lab Control Sample Analysis

Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCS %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits	Column
Organochlorine Pesticides by GC - Westborough Lab Associated sample(s): 01-06 Batch: WG2138168-2 WG2138168-3									
Delta-BHC	96		92		30-150	5		20	A
Lindane	120		115		30-150	4		20	A
Alpha-BHC	115		112		30-150	3		20	A
Beta-BHC	121		116		30-150	4		20	A
Heptachlor	118		113		30-150	4		20	A
Aldrin	114		110		30-150	4		20	A
Heptachlor epoxide	123		119		30-150	3		20	A
Endrin	124		119		30-150	4		20	A
Endrin aldehyde	120		114		30-150	5		20	A
Endrin ketone	130		126		30-150	3		20	A
Dieldrin	126		121		30-150	4		20	A
4,4'-DDE	120		116		30-150	3		20	A
4,4'-DDD	128		121		30-150	6		20	A
4,4'-DDT	130		125		30-150	4		20	A
Endosulfan I	125		119		30-150	5		20	A
Endosulfan II	150		144		30-150	4		20	A
Endosulfan sulfate	120		115		30-150	4		20	A
Methoxychlor	142		138		30-150	3		20	A
cis-Chlordane	112		109		30-150	3		20	A

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Parameter	LCS %Recovery	Qual	LCSD %Recovery	Qual	%Recovery Limits	RPD	Qual	RPD Limits
Organochlorine Pesticides by GC - Westborough Lab Associated sample(s): 01-06 Batch: WG2138168-2 WG2138168-3								
trans-Chlordane	125		121		30-150	3		20 A

Surrogate	LCS %Recovery	Qual	LCSD %Recovery	Qual	Acceptance Criteria	Column
2,4,5,6-Tetrachloro-m-xylene	114		110		30-150	A
Decachlorobiphenyl	109		110		30-150	A
2,4,5,6-Tetrachloro-m-xylene	106		101		30-150	B
Decachlorobiphenyl	115		111		30-150	B



METALS



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Total Metals - Mansfield Lab											
Aluminum, Total	0.247		mg/l	0.0100	0.00327	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Antimony, Total	ND		mg/l	0.00400	0.00042	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Arsenic, Total	0.00357		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Barium, Total	0.1781		mg/l	0.00050	0.00017	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Beryllium, Total	ND		mg/l	0.00050	0.00010	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Cadmium, Total	0.00007	J	mg/l	0.00020	0.00005	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Calcium, Total	182.		mg/l	0.100	0.0394	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Chromium, Total	0.00128		mg/l	0.00100	0.00017	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Cobalt, Total	0.00148		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Copper, Total	0.00394		mg/l	0.00100	0.00038	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Iron, Total	23.0		mg/l	0.0500	0.0191	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Lead, Total	0.00412		mg/l	0.00100	0.00034	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Magnesium, Total	21.6		mg/l	0.0700	0.0242	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Manganese, Total	1.814		mg/l	0.00100	0.00044	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Mercury, Total	ND		mg/l	0.00020	0.00009	1	11/06/25 14:52	11/10/25 14:21	EPA 7470A	1,7470A	JWN
Nickel, Total	0.00358		mg/l	0.00200	0.00055	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Potassium, Total	19.5		mg/l	0.100	0.0309	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Selenium, Total	ND		mg/l	0.00500	0.00173	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Silver, Total	ND		mg/l	0.00040	0.00016	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Sodium, Total	42.3		mg/l	0.500	0.0293	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Thallium, Total	ND		mg/l	0.00100	0.00014	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Vanadium, Total	ND		mg/l	0.00500	0.00157	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Zinc, Total	0.01749		mg/l	0.01000	0.00341	1	11/06/25 14:19	11/11/25 10:58	EPA 3005A	1,6020B	SMV
Dissolved Metals - Mansfield Lab											
Aluminum, Dissolved	0.00431	J	mg/l	0.0100	0.00327	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Antimony, Dissolved	ND		mg/l	0.00400	0.00042	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Arsenic, Dissolved	0.00067		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Barium, Dissolved	0.1390		mg/l	0.00050	0.00017	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Beryllium, Dissolved	ND		mg/l	0.00050	0.00010	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-01

Date Collected: 11/04/25 09:30

Client ID: GZM-2

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Cadmium, Dissolved	ND		mg/l	0.00020	0.00005	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Calcium, Dissolved	192.		mg/l	0.100	0.0394	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Chromium, Dissolved	0.00022	J	mg/l	0.00100	0.00017	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Cobalt, Dissolved	0.00120		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Copper, Dissolved	ND		mg/l	0.00100	0.00038	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Iron, Dissolved	0.333		mg/l	0.0500	0.0191	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Lead, Dissolved	ND		mg/l	0.00100	0.00034	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Magnesium, Dissolved	22.6		mg/l	0.0700	0.0242	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Manganese, Dissolved	1.855		mg/l	0.00100	0.00044	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Mercury, Dissolved	ND		mg/l	0.00020	0.00009	1	11/10/25 11:45	11/11/25 07:29	EPA 7470A	1,7470A	JWN
Nickel, Dissolved	0.00227		mg/l	0.00200	0.00055	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Potassium, Dissolved	22.2		mg/l	0.100	0.0309	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Selenium, Dissolved	ND		mg/l	0.00500	0.00173	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Silver, Dissolved	ND		mg/l	0.00040	0.00016	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Sodium, Dissolved	43.1		mg/l	0.500	0.0293	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Thallium, Dissolved	ND		mg/l	0.00100	0.00014	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Vanadium, Dissolved	ND		mg/l	0.00500	0.00157	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV
Zinc, Dissolved	0.00433	J	mg/l	0.01000	0.00341	1	11/07/25 16:13	11/11/25 10:23	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-02

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Total Metals - Mansfield Lab											
Aluminum, Total	1.89		mg/l	0.0100	0.00327	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Antimony, Total	ND		mg/l	0.00400	0.00042	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Arsenic, Total	0.00311		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Barium, Total	0.4363		mg/l	0.00050	0.00017	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Beryllium, Total	ND		mg/l	0.00050	0.00010	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Cadmium, Total	ND		mg/l	0.00020	0.00005	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Calcium, Total	127.		mg/l	0.100	0.0394	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Chromium, Total	0.00475		mg/l	0.00100	0.00017	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Cobalt, Total	0.00195		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Copper, Total	0.01884		mg/l	0.00100	0.00038	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Iron, Total	21.2		mg/l	0.0500	0.0191	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Lead, Total	0.00389		mg/l	0.00100	0.00034	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Magnesium, Total	26.6		mg/l	0.0700	0.0242	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Manganese, Total	3.825		mg/l	0.00100	0.00044	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Mercury, Total	ND		mg/l	0.00020	0.00009	1	11/06/25 14:52	11/10/25 14:36	EPA 7470A	1,7470A	JWN
Nickel, Total	0.00461		mg/l	0.00200	0.00055	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Potassium, Total	19.6		mg/l	0.100	0.0309	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Selenium, Total	ND		mg/l	0.00500	0.00173	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Silver, Total	ND		mg/l	0.00040	0.00016	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Sodium, Total	229.		mg/l	2.50	0.146	5	11/06/25 14:19	11/11/25 12:13	EPA 3005A	1,6020B	SMV
Thallium, Total	ND		mg/l	0.00100	0.00014	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Vanadium, Total	0.00549		mg/l	0.00500	0.00157	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Zinc, Total	0.01022		mg/l	0.01000	0.00341	1	11/06/25 14:19	11/11/25 11:32	EPA 3005A	1,6020B	SMV
Dissolved Metals - Mansfield Lab											
Aluminum, Dissolved	ND		mg/l	0.0100	0.00327	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Antimony, Dissolved	ND		mg/l	0.00400	0.00042	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Arsenic, Dissolved	0.00060		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Barium, Dissolved	0.3149		mg/l	0.00050	0.00017	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Beryllium, Dissolved	ND		mg/l	0.00050	0.00010	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-02

Date Collected: 11/04/25 10:30

Client ID: GZM-1

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Cadmium, Dissolved	ND		mg/l	0.00020	0.00005	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Calcium, Dissolved	121.		mg/l	0.100	0.0394	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Chromium, Dissolved	ND		mg/l	0.00100	0.00017	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Cobalt, Dissolved	0.00029	J	mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Copper, Dissolved	ND		mg/l	0.00100	0.00038	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Iron, Dissolved	0.0358	J	mg/l	0.0500	0.0191	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Lead, Dissolved	0.00060	J	mg/l	0.00100	0.00034	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Magnesium, Dissolved	25.1		mg/l	0.0700	0.0242	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Manganese, Dissolved	3.573		mg/l	0.00100	0.00044	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Mercury, Dissolved	ND		mg/l	0.00020	0.00009	1	11/10/25 11:45	11/11/25 07:33	EPA 7470A	1,7470A	JWN
Nickel, Dissolved	0.00071	J	mg/l	0.00200	0.00055	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Potassium, Dissolved	18.8		mg/l	0.100	0.0309	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Selenium, Dissolved	ND		mg/l	0.00500	0.00173	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Silver, Dissolved	ND		mg/l	0.00040	0.00016	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Sodium, Dissolved	244.		mg/l	0.500	0.0293	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Thallium, Dissolved	ND		mg/l	0.00100	0.00014	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Vanadium, Dissolved	ND		mg/l	0.00500	0.00157	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV
Zinc, Dissolved	ND		mg/l	0.01000	0.00341	1	11/07/25 16:13	11/11/25 10:28	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-03

