BROWNFIELD CLEANUP PROGRAM

REMEDIAL ACTION WORK PLAN
(RAWP)

FOR

HNJ REALTY LLC.
FOR
SCHMUKLERS CLEANERS
358 - 364 North Avenue, New Rochelle, New York
Site No.: C360088
Index No.: A3-0542-0306

PREPARED FOR
NEW YORK STATE DEPARTMENT OF
ENVIRONMENTAL CONSERVATION
625 BROADWAY
ALBANY, NEW YORK 12233-7016



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August 2026

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TABLE OF CONTENTS

P.E. CERTIFICATION

LIST OF ACRONYMS

1.0 INTRODUCTION
2.0 SITE DESCRIPTION AND HISTORY
- 2.1 Site Location
- 2.2 Public Water Supply
- 2.3 Surface Water and Hydrogeology
- 2.4 Institutional Controls

3.0 SUMMARY OF REMEDIAL INVESTIGATION
- 3.1 Current Environmental Conditions

4.0 REMEDIAL GOALS AND REMEDIAL ACTION OBJECTIVES
5.0 RECOMMENDED REMEDY
6.0 REMEDIAL ACTION WORK PLAN (RAWP)
- 6.1 Enhanced Biological Remediation
  - *6.1.1 Application Well Locations and Specifications*
  - *6.1.2 Dosage Quantities*
  - *6.1.3 Application Procedure*
  - *6.1.4 Application Monitoring*
  - *6.1.5 Chemical Handling, Staging and Storage*
  - *6.1.6 Project Schedule*
  - *6.1.7 Daily Reporting Requirements*

7.0 GREEN AND SUSTAINABLE REMEDIATION
8.0 POST REMEDY EFFECTIVENESS
9.0 HEALTH AND SAFETY
- 9.1 Community Air Monitoring Plan (CAMP)

10.0 CONCLUSION

FIGURES

Figure-1 Site Location
Figure-2 Site Survey

Figure-3 Historic Data
Figure-4 Application/Monitoring Well Locations
Figure-5 Groundwater Sampling Post map

ATTACHMENTS

Attachment-A Micro-Bac Brochure
Attachment-B Lab Case Study
Attachment-C M1000H Product Information
Attachment-D Tri-Phasic 12 Product Information
Attachment-E M1000H Material Safety
Attachment-F Community Air Monitoring Plan (CAMP)
Attachment-G As-built SVE/SSD system drawings
Attachment-H Remedial Investigation validated lab data

APPENDICES

Appendix-A Professional staff resumes

Certification

I, Nicholas A. Andrianas certify that I am currently a NYS registered professional engineer as defined in 6 NYCRR Part 375 and that this Remedial Action Work Plan was prepared in accordance with all applicable statutes and regulations and in substantial conformance with the DER Technical Guidance for Site Investigation and Remediation (DER-10).



Nicholas A. Andrianas, P.E.

Date: 8/12/26

List of Acronyms

AS	Air Sparging
ASP	Analytical Services Protocol
BCA	Brownfield Cleanup Agreement
BCP	Brownfield Cleanup Program
CERCLA	Comprehensive Environmental Response, Compensation and Liability Act
CAMP	Community Air Monitoring Plan
C/D	Construction and Demolition
CFR	Code of Federal Regulation
CLP	Contract Laboratory Program
COC	Certificate of Completion
CO2	Carbon Dioxide
CP	Commissioner Policy
DER	Division of Environmental Remediation
EC	Engineering Control
ECL	Environmental Conservation Law
ELAP	Environmental Laboratory Approval Program
ERP	Environmental Restoration Program
EWP	Excavation Work Plan
GHG	Green House Gas
GWE&T	Groundwater Extraction and Treatment
HASP	Health and Safety Plan
IC	Institutional Control
NYSDEC	New York State Department of Environmental Conservation
NYSDOH	New York State Department of Health
NYCRR	New York Codes, Rules and Regulations
O&M	Operation and Maintenance
OM&M	Operation, Maintenance and Monitoring
OSHA	Occupational Safety and Health Administration
OU	Operable Unit
PID	Photoionization Detector
PRP	Potentially Responsible Party
PRR	Periodic Review Report
QA/QC	Quality Assurance/Quality Control
QAPP	Quality Assurance Project Plan
RAO	Remedial Action Objective
RAWP	Remedial Action Work Plan
RCRA	Resource Conservation and Recovery Act
RI/FS	Remedial Investigation/Feasibility Study
ROD	Record of Decision
RP	Remedial Party
RSO	Remedial System Optimization
SAC	State Assistance Contract
SCG	Standards, Criteria and Guidelines

SCO	Soil Cleanup Objective
SMP	Site Management Plan
SOP	Standard Operating Procedures
SOW	Statement of Work
SPDES	State Pollutant Discharge Elimination System
SSD	Sub-slab Depressurization
SVE	Soil Vapor Extraction
SVI	Soil Vapor Intrusion
TAL	Target Analyte List
TCL	Target Compound List
TCLP	Toxicity Characteristic Leachate Procedure
USEPA	United States Environmental Protection Agency
UST	Underground Storage Tank
VCA	Voluntary Cleanup Agreement
VCP	Voluntary Cleanup Program

1.0 INTRODUCTION

The following document is a Remedial Action Work Plan (RAWP) prepared by BEI Consulting LLC and Nicholas Andrianas P.E. on behalf of HNJ Realty LLC, pursuant to the requirements of an executed Brownfield Cleanup Program Agreement (BCA) (dated February 27, 2006), between the New York State Department of Environmental Conservation (NYSDEC), Division of Environmental Remediation (DER) and HNJ Realty, LLC, *the Volunteer*. The Site is a commercial property located at 358 through 364 North Avenue, New Rochelle, New York (Figures 1 and 2), fully described as Section 4 - Block 1206 - Lot 19 of the tax maps of City of New Rochelle.

The purpose of the selected RAWP is to protect human health and the environment, attain the Remedial Action Objectives (RAOs) selected for the site to the extent feasible and prevent migration of on-site groundwater contamination.

2.0 SITE DESCRIPTION AND HISTORY

According to review of the City of New Rochelle records, as well as a review of historical Sanborn Fire Insurance Maps, the Site was developed as early as 1891 with two residential-use buildings. The property was subsequently redeveloped with a multi-tenant two-story commercial building. The first floor and basement of the building have been utilized by Schmukler's Dry Cleaners since around 1914. This business has historically operated on the first floor and in the basement of the building. Currently, the first floor of the building is being rented by United Community Center owned by Jackie A. The southern portion of the building is currently being rented by Comfort Homes. The second floor of the building provides professional space including: a spa and HNJ Realty office.

Physical Site Description

Site Name: Schmukler's Dry Cleaners
358 through 364 North Avenue, New Rochelle, New York
Owner: HNJ Realty, LLC
Location: 358 through 364 North Avenue, New Rochelle, New York
Latitude -73.784480", Longitude "40.914176"
Brownfield Cleanup Agreement Site No.: C360088
Index No.: A3-0542-0306

2.1 Site Location

The Site is located at an elevation of approximately 80 feet above mean sea level according to review of United States Geological Survey (USGS) Topographic Map, Mount Vernon Quadrangle (Figure 1). The Site consists of a partial two-story/part one-story commercial building on a 0.21-acre parcel identified as the street addresses 358 through 364 North Avenue, New Rochelle, New York. The subject parcel (Site) is located in the City of New Rochelle, County of Westchester. The property is assessed as 0.21 acres in size and improved with a commercial building constructed in 1914. The owner of record of the Site is HNJ

Realty, LLC[1]. An addition was made to the western portion of the building circa 1937. An addition was also made to the southwestern portion of the building sometime prior to 1987. A final addition was made to the northwestern portion of the building (which includes the steam boiler) in 1987.

The Site is located within a mixed commercial/residential use area. Properties along North Avenue are predominantly occupied for commercial use, with some maintaining residential apartments above. Commercial uses dominate areas within one block east and west of North Avenue. Beyond this one block area, residences are the prevailing land-use. The following businesses are located to the north of the subject site on the west side of North Avenue: 366 Real Estate Company; 360 Family Dentistry. Located on the southern side of Lockwood Avenue heading to the west the following businesses are noted: 2 Lockwood, Mattress Dealer; 6 Lockwood, Cabinet Store; 8 Lockwood, LB Inc Swimming Pools.; 10 Lockwood, Modern Press; 12 Lockwood, LBI. Located to the direct west is 17 May Street which is mainly a residential complex. To the south on the west side of North Avenue is Love Music Inc.

2.2 Public Water Supply

Public water is provided to the Site by United Water New Rochelle. No on-site potable or dry cleaning or washing make-up supply wells, active or inactive, have been observed at the site. According to information provided by United Water, the supply source is surface water that is purchased from the New York City Department of Environmental Protection (NYCDEP). The three sources of the New York City supply that is utilized includes the Croton, Catskill and Delaware Aqueducts. The Central Avenue and little Catskill pump stations supply the day to day demands to the system.

2.3 Surface Water and Hydrogeology

According to the Surficial Geology Map of New York Lower Hudson Sheet produced by the

[1]According to the information provided by the client.

University of the State of New York; State Education Department, 1989, the Site is located within an area of New York where the surficial geology is defined as glacial till. The till is described as variable texture (e.g., clay, silt-clay, boulder clay), usually poorly sorted, which was deposited beneath glacial ice. The till is generally characterized by mixtures of relatively impermeable loamy matrix-to-sandy in areas underlain by gneiss or sandstone. The thickness of the till varies between one and 50 meters.

The Site is located in the eastern portion of the City of New Rochelle and groundwater flow direction could not be determined from published information. As groundwater is not used as a potable water source in the Site vicinity, current groundwater quality data are very limited. No water table elevation maps are available for the City of New Rochelle. However, given both the surface and bedrock topography in the area (which generally slopes down toward the Long Island Sound), groundwater flow in unconsolidated deposits and/or upper weathered bedrock is likely easterly-southeasterly, toward the Long Island Sound. A groundwater flow survey was conducted at the site as part of the supplemental remedial investigation activities utilizing all available overburden wells and bedrock wells. The results of the survey concluded that groundwater appears to be moving in a south-easterly direction.

Due to the heavily developed nature of the immediate surrounding area, groundwater quality is expected to be regionally degraded. No surface water bodies are located on, adjoining or proximate to the Site. Ferris Creek and The Long Island Sound are located approximately 3,250 feet to the east/southeast of the Site. According to Freshwater Wetland, National Wetland Inventory, Westchester Wetlands and Tidal Wetlands data available on the Westchester County GIS System, there are no regulated wetlands on or adjoining the Site.

2.4 Institutional Controls (ICs)

The ICs for the site implemented during the RAWP are outlined in the Site Management Plan (SMP) for the site and will remain as follows:

- Controlled property may be used for: Restricted Residential use as described in 6 NYCRR Part 375-1.8(g)(2)(ii), Commercial use as described in 6 NYCRR Part 375-1.8(g)(2)(iii) and Industrial use as described in NYCRR Part 375-1.8(g)(2)(iv);

- All ECs must be operated and maintained as specified in this Interim Site Management Plan (ISMP);

- All ECs must be inspected at a frequency and in a manner defined in the ISMP.

- The use of groundwater underlying the property is prohibited without necessary water quality treatment as determined by the NYSDOH or the Westchester Department of Health to render it safe for use as drinking water or for industrial purposes, and the user must first notify and obtain written approval to do so from the Department.

- Groundwater and other environmental or public health monitoring must be performed to include: annual monitoring and sampling of subject site monitoring wells

- Data and information pertinent to site management must be reported at the frequency and in a manner as defined in this ISMP;

- All future activities that will disturb remaining contaminated material must be conducted in accordance with this ISMP;

- Monitoring to assess the performance and effectiveness of the remedy must be performed as defined in this ISMP;

- Operation, maintenance, monitoring, inspection, and reporting of any mechanical or physical component of the remedy shall be performed as defined in this ISMP;

- Access to the site must be provided to agents, employees or other representatives of the State of New York with reasonable prior notice to the property owner to assure compliance with the restrictions identified by the Environmental Easement (EE).

- The potential for vapor intrusion must be evaluated for any buildings developed in the area within the IC boundaries noted on Figure 4, and any potential impacts that are identified must be monitored or mitigated; and

- Vegetable gardens and farming on the site are prohibited;

3.0 SUMMARY OF REMEDIAL INVESTIGATION

Historic data collected over the past several years has indicated the presence of chlorinated Volatile Organic Compounds (VOCs) and petroleum related VOCs at the subject site in soil and groundwater. The former dry-cleaning equipment room has been the main source area for

chlorinated contamination and continues to be the main focus of our remedial efforts. A Soil Vapor Extraction (SVE)/ Sub-slab Depressurization System (SSDS) combo remediation system has been in operation since approximately the winter of 2009 and has been removing volatiles from the sub surface around the clock. The SVE/SSDS system is geared heavily towards the dry-cleaning equipment room and the basement. The system also extends to the rear of the building focusing on the former dry-well where substantial Perchloroethylene (PCE) was found in GW-9 at 25,000 ppb. Please refer to Attachment-G for the as-built engineering drawings. Concentrations of PCE have been discovered in GW-4 as high as 830,000 ppb and Trichlorethylene (TCE) as high as 58,000 ppb in groundwater beneath the dry-cleaning equipment room. (note: These concentrations are from the August 2007 RI). The active SVE/SSDS system has likely reduced these numbers since the system's startup during the winter of 2009. Please see Figure-3 for historical data discussed above and Attachment-H for the validated lab data package. In December of 2008 MW-3 (located in the equipment room) was sampled and lab data indicated that PCE had a concentration of 26,000 ppb, TCE at 6,000 ppb, DCE at 2,400 ppb and Vinyl Chloride (VC) at 430 ppb. Samples were also collected during November of 2009 from MW-3, just prior to the start-up of the SSDS/SVE system, and PCE was found to be at 75,000 ppb, TCE at 4,000, DCE at 4,100 ppb and VC at 250. Nearly four years later, during August of 2012, PCE concentrations at MW-3 were reduced to 18,000 ppb, TCE was significantly reduced to 810 ppb, DCE was recorded at 1,400 ppb and VC at just 61 ppb. This is most likely attributed to the active and ongoing SVE/SSDS system installed during the winter 2009 volatilizing VOCs from the shallow groundwater. A total of three (3) groundwater samples (GW-1, GW-5 and GW-6) were collected from the basement. The PCE concentration in the sample collected at GW-1was non-detect, the PCE concentration in the GW-6 sample to the west of this location detected PCE at 310 ppb and the PCE concentration in the GW-5 sample collected at the center of the southern basement wall was 150 ppb. Chlorinated VOC levels totaled approximately 10,000 ppb in the basement with the majority of the contamination consisting of DCE (8,700 ppb) in GW-6. Petroleum related contamination has been found throughout the site in groundwater, mainly in the basement and the former dry-cleaning equipment room. Petroleum constituent levels have been detected as high as 890 ppb (1, 3, 5 trimethylbenzene) in the equipment room and 670 ppb (naphthalene). The source of these impacts in unknown.