Date Collected: 11/04/25 11:30

Client ID: GZM-3

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Total Metals - Mansfield Lab											
Aluminum, Total	1.17		mg/l	0.0100	0.00327	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Antimony, Total	0.00055	J	mg/l	0.00400	0.00042	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Arsenic, Total	0.00239		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Barium, Total	0.1324		mg/l	0.00050	0.00017	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Beryllium, Total	ND		mg/l	0.00050	0.00010	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Cadmium, Total	ND		mg/l	0.00020	0.00005	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Calcium, Total	69.1		mg/l	0.100	0.0394	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Chromium, Total	0.00266		mg/l	0.00100	0.00017	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Cobalt, Total	0.00142		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Copper, Total	0.01630		mg/l	0.00100	0.00038	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Iron, Total	1.62		mg/l	0.0500	0.0191	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Lead, Total	0.00294		mg/l	0.00100	0.00034	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Magnesium, Total	9.61		mg/l	0.0700	0.0242	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Manganese, Total	0.4893		mg/l	0.00100	0.00044	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Mercury, Total	ND		mg/l	0.00020	0.00009	1	11/06/25 14:52	11/10/25 14:45	EPA 7470A	1,7470A	JWN
Nickel, Total	0.00441		mg/l	0.00200	0.00055	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Potassium, Total	27.5		mg/l	0.100	0.0309	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Selenium, Total	ND		mg/l	0.00500	0.00173	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Silver, Total	ND		mg/l	0.00040	0.00016	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Sodium, Total	248.		mg/l	2.50	0.146	5	11/06/25 14:19	11/11/25 12:19	EPA 3005A	1,6020B	SMV
Thallium, Total	ND		mg/l	0.00100	0.00014	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Vanadium, Total	0.00527		mg/l	0.00500	0.00157	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Zinc, Total	0.00620	J	mg/l	0.01000	0.00341	1	11/06/25 14:19	11/11/25 11:37	EPA 3005A	1,6020B	SMV
Dissolved Metals - Mansfield Lab											
Aluminum, Dissolved	0.0551		mg/l	0.0100	0.00327	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Antimony, Dissolved	0.00080	J	mg/l	0.00400	0.00042	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Arsenic, Dissolved	0.00212		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Barium, Dissolved	0.1109		mg/l	0.00050	0.00017	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Beryllium, Dissolved	ND		mg/l	0.00050	0.00010	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-03

Date Collected: 11/04/25 11:30

Client ID: GZM-3

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Cadmium, Dissolved	ND		mg/l	0.00020	0.00005	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Calcium, Dissolved	70.6		mg/l	0.100	0.0394	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Chromium, Dissolved	0.00038	J	mg/l	0.00100	0.00017	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Cobalt, Dissolved	0.00037	J	mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Copper, Dissolved	0.00089	J	mg/l	0.00100	0.00038	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Iron, Dissolved	0.0197	J	mg/l	0.0500	0.0191	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Lead, Dissolved	ND		mg/l	0.00100	0.00034	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Magnesium, Dissolved	9.56		mg/l	0.0700	0.0242	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Manganese, Dissolved	0.3445		mg/l	0.00100	0.00044	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Mercury, Dissolved	ND		mg/l	0.00020	0.00009	1	11/10/25 11:45	11/11/25 07:36	EPA 7470A	1,7470A	JWN
Nickel, Dissolved	0.00223		mg/l	0.00200	0.00055	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Potassium, Dissolved	28.2		mg/l	0.100	0.0309	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Selenium, Dissolved	ND		mg/l	0.00500	0.00173	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Silver, Dissolved	ND		mg/l	0.00040	0.00016	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Sodium, Dissolved	265.		mg/l	2.50	0.146	5	11/07/25 16:13	11/11/25 11:05	EPA 3005A	1,6020B	SMV
Thallium, Dissolved	ND		mg/l	0.00100	0.00014	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Vanadium, Dissolved	0.00272	J	mg/l	0.00500	0.00157	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV
Zinc, Dissolved	ND		mg/l	0.01000	0.00341	1	11/07/25 16:13	11/11/25 10:33	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-04

Date Collected: 11/04/25 12:30

Client ID: GZM-4

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Total Metals - Mansfield Lab											
Aluminum, Total	2.60		mg/l	0.0100	0.00327	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Antimony, Total	0.00212	J	mg/l	0.00400	0.00042	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Arsenic, Total	0.00557		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Barium, Total	0.1064		mg/l	0.00050	0.00017	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Beryllium, Total	0.00018	J	mg/l	0.00050	0.00010	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Cadmium, Total	0.00035		mg/l	0.00020	0.00005	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Calcium, Total	177.		mg/l	0.100	0.0394	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Chromium, Total	0.00486		mg/l	0.00100	0.00017	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Cobalt, Total	0.00282		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Copper, Total	0.01367		mg/l	0.00100	0.00038	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Iron, Total	4.42		mg/l	0.0500	0.0191	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Lead, Total	0.08369		mg/l	0.00100	0.00034	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Magnesium, Total	29.6		mg/l	0.0700	0.0242	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Manganese, Total	0.4111		mg/l	0.00100	0.00044	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Mercury, Total	0.00009	J	mg/l	0.00020	0.00009	1	11/06/25 14:52	11/10/25 14:47	EPA 7470A	1,7470A	JWN
Nickel, Total	0.00907		mg/l	0.00200	0.00055	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Potassium, Total	12.8		mg/l	0.100	0.0309	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Selenium, Total	ND		mg/l	0.00500	0.00173	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Silver, Total	ND		mg/l	0.00040	0.00016	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Sodium, Total	24.6		mg/l	0.500	0.0293	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Thallium, Total	ND		mg/l	0.00100	0.00014	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Vanadium, Total	0.00875		mg/l	0.00500	0.00157	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Zinc, Total	0.09718		mg/l	0.01000	0.00341	1	11/06/25 14:19	11/11/25 12:02	EPA 3005A	1,6020B	SMV
Dissolved Metals - Mansfield Lab											
Aluminum, Dissolved	0.00551	J	mg/l	0.0100	0.00327	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Antimony, Dissolved	0.00201	J	mg/l	0.00400	0.00042	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Arsenic, Dissolved	0.00272		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Barium, Dissolved	0.03889		mg/l	0.00050	0.00017	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Beryllium, Dissolved	ND		mg/l	0.00050	0.00010	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES**Lab Number:** L2569921**Project Number:** 41.0163256.00**Report Date:** 11/11/25**SAMPLE RESULTS**

Lab ID: L2569921-04

Date Collected: 11/04/25 12:30

Client ID: GZM-4

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Cadmium, Dissolved	ND		mg/l	0.00020	0.00005	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Calcium, Dissolved	121.		mg/l	0.100	0.0394	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Chromium, Dissolved	ND		mg/l	0.00100	0.00017	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Cobalt, Dissolved	0.00085		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Copper, Dissolved	0.00066	J	mg/l	0.00100	0.00038	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Iron, Dissolved	ND		mg/l	0.0500	0.0191	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Lead, Dissolved	0.00041	J	mg/l	0.00100	0.00034	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Magnesium, Dissolved	22.2		mg/l	0.0700	0.0242	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Manganese, Dissolved	0.2026		mg/l	0.00100	0.00044	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Mercury, Dissolved	ND		mg/l	0.00020	0.00009	1	11/10/25 11:45	11/11/25 07:39	EPA 7470A	1,7470A	JWN
Nickel, Dissolved	0.00373		mg/l	0.00200	0.00055	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Potassium, Dissolved	12.0		mg/l	0.100	0.0309	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Selenium, Dissolved	ND		mg/l	0.00500	0.00173	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Silver, Dissolved	ND		mg/l	0.00040	0.00016	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Sodium, Dissolved	21.0		mg/l	0.500	0.0293	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Thallium, Dissolved	ND		mg/l	0.00100	0.00014	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Vanadium, Dissolved	ND		mg/l	0.00500	0.00157	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV
Zinc, Dissolved	0.02183		mg/l	0.01000	0.00341	1	11/07/25 16:13	11/11/25 10:55	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-05

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Total Metals - Mansfield Lab											
Aluminum, Total	0.370		mg/l	0.0100	0.00327	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Antimony, Total	ND		mg/l	0.00400	0.00042	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Arsenic, Total	0.00399		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Barium, Total	0.1873		mg/l	0.00050	0.00017	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Beryllium, Total	ND		mg/l	0.00050	0.00010	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Cadmium, Total	0.00009	J	mg/l	0.00020	0.00005	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Calcium, Total	192.		mg/l	0.100	0.0394	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Chromium, Total	0.00103		mg/l	0.00100	0.00017	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Cobalt, Total	0.00161		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Copper, Total	0.00434		mg/l	0.00100	0.00038	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Iron, Total	25.4		mg/l	0.0500	0.0191	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Lead, Total	0.00514		mg/l	0.00100	0.00034	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Magnesium, Total	22.3		mg/l	0.0700	0.0242	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Manganese, Total	1.984		mg/l	0.00100	0.00044	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Mercury, Total	ND		mg/l	0.00020	0.00009	1	11/06/25 14:52	11/10/25 14:50	EPA 7470A	1,7470A	JWN
Nickel, Total	0.00357		mg/l	0.00200	0.00055	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Potassium, Total	20.0		mg/l	0.100	0.0309	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Selenium, Total	ND		mg/l	0.00500	0.00173	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Silver, Total	ND		mg/l	0.00040	0.00016	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Sodium, Total	42.8		mg/l	0.500	0.0293	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Thallium, Total	ND		mg/l	0.00100	0.00014	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Vanadium, Total	0.00164	J	mg/l	0.00500	0.00157	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Zinc, Total	0.01988		mg/l	0.01000	0.00341	1	11/06/25 14:19	11/11/25 12:08	EPA 3005A	1,6020B	SMV
Dissolved Metals - Mansfield Lab											
Aluminum, Dissolved	0.0108		mg/l	0.0100	0.00327	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Antimony, Dissolved	ND		mg/l	0.00400	0.00042	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Arsenic, Dissolved	0.00075		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Barium, Dissolved	0.1450		mg/l	0.00050	0.00017	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Beryllium, Dissolved	ND		mg/l	0.00050	0.00010	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-05

Date Collected: 11/04/25 00:00

Client ID: GZM-DUP

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Cadmium, Dissolved	ND		mg/l	0.00020	0.00005	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Calcium, Dissolved	193.		mg/l	0.100	0.0394	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Chromium, Dissolved	0.00062	J	mg/l	0.00100	0.00017	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Cobalt, Dissolved	0.00128		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Copper, Dissolved	ND		mg/l	0.00100	0.00038	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Iron, Dissolved	0.626		mg/l	0.0500	0.0191	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Lead, Dissolved	ND		mg/l	0.00100	0.00034	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Magnesium, Dissolved	22.0		mg/l	0.0700	0.0242	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Manganese, Dissolved	1.942		mg/l	0.00100	0.00044	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Mercury, Dissolved	ND		mg/l	0.00020	0.00009	1	11/10/25 11:45	11/11/25 07:49	EPA 7470A	1,7470A	JWN
Nickel, Dissolved	0.00220		mg/l	0.00200	0.00055	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Potassium, Dissolved	21.5		mg/l	0.100	0.0309	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Selenium, Dissolved	ND		mg/l	0.00500	0.00173	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Silver, Dissolved	ND		mg/l	0.00040	0.00016	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Sodium, Dissolved	42.1		mg/l	0.500	0.0293	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Thallium, Dissolved	ND		mg/l	0.00100	0.00014	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Vanadium, Dissolved	ND		mg/l	0.00500	0.00157	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV
Zinc, Dissolved	ND		mg/l	0.01000	0.00341	1	11/07/25 16:13	11/11/25 11:00	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-06

Date Collected: 11/04/25 14:00

Client ID: FIELD BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Total Metals - Mansfield Lab											
Aluminum, Total	ND		mg/l	0.0100	0.00327	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Antimony, Total	ND		mg/l	0.00400	0.00042	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Arsenic, Total	ND		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Barium, Total	0.00018	JB	mg/l	0.00050	0.00017	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Beryllium, Total	ND		mg/l	0.00050	0.00010	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Cadmium, Total	ND		mg/l	0.00020	0.00005	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Calcium, Total	ND		mg/l	0.100	0.0394	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Chromium, Total	ND		mg/l	0.00100	0.00017	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Cobalt, Total	ND		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Copper, Total	ND		mg/l	0.00100	0.00038	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Iron, Total	ND		mg/l	0.0500	0.0191	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Lead, Total	ND		mg/l	0.00100	0.00034	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Magnesium, Total	ND		mg/l	0.0700	0.0242	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Manganese, Total	ND		mg/l	0.00100	0.00044	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Mercury, Total	ND		mg/l	0.00020	0.00009	1	11/06/25 14:52	11/10/25 14:53	EPA 7470A	1,7470A	JWN
Nickel, Total	ND		mg/l	0.00200	0.00055	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Potassium, Total	ND		mg/l	0.100	0.0309	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Selenium, Total	ND		mg/l	0.00500	0.00173	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Silver, Total	ND		mg/l	0.00040	0.00016	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Sodium, Total	0.180	J	mg/l	0.500	0.0293	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Thallium, Total	ND		mg/l	0.00100	0.00014	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Vanadium, Total	ND		mg/l	0.00500	0.00157	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Zinc, Total	ND		mg/l	0.01000	0.00341	1	11/06/25 14:19	11/11/25 11:57	EPA 3005A	1,6020B	SMV
Dissolved Metals - Mansfield Lab											
Aluminum, Dissolved	ND		mg/l	0.0100	0.00327	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Antimony, Dissolved	ND		mg/l	0.00400	0.00042	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Arsenic, Dissolved	ND		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Barium, Dissolved	ND		mg/l	0.00050	0.00017	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Beryllium, Dissolved	ND		mg/l	0.00050	0.00010	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

SAMPLE RESULTS

Lab ID: L2569921-06

Date Collected: 11/04/25 14:00

Client ID: FIELD BLANK

Date Received: 11/04/25

Sample Location: BRONX, NY

Field Prep: Not Specified

Sample Depth:

Matrix: Water

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Prep Method	Analytical Method	Analyst
Cadmium, Dissolved	ND		mg/l	0.00020	0.00005	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Calcium, Dissolved	0.0614	J	mg/l	0.100	0.0394	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Chromium, Dissolved	ND		mg/l	0.00100	0.00017	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Cobalt, Dissolved	ND		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Copper, Dissolved	0.00350		mg/l	0.00100	0.00038	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Iron, Dissolved	ND		mg/l	0.0500	0.0191	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Lead, Dissolved	ND		mg/l	0.00100	0.00034	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Magnesium, Dissolved	ND		mg/l	0.0700	0.0242	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Manganese, Dissolved	ND		mg/l	0.00100	0.00044	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Mercury, Dissolved	ND		mg/l	0.00020	0.00009	1	11/10/25 11:45	11/11/25 07:53	EPA 7470A	1,7470A	JWN
Nickel, Dissolved	0.00305		mg/l	0.00200	0.00055	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Potassium, Dissolved	ND		mg/l	0.100	0.0309	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Selenium, Dissolved	ND		mg/l	0.00500	0.00173	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Silver, Dissolved	ND		mg/l	0.00040	0.00016	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Sodium, Dissolved	0.184	J	mg/l	0.500	0.0293	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Thallium, Dissolved	ND		mg/l	0.00100	0.00014	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Vanadium, Dissolved	ND		mg/l	0.00500	0.00157	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV
Zinc, Dissolved	ND		mg/l	0.01000	0.00341	1	11/07/25 16:13	11/11/25 10:51	EPA 3005A	1,6020B	SMV



Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis Batch Quality Control

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Analytical Method	Analyst
Total Metals - Mansfield Lab for sample(s): 01-06 Batch: WG2138162-1										
Aluminum, Total	0.00343	J	mg/l	0.0100	0.00327	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Antimony, Total	ND		mg/l	0.00400	0.00042	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Arsenic, Total	0.00017	J	mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Barium, Total	0.00065		mg/l	0.00050	0.00017	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Beryllium, Total	ND		mg/l	0.00050	0.00010	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Cadmium, Total	ND		mg/l	0.00020	0.00005	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Calcium, Total	ND		mg/l	0.100	0.0394	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Chromium, Total	0.00026	J	mg/l	0.00100	0.00017	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Cobalt, Total	ND		mg/l	0.00050	0.00016	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Copper, Total	ND		mg/l	0.00100	0.00038	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Iron, Total	ND		mg/l	0.0500	0.0191	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Lead, Total	ND		mg/l	0.00100	0.00034	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Magnesium, Total	ND		mg/l	0.0700	0.0242	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Manganese, Total	ND		mg/l	0.00100	0.00044	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Nickel, Total	ND		mg/l	0.00200	0.00055	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Potassium, Total	ND		mg/l	0.100	0.0309	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Selenium, Total	ND		mg/l	0.00500	0.00173	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Silver, Total	ND		mg/l	0.00040	0.00016	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Sodium, Total	ND		mg/l	0.500	0.0293	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Thallium, Total	ND		mg/l	0.00100	0.00014	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Vanadium, Total	ND		mg/l	0.00500	0.00157	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV
Zinc, Total	ND		mg/l	0.01000	0.00341	1	11/06/25 14:19	11/11/25 10:47	1,6020B	SMV

Prep Information

Digestion Method: EPA 3005A

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Analytical Method	Analyst
Total Metals - Mansfield Lab for sample(s): 01-06 Batch: WG2138164-1										
Mercury, Total	ND		mg/l	0.00020	0.00009	1	11/06/25 14:52	11/10/25 14:15	1,7470A	JWN



Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Method Blank Analysis Batch Quality Control

Prep Information

Digestion Method: EPA 7470A

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Analytical Method	Analyst
Dissolved Metals - Mansfield Lab for sample(s): 01-06 Batch: WG2138719-1										
Aluminum, Dissolved	ND		mg/l	0.0100	0.00327	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Antimony, Dissolved	0.00063	J	mg/l	0.00400	0.00042	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Arsenic, Dissolved	ND		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Barium, Dissolved	0.00029	J	mg/l	0.00050	0.00017	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Beryllium, Dissolved	ND		mg/l	0.00050	0.00010	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Cadmium, Dissolved	ND		mg/l	0.00020	0.00005	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Calcium, Dissolved	ND		mg/l	0.100	0.0394	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Chromium, Dissolved	ND		mg/l	0.00100	0.00017	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Cobalt, Dissolved	ND		mg/l	0.00050	0.00016	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Copper, Dissolved	ND		mg/l	0.00100	0.00038	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Iron, Dissolved	ND		mg/l	0.0500	0.0191	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Lead, Dissolved	ND		mg/l	0.00100	0.00034	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Magnesium, Dissolved	ND		mg/l	0.0700	0.0242	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Manganese, Dissolved	ND		mg/l	0.00100	0.00044	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Nickel, Dissolved	ND		mg/l	0.00200	0.00055	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Potassium, Dissolved	ND		mg/l	0.100	0.0309	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Selenium, Dissolved	ND		mg/l	0.00500	0.00173	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Silver, Dissolved	ND		mg/l	0.00040	0.00016	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Sodium, Dissolved	ND		mg/l	0.500	0.0293	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Thallium, Dissolved	ND		mg/l	0.00100	0.00014	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Vanadium, Dissolved	ND		mg/l	0.00500	0.00157	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV
Zinc, Dissolved	ND		mg/l	0.01000	0.00341	1	11/07/25 16:13	11/11/25 09:48	1,6020B	SMV

Prep Information

Digestion Method: EPA 3005A



Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Method Blank Analysis Batch Quality Control

Parameter	Result	Qualifier	Units	RL	MDL	Dilution Factor	Date Prepared	Date Analyzed	Analytical Method	Analyst
Dissolved Metals - Mansfield Lab for sample(s): 01-06 Batch: WG2139405-1										
Mercury, Dissolved	ND		mg/l	0.00020	0.00009	1	11/10/25 11:45	11/11/25 07:13	1,7470A	JWN

Prep Information

Digestion Method: EPA 7470A



Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS		LCSD		%Recovery Limits	RPD	Qual	RPD Limits
	%Recovery	Qual	%Recovery	Qual				
Total Metals - Mansfield Lab Associated sample(s): 01-06 Batch: WG2138162-2								
Aluminum, Total	105		-		80-120	-		
Antimony, Total	101		-		80-120	-		
Arsenic, Total	105		-		80-120	-		
Barium, Total	103		-		80-120	-		
Beryllium, Total	102		-		80-120	-		
Cadmium, Total	101		-		80-120	-		
Calcium, Total	101		-		80-120	-		
Chromium, Total	103		-		80-120	-		
Cobalt, Total	101		-		80-120	-		
Copper, Total	103		-		80-120	-		
Iron, Total	106		-		80-120	-		
Lead, Total	103		-		80-120	-		
Magnesium, Total	103		-		80-120	-		
Manganese, Total	104		-		80-120	-		
Nickel, Total	103		-		80-120	-		
Potassium, Total	103		-		80-120	-		
Selenium, Total	101		-		80-120	-		
Silver, Total	102		-		80-120	-		
Sodium, Total	106		-		80-120	-		
Thallium, Total	104		-		80-120	-		
Vanadium, Total	105		-		80-120	-		