3.1 Current Environmental Conditions

The current groundwater conditions at the site are based on the latest groundwater sampling data (July 2023-July 2025) collected as part of the bi-annual sampling program. The main source area at the site has been and remains to be the area of MW-3, which is located within the former dry-cleaning equipment room. PCE has ranged in concentration levels from approximately 13,000 ppb -32,000 ppb over the last two (2) years. PCE daughter breakdown products such as TCE and cis/trans 1,2-dichloroethene (total DCE) have also fluctuated over the years. TCE concentrations have ranged between 5,000 ppb - 40,000 ppb, while total DCE concentrations ranged between 1,600 ppb -10,000 ppb. VC has remained at levels ranging from 75 ppb - 230ppb. The remaining monitoring wells at the site including off-site bedrock wells down gradient of the MW-3 location continue to exhibit very minor to non-detect readings for all chlorinated contamination. Please see Figure-5 for a post map with the latest monitoring well data.

Soil gas concentrations have been documented on a quarterly basis while sampling the SVE/SSD system under the WCDOH air permit. Influent sub-slab vapor concentrations for PCE were documented at a level of 7,050 ug/m3, TCE at 42 ug/m3, total DCE at 24 ug/m3 and VC was non-detect. These detections were recorded during the July 2025 SVE/SSD system quarterly sampling event. Please refer to Attachment-G for the as-built engineering drawings.

The SVE/SSDS was due for a three (3) year inspection by the Westchester County Department of Health (WCDOH) during October of 2025. Upon arrival to perform the system inspection the engineering control (vacuum blower) was not operational. The engineering control for the system has since been replaced during August of 2026 and is fully operational. A new Airtech® model vacuum blower was installed and sub-slab vacuum was confirmed at all sample point locations. (PV-1-PV-6) The re-inspection of the WCDOH air discharge permit is currently in the process of being scheduled.

4.0 REMEDIAL GOALS AND REMEDIAL ACTION OBJECTIVES

Based upon contamination discovered above regulatory standards for groundwater the following Remedial Action Objectives (RAOs) will apply during the remedial phase in order to conform with Track-4 clean-up requirements in order to protect the environment and public health:

- prevent migration of groundwater related contamination that would result in further off-site groundwater or surface water contamination
- restore groundwater aquifer to pre-disposal/pre-release conditions, to the extent practicable (Division of Water TOGS 1.1.1) with provisions for ICs/ECs in order to prevent on-site potable groundwater use exceeding drinking water standards:
- prevent ingestion of groundwater with contaminant levels exceeding drinking water standards
- remove source of groundwater contamination
- prevent contact with, or inhalation of volatiles, from all contaminated media via the current on-going Interim Remedial Measure (IRM).

Based upon the potential for Soil Vapor Intrusion (SVI) due to groundwater and soil contamination, the following RAOs apply for the protection of the environment and public health:

- mitigate potential impacts to public health resulting from existing, or the potential for, soil vapor intrusion through the use of the SVE/SSDS (Guidance for Evaluating Soil Vapor Intrusion - October 2006)

An IRM to address vapor intrusion in the form of an SVE/SSDS in order to protect the environment, public health and on-site workers was developed and approved by the Department on July 23, 2015. An IRM Construction Completion Report (CCR) was issued to the Department in February of 2016.

5.0 RECOMMENDED REMEDY

The following section provides information on the selected remedy (Enhanced Bioremediation) for the site. The selected remedial measure is expected to meet the RAOs established for the site, to the extent feasible, reduce the potential for human exposure to contaminated media and prevent migration of on-site contamination.

5.1 Microbial (Bac-t) and Nutrients

The proposed remedial measure to be undertaken at the subject property involves treatment of chlorinated VOCs through the process of enhanced biodegradation. The use of bacteria (microbes/bac-t or M-1000H®) and Tri-Phasic®12 (nutrients), is proposed to help stimulate the degradation process where the proper naturally occurring microbes may be lackluster or nonexistent. The applied microbes will produce enzymes that break down petroleum products and halogenated compounds, and utilize the compounds as a food source. This remedial method of introducing bac-t and nutrients to a contaminated aquifer has been widely recognized by the environmental industry throughout the United States when treating hydrocarbon and chlorinated solvent spill incidents. The biological mixture will be applied to the sub-surface, at available monitoring wells in the areas of concern MW-9, MW-10 and MW-12. A Pilot Study using the existing monitoring wells was previously performed at the Site, which included the use of the above-mentioned biological solution focusing on the treatment of both chlorinated VOCs and petroleum related SVOCs via enhanced bioremediation.

5.2 Light Non-aqueous Phase Liquid (LNAPL)

Down gradient bedrock well BW-4 has historically contained LNAPL contamination in the form of #2 fuel oil. LNAPL and water/product mixture will be contained in DOT approved 55-gallon drums and stored on-site for proper disposal. The amount of product recovered and manifests for transportation to an approved disposal facility will be documented within bi-annual sampling Reports. In addition, more aggressive approaches such as vacuum enhanced fluid recovery (VEFR) will be considered as the selected remedy.

6.0 REMEDIAL ACTION WORK PLAN (RAWP)

6.1 Enhanced Biological Remediation

The following section discusses the implementation of the proposed remedial action for remediation of groundwater, which includes the following information: well point locations, application procedure, dosage amounts, product information and health and safety.

6.1.1 Application Well Locations and Specifications

Monitoring wells MW-9, MW-10 and MW-12 will be utilized for the application procedure. Down gradient monitoring wells (MW-3 and MW-6) will be utilized to monitor the application process discussed in 6.1.3.

Specifications on the following wells utilized as application wells are as follows: MW-9=10' of 2" diameter PVC riser pipe with 5' of 2" diameter PVC slot screen; MW-10= 5' of 2" PVC 0.02" slot screen, 10' of 2" PVC riser to grade and MW-12- 4' of 2" PVC 0.02" slot screen, 2' of 2" PVC riser to grade. Each well was installed with a 2' bentonite seal above the screened interval and finished at grade surface with a 5" stainless steel manhole cover, cement sealed to prevent surface water infiltration. Please refer to Figure- 4 for the locations of the intended application wells and monitoring wells.

6.1.2 Dosage Quantities

The bac-t and nutrients are proposed to be applied in multiple areas including: the former dry cleaning equipment area (source area), and the southwestern basement area.

Treatment Area	M1000H (bac-t)	Nutrient/H2O Mix	Number of Applications
Upgradient (MW-9)	41.25 gallons M1000H	41.25 gallons nut/h2O	1 application event
Upgradient (MW-10)	41.25 gallons M1000H	41.25 gallons nut/h2O	1 application event

Down gradient in Basement (MW-12)	27.5 gallons M1000H	27.5 gallons nut/h2O	1 application event

- Note 1 gallon of nutrients is mixed with 54 gallons of water
- 55 gallons (1 drum) of bacteria requires 55 gallons of nutrient/water

6.1.3 Application Procedure

In order to apply the bac-t and nutrient mix to the MW-9, 10 and 12 areas, a low flow peristaltic pump will be utilized to introduce the bio solution to the impacted sub-surface area. A small submersible pump will be connected to a clean 3/8" piece of poly-tubing, which will be used to draw the bacteria solution from a small holding container. The tubing will be lowered to the bottom of the well at which point the pump is turned on subsequently dispersing the solution into the sub-surface at a very low flow rate. The minimal pressure generated from the pump allowed for the solution to disperse vertically and laterally throughout the contaminated aquifer and vadose zone. The nutrient mix is prepared separately (1 gallon of nutrients mixed with 55-gallons of water) and applied in the same manner as the bacteria, at a low flow rate. *The nutrient mixture will be applied prior to the bacteria solution to ensure the appropriate conditions are present for optimal dichlorination.* Please see Figure-4 for the proposed application areas. During the application event the SSDS/SVE system will be temporarily shut-down.

6.1.4 Application Monitoring

Before, during and post application of the bacteria and nutrient installment, nearby down gradient monitoring wells (MW-3 and BW-4) will be gauged for Depth to Water (DTW). DTW readings will be gauged pre, during and post application for changes in elevation.

6.1.5 Chemical Handling, Staging and Storage

The M1000H and nutrients will be stored off-site prior to the application event. These products are natural, non-pathogenic, non-engineered biological products that meet the Environmental Protection Agency (EPA) requirements for release into the environment. Special clothing or equipment is not required for handling M1000H and nutrients. Please see Attachments-A, C and D for the company information and specification sheets on the proposed product.

6.1.6 Project Schedule

The RAWP is anticipated to be implemented within 120 days of the approval. As discussed below, dates for submissions of deliverables submitted to the DER will comply with the bi-annual groundwater sampling events utilized to evaluate pre and post-application remedy effectiveness. Please see table below for scheduling dates and milestones:

Milestone	**Date**
RAWP acceptance	October 2026
RAWP field mobilization upon RAWP approval (including ordering of materials, delivery of product and securing a sub-contractor	February 2027
Final Engineering Report (FER)	October 2027
Bi-annual groundwater sampling	October 2026 (second event of 2026)

The Final Engineering Report (FER) will be revised and updated upon completion of the final post round sampling event for the site.

6.1.7 Daily Reporting Requirements

Daily Reporting for the RAWP field mobilization will include at a minimum the following items:

- Date of the Reporting period
- Work start and end times
- Weather conditions
- Work activities summary for the Reporting period
- Samples collected
- CAMP summary with map of sampling locations
- Problems or delays encountered during the Reporting period
- Planned activities for the next Reporting period; and
- Relevant site photos

7.0 GREEN AND SUSTAINABLE REMEDIATION

7.1 Green Remediation Evaluation

NYSDEC's DER-31 Green Remediation requires that green remediation concepts and techniques be considered during all stages of the remedial program including site management, with the goal of improving the sustainability of the cleanup and summarizing the net environmental benefit of any implemented green technology. This section of the RAWP provides a summary of any green remediation evaluations to be completed for the site during the remedial phase of work.

It is anticipated that there will be very minimal energy consumption needed to perform the proposed remedial actions. A low flow peristaltic pump that utilizes 110-volt power will be used to deliver the remediation fluids into the proposed remediation wells. All other instrumentation utilized to monitor for the CAMP and remedial effectiveness requires rechargeable/replaceable battery-operated equipment.

The proposed bioremediation remediation method is consistent with NYSDEC's "DER-31 Green and Sustainable" remediation guidelines based on the following:

1. Uses enhanced natural environmental processes to effect remediation;
2. Enhanced bioremediation is a low energy alternative;
3. Bioremediation as source removal with monitored natural attenuation is a green remediation method;
4. Bioremediation permanently destroys contamination to reduce environmental impacts associated with long-term site management;
5. Addresses the source more aggressively to reduce long-term operation and maintenance of a more energy consuming remediation such as a pump and treat technology;
6. Minimizes waste generation;
7. Integrates with contemplated reuse of site without the need to design and construct new buildings and structures that would be required for traditional pump and treat systems;
8. Poses fewer short-term and long-term ancillary impacts to the environment;
9. Implementation of bioremediation results in fewer GHG emissions than traditional pump and treat technologies;
10. The remedy requires a small environmental footprint;
11. Achieves the remedial action objectives more sustainably;
12. Allows for the greenest site reuse with new buildings and structures;
13. Achieves a permanent cleanup; and
14. Permanently and significantly reduces the toxicity, mobility or volume of contamination.

8.0 POST REMEDY EFFECTIVENESS

8.1 Pre/Post Application Sampling Results

Pre application sampling data from the bi-annual sampling Reports will be used in order to evaluate the effectiveness of the remedy. The sampling data from the bi-annual sampling events will also be used as post remedy effectiveness evaluation. Specifically, monitoring wells that are included in the routine bi-annual sampling events will be used to measure effectiveness. This would include MW-3, which is the only monitoring well that exhibits elevated chlorinated contamination.

8.2 Contaminant Migration Prevention

The implementation of the down gradient application at well MW-12 will ensure the prevention of off-site contaminant migration. This well is located directly down gradient of the main source area at MW-3 and the application of microbes/nutrients in this area will create a biological curtain in order to prevent the potential for off-site impacts.

9.0 HEALTH AND SAFETY

All Micro-Bac® products are natural, non-pathogenic, and are not genetically engineered. They were developed by Micro® Bac's research and development laboratories and are manufactured under strict quality guidelines with batch and lot control. No special clothing or equipment is necessary to handle the product. Micro-Bac® products meet United States Environmental Protection Agency (USEPA) requirements for release into the environment.