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	LCSD %Recovery	%Recovery Limits	RPD	RPD Limits
Total Metals - Mansfield Lab Associated sample(s): 01-06 Batch: WG2138162-2					
Zinc, Total	107	-	80-120	-	
Total Metals - Mansfield Lab Associated sample(s): 01-06 Batch: WG2138164-2					
Mercury, Total	96	-	80-120	-	20

Lab Control Sample Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	LCSD %Recovery	%Recovery Limits	RPD	RPD Limits
Dissolved Metals - Mansfield Lab Associated sample(s): 01-06 Batch: WG2138719-2					
Aluminum, Dissolved	94	-	80-120	-	
Antimony, Dissolved	86	-	80-120	-	
Arsenic, Dissolved	94	-	80-120	-	
Barium, Dissolved	96	-	80-120	-	
Beryllium, Dissolved	99	-	80-120	-	
Cadmium, Dissolved	93	-	80-120	-	
Calcium, Dissolved	92	-	80-120	-	
Chromium, Dissolved	95	-	80-120	-	
Cobalt, Dissolved	98	-	80-120	-	
Copper, Dissolved	94	-	80-120	-	
Iron, Dissolved	101	-	80-120	-	
Lead, Dissolved	98	-	80-120	-	
Magnesium, Dissolved	100	-	80-120	-	
Manganese, Dissolved	99	-	80-120	-	
Nickel, Dissolved	94	-	80-120	-	
Potassium, Dissolved	100	-	80-120	-	
Selenium, Dissolved	93	-	80-120	-	
Silver, Dissolved	90	-	80-120	-	
Sodium, Dissolved	100	-	80-120	-	
Thallium, Dissolved	92	-	80-120	-	
Vanadium, Dissolved	94	-	80-120	-	

Lab Control Sample Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	LCS %Recovery	LCSD %Recovery	%Recovery Limits	RPD	RPD Limits
Dissolved Metals - Mansfield Lab Associated sample(s): 01-06 Batch: WG2138719-2					
Zinc, Dissolved	96	-	80-120	-	
Dissolved Metals - Mansfield Lab Associated sample(s): 01-06 Batch: WG2139405-2					
Mercury, Dissolved	95	-	80-120	-	

Matrix Spike Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Parameter	Native Sample	MS Added	MS Found	MS %Recovery	Qual	MSD Found	MSD %Recovery	Qual	Recovery Limits	RPD	Qual	RPD Limits
Total Metals - Mansfield Lab Associated sample(s): 01-06 QC Batch ID: WG2138162-3 QC Sample: L2569921-01 Client ID: GZM-2												
Aluminum, Total	0.247	2	2.31	103		-	-		75-125	-		
Antimony, Total	ND	0.5	0.5261	105		-	-		75-125	-		
Arsenic, Total	0.00357	0.12	0.1326	108		-	-		75-125	-		
Barium, Total	0.1781	2	2.282	105		-	-		75-125	-		
Beryllium, Total	ND	0.05	0.05327	106		-	-		75-125	-		
Cadmium, Total	0.00007J	0.053	0.05513	104		-	-		75-125	-		
Calcium, Total	182.	10	202	200	Q	-	-		75-125	-		
Chromium, Total	0.00128	0.2	0.2107	105		-	-		75-125	-		
Cobalt, Total	0.00148	0.5	0.5158	103		-	-		75-125	-		
Copper, Total	0.00394	0.25	0.2692	106		-	-		75-125	-		
Iron, Total	23.0	1	25.4	240	Q	-	-		75-125	-		
Lead, Total	0.00412	0.53	0.5683	106		-	-		75-125	-		
Magnesium, Total	21.6	10	32.9	113		-	-		75-125	-		
Manganese, Total	1.814	0.5	2.471	131	Q	-	-		75-125	-		
Nickel, Total	0.00358	0.5	0.5324	106		-	-		75-125	-		
Potassium, Total	19.5	10	30.6	111		-	-		75-125	-		
Selenium, Total	ND	0.12	0.123	102		-	-		75-125	-		
Silver, Total	ND	0.05	0.05172	103		-	-		75-125	-		
Sodium, Total	42.3	10	55.0	127	Q	-	-		75-125	-		

Matrix Spike Analysis
Batch Quality Control

Project Name: BLONDELL EQUITIES

Lab Number: L2569921

Project Number: 41.0163256.00

Report Date: 11/11/25

Parameter	Native Sample	MS Added	MS Found	MS %Recovery	Qual	MSD Found	MSD %Recovery	Qual	Recovery Limits	RPD	Qual	RPD Limits
Total Metals - Mansfield Lab Associated sample(s): 01-06			QC Batch ID: WG2138162-3			QC Sample: L2569921-01			Client ID: GZM-2			
Thallium, Total	ND	0.12	0.1285	107		-	-		75-125	-		
Vanadium, Total	ND	0.5	0.5392	108		-	-		75-125	-		
Zinc, Total	0.01749	0.5	0.5457	106		-	-		75-125	-		
Total Metals - Mansfield Lab Associated sample(s): 01-06			QC Batch ID: WG2138164-3			QC Sample: L2569921-01			Client ID: GZM-2			
Mercury, Total	ND	0.005	0.00498	100		-	-		75-125	-		

Matrix Spike Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Parameter	Native Sample	MS Added	MS Found	MS %Recovery	MSD Qual	MSD Found	MSD %Recovery	MSD Qual	Recovery Limits	RPD	RPD Qual	RPD Limits
Dissolved Metals - Mansfield Lab Associated sample(s): 01-06 QC Batch ID: WG2138719-3 WG2138719-4 QC Sample: L2569884-01 Client ID: MS Sample												
Aluminum, Dissolved	0.004J	2	1.99	100		1.88	94		75-125	6		20
Antimony, Dissolved	ND	0.5	0.4605	92		0.4421	88		75-125	4		20
Arsenic, Dissolved	0.00253	0.12	0.1226	100		0.1137	93		75-125	8		20
Barium, Dissolved	0.1653	2	2.111	97		2.048	94		75-125	3		20
Beryllium, Dissolved	ND	0.05	0.05019	100		0.04908	98		75-125	2		20
Cadmium, Dissolved	ND	0.053	0.05201	98		0.04895	92		75-125	6		20
Calcium, Dissolved	73.7	10	87.2	135	Q	80.4	67	Q	75-125	8		20
Chromium, Dissolved	0.0002J	0.2	0.1911	96		0.1878	94		75-125	2		20
Cobalt, Dissolved	0.0002J	0.5	0.4798	96		0.4798	96		75-125	0		20
Copper, Dissolved	0.0004J	0.25	0.2357	94		0.2348	94		75-125	0		20
Iron, Dissolved	ND	1	0.989	99		0.955	96		75-125	3		20
Lead, Dissolved	0.0013	0.53	0.5314	100		0.5064	95		75-125	5		20
Magnesium, Dissolved	25.4	10	37.3	119		36.4	110		75-125	2		20
Manganese, Dissolved	0.1948	0.5	0.6982	101		0.6744	96		75-125	3		20
Nickel, Dissolved	0.0006J	0.5	0.4844	97		0.4516	90		75-125	7		20
Potassium, Dissolved	6.67	10	17.0	103		16.9	102		75-125	1		20
Selenium, Dissolved	ND	0.12	0.117	98		0.117	98		75-125	0		20
Silver, Dissolved	ND	0.05	0.04761	95		0.04537	91		75-125	5		20

Matrix Spike Analysis Batch Quality Control

Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Lab Number: L2569921
Report Date: 11/11/25

Parameter	Native Sample	MS Added	MS Found	MS %Recovery	MSD Found	MSD %Recovery	Recovery Limits	RPD	RPD Limits
Dissolved Metals - Mansfield Lab Associated sample(s): 01-06 QC Batch ID: WG2138719-3 WG2138719-4 QC Sample: L2569884-01 Client ID: MS Sample									
Sodium, Dissolved	63.7	10	76.1	124	71.9	82	75-125	6	20
Thallium, Dissolved	ND	0.12	0.1112	93	0.1094	91	75-125	2	20
Vanadium, Dissolved	ND	0.5	0.5156	103	0.4729	94	75-125	9	20
Zinc, Dissolved	ND	0.5	0.4833	97	0.4630	93	75-125	4	20
Dissolved Metals - Mansfield Lab Associated sample(s): 01-06 QC Batch ID: WG2139405-3 WG2139405-4 QC Sample: L2569923-11 Client ID: MS Sample									
Mercury, Dissolved	ND	0.005	0.00471	94	0.00473	95	75-125	0	20

Lab Duplicate Analysis

Batch Quality Control

Project Name: BLONDELL EQUITIES

Project Number: 41.0163256.00

Lab Number: L2569921

Report Date: 11/11/25

Parameter	Native Sample	Duplicate Sample	Units	RPD	Qual	RPD Limits
Total Metals - Mansfield Lab Associated sample(s): 01-06 QC Batch ID: WG2138162-4 QC Sample: L2569921-01 Client ID: GZM-2						
Aluminum, Total	0.247	0.257	mg/l	4		20
Antimony, Total	ND	ND	mg/l	NC		20
Arsenic, Total	0.00357	0.00371	mg/l	4		20
Barium, Total	0.1781	0.1832	mg/l	3		20
Beryllium, Total	ND	ND	mg/l	NC		20
Cadmium, Total	0.00007J	0.00008J	mg/l	NC		20
Calcium, Total	182.	187	mg/l	3		20
Chromium, Total	0.00128	0.00075J	mg/l	NC		20
Cobalt, Total	0.00148	0.00156	mg/l	5		20
Copper, Total	0.00394	0.00466	mg/l	17		20
Iron, Total	23.0	23.7	mg/l	3		20
Lead, Total	0.00412	0.00434	mg/l	5		20
Magnesium, Total	21.6	22.9	mg/l	6		20
Manganese, Total	1.814	1.875	mg/l	3		20
Nickel, Total	0.00358	0.00415	mg/l	15		20
Potassium, Total	19.5	20.2	mg/l	4		20
Selenium, Total	ND	ND	mg/l	NC		20

Lab Duplicate Analysis

Batch Quality Control

Project Name: BLONDELL EQUITIES

Project Number: 41.0163256.00

Lab Number: L2569921

Report Date: 11/11/25

Parameter	Native Sample	Duplicate Sample	Units	RPD	Qual	RPD Limits
Total Metals - Mansfield Lab Associated sample(s): 01-06 QC Batch ID: WG2138162-4 QC Sample: L2569921-01 Client ID: GZM-2						
Silver, Total	ND	ND	mg/l	NC		20
Sodium, Total	42.3	44.7	mg/l	6		20
Thallium, Total	ND	ND	mg/l	NC		20
Vanadium, Total	ND	ND	mg/l	NC		20
Zinc, Total	0.01749	0.01836	mg/l	5		20
Total Metals - Mansfield Lab Associated sample(s): 01-06 QC Batch ID: WG2138164-4 QC Sample: L2569921-01 Client ID: GZM-2						
Mercury, Total	ND	ND	mg/l	NC		20

**Lab Serial Dilution
Analysis
Batch Quality Control**

Project Name: BLONDELL EQUITIES

Project Number: 41.0163256.00

Lab Number: L2569921

Report Date: 11/11/25

Parameter	Native Sample	Serial Dilution	Units	% D	Qual	RPD Limits
Dissolved Metals - Mansfield Lab Associated sample(s): 01-06 QC Batch ID: WG2138719-6 QC Sample: L2569884-01 Client ID: DUP Sample						
Manganese, Dissolved	0.1948	0.2004	mg/l	3		20



Project Name: BLONDELL EQUITIES
Project Number: 41.0163256.00

Serial_No:11112516:58
Lab Number: L2569921
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Sample Receipt and Container Information

Were project specific reporting limits specified?

YES

Cooler Information

Cooler **Custody Seal**

Container Information

Container ID	Container Type	Cooler	Initial pH	Final pH	Temp deg C	Pres	Seal	Frozen Date/Time	Analysis(*)
L2569921-01A	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-01B	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-01C	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-01D	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-01E	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-01F	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-01G	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-01H	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-01I	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-01J	Plastic 250ml unpreserved	NA	NA			Y	Absent		-
L2569921-01K	Plastic 250ml HNO3 preserved	NA	NA			Y	Absent		SE-6020T(180),TL-6020T(180),FE-6020T(180),BA-6020T(180),CR-6020T(180),CA-6020T(180),K-6020T(180),NI-6020T(180),ZN-6020T(180),CU-6020T(180),NA-6020T(180),PB-6020T(180),BE-6020T(180),MN-6020T(180),SB-6020T(180),AS-6020T(180),V-6020T(180),AG-6020T(180),CD-6020T(180),MG-6020T(180),HG-T(28),AL-6020T(180),CO-6020T(180)
L2569921-01L	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)
L2569921-01M	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)



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Container Information

Container ID	Container Type	Cooler	Initial pH	Final pH	Temp deg C	Pres	Seal	Frozen Date/Time	Analysis(*)
L2569921-01X	Plastic 120ml HNO3 preserved Filtrates	NA	NA			Y	Absent		V-6020S(180),K-6020S(180),SE-6020S(180),CU-6020S(180),MN-6020S(180),BE-6020S(180),CO-6020S(180),MG-6020S(180),ZN-6020S(180),FE-6020S(180),CA-6020S(180),CR-6020S(180),NI-6020S(180),TL-6020S(180),NA-6020S(180),BA-6020S(180),PB-6020S(180),AG-6020S(180),SB-6020S(180),AS-6020S(180),CD-6020S(180),AL-6020S(180),HG-S(28)
L2569921-02A	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-02B	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-02C	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-02D	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-02E	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-02F	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-02G	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-02H	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-02I	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-02J	Plastic 250ml unpreserved	NA	NA			Y	Absent		-
L2569921-02K	Plastic 250ml HNO3 preserved	NA	NA			Y	Absent		FE-6020T(180),TL-6020T(180),BA-6020T(180),SE-6020T(180),NI-6020T(180),CA-6020T(180),CR-6020T(180),K-6020T(180),NA-6020T(180),CU-6020T(180),ZN-6020T(180),PB-6020T(180),BE-6020T(180),MN-6020T(180),AS-6020T(180),V-6020T(180),SB-6020T(180),AL-6020T(180),AG-6020T(180),CD-6020T(180),HG-T(28),MG-6020T(180),CO-6020T(180)
L2569921-02L	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)
L2569921-02M	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)



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Container Information

Container ID	Container Type	Cooler	Initial pH	Final pH	Temp deg C	Pres	Seal	Frozen Date/Time	Analysis(*)
L2569921-02X	Plastic 120ml HNO3 preserved Filtrates	NA	NA			Y	Absent		K-6020S(180),SE-6020S(180),CU-6020S(180),V-6020S(180),MN-6020S(180),MG-6020S(180),ZN-6020S(180),CO-6020S(180),BE-6020S(180),FE-6020S(180),CA-6020S(180),CR-6020S(180),PB-6020S(180),BA-6020S(180),NA-6020S(180),NI-6020S(180),TL-6020S(180),AG-6020S(180),SB-6020S(180),AS-6020S(180),CD-6020S(180),HG-S(28),AL-6020S(180)
L2569921-03A	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-03B	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-03C	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-03D	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-03E	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-03F	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-03G	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-03H	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-03I	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-03J	Plastic 250ml unpreserved	NA	NA			Y	Absent		-
L2569921-03K	Plastic 250ml HNO3 preserved	NA	NA			Y	Absent		SE-6020T(180),TL-6020T(180),BA-6020T(180),FE-6020T(180),K-6020T(180),CA-6020T(180),NI-6020T(180),CR-6020T(180),ZN-6020T(180),NA-6020T(180),CU-6020T(180),PB-6020T(180),MN-6020T(180),BE-6020T(180),AS-6020T(180),SB-6020T(180),V-6020T(180),CD-6020T(180),AG-6020T(180),HG-T(28),AL-6020T(180),MG-6020T(180),CO-6020T(180)
L2569921-03L	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)
L2569921-03M	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)



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Container Information

Container ID	Container Type	Cooler	Initial pH	Final pH	Temp deg C	Pres	Seal	Frozen Date/Time	Analysis(*)
L2569921-03X	Plastic 120ml HNO3 preserved Filtrates	NA	NA			Y	Absent		CU-6020S(180),SE-6020S(180),K-6020S(180),V-6020S(180),MN-6020S(180),CO-6020S(180),ZN-6020S(180),MG-6020S(180),BE-6020S(180),CA-6020S(180),CR-6020S(180),FE-6020S(180),PB-6020S(180),TL-6020S(180),NI-6020S(180),NA-6020S(180),BA-6020S(180),AG-6020S(180),AS-6020S(180),SB-6020S(180),HG-S(28),AL-6020S(180),CD-6020S(180)
L2569921-04A	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-04B	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-04D	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-04E	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-04F	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-04G	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-04H	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-04I	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-04J	Plastic 250ml unpreserved	NA	NA			Y	Absent		-
L2569921-04K	Plastic 250ml HNO3 preserved	NA	NA			Y	Absent		FE-6020T(180),SE-6020T(180),TL-6020T(180),BA-6020T(180),NI-6020T(180),K-6020T(180),CA-6020T(180),CR-6020T(180),NA-6020T(180),ZN-6020T(180),CU-6020T(180),PB-6020T(180),MN-6020T(180),BE-6020T(180),SB-6020T(180),V-6020T(180),AS-6020T(180),AL-6020T(180),HG-T(28),MG-6020T(180),CD-6020T(180),AG-6020T(180),CO-6020T(180)
L2569921-04L	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)
L2569921-04M	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)



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Container Information

Container ID	Container Type	Cooler	Initial pH	Final pH	Temp deg C	Pres	Seal	Frozen Date/Time	Analysis(*)
L2569921-04X	Plastic 120ml HNO3 preserved Filtrates	NA	NA			Y	Absent		CU-6020S(180),V-6020S(180),K-6020S(180),SE-6020S(180),MN-6020S(180),MG-6020S(180),ZN-6020S(180),CO-6020S(180),BE-6020S(180),CR-6020S(180),FE-6020S(180),CA-6020S(180),TL-6020S(180),BA-6020S(180),NA-6020S(180),PB-6020S(180),NI-6020S(180),AS-6020S(180),AG-6020S(180),SB-6020S(180),HG-S(28),AL-6020S(180),CD-6020S(180)
L2569921-05A	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-05B	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-05C	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-05D	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-05E	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-05F	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-05G	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-05H	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-05I	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-05J	Plastic 250ml unpreserved	NA	NA			Y	Absent		-
L2569921-05K	Plastic 250ml HNO3 preserved	NA	NA			Y	Absent		SE-6020T(180),TL-6020T(180),BA-6020T(180),FE-6020T(180),CA-6020T(180),CR-6020T(180),K-6020T(180),NI-6020T(180),ZN-6020T(180),NA-6020T(180),CU-6020T(180),PB-6020T(180),BE-6020T(180),MN-6020T(180),SB-6020T(180),V-6020T(180),AS-6020T(180),AG-6020T(180),AL-6020T(180),HG-T(28),CD-6020T(180),MG-6020T(180),CO-6020T(180)
L2569921-05L	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)
L2569921-05M	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)



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Container Information

Container ID	Container Type	Cooler	Initial pH	Final pH	Temp deg C	Pres	Seal	Frozen Date/Time	Analysis(*)
L2569921-05X	Plastic 120ml HNO3 preserved Filtrates	NA	NA			Y	Absent		V-6020S(180),K-6020S(180),CU-6020S(180),SE-6020S(180),MN-6020S(180),BE-6020S(180),CO-6020S(180),MG-6020S(180),ZN-6020S(180),FE-6020S(180),CR-6020S(180),CA-6020S(180),NI-6020S(180),PB-6020S(180),NA-6020S(180),TL-6020S(180),BA-6020S(180),AS-6020S(180),SB-6020S(180),AG-6020S(180),HG-S(28),CD-6020S(180),AL-6020S(180)
L2569921-06A	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-06B	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-06C	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-06D	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-06E	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8270-RVT(7),NYTCL-8270-SIM-RVT(7)
L2569921-06F	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-06G	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8081-RVT(7)
L2569921-06H	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-06I	Amber 100ml unpreserved	NA	NA			Y	Absent		NYTCL-8082-RVT(365)
L2569921-06J	Plastic 250ml unpreserved	NA	NA			Y	Absent		-
L2569921-06K	Plastic 250ml HNO3 preserved	NA	NA			Y	Absent		BA-6020T(180),TL-6020T(180),SE-6020T(180),FE-6020T(180),CR-6020T(180),K-6020T(180),CA-6020T(180),NI-6020T(180),NA-6020T(180),CU-6020T(180),ZN-6020T(180),PB-6020T(180),MN-6020T(180),BE-6020T(180),AS-6020T(180),SB-6020T(180),V-6020T(180),CD-6020T(180),MG-6020T(180),HG-T(28),AG-6020T(180),AL-6020T(180),CO-6020T(180)
L2569921-06L	Plastic 250ml unpreserved	NA	NA			Y	Absent		A2-NY-1633(28)