In order to ensure the safety of our workers and staff involved with the treatment procedure BEI adheres to the Occupational Safety and Health Administration (OSHA) guidelines. During the application procedure BEI field technicians and staff will be equipped with Level D personal protective equipment (PPE). Level D OSHA guidelines require the following PPE:

- Eye protection- wear goggles or face shield (splash prevention)
- Head- hard hat when required

- Respiratory- Use dust respirator approved by NIOSH/MSA
- Hands- Wear neoprene gloves
- Feet- Wear steel-toe boots with chemical resistant soles or neoprene covers
- Clothing- Long sleeve shirts and long pant legs. Consider using Tyvek body suit or coveralls

The Safety Data Sheet (SDS) for the M1000H product is included as Attachment-E.

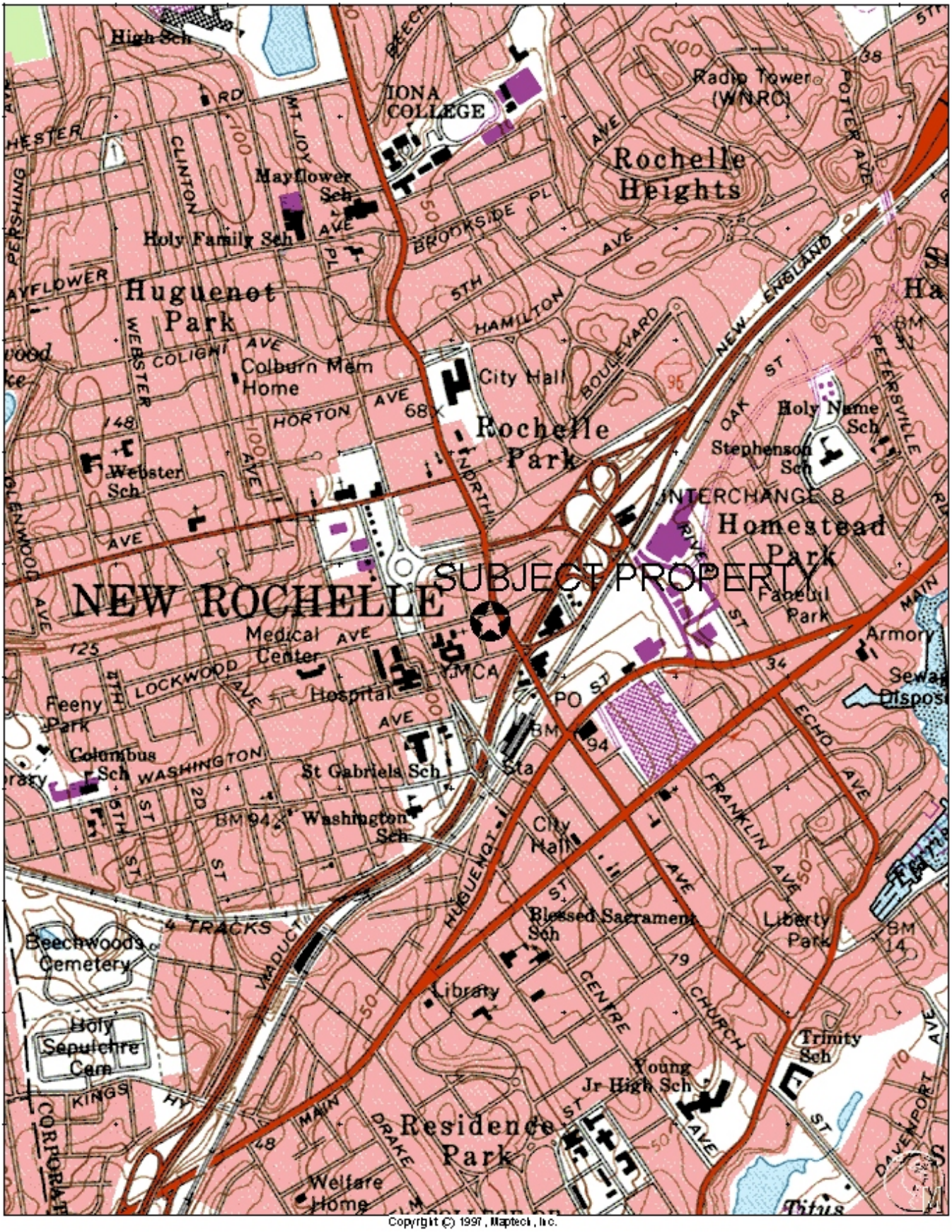
9.1 Community Air Monitoring Plan (CAMP)

A CAMP has been developed and is included as Attachment-F. The CAMP discusses established monitoring requirements that are employed during all ground intrusive activities in order to protect off-site receptors including: residences and businesses, and on-site workers not directly involved with the subject work activities.

10.0 CONCLUSION

The overall goal of the remedy is to meet the RAO's established for the site and document a continued downward trend in groundwater contamination. Pre application sampling data from the bi-annual sampling Reports will be used in order to evaluate the effectiveness of the remedy. The sampling data from the bi-annual sampling events will also be used as post remedy effectiveness evaluation. Upon completion of this Work Plan, if post-monitoring does not demonstrate groundwater contamination has been contained to the site boundary and demonstrate a reduction of groundwater contamination on-site, further remedial action will be required.

FIGURES



Remedial Investigation Report November 2007

Figure 1- Site Location	**Schmuklers Cleaners** **358 - 364 North Avenue** **New Rochelle, NY** **Site #C360088** **Index# A3-0542-0306**	BEI Consulting LLC C. Moriches, NY

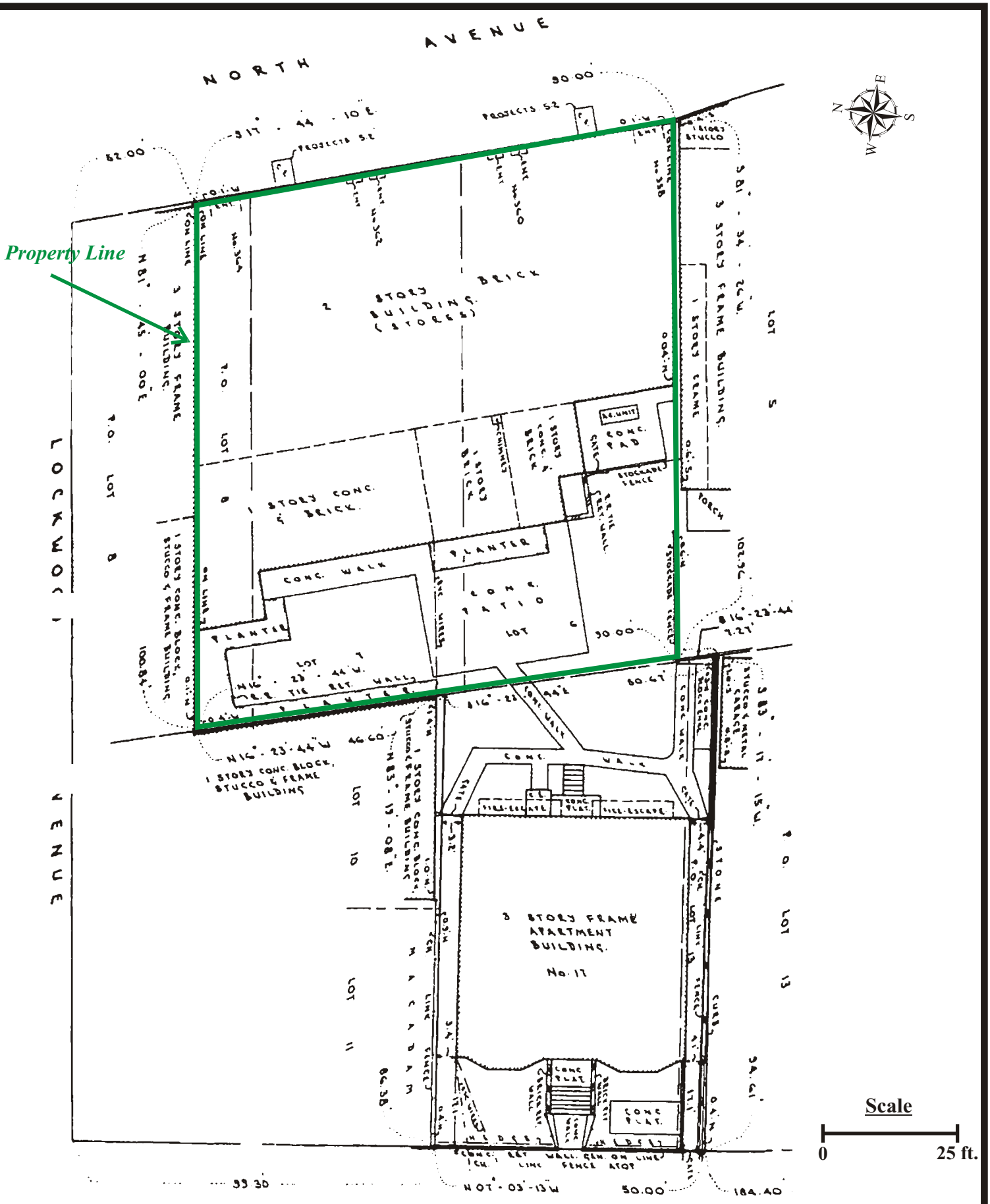
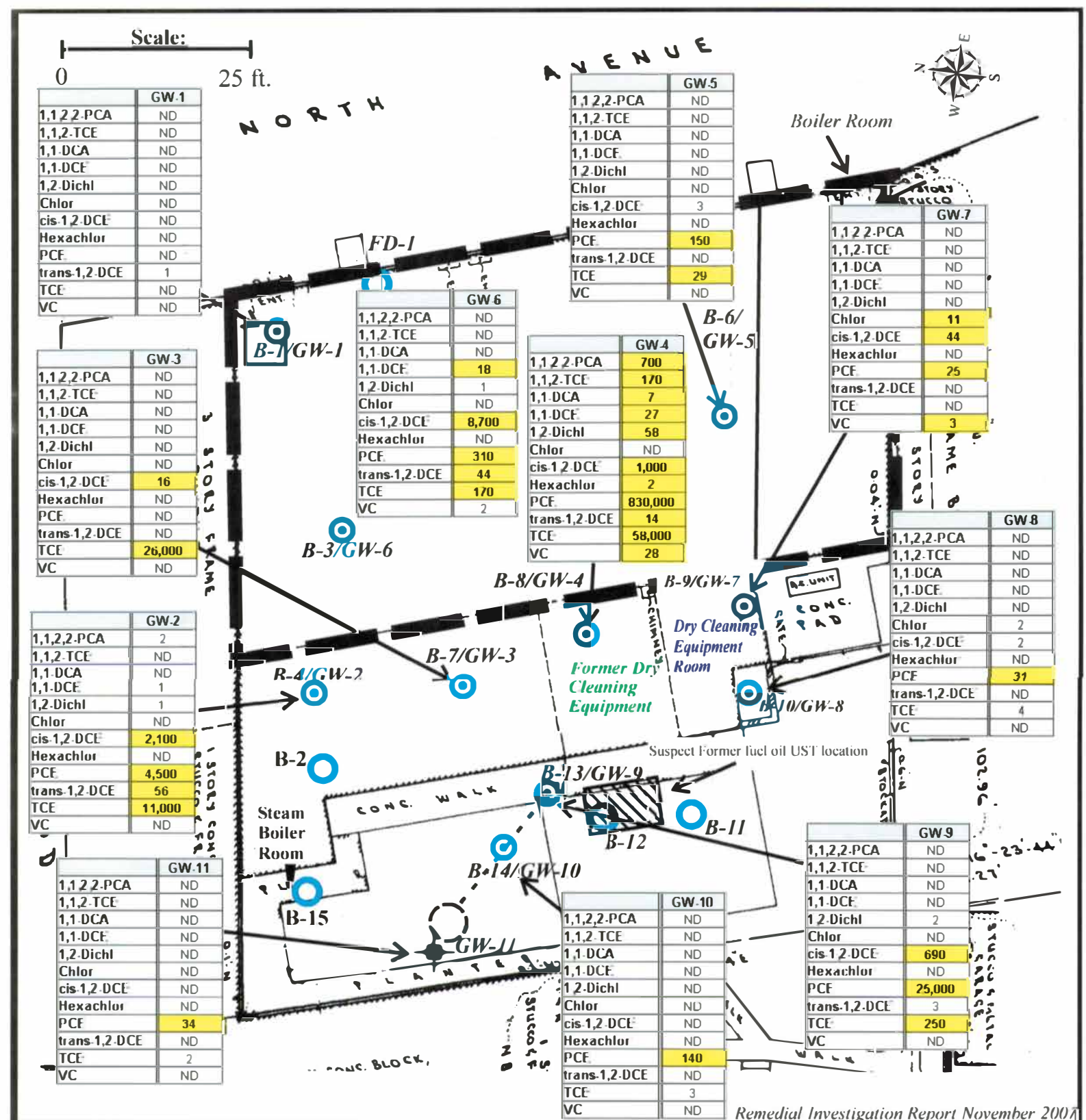


Figure 2 - Site Survey *survey performed by L.S not listed	**Schmuklers Cleaners** **358 - 364 North Avenue** **New Rochelle, NY** **Site #C360088** **Index# A3-0542-0306**	BEI Consulting LLC C. Moriches NY



Remedial Investigation Report November 2007

1,1,2,2-PCA- 1,1,2,2-Tetrachloroethane; **1,1,2-TCE**- 1,1,2-Trichloroethene; **1,1 - DCA** - 1,1-Dichloroethane; **1,1-DCE**- 1,1-Dichloroethene; **1,2-Dichl**- 1,2-Dichlorobenzene; **Chlor** - Chloroethene; **cis-1,2-DCE** - cis-1,2-Dichloroethene; **Hexachlor**- Hexachlorobutadine; **PCE**- Tetrachloroethene; **TCE** - Trichloroethene; **VC** - Vinyl chloride