Project Name: BLONDELL EQUITIES

Project Number: 41.0163256.00

Serial_No:11112516:58

Lab Number: L2569921

Report Date: 11/11/25

Container Information

Container ID	Container Type	Cooler	Initial pH	Final pH	Temp deg C	Pres	Seal	Frozen Date/Time	Analysis(*)
L2569921-06X	Plastic 120ml HNO3 preserved Filtrates	NA	NA			Y	Absent		K-6020S(180),V-6020S(180),SE-6020S(180),CU-6020S(180),MN-6020S(180),ZN-6020S(180),MG-6020S(180),CO-6020S(180),BE-6020S(180),CR-6020S(180),FE-6020S(180),CA-6020S(180),TL-6020S(180),NI-6020S(180),PB-6020S(180),BA-6020S(180),NA-6020S(180),SB-6020S(180),AG-6020S(180),AS-6020S(180),CD-6020S(180),AL-6020S(180),HG-S(28)
L2569921-07A	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)
L2569921-07B	Vial HCl preserved	NA	NA			Y	Absent		NYTCL-8260(14)

PFAS PARAMETER SUMMARY

Parameter	Acronym	CAS Number
PERFLUOROALKYL CARBOXYLIC ACIDS (PFCAs)		
Perfluorooctadecanoic Acid	PFODA	16517-11-6
Perfluorohexadecanoic Acid	PFHxDA	67905-19-5
Perfluorotetradecanoic Acid	PFTA/PFTeDA	376-06-7
Perfluorotridecanoic Acid	PFTrDA	72629-94-8
Perfluorododecanoic Acid	PFDoA	307-55-1
Perfluoroundecanoic Acid	PFUnA	2058-94-8
Perfluorodecanoic Acid	PFDA	335-76-2
Perfluorononanoic Acid	PFNA	375-95-1
Perfluorooctanoic Acid	PFOA	335-67-1
Perfluoroheptanoic Acid	PFHpA	375-85-9
Perfluorohexanoic Acid	PFHxA	307-24-4
Perfluoropentanoic Acid	PFPeA	2706-90-3
Perfluorobutanoic Acid	PFBA	375-22-4
PERFLUOROALKYL SULFONIC ACIDS (PFSAs)		
Perfluorododecanesulfonic Acid	PFDoDS/PFDoS	79780-39-5
Perfluorodecanesulfonic Acid	PFDS	335-77-3
Perfluorononanesulfonic Acid	PFNS	68259-12-1
Perfluorooctanesulfonic Acid	PFOS	1763-23-1
Perfluoroheptanesulfonic Acid	PFHpS	375-92-8
Perfluorohexanesulfonic Acid	PFHxS	355-46-4
Perfluoropentanesulfonic Acid	PFPeS	2706-91-4
Perfluorobutanesulfonic Acid	PFBS	375-73-5
Perfluoropropanesulfonic Acid	PFPrS	423-41-6
FLUOROTELOMERS		
1H,1H,2H,2H-Perfluorododecanesulfonic Acid	10:2FTS	120226-60-0
1H,1H,2H,2H-Perfluorodecanesulfonic Acid	8:2FTS	39108-34-4
1H,1H,2H,2H-Perfluorooctanesulfonic Acid	6:2FTS	27619-97-2
1H,1H,2H,2H-Perfluorohexanesulfonic Acid	4:2FTS	757124-72-4
PERFLUOROALKANE SULFONAMIDES (FASAs)		
Perfluorooctanesulfonamide	FOSA/PFOSA	754-91-6
N-Ethyl Perfluorooctane Sulfonamide	NEtFOSA	4151-50-2
N-Methyl Perfluorooctane Sulfonamide	NMeFOSA	31506-32-8
PERFLUOROALKANE SULFONYL SUBSTANCES		
N-Ethyl Perfluorooctanesulfonamido Ethanol	NEtFOSE	1691-99-2
N-Methyl Perfluorooctanesulfonamido Ethanol	NMeFOSE	24448-09-7
N-Ethyl Perfluorooctanesulfonamidoacetic Acid	NEtFOSAA	2991-50-6
N-Methyl Perfluorooctanesulfonamidoacetic Acid	NMeFOSAA	2355-31-9
PER- and POLYFLUOROALKYL ETHER CARBOXYLIC ACIDS		
2,3,3,3-Tetrafluoro-2-[1,1,2,2,3,3,3-Heptafluoropropoxy]-Propanoic Acid	HFPO-DA	13252-13-6
4,8-Dioxa-3h-Perfluorononanoic Acid	ADONA	919005-14-4
CHLORO-PERFLUOROALKYL SULFONIC ACIDS		
11-Chloroeicosafuoro-3-Oxaundecane-1-Sulfonic Acid	11Cl-PF3OUdS	763051-92-9
9-Chlorohexadecafluoro-3-Oxanone-1-Sulfonic Acid	9Cl-PF3ONS	756426-58-1
PERFLUOROETHER SULFONIC ACIDS (PFESAs)		
Perfluoro(2-Ethoxyethane)Sulfonic Acid	PFEESA	113507-82-7
PERFLUOROETHER/POLYETHER CARBOXYLIC ACIDS (PFPCAs)		
Perfluoro-3-Methoxypropanoic Acid	PFMPA	377-73-1
Perfluoro-4-Methoxybutanoic Acid	PFMBA	863090-89-5
Nonafluoro-3,6-Dioxaheptanoic Acid	NFDHA	151772-58-6

Project Name: BLONDELL EQUITIES
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PFAS PARAMETER SUMMARY

Parameter	Acronym	CAS Number
FLUOROTELOMER CARBOXYLIC ACIDS (FTCAs)		
3-Perfluoroheptyl Propanoic Acid	7:3FTCA	812-70-4
2H,2H,3H,3H-Perfluorooctanoic Acid	5:3FTCA	914637-49-3
3-Perfluoropropyl Propanoic Acid	3:3FTCA	356-02-5

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GLOSSARY

Acronyms

DL	- Detection Limit: This value represents the level to which target analyte concentrations are reported as estimated values, when those target analyte concentrations are quantified below the limit of quantitation (LOQ). The DL includes any adjustments from dilutions, concentrations or moisture content, where applicable. (DoD report formats only.)
EDL	- Estimated Detection Limit: This value represents the level to which target analyte concentrations are reported as estimated values, when those target analyte concentrations are quantified below the reporting limit (RL). The EDL includes any adjustments from dilutions, concentrations or moisture content, where applicable. The use of EDLs is specific to the analysis of PAHs using Solid-Phase Microextraction (SPME).
EMPC	- Estimated Maximum Possible Concentration: The concentration that results from the signal present at the retention time of an analyte when the ions meet all of the identification criteria except the ion abundance ratio criteria. An EMPC is a worst-case estimate of the concentration.
EPA	- Environmental Protection Agency.
LCS	- Laboratory Control Sample: A sample matrix, free from the analytes of interest, spiked with verified known amounts of analytes or a material containing known and verified amounts of analytes.
LCSD	- Laboratory Control Sample Duplicate: Refer to LCS.
LFB	- Laboratory Fortified Blank: A sample matrix, free from the analytes of interest, spiked with verified known amounts of analytes or a material containing known and verified amounts of analytes.
LOD	- Limit of Detection: This value represents the level to which a target analyte can reliably be detected for a specific analyte in a specific matrix by a specific method. The LOD includes any adjustments from dilutions, concentrations or moisture content, where applicable. (DoD report formats only.)
LOQ	- Limit of Quantitation: The value at which an instrument can accurately measure an analyte at a specific concentration. The LOQ includes any adjustments from dilutions, concentrations or moisture content, where applicable. (DoD report formats only.) Limit of Quantitation: The value at which an instrument can accurately measure an analyte at a specific concentration. The LOQ includes any adjustments from dilutions, concentrations or moisture content, where applicable. (DoD report formats only.)
MDL	- Method Detection Limit: This value represents the level to which target analyte concentrations are reported as estimated values, when those target analyte concentrations are quantified below the reporting limit (RL). The MDL includes any adjustments from dilutions, concentrations or moisture content, where applicable.
MS	- Matrix Spike Sample: A sample prepared by adding a known mass of target analyte to a specified amount of matrix sample for which an independent estimate of target analyte concentration is available. For Method 332.0, the spike recovery is calculated using the native concentration, including estimated values.
MSD	- Matrix Spike Sample Duplicate: Refer to MS.
NA	- Not Applicable.
NC	- Not Calculated: Term is utilized when one or more of the results utilized in the calculation are non-detect at the parameter's reporting unit.
NDPA/DPA	- N-Nitrosodiphenylamine/Diphenylamine.
NI	- Not Ignitable.
NP	- Non-Plastic: Term is utilized for the analysis of Atterberg Limits in soil.
NR	- No Results: Term is utilized when 'No Target Compounds Requested' is reported for the analysis of Volatile or Semivolatile Organic TIC only requests.
RL	- Reporting Limit: The value at which an instrument can accurately measure an analyte at a specific concentration. The RL includes any adjustments from dilutions, concentrations or moisture content, where applicable.
RPD	- Relative Percent Difference: The results from matrix and/or matrix spike duplicates are primarily designed to assess the precision of analytical results in a given matrix and are expressed as relative percent difference (RPD). Values which are less than five times the reporting limit for any individual parameter are evaluated by utilizing the absolute difference between the values; although the RPD value will be provided in the report.
SRM	- Standard Reference Material: A reference sample of a known or certified value that is of the same or similar matrix as the associated field samples.
STLP	- Semi-dynamic Tank Leaching Procedure per EPA Method 1315.
TEF	- Toxic Equivalency Factors: The values assigned to each dioxin and furan to evaluate their toxicity relative to 2,3,7,8-TCDD.
TEQ	- Toxic Equivalent: The measure of a sample's toxicity derived by multiplying each dioxin and furan by its corresponding TEF and then summing the resulting values.
TIC	- Tentatively Identified Compound: A compound that has been identified to be present and is not part of the target compound list (TCL) for the method and/or program. All TICs are qualitatively identified and reported as estimated concentrations.