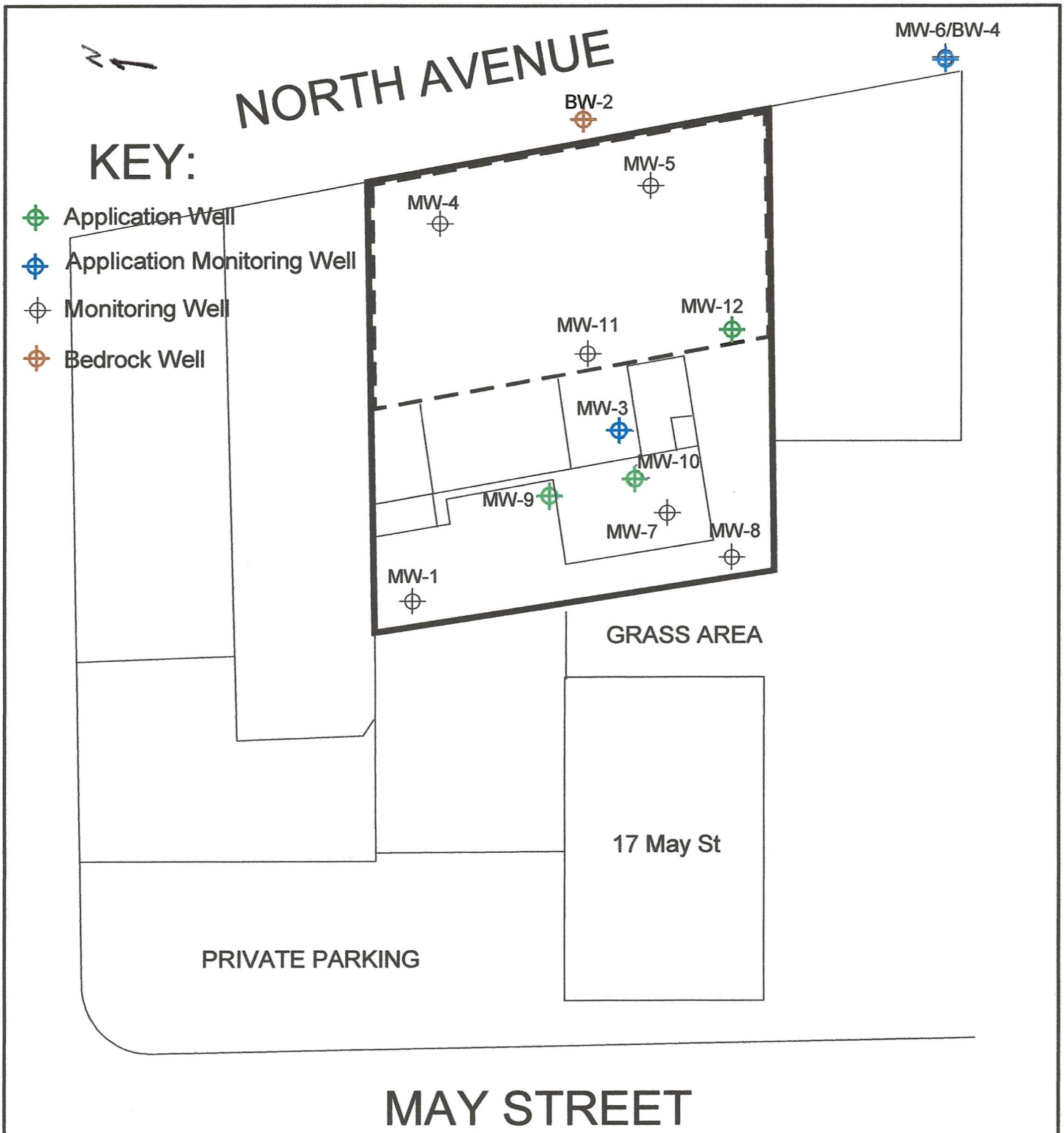
Bolded and highlighted concentrations are indicative of VOC detected at concentration exceeding applicable NYSDEC Class GA Groundwater Standards and/or Guidance Values

- Basement Portion of the Building - Soil Only Sampling Location - Soil and groundwater sampling locations - Groundwater Only Sample Location

Figure 3 - Halogenated VOCs detected in Groundwater Samples in micrograms per liter (ug/L)

Schmuklers Cleaners
358 - 364 North Avenue
New Rochelle, NY
Site #C360088
Index# A3-0542-0306

BEI Consulting LLC.
C.Moriches New York

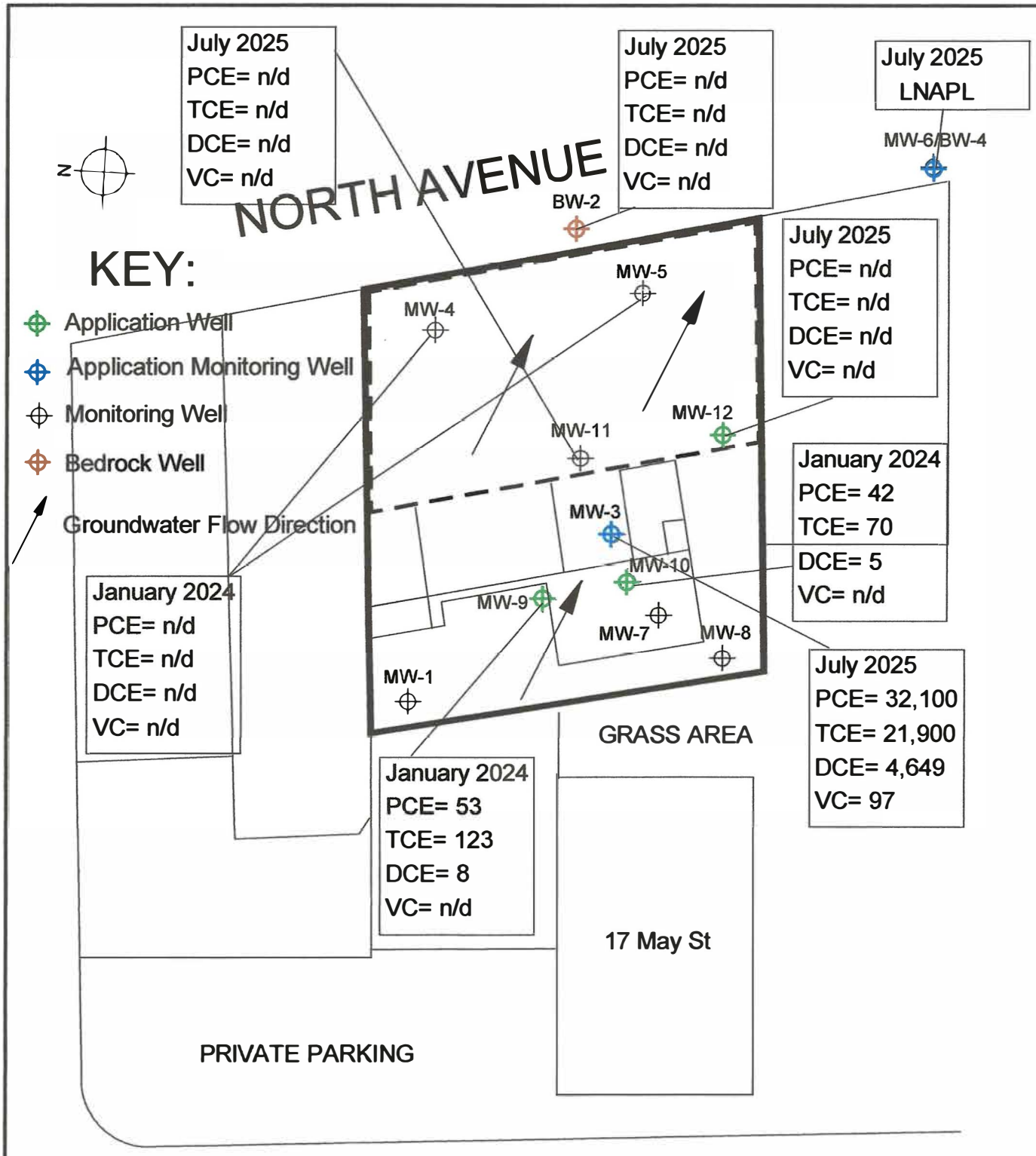


Application/Monitoring Well Locations

FIGURE-4

Former Schmuklers Cleaners
358-364 North Avenue
New Rochelle, NY

BEI Consulting LLC
Center Moriches, NY



* groundwater concentrations in ppb

Application/Monitoring Well Locations	Former Schmuklers Cleaners	BEI Consulting LLC
	358-364 North Avenue	Center Moriches, NY
FIGURE-5	New Rochelle, NY	

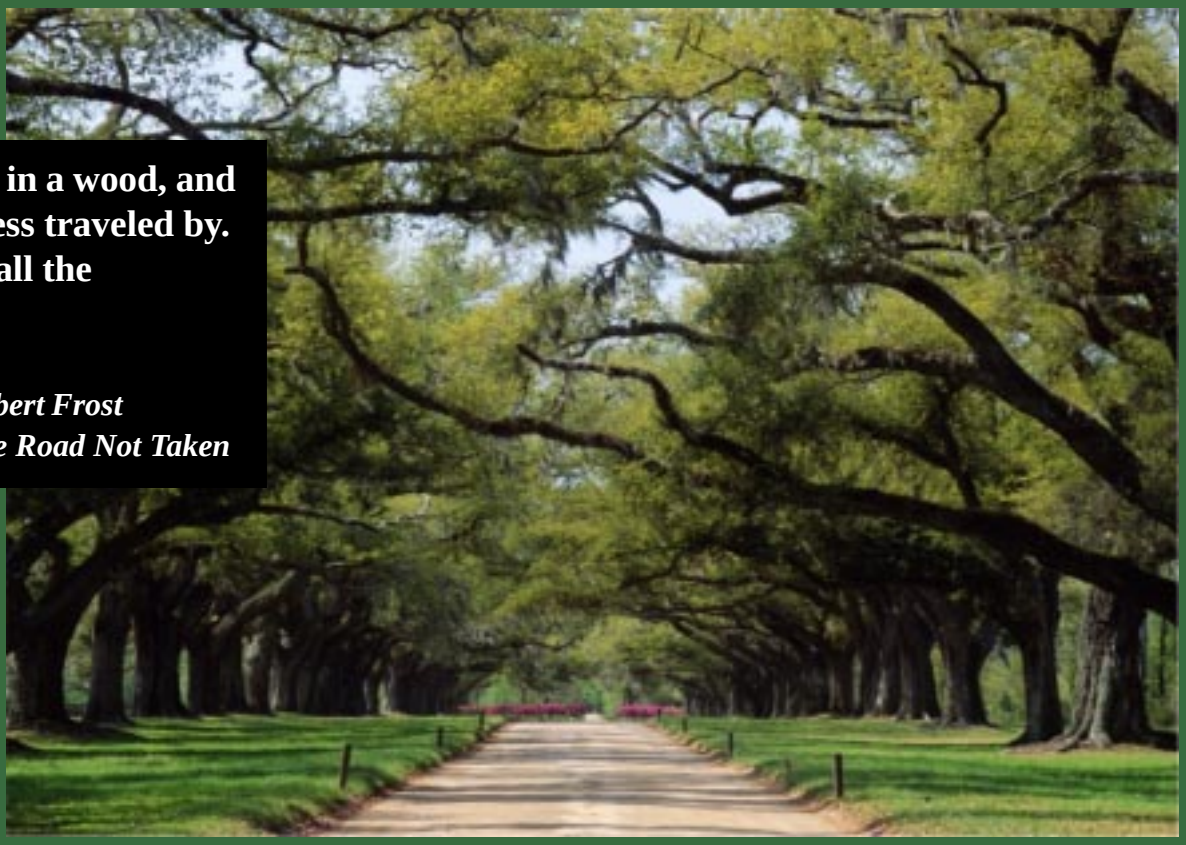
Attachment-A

Micro Bac International Brochure

Leading The New Direction



Micro-Bac International Inc.®



"Two roads diverged in a wood, and I -- I took the one less traveled by. And that has made all the difference."

Robert Frost
The Road Not Taken

When Micro-Bac International Inc.® set out in 1979 to improve the environment and enhance the accessibility to the world's energy reserves, they had little more than a conscience and a vision. Today, Micro-Bac continues to create global solutions for our resource-strapped planet. From the critical issues with water and wastewater, to the detrimental effects of environmental contamination, to the challenges of efficient petroleum production, Micro-Bac is committed to solving these problems with innovative and environmentally friendly solutions. And that first step has made all the difference.



A Commitment to Innovation

Developing solutions for a marketplace that is fast approaching $1 trillion requires not only constant innovation, but a dedication to a new way of thinking. Toward this goal, Micro-Bac R & D teams are firmly committed to new ideas such as Green Integrated Technologies. By recognizing the value of integrating other key technologies, Micro-Bac ensures that new product development creates compatible and synergistic solutions for us all.

Unlocking the Secrets of the Universe

The key to solving the world's environmental and energy problems is apparent to those researchers who truly understand the dynamics of environmental biotechnology. Micro-Bac has spent decades compiling and maintaining one of the most extensive environmental culture collections in the world. Micro-Bac scientists continue to discover new biological solutions from the myriad that can be expressed under the diverse conditions that exist in the environment.









"The secrets to environmental solutions are ours for the taking."