Report Format: DU Report with 'J' Qualifiers



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Footnotes

- 1 - The reference for this analyte should be considered modified since this analyte is absent from the target analyte list of the original method.

Terms

Analytical Method: Both the document from which the method originates and the analytical reference method. (Example: EPA 8260B is shown as 1,8260B.) The codes for the reference method documents are provided in the References section of the Addendum.

Chlordane: The target compound Chlordane (CAS No. 57-74-9) is reported for GC ECD analyses. Per EPA, this compound "refers to a mixture of chlordane isomers, other chlorinated hydrocarbons and numerous other components." (Reference: USEPA Toxicological Review of Chlordane, In Support of Summary Information on the Integrated Risk Information System (IRIS), December 1997.)

Difference: With respect to Total Oxidizable Precursor (TOP) Assay analysis, the difference is defined as the Post-Treatment value minus the Pre-Treatment value.

Final pH: As it pertains to Sample Receipt & Container Information section of the report, Final pH reflects pH of container determined after adjustment at the laboratory, if applicable. If no adjustment required, value reflects Initial pH.

Frozen Date/Time: With respect to Volatile Organics in soil, Frozen Date/Time reflects the date/time at which associated Reagent Water-preserved vials were initially frozen. Note: If frozen date/time is beyond 48 hours from sample collection, value will be reflected in 'bold'.

Gasoline Range Organics (GRO): Gasoline Range Organics (GRO) results include all chromatographic peaks eluting from Methyl tert butyl ether through Naphthalene, with the exception of GRO analysis in support of State of Ohio programs, which includes all chromatographic peaks eluting from Hexane through Dodecane.

Initial pH: As it pertains to Sample Receipt & Container Information section of the report, Initial pH reflects pH of container determined upon receipt, if applicable.

PAH Total: With respect to Alkylated PAH analyses, the 'PAHs, Total' result is defined as the summation of results for all or a subset of the following compounds: Naphthalene, C1-C4 Naphthalenes, 2-Methylnaphthalene, 1-Methylnaphthalene, Biphenyl, Acenaphthylene, Acenaphthene, Fluorene, C1-C3 Fluorenes, Phenanthrene, C1-C4 Phenanthrenes/Anthracenes, Anthracene, Fluoranthene, Pyrene, C1-C4 Fluoranthenes/Pyrenes, Benz(a)anthracene, Chrysene, C1-C4 Chrysenes, Benzo(b)fluoranthene, Benzo(j)+(k)fluoranthene, Benzo(e)pyrene, Benzo(a)pyrene, Perylene, Indeno(1,2,3-cd)pyrene, Dibenz(ah)+(ac)anthracene, Benzo(g,h,i)perylene. If a 'Total' result is requested, the results of its individual components will also be reported.

PFAS Total: With respect to PFAS analyses, the 'PFAS, Total (5)' result is defined as the summation of results for: PFHpA, PFHxS, PFOA, PFNA and PFOS. In addition, the 'PFAS, Total (6)' result is defined as the summation of results for: PFHpA, PFHxS, PFOA, PFNA, PFDA and PFOS. For MassDEP DW compliance analysis only, the 'PFAS, Total (6)' result is defined as the summation of results at or above the RL. Note: If a 'Total' result is requested, the results of its individual components will also be reported.

Total: With respect to Organic analyses, a 'Total' result is defined as the summation of results for individual isomers or Aroclors. If a 'Total' result is requested, the results of its individual components will also be reported. This is applicable to 'Total' results for methods 8260, 8081 and 8082.

Data Qualifiers

- A** - Spectra identified as "Aldol Condensates" are byproducts of the extraction/concentration procedures when acetone is introduced in the process.
- B** - The analyte was detected above the reporting limit in the associated method blank. Flag only applies to associated field samples that have detectable concentrations of the analyte at less than ten times (10x) the concentration found in the blank. For MCP-related projects, flag only applies to associated field samples that have detectable concentrations of the analyte at less than ten times (10x) the concentration found in the blank. For DOD-related projects, flag only applies to associated field samples that have detectable concentrations of the analyte at less than ten times (10x) the concentration found in the blank AND the analyte was detected above one-half the reporting limit (or above the reporting limit for common lab contaminants) in the associated method blank. For NJ-Air-related projects, flag only applies to associated field samples that have detectable concentrations of the analyte above the reporting limit. For NJ-related projects (excluding Air), flag only applies to associated field samples that have detectable concentrations of the analyte, which was detected above the reporting limit in the associated method blank or above five times the reporting limit for common lab contaminants (Phthalates, Acetone, Methylene Chloride, 2-Butanone).
- C** - Co-elution: The target analyte co-elutes with a known lab standard (i.e. surrogate, internal standards, etc.) for co-extracted analyses.
- D** - Concentration of analyte was quantified from diluted analysis. Flag only applies to field samples that have detectable concentrations of the analyte.
- E** - Concentration of analyte exceeds the range of the calibration curve and/or linear range of the instrument.
- F** - The ratio of quantifier ion response to qualifier ion response falls outside of the laboratory criteria. Results are considered to be an estimated maximum concentration.
- G** - The concentration may be biased high due to matrix interferences (i.e. co-elution) with non-target compound(s). The result should be considered estimated.
- H** - The analysis of pH was performed beyond the regulatory-required holding time of 15 minutes from the time of sample collection.
- I** - The lower value for the two columns has been reported due to obvious interference.
- J** - Estimated value. The Target analyte concentration is below the quantitation limit (RL), but above the Method Detection Limit (MDL) or Estimated Detection Limit (EDL) for SPME-related analyses. This represents an estimated concentration for Tentatively Identified Compounds (TICs). For calculated parameters, this represents that one or more values used in the calculation were

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estimated.

- M** - Reporting Limit (RL) exceeds the MCP CAM Reporting Limit for this analyte.
- ND** - Not detected at the method detection limit (MDL) for the sample, or estimated detection limit (EDL) for SPME-related analyses.
- NJ** - Presumptive evidence of compound. This represents an estimated concentration for Tentatively Identified Compounds (TICs), where the identification is based on a mass spectral library search.
- P** - The RPD between the results for the two columns exceeds the method-specified criteria.
- Q** - The quality control sample exceeds the associated acceptance criteria. For DOD-related projects, LCS and/or Continuing Calibration Standard exceedences are also qualified on all associated sample results. Note: This flag is not applicable for matrix spike recoveries when the sample concentration is greater than 4x the spike added or for batch duplicate RPD when the sample concentrations are less than 5x the RL. (Metals only.)
- R** - Analytical results are from sample re-analysis.
- RE** - Analytical results are from sample re-extraction.
- S** - Analytical results are from modified screening analysis.
- V** - The surrogate associated with this target analyte has a recovery outside the QC acceptance limits. (Applicable to MassDEP DW Compliance samples only.)
- Z** - The batch matrix spike and/or duplicate associated with this target analyte has a recovery/RPD outside the QC acceptance limits. (Applicable to MassDEP DW Compliance samples only.)

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Report Date: 11/11/25

REFERENCES

- 1 Test Methods for Evaluating Solid Waste: Physical/Chemical Methods. EPA SW-846. Third Edition. Updates I - VI, 2018.
- 168 Analysis of Per- and Polyfluoroalkyl Substances (PFAS) in Aqueous, Solid, Biosolids, and Tissue Samples by LC-MS/MS. EPA Method 1633A, EPA 820-R-24-007, December 2024. For Massachusetts Contingency Plan, WSC-CAM-XA, September 2025.

LIMITATION OF LIABILITIES

Pace Analytical Services performs services with reasonable care and diligence normal to the analytical testing laboratory industry. In the event of an error, the sole and exclusive responsibility of Pace Analytical Services shall be to re-perform the work at it's own expense. In no event shall Pace Analytical Services be held liable for any incidental, consequential or special damages, including but not limited to, damages in any way connected with the use of, interpretation of, information or analysis provided by Pace Analytical Services.

We strongly urge our clients to comply with EPA protocol regarding sample volume, preservation, cooling, containers, sampling procedures, holding time and splitting of samples in the field.



Certification Information

The following analytes are not included in our Primary NELAP Scope of Accreditation:

Westborough Facility – 8 Walkup Dr. Westborough, MA 01581

EPA 624.1: m/p-xylene, o-xylene, Naphthalene

EPA 625.1: alpha-Terpineol

EPA 8260D: NPW: 1,2,4,5-Tetramethylbenzene; 4-Ethyltoluene; SCM: Iodomethane (methyl iodide), 1,2,4,5-Tetramethylbenzene; 4-Ethyltoluene.

EPA 8270E: NPW: Dimethylnaphthalene, 1,4-Diphenylhydrazine, alpha-Terpineol, Azobenzene; SCM: Dimethylnaphthalene, 1,4-Diphenylhydrazine.

SM4500: NPW: Amenable Cyanide; SCM: Total Phosphorus, TKN, NO₂, NO₃.

Mansfield Facility – 320 Forbes Blvd. Mansfield, MA 02048

SM 2540D: TSS.

EPA TO-15: Halothane, 2,4,4-Trimethyl-2-pentene, 2,4,4-Trimethyl-1-pentene, Thiophene, 2-Methylthiophene,

3-Methylthiophene, 2-Ethylthiophene, 1,2,3-Trimethylbenzene, Indan, Indene, 1,2,4,5-Tetramethylbenzene, Benzothiophene, 1-Methylnaphthalene.

MADEP-APH.

Nonpotable Water: EPA RSK-175 Dissolved Gases

Biological Tissue Matrix: EPA 3050B

Mansfield Facility – 120 Forbes Blvd. Mansfield, MA 02048

EPA TO-15: Halothane, 2,4,4-Trimethyl-2-pentene, 2,4,4-Trimethyl-1-pentene, Thiophene, 2-Methylthiophene,

3-Methylthiophene, 2-Ethylthiophene, 1,2,3-Trimethylbenzene, Indan, Indene, 1,2,4,5-Tetramethylbenzene, Benzothiophene, 1-Methylnaphthalene.

Nonpotable Water: EPA RSK-175 Dissolved Gases

The following test method is not included in our New Jersey Secondary NELAP Scope of Accreditation:

Mansfield Facility – 320 Forbes Blvd. Mansfield, MA 02048

Determination of Selected Perfluorinated Alkyl Substances by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry Isotope Dilution (via Alpha SOP 23528)

The following analytes are included in our Massachusetts DEP Scope of Accreditation

Westborough Facility – 8 Walkup Dr. Westborough, MA 01581

Drinking Water

EPA 300.0: Chloride, Nitrate-N, Fluoride, Sulfate; **EPA 353.2:** Nitrate-N, Nitrite-N; **SM4500NO3-F:** Nitrate-N, Nitrite-N; **SM4500F-C, SM4500CN-CE,**

EPA 180.1, SM2130B, SM4500Cl-D, SM2320B, SM2540C, SM4500H-B, SM4500NO2-B

EPA 524.2: THMs and VOCs; **EPA 504.1:** EDB, DBCP.

Microbiology: **SM9215B; SM9223-P/A, SM9223B-Colilert-QT, SM9222D.**

Non-Potable Water

SM4500H,B, EPA 120.1, SM2510B, SM2540C, SM2320B, SM4500CL-E, SM4500F-BC, SM4500NH3-BH: Ammonia-N and Kjeldahl-N, **EPA 350.1:**

Ammonia-N, **LACHAT 10-107-06-1-B:** Ammonia-N, **EPA 351.1, SM4500NO3-F, EPA 353.2:** Nitrate-N, **SM4500P-E, SM4500P-B, E, SM4500SO4-E,**

SM5220D, EPA 410.4, SM5210B, SM5310C, SM4500CL-D, SM4500CL-G, EPA 1664, EPA 420.1, SM4500-CN-CE, SM2540D, EPA 300: Chloride,

Sulfate, Nitrate.

EPA 624.1: Volatile Halocarbons & Aromatics,

EPA 608.3: Chlordane, Toxaphene, Aldrin, alpha-BHC, beta-BHC, gamma-BHC, delta-BHC, Dieldrin, DDD, DDE, DDT, Endosulfan I, Endosulfan II,

Endosulfan sulfate, Endrin, Endrin Aldehyde, Heptachlor, Heptachlor Epoxide, PCBs

EPA 625.1: SVOC (Acid/Base/Neutral Extractables).

Microbiology: **SM9223B-Colilert-QT; Enterolert-QT.**

Mansfield Facility – 320 Forbes Blvd. Mansfield, MA 02048

Drinking Water

EPA 200.7: Al, Ba, Cd, Cr, Cu, Fe, Mn, Ni, Na, Ag, Ca, Zn. **EPA 200.8:** Al, Sb, As, Ba, Be, Cd, Cr, Cu, Pb, Mn, Ni, Se, Ag, TL, Zn. **EPA 245.1 Hg.**

EPA 522, EPA 537.1.

Non-Potable Water

EPA 200.7: Al, Sb, As, Be, Cd, Ca, Cr, Co, Cu, Fe, Pb, Mg, Mn, Mo, Ni, K, Se, Ag, Na, Sr, TL, Ti, V, Zn.

EPA 200.8: Al, Sb, As, Be, Cd, Ca, Cr, Cu, Fe, Pb, Mg, Mn, Ni, K, Se, Ag, Na, TL, Zn.

EPA 245.1: Hg. **EPA 245.7:** Hg.

SM2340B

Pace Analytical Services LLCID No.:**17873**Facility: **Northeast**

Revision 28

Department: **Quality Assurance**

Published Date: 07/25/2025

Title: **Certificate/Approval Program Summary**

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Certification IDs:**Westborough Facility – 8 Walkup Dr. Westborough, MA 01581**

CT PH-0826, IL 200077, IN C-MA-03, KY KY98045, ME MA00086, MD 348, MA M-MA086, NH 2064, NJ MA935, NY 11148, NC (DW) 25700, NC (NPW/SCM) 666, OR MA-1316, PA 68-03671, RI LAO00065, TX T104704476, VT VT-0935, VA 460195

Mansfield Facility – 320 Forbes Blvd. Mansfield, MA 02048

MA M-MA00030, CT PH-0825, ANAB/DoD L2474, IL 200081, IN C-MA-04, KY KY98046, LA 85084, ME MA00030, MI 9110, MN 025-999-495, NH 2062, NJ MA015, NY 11627, NC (NPW/SCM) 685, OR MA-0262, PA 68-02089, RI LAO00299, TX T-104704419, VT VT-0015, VA 460194, WA C954

Mansfield Facility – 120 Forbes Blvd. Mansfield, MA 02048

ANAB/DoD L2474, LA 245052, ME MA01156, MN 025-999-498, NH 2249, NJ MA025, NY 12191, OR 4203, TX T104704583, VA 460311, WA C1104.

For a complete listing of analytes and methods, please contact your Project Manager.

 Pace <small>ANALYTICAL SERVICES</small>	NEW YORK CHAIN OF CUSTODY	Service Centers Woodcliff Lake, NJ 07677: 123 Tice Blvd, Suite 101 Albany, NY 12205: 14 Walker Way Tonawanda, NY 14150: 275 Cooper Ave, Suite 105	Page	Date Rec'd in Lab	Pace Job #		
			1 of 1	11/05/25	L2569921		
Westborough, MA 01581 8 Walkup Dr. TEL: 508-898-9220 FAX: 508-898-9193	Mansfield, MA 02048 320 Forbes Blvd TEL: 508-822-9300 FAX: 508-822-3288	Project Information Project Name: <u>Blondell</u> Project Location: <u>Bronx, NY</u> Project # <u>41-0163256.00</u> (Use Project name as Project #) <input type="checkbox"/>		Deliverables <input type="checkbox"/> ASP-A <input type="checkbox"/> ASP-B <input checked="" type="checkbox"/> EQulS (1 File) <input type="checkbox"/> EQulS (4 File) <input type="checkbox"/> Other			
Client Information Client: <u>GZA Geo Environmental</u> Address: <u>104 West 29th Street</u> Phone: _____ Fax: _____ Email: <u>Zachary.Landis@GZA.com</u>		Project Manager: <u>Zach Landis</u> PACE Quote #: _____ Turn-Around Time Standard <input checked="" type="checkbox"/> <input checked="" type="checkbox"/> Due Date: <u>11/11/25</u> Rush (only if pre approved) <input type="checkbox"/> # of Days: <u>1 Week TAT</u>		Billing Information <input checked="" type="checkbox"/> Same as Client Info PO # _____			
These samples have been previously analyzed by Pace <input type="checkbox"/>		Regulatory Requirement <input type="checkbox"/> NY TOGS <input type="checkbox"/> NY Part 375 <input type="checkbox"/> AWQ Standards <input type="checkbox"/> NY CP-51 <input type="checkbox"/> NY Restricted Use <input type="checkbox"/> Other <input type="checkbox"/> NY Unrestricted Use <input type="checkbox"/> NYC Sewer Discharge		Disposal Site Information Please identify below location of applicable disposal facilities. Disposal Facility: <input type="checkbox"/> NJ <input type="checkbox"/> NY <input type="checkbox"/> Other: _____			
Other project specific requirements/comments: <u>*GZ-4 Sample has 2/3 Vials, Cynthia Pagot approved analysis with the 2 vials*</u> Please specify Metals or TAL.		ANALYSIS TCL Volatiles - 8260 NYCL Semi-volatiles - 8210 TCL Pesticides TCL PCBs Total TAL Metals - 6020/4g Dissolved TAL Metals - 6020/4g PFAS - 1633		Sample Filtration <input type="checkbox"/> Done <input type="checkbox"/> Lab to do Preservation <input type="checkbox"/> Lab to do (Please Specify below)			
PACE Lab ID (Lab Use Only)	Sample ID	Collection Date Time	Sample Matrix	Sampler's Initials	TALL NYCL TCL Pesticides TCL PCBs Total TAL Metals Dissolved TAL Metals PFAS	Sample Specific Comments	
<u>69921-01</u>	<u>GZM-2</u>	<u>11/4/25</u>	<u>9:30</u>	<u>Water</u>	<u>MD</u>	<u>X X X X X X X</u>	
<u>(MR) -02</u>	<u>GZM-1</u>	<u>↓</u>	<u>10:30</u>	<u>Water</u>	<u>↓</u>	<u>X X X X X X X</u>	
<u>-03</u>	<u>GZM-3</u>	<u>↓</u>	<u>11:30</u>	<u>Water</u>	<u>↓</u>	<u>X X X X X X X</u>	
<u>-04</u>	<u>GZM-4</u>	<u>↓</u>	<u>12:30</u>	<u>Water</u>	<u>↓</u>	<u>X X X X X X X</u>	
<u>-05</u>	<u>GZM-DUP</u>	<u>↓</u>	<u>-</u>	<u>Water</u>	<u>↓</u>	<u>X X X X X X X</u>	
<u>-06</u>	<u>Field Blank</u>	<u>↓</u>	<u>14:00</u>	<u>Water</u>	<u>-</u>	<u>X X X X X X X</u>	
<u>-07</u>	<u>Trip Blank</u>	<u>11/4/25</u>	<u>-</u>	<u>Water</u>	<u>-</u>	<u>X</u>	
Preservative Code: A = None B = HCl C = HNO ₃ D = H ₂ SO ₄ E = NaOH F = MeOH G = NaHSO ₄ H = Na ₂ S ₂ O ₃ K/E = Zn Ac/NaOH O = Other		Container Code: P = Plastic A = Amber Glass V = Vial G = Glass B = Bacteria Cup C = Cube O = Other E = Encore D = BOD Bottle		Westboro: Certification No: MA935 Mansfield: Certification No: MA015		Container Type Preservative	
Relinquished By: <u>[Signature]</u> (GZA)		Date/Time: <u>11/4/25 at 5:00pm</u>		Received By: <u>SSM</u>		Date/Time: <u>11-4-25 1708</u>	
[Signature] NJSC SS		11-4-25 1509		[Signature] NJSC SS		11-4-25 1607	
[Signature]		11/5 020		[Signature]		11/5 020	

Please print clearly, legibly and completely. Samples can not be logged in and turnaround time clock will not start until any ambiguities are resolved. BY EXECUTING THIS COC, THE CLIENT HAS READ AND AGREES TO BE BOUND BY PACE'S TERMS & CONDITIONS. (See reverse side.)