Biotechnologies

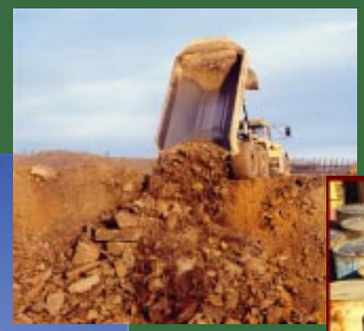
Micro-Bac technologies are successfully being used in the following markets:*

- Oil Production
- Reservoir Stimulation
- Refinery and Terminal Services
- Paraffin and Scale Control
- Fracture Damage Repair
- Tank Cleaning
- Bioremediation
- Municipal Wastewater Treatment
- Industrial Wastewater Treatment
- Grease Trap Maintenance
- Food Processing Waste
- Waste Odor Control
- Animal Waste Degradation
- Animal Health

* Box color represents product line color

SOLUTIONS WITH A CONSCIENCE

Soil & Groundwater Clean-up



Oil Production



Bioremediation



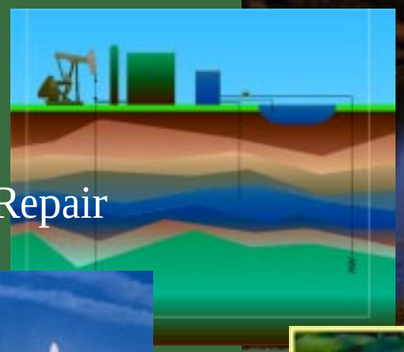
Paraffin & Scale Control



Tank Cleaning

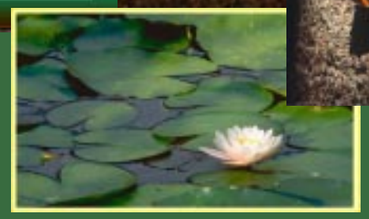


Terminal Services



Wastewater Remedies

Frac Damage Repair





Grease Digestion and Trap Maintenance



Animal Waste Solutions

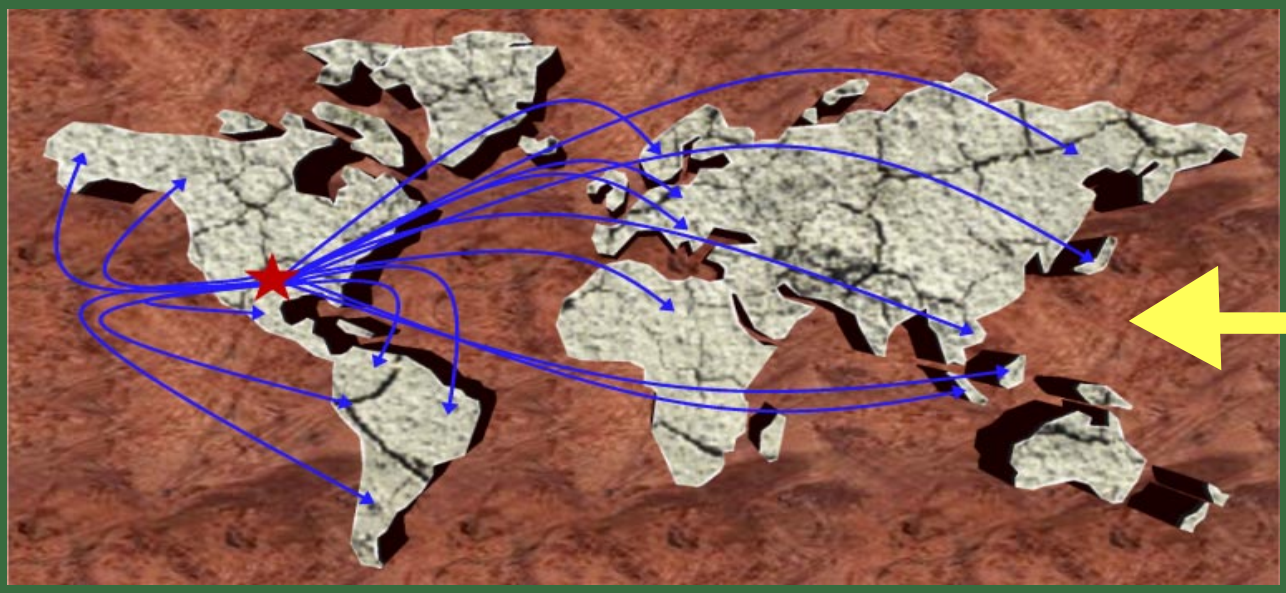


NASA Commercial Technology Spinoff



Degradation of Food Processing Wastes

A Global Marketplace



Strategically located business interests throughout the world.

Micro-Bac has developed an impressive list of partnerships around the world that share our vision. It is the mission of the business development group to seek out those individuals which exemplify the entrepreneurial spirit and those businesses that realize the value in sharing strategies, time and resources.

Information

As advances in Information Technology rapidly unfold, Micro-Bac will continue to develop its seamless business and distribution channels. Using both internet and intranet capabilities, our goal is to link partners, distributors, industries and customers with the latest information, the best products and the most innovative platform from which to build business.



A Formula for Success

Today more than ever, businesses need to build strong relationships to stay competitive.

From distributors to joint ventures to strategic alliances, Micro-Bac continues to refine its business models for success. All Micro-Bac distributors are trained both technically and in business development to ensure long term success and greater market growth.

A Full Service Company



Analytical Testing and Services

Project Management

Business Development

Oil Field Engineering Services

Product Application Support

Marketing Support

Leading the New Direction

The new direction requires us to address the Earth's diminishing resources with a dynamic consortium of science, business and vision. Micro-Bac is at the forefront of this revolution by combining biological innovation and information technology to develop safe, alternative solutions for cleaning up the environment.

The new direction has just begun and Micro-Bac is leading the way.



Micro-Bac International Inc.
3200 N. I H 35 • Round Rock, TX 78681
Ph: 512-310-9000 • Fax: 512-310-8800
Email: mail@micro-bac.com • Web: www.micro-bac.com

Attachment-B

Lab Case Study

M-1000H★
Biodegradation Lab Study

SUBSTANCE: PCE Contaminated water

TESTING: Groundwater samples were obtained to perform a four week biodegradation study. The study was conducted such that each sampling point had a sterilized sample, a control sample, and test samples with bacteria only, with nutrients only, and with bacteria and nutrients.

The samples were incubated at 35°C in containers sealed to minimize any volatilization. Samples were analyzed by U.S. EPA method 8260 for volatile organics.

RESULTS: The following compounds were found in the samples: 2-butanone, acetone, benzene, ethylbenzene, m,p xylenes, methylene chloride, 0-xylenes, t1,2 dichloroethene, tetrachloroethene (PCE), toluene, and trichloroethene. No vinyl chloride was detected in any sample prior to treatment.

The principal contaminants found were 2-butanone (methyl ethyl ketone) at 12,000 micrograms per liter and tetrachlorethene (PCE) at 49,000 micrograms per liter. All other contaminants were in the in the 200 micrograms per liter or less range. Volatilization appeared to be minimal as levels of 2-butanone and methylene chloride in the control were maintained throughout the incubation period.

Good evidence for biodegradation of the various pollutants was found. The M-1000H★ treated samples consistently produced levels lower than the uninoculated controls. The level of biodegradation ranged from >99% for 2-butanone to >97% for tetrachloroethene by week four. (see table below)

		2-butanone	tetrachloroethene
	Starting concentrations	12,000	49,000
Day 7	Control	10,000	17,000
	Bacteria	<100	4,000
Day 14	Control	14,000	270
	Bacteria	<100	11
Day 21	Control	9,200	27
	Bacteria	<100	<5
Day 28	Control	12,000	210
	Bacteria	<100	<5

Methylene chloride exhibited the least amount of biodegradation (19% decline), and by week four was the only pollutant out of eleven still detected in the samples. Vinyl chloride was not detected in any of the samples following treatment.

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Attachment-C

M-1000H Product Information

MICRO-BAC INTERNATIONAL, INC.
3200 N. IH-35
ROUND ROCK, TX 78681-2410
(512) 310-9000 fax (512)310-8800

MATERIAL SAFETY DATA SHEET

Section 1. Identification	Tri-Phasic Nutrient #12
Section 2. Identity Information / Composition	
Hazardous Components	Phosphates, Potassium, Nitrogen Compounds
Common Name	Same
Section 3. Physical Characteristics	
Specific Gravity	1.35-1.50
Boiling Point	>100°C
Melting Point	N/A
Solubility in Water	Water Soluble
Appearance and Odor	Rich chocolate brown liquid when suspended. Product settled in three phases after prolonged storage, faint chemical odor
Section 4. Fire and Explosion Hazard	
Flash Point	N/A
Flammable Limits	N/A
Extinguishing Media	N/A
Special Fire Fighting Procedures	N/A
Unusual Fire and Explosion Hazards	None
Section 5. Reactivity Data	
Stability	Stable
Incompatibility	Reducing Agents
Conditions to Avoid	Extreme Conditions
Hazardous Decomposition or Byproducts	Nitrogen Oxides
Section 6. Health Hazard Data	
Routes of Entry	Skin, Ingestion
Carcinogenicity (NTP or IARC)	N/A
Signs and Symptoms of Exposure	If ingested, may cause gastric and/or intestinal upset, may cause irritation to skin or eyes
Emergency and First Aid Procedures	**Eye or Skin Contact**: Flush with clean water for 10 minutes **Ingestion:** Do not induce vomiting; Drink plenty of water
Section 7. Precautions for Safe Handling	
Material Release or Spillage	Clean with soap and water
Waste Disposal Method	Flush with clean water
Handling and Storage Precautions	Avoid contact with skin and eyes
Ventilation Required	None
Respiratory Protection	None
Personal Protection	Rubber, plastic, or cloth gloves
Work/Hygienic Practices	Routine

MICRO-BAC INTERNATIONAL, INC.
3200 N. IH-35
ROUND ROCK, TX 78681-2410
(512) 310-9000 FAX (512)310-8800

MATERIAL SAFETY DATA SHEET

Section 1. Identification — **M-1000H★**

Section 2. Identity Information / Composition

Hazardous Components	None (TSCA 40 CFR 710.4 b)
Common Name	Microbial Product: Naturally occurring microorganisms

Section 3. Physical Characteristics

Specific Gravity	1.02
Boiling Point	100°C
Melting Point	0°C
Solubility in Water	Water Soluble
Appearance and Odor	Light Pink to Tan Liquid with Moderate Odor

Section 4. Fire and Explosion Hazard

Flash Point	NA
Flammable Limits	NA
Extinguishing Media	NA
Special Fire Fighting Procedures	None
Unusual Fire and Explosion Hazards	None

Section 5. Reactivity Data

Stability	Stable
Incompatibility	None
Conditions to Avoid	Extreme Conditions
Hazardous Decomposition or Byproducts	None

Section 6. Health Hazard Data

Routes of Entry	Ingestion; Eye Contact
Carcinogenicity (NTP or IARC)	None
Signs and Symptoms of Exposure	May Cause Gastric and/or Intestinal Upset
Emergency and First Aid Procedures	Ingestion: Do Not Induce Vomiting; Drink plenty of water Eye Contact: Flush with clean water for 10 minutes

Section 7. Precautions for Safe Handling

Material Release or Spillage	Clean up with soap and water or with disinfectant
Waste Disposal Method	Flush with clean water
Handling and Storage Precautions	Store in tightly closed original container at temperatures between 13°C to 32°C
Ventilation Required	None Required
Respiratory Protection	None Required
Personal Protection	None required; use of gloves and safety glasses suggested
Work/Hygienic Practices	Routine

1-17-94

Attachment-D

Tri-Phasic-12(Nutrients)

TECHNICAL DATA
TRI-PHASIC 12™

BIOLOGICAL STIMULANT

General Description

TRI-PHASIC 12™ is a proprietary blend of vitamins, minerals and organic nutrients used to stimulate microbial activity in product applications for Micro-Bac® International biological products where indigenous nutrients are insufficient to support optimal microbial activity. Such applications include bioremediation of contaminated soil and water. TRI-PHASIC 12™ may also be used in the remediation of oligotrophic waters and tank bottoms. By lowering the carbon/nitrogen ratio, biodegradation is enhanced and mineralization is promoted. TRI-PHASIC 12™ is supplied in an easy-to-use colloidal liquid form that flows and is freely soluble in water. It is designed to be used in conjunction with Micro-Bac® International products such as M-1000H*™.

Product Formula

TRI-PHASIC 12™ contains all the major and minor mineral compounds as well as defined vitamins required by the specific microorganisms used in the Micro-Bac® International products. It supports maximal activity of these microorganisms under both aerobic and anaerobic conditions.

Product Specifications

Color	dark brown layers
pH concentrated	circa 4.3
pH diluted	5-8
Shipping Size	1 gal. container
Odor	mild
Wt. per Gal.	11.4 lbs.
Specific Gravity	1.30
Solubility	> 2 gal. per 100 gal water

Dosage

Micro-Bac's® technical staff can provide dosage recommendations based on the design and volumes of the project site. For example one container of Tri-Phasic 12™ nutrient is generally used with 50 cubic yards of contaminated soil.

Application

TRI-PHASIC 12™ is generally dissolved in water at a rate of 1 container per 55 gallons. It is generally applied after and occasionally simultaneous with microbial treatments.

Handling & Safety

TRI-PHASIC 12™ is a natural product that meets EPA requirements for release into the environment. Special clothing and equipment are not required for handling TRI-PHASIC 12™ although contact by inhalation or touch should be avoided and the use of gloves and a dust respirator is suggested. Routine hygiene should be observed.

Shipping

TRI-PHASIC 12™ is shipped in a one gallon container, F.O.B. Round Rock, Texas

Service

When the use of TRI-PHASIC 12™ is indicated through testing, Micro-Bac® provides technical support and treatment design to its customers.

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(512)310-9000 • Toll-Free (877)559-1800 • Fax (512)310-8800 • E-mail: mail@micro-bac.com • Website: http://www.micro-bac.com

Attachment-E

M-1000H MS/MSD

TECHNICAL DATA

M-1000H*™

FOR THE DEGRADATION OF CONTAMINATED SUBSTANCES

General Description

M-1000H*™ is a biological product designed and formulated for the broad spectrum degradation of a variety of compounds found in contaminated and/or hazardous wastes. These compounds include simple aromatics such as benzene, toluene, ethyl benzene, and xylenes, as well as more complex aromatics like naphthalene, chlorinated compounds, and benzo-a-pyrene. Alkane mixtures, including transport fluids (gasoline and diesel), are also degraded. A variety of chlorinated aliphatic and aromatic compounds such as trichlorethene and chlorinated benzenes and biphenyls (PCBs) are also degraded. This product has been used successfully in a variety of *in-situ* and *ex-situ* applications.

M-1000H*™ consists of live, specially selected, biologicals and biochemicals, along with a supply of balanced nutrients in a ready-to-use liquid medium.

These microorganisms thrive in a variety of site conditions with diverse soils and various water chemistries. They are capable of using many of the listed hazardous waste chemicals as a carbon source.

Specifications

Color	light pink to tan
pH	6.5-8
Weight per gallon	8.51 lbs.
Specific gravity	1.02
Freeze point	32°F
Viscosity	1.30 cps - 60°F
Odor	mild organic

Application

M-1000H*™ can be applied to contaminated or hazardous substances in numerous ways. For soil applications, the product can be sprayed, or the soil can be reduced to a slurry and circulated with added product. For vadose zone or groundwater applications, the product can be applied into wells or infiltration galleries. Often the addition of specially-formulated nutrients can be used to augment the activity of the product in conditions where macro nutrients such as carbon, nitrogen, or phosphate are limited. A good monitoring program is critical to the success of any bioremediation project.

Handling and Safety

M-1000H*™ is a natural, non-pathogenic, non-engineered biological product that meets EPA requirements for release into the environment. Special clothing or equipment are not required for handling M-1000H*™. Routine hygiene should be observed.

Shipping

M-1000H*™ is shipped in two sizes: in 5-gallon containers (45 lbs), and in 55-gallon containers (495 lbs), F.O.B. Round Rock, Texas.

Service

When the use of M-1000H*™ is indicated, Micro-Bac International provides technical support services to its customers.

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(512)310-9000 • Toll-Free (877)559-1800 • Fax (512)310-8800 • E-mail: mail@micro-bac.com • Website: www.micro-bac.com

Attachment-F
CAMP

BROWNFIELD C L E A N U P P R O G R A M

COMMUNITY AIR MONITORING PLAN

FOR

HNJ REALTY LLC.

FOR
FORMER SCHMUKLERS CLEANERS
358 - 364 North Avenue, New Rochelle, New York
Site No.: C360088
Index No.: A3-0542-0306

PREPARED BY

BEI Consulting LLC
29 Beachfern Road
Center Moriches, NY

August 2026

TABLE OF CONTENTS

1.0 INTRODUCTION. . . . . Page 01
1.1 Regulatory Requirements.. . . . . Page 04

2.0 AIR MONITORING. . . . . Page 02
2.1 Meteorological Data.. . . . . Page 02
2.2 Community Air Monitoring Requirements. . . . . Page 02

3.0 VOC MONITORING, RESPONSE LEVELS, AND ACTIONS. . . . . Page 02
3.1 Potential Corrective Measures and VOC Suppression Techniques.. . . . . Page 03

4.0 PARTICULATE MONITORING. . . . . Page 04
4.1 Potential Particulate Suppression Techniques. . . . . Page 05

5.0 DATA QUALITY ASSURANCE.. . . . . Page 06
5.1 Calibration. . . . . Page 06
5.2 Operations. . . . . Page 06
5.3 Data Review.. . . . . Page 06

6.0 RECORDS AND REPORTING.. . . . . Page 06

1.0 INTRODUCTION

This Community Air Monitoring Plan (CAMP) has been prepared for work activities to be performed under the Remedial Action Work Plan at the Former Schmuklers Cleaners site located at 358-364 North Avenue in New Rochelle, New York. The CAMP provides measures for protection for the downwind community (i.e., off-site receptors including residences, businesses, and on-site workers not directly involved in the remedial work) from potential airborne contaminant releases resulting from investigative activities at the site.

Compliance with this CAMP is required during all ground intrusive activities that have the potential to generate airborne particulate matter and volatile organic compounds (VOCs). These activities include, but are not limited to; boring, remedial, soil and groundwater sampling activities. This CAMP has been prepared to ensure that the work activities do not adversely affect passers by, residents, or workers in the area immediately surrounding the Site and to preclude or minimize airborne migration of work-related contaminants to off-site areas.

1.1 Regulatory Requirements

This CAMP was established in accordance with the following requirements:

- New York State Department of Health's (NYSDOH) Generic Community Air Monitoring Plan as presented in DER-10 Technical Guidance for Site Investigation and Remediation (NYSDEC May 3, 2010). This guidance specifies that a community air-monitoring program shall be implemented to protect the surrounding community and to confirm that the work does not spread contamination off-site through the air;

- New York State Department of Environmental Conservation Fugitive Dust Suppression and Particulate Monitoring Program at Inactive Hazardous Waste Sites: This guidance provides a basis for developing and implementing a fugitive dust suppression and particulate monitoring program as an element of a hazardous waste site's health and safety program.

2.0 AIR MONITORING

Chlorinated and Petroleum related volatile organic compounds (VOCs) are the constituents of concern at the Site. The appropriate method to monitor air for these constituents during the investigation activities is through real-time VOC and air particulate (dust) monitoring. Since work areas will be within 20 feet of potentially exposed populations or occupied structures, the continuous monitoring locations for VOCs and particulates must reflect the nearest potentially exposed individuals and the location of ventilation system intakes for nearby structures. In order to rule out any potential exposures to nearby receptors, the immediate work area will be isolated and monitoring points will be established outside of the work area where the potential for exposure exists.

2.1 Meteorological Data

At a minimum, wind direction will be evaluated at the start of each workday, noon of each workday, and the end of each workday. These readings will be utilized to position the monitoring equipment in appropriate upwind and downwind locations.

2.2 Community Air Monitoring Requirements

To establish ambient air background concentrations, air will be monitored at several locations around the site perimeter before activities begin. These points will be monitored periodically in series during the site work. In this case, the work area is within 20 feet of potentially exposed populations and occupied structures, so perimeter monitoring points will be located to represent the nearest potentially exposed individuals within the building. One upwind and two downwind air monitoring stations will be deployed on-site for all ground intrusive activities at the site.

Fugitive respirable dust will be monitored using a MiniRae Model PDM-3 aerosol monitor (or equivalent). Air will be monitored for VOCs with a portable Mini Rae photoionization detector (PID), or equivalent. All air monitoring data will be documented in a site log book by the designated site safety officer. The site safety officer or delegate must ensure that air monitoring instruments are calibrated and maintained in accordance with manufacturer's specifications. All instruments will be zeroed daily and checked for accuracy. A daily log will be kept. If additional monitoring is required, the protocols will be developed and appended to this plan.

3.0 VOC MONITORING, RESPONSE LEVELS, AND ACTIONS

Volatile organic compounds (VOCs) will be monitored at the downwind perimeter of the immediate work area (i.e., the exclusion zone) on a continuous basis or as otherwise specified. Upwind concentrations should be measured at the start of each workday and periodically thereafter to establish background conditions. The monitoring work should be performed using equipment

appropriate to measure the types of contaminants known or suspected to be present.

The equipment should be calibrated at least daily for the contaminant(s) of concern or for an appropriate surrogate. The equipment should be capable of calculating 15-minute running average concentrations, which will be compared to the levels specified below:

- If the ambient air concentration of total organic vapors at the downwind perimeter of the work area or exclusion zone exceeds 5 parts per million (ppm) above background for the 15-minute average, work activities must be temporarily halted and monitoring continued. If the total organic vapor level readily decreases (per instantaneous readings) below 5 ppm over background, work activities can resume with continued monitoring.

- If total organic vapor levels at the downwind perimeter of the work area or exclusion zone persist at levels in excess of 5 ppm over background but less than 25 ppm, work activities must be halted, the source of vapors identified, corrective actions taken to abate emissions, and monitoring continued. After these steps, work activities can resume provided that the total organic vapor level 200 feet downwind of the exclusion zone or half the distance to the nearest potential receptor or residential/commercial structure, whichever is less - but in no case less than 20 feet, is below 5 ppm over background for the 15-minute average.

- If the organic vapor level is above 25 ppm at the perimeter of the work area, activities must be shutdown. All 15-minute readings must be recorded and be available for State (DEC and DOH) personnel to review. Instantaneous readings, if any, used for decision purposes should also be recorded.

All readings will be recorded and made available for NYSDEC and NYSDOH personnel to review. If an exceedance of the Action Limits occurs, an Action Limit Report will be completed.

3.1 Potential Corrective Measures and VOC Suppression Techniques

If the 15-minute integrated VOC level at the downwind location persists at a concentration that exceeds the upwind level by more than 5 ppm but less than 25 ppm during remediation activities, then vapor suppression techniques will be employed. The following techniques, or others, may be employed to mitigate the generation and migration of fugitive organic vapors:

- collection of purge water in covered containers ;
- storage of excess samples and soils in drums or covering with plastic;

4.0 PARTICULATE MONITORING

Air monitoring for particulates (i.e., dust) will be performed continuously during boring activities using both air monitoring equipment and visual observation at upwind and downwind locations. Monitoring equipment capable of measuring particulate matter smaller than 10 microns (PM_{10}) and capable of integrating (averaging) over periods of 15 minutes or less will be set up at upwind (i.e., background) and downwind locations, at heights approximately four to five feet above land surface (i.e., the breathing zone). Monitoring equipment will be MIE Data Ram monitors, or equivalent. The audible alarm on the particulate monitoring device will be set at 90 micrograms per cubic meter ($\mu g/m^3$). This setting will allow proactive evaluation of worksite conditions prior to reaching the action level of 100 $\mu g/m^3$ above background. The monitors will be calibrated at least once per day prior to work activities and recalibrated as needed thereafter. In addition, fugitive dust migration will be visually assessed during all intrusive work activities.

The following summarizes particulate action levels and the appropriate responses:

- If the downwind PM-10 particulate level is 100 $\mu g/m^3$ greater than background (upwind perimeter) for the 15-minute period, or if airborne dust is observed leaving the work area, then dust suppression techniques must be employed. Work may continue with dust suppression techniques provided that downwind PM-10 particulate levels do not exceed 150 $\mu g/m^3$ above the upwind level and provided that no visible dust is migrating from the work area.

- If, after implementation of dust suppression techniques, downwind PM-10 particulate levels are greater than 150 $\mu g/m^3$ above the upwind level, work must be stopped and an evaluation of activities initiated. Work can resume provided that dust suppression measures (as described in Section 2.3.1 below) and other controls are successful in reducing the downwind PM-10 particulate concentration to within 150 $\mu g/m^3$ of the upwind level and in preventing visible dust migration.

All readings will be recorded and be available for NYSDEC and NYSDOH personnel to review. If an exceedance of the Action Limits occurs, an Action Limit Report will be completed.

4.1 Potential Particulate Suppression Techniques

If the integrated particulate level at the downwind location exceeds the upwind level by more than 100 μg/m3 at any time during boring activities, then dust suppression techniques will be employed. The following techniques, or others, may be employed to mitigate the generation and migration of fugitive dusts:

- Placement of soils in drums or covering stockpiles with plastic;
- Misting of the boring area with a fine water spray from a hand-held spray bottle

Work may continue with dust suppression techniques provided that downwind PM_{10} levels are not more than 150 μg/m3 greater than the upwind levels.

There may also be situations where the dust is generated by boring activities and migrates to downwind locations, but is not detected by the monitoring equipment at or above the action level. Therefore, if dust is observed leaving the working area, dust suppression techniques such as those listed above will be employed.

If dust suppression techniques do not lower particulates to below 150 μg/m3, or visible dust persists, work will be suspended until appropriate corrective measures are identified and implemented to remedy the situation.

All air monitoring readings will be recorded in the field logbook and will be available for the NYSDEC and NYSDOH personnel to review.

5.0 DATA QUALITY ASSURANCE

5.1 Calibration

Instrument calibration shall be documented on instrument calibration and maintenance sheets or in the designated field logbook. All instruments shall be calibrated as required by the manufacturer. Calibration checks may be used during the day to confirm instrument accuracy. Duplicate readings may be taken to confirm individual instrument response.

5.2 Operations

All instruments shall be operated in accordance with the manufacturer's specifications. Manufacturers' literature, including an operations manual for each piece of monitoring equipment will be maintained on-site by the SSO for reference.

5.3 Data Review

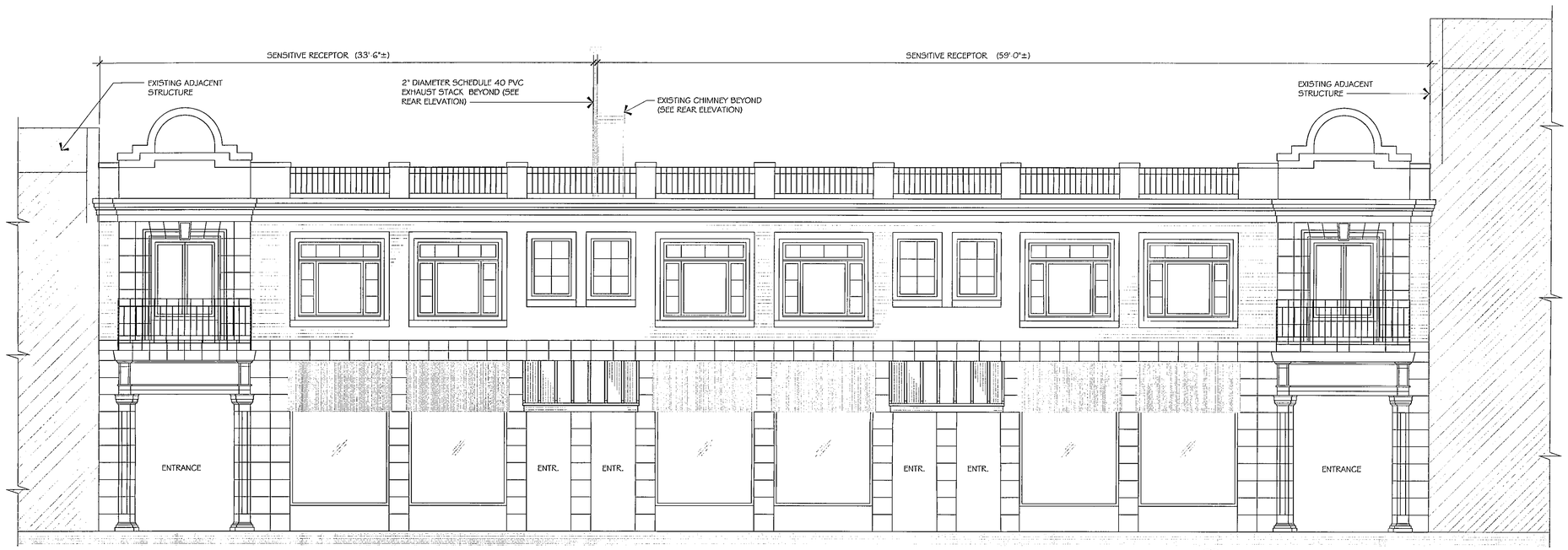
The SSO will interpret all monitoring data based upon the established criteria and his/her professional judgment. The SSO shall review the data with the PM to evaluate the potential for worker exposure, upgrades/downgrades in level of protection, comparison to direct reading instrumentation and changes in the integrated monitoring strategy.

Monitoring and sampling data, along with all sample documentation will be periodically reviewed by the PM.

6.0 RECORDS AND REPORTING

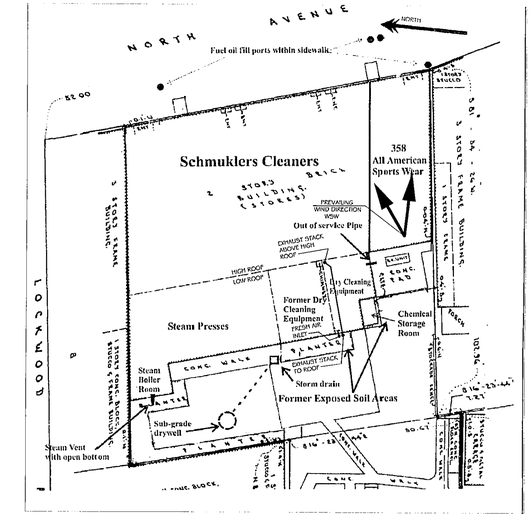
All air readings must be recorded on daily air monitoring log sheets and made available for review by personnel from NYSDEC and NYSDOH.

Attachment-G
P.E As-built SVE/SSDS
Drawings



FRONT ELEVATION - FACING NORTH AVENUE (LOOKING WEST)

SCALE: 1/4" = 1'-0"



SCHEMATIC FLOOR PLAN AND SITE FEATURES

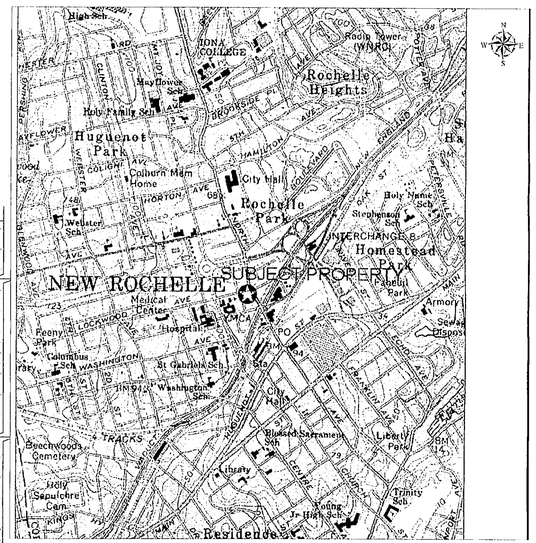
NO SCALE

INFORMATION SHOWN ON THIS PLAN IS TAKEN FROM 'BERNINGER ENVIRONMENTAL INC.' AIR PERMIT APPLICATION FIGURE 2b.



REAR ELEVATION - LOOKING EAST

SCALE: 1/4" = 1'-0"



LOCATION PLAN

NO SCALE

DRAWING ALTERATION

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Issued for Approval

Berninger Environmental Inc.
90 Knickerbocker Avenue
Bohemia, New York 11716

Michael W. Mckeown, P.E.
6 Oak Ridge Court
Manorville, New York, 11949

STATE OF NEW YORK · MICHAEL W. McKEOWN · LICENSED PROFESSIONAL ENGINEER · No. 062149
MICHAEL W. McKEOWN, P.E.

NO.	DATE	REVISION	BY
–	6-12-09	ISSUED FOR DEPARTMENT OF HEALTH APPROVAL	W.L.

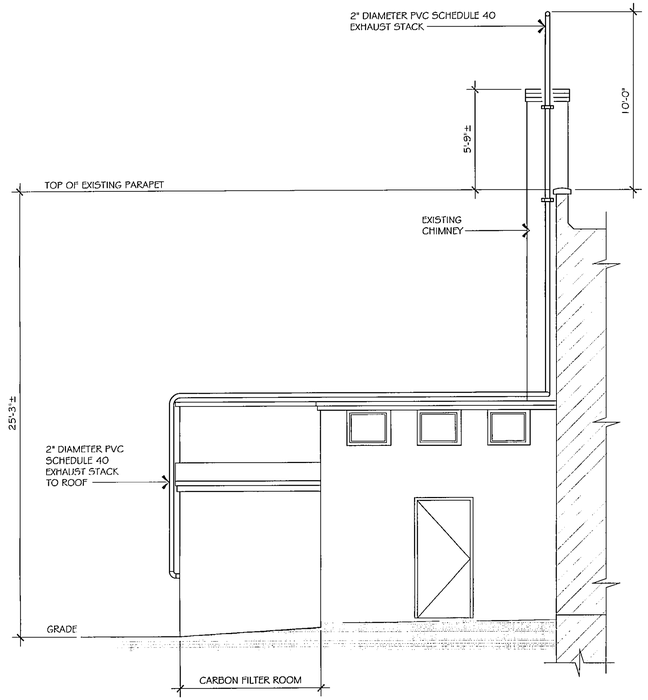
Air Permit Application for :
Schmuklers Cleaners
358-364 North Ave., New Rochelle, NY
Site # C360088

PROJECT NO.	052009
SCALE	AS NOTED
DATE	6-12-09
DRAWN BY	W.L.
CHECKED BY	M.W.M.

TITLE: Location Plan, Schematic Floor Plan and Building Elevations

DRAWING NO. AP-1

SH. 1 OF 4



PARTIAL LEFT SIDE ELEVATION - LOOKING NORTH

SCALE : ¼" = 1'-0"

Issued for Approval

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MICHAEL W. McKEOWN
LICENSED PROFESSIONAL ENGINEER
NO. 082149
MICHAEL W. McKEOWN, P.E.

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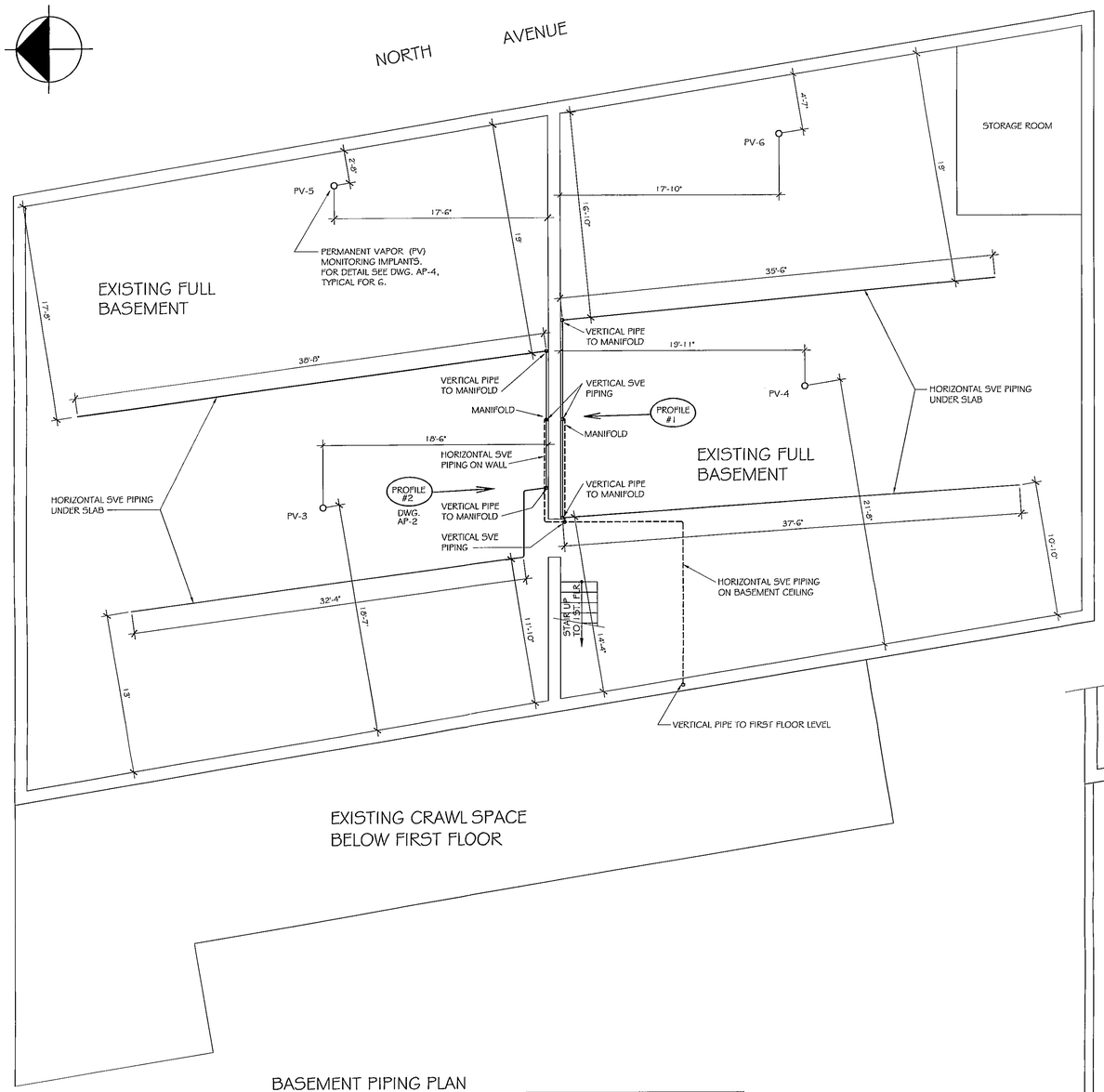
Air Permit Application for :
Schmuklers Cleaners
358-364 North Ave., New Rochelle, NY
Site # C360088

PROJECT NO. 052009
SCALE AS NOTED
DATE 6-12-09
DRAWN BY W.L.
CHECKED BY M.W.M.

TITLE
Building Elevations

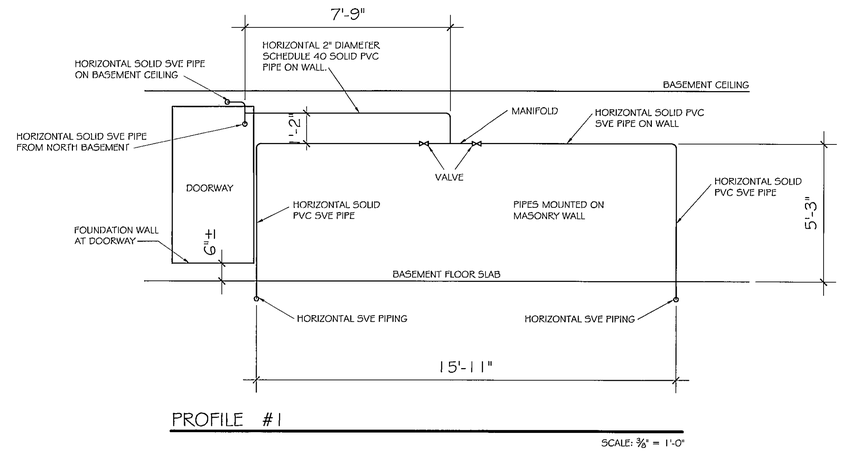
DRAWING NO.
AP-2

SH. 2 OF 4



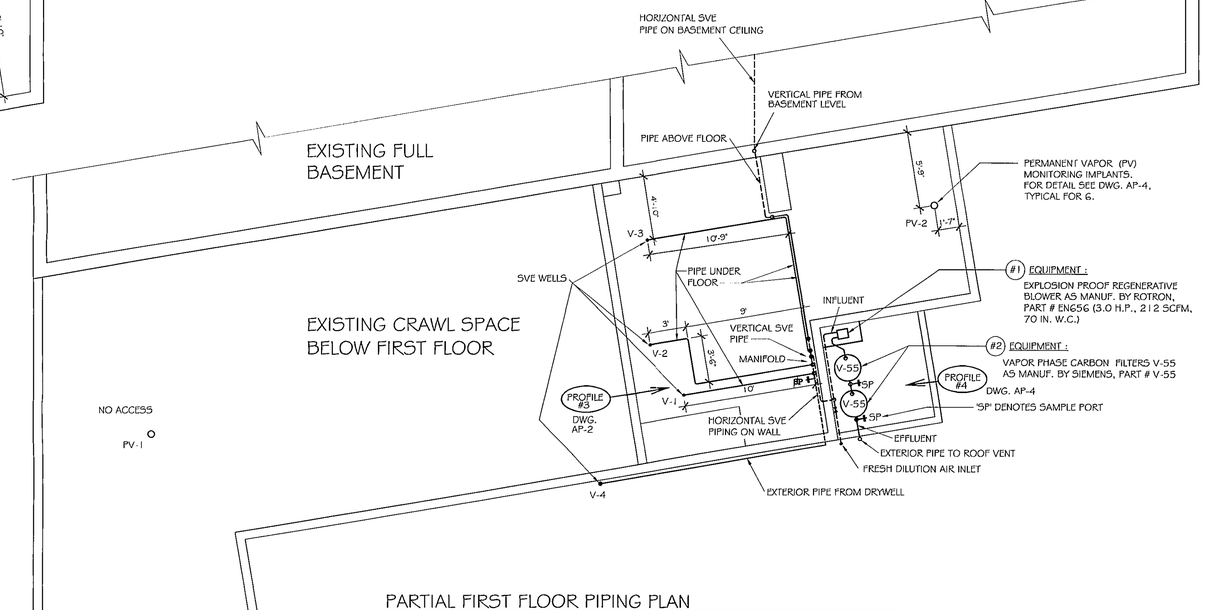
BASEMENT PIPING PLAN

SCALE: 3/16" = 1'-0"



PROFILE #1

SCALE: 3/8" = 1'-0"



PARTIAL FIRST FLOOR PIPING PLAN

SCALE: 3/16" = 1'-0"

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STATE OF NEW YORK · MICHAEL W. McKEOWN · No. 082149 · LICENSED PROFESSIONAL ENGINEER

MICHAEL W. McKEOWN, P.E.

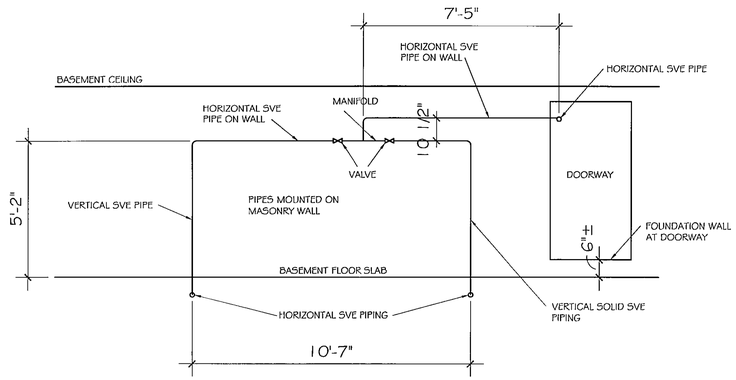
NO.	DATE	REVISION	BY
--	6.12.09	ISSUED FOR DEPARTMENT OF HEALTH APPROVAL	W.L.

Air Permit Application for :
Schmuklers Cleaners
358-364 North Ave., New Rochelle, NY
Site # C360088

PROJECT NO.	052009
SCALE	AS NOTED
DATE	6.12.09
DRAWN BY	SM
CHECKED BY	M.W.M.

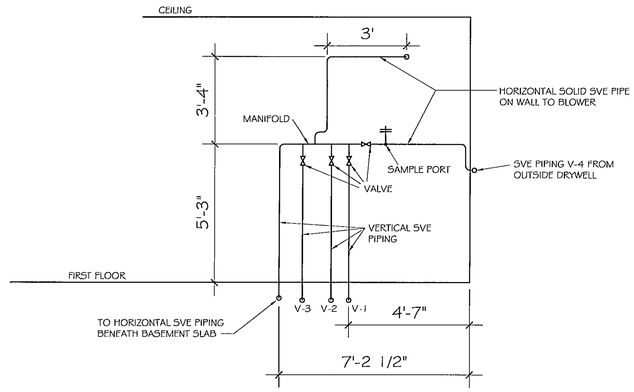
TITLE
Part Plans and Schematic Piping Elevations

DRAWING NO.
AP-3
SH. 3 OF 4



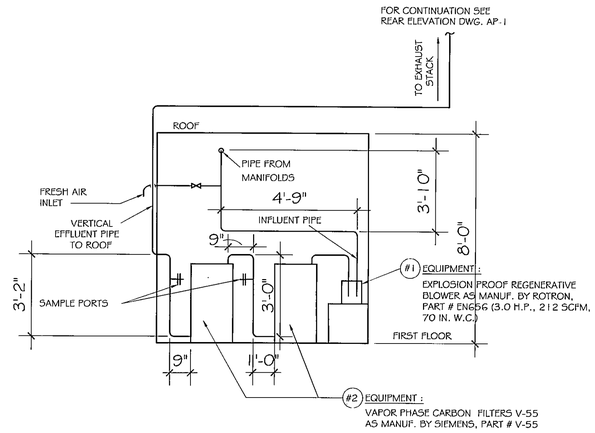
PROFILE #2

SCALE: 3/8" = 1'-0"



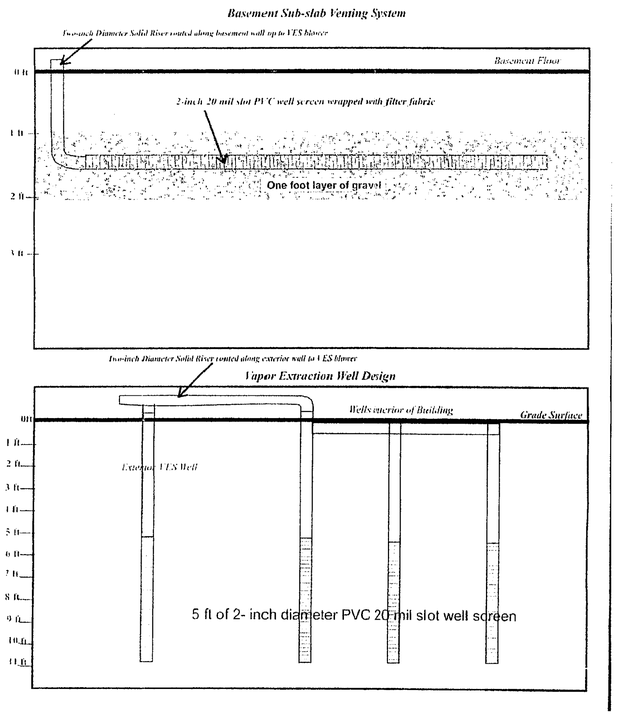
PROFILE #3

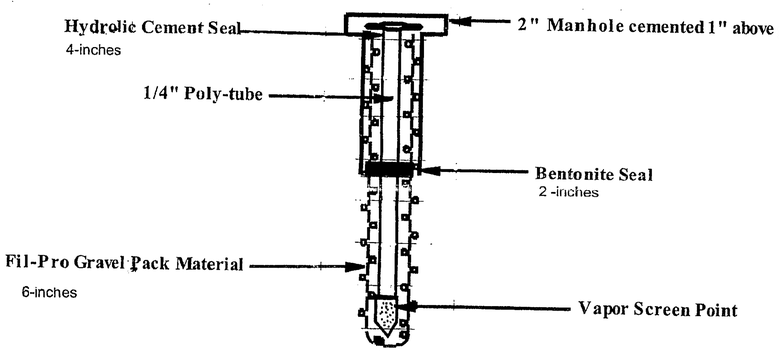
SCALE: 3/8" = 1'-0"



PROFILE #4

SCALE: 3/8" = 1'-0"





PERMANENT VAPOR (VP) MONITORING IMPLANTS

SCALE: NONE

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Manorville, New York, 11949

STATE OF NEW YORK MICHAEL W. McKEOWN LICENSED PROFESSIONAL ENGINEER No. 082148
MICHAEL W. McKEOWN, P.E.

NO.	DATE	REVISION	BY
–	6.12.09	ISSUED FOR DEPARTMENT OF HEALTH APPROVAL	W.L.

Air Permit Application for :
Schmuklers Cleaners
358-364 North Ave., New Rochelle, NY
Site # C360088

PROJECT NO.	052009
SCALE	SCALE: 3/8"= 1'-0"
DATE	6.12.09
DRAWN BY	SM
CHECKED BY	M.W.M.

TITLE: Piping Profiles

DRAWING NO.: AP-4

SH. 4 OF 4

Attachment-H
Remedial Investigation
Lab Data

L.A.B. Validation Corp. 14 West Point Drive, East Northport, N.Y. 11731 (516) 523-7891

DATA VALIDATION REPORT

ORGANIC/INORGANIC ANALYES

VOLATILES BY GC/MS
SEMIVOLATILES BY GC/MS
PESTICIDES/PCBs BY GC/ECD
TARGET ANALYTE LIST (TAL) METALS BY ICP/CV

For Groundwater and Soil Samples Collected
August 07, 2007, August 08, 2007, August 09, 2007 and August 13, 2007
From Schmuckler's Cleaners – New Rochelle, New York
By Berninger Environmental, Inc.

SAMPLE DELIVERY GROUP NUMBERS: BER042 and BER043
By H2M Labs, Inc.

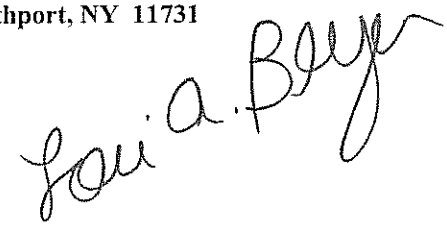
SUBMITTED TO:

Mr. Walter Berninger/President
Berninger Environmental
90 Knickerbocker Avenue
Bohemia, New York 11716

October 22, 2007

PREPARED BY:

Lori A. Beyer/President
L.A.B. Validation Corp.
14 West Point Drive
East Northport, NY 11731



LABValidation@aol.com

Schmuckler's Cleaners – August 2007 Sampling Event – Groundwater/Soil
Data Validation Report: Volatile, Semivolatile, Pesticides/PCBs, Target Analyte List (TAL) Metals

Table of Contents:

Introduction
Data Qualifier Definitions
Sample Receipt

1.0 Volatile Organics by GC/MS SW846 Method 8260
- 1.1 Holding Time
- 1.2 System Monitoring Compound (Surrogate) Recovery
- 1.3 Matrix Spikes (MS), Matrix Spike Duplicates (MSD), Matrix Spike Blank (MSB)
- 1.4 Laboratory Control Sample/Laboratory Fortified Blank
- 1.5 Blank Contamination
- 1.6 GC/MS Instrument Performance Check
- 1.7 Initial and Continuing Calibrations
- 1.8 Internal Standards
- 1.9 Target/Non Target Compound List Identification
- 1.10 Compound Quantitation and Reported Detection Limits
- 1.11 Overall System Performance

2.0 Semivolatile Organics by GC/MS SW846 Method 8270
- 2.1 Holding Time
- 2.2 Surrogate Recovery
- 2.3 Matrix Spikes (MS), Matrix Spike Duplicates (MSD), Matrix Spike Blank (MSB)
- 2.4 Laboratory Control Sample/Laboratory Fortified Blank
- 2.5 Method Blanks
- 2.6 GC/MS Instrument Performance Check
- 2.7 Initial and Continuing Calibrations
- 2.8 Internal Standards
- 2.9 Target/Non Target Compound List Identification
- 2.10 Compound Quantitation and Reported Detection Limits
- 2.11 Overall System Performance

3.0 Pesticides/PCBs by GC-ECD SW846 Methods 8081/8082
- 3.1 Holding Time
- 3.2 Surrogate Recovery
- 3.3 Matrix Spikes (MS), Matrix Spike Duplicates (MSD)
- 3.4 Laboratory Control Samples
- 3.5 Blanks
- 3.6 Calibration Verification
- 3.7 Target Compound Identification
- 3.8 Compound Quantification and Reported Detection Limits
- 3.9 Overall Assessment of Data

4.0 Target Analyte List Metals by ICP/Cold Vapor SW846 Method 6010/7471
- 4.1 Holding Times
- 4.2 Calibration (Initial and Continuing Calibration Verifications)
- 4.3 Blanks
- 4.4 Spiked Sample Recovery
- 4.5 Laboratory Duplicates
- 4.6 Laboratory Control Sample
- 4.7 Interference Check Sample
- 4.8 ICP Serial Dilution
- 4.9 Sample Results Verification
- 4.10 Overall Assessment of Data

APPENDICES:
- A. NYSDEC ASP Summary Forms
- B. Chain of Custody Documents
- C. SDG Narratives
- D. Tentatively Identified Compounds (TICs) for VOA/SVOA
- E. Data Summary Tables with Qualifications

Introduction:

A validation was performed on groundwater and soil samples and the associated quality control samples for organic/inorganic analysis for samples collected under chain of custody documentation by Berninger, Inc. and submitted to H2M Labs, Inc. for subsequent analysis. This report contains the laboratory and validation results for the field samples itemized on the following pages. The samples were collected on August 7, 2007, August 08, 2007, August 09, 2007 and August 13, 2007.

The samples were analyzed by H2M Labs, Inc. utilizing SW846 Methods and submitted under NYSDEC ASP Category B equivalent deliverable requirements for the associated analytical methodologies employed. The analytical testing consisted of the Priority Pollutant List (PPL) of analytes for Volatile Organics, the Priority Pollutant List (PPL) for Semivolatiles, Pesticides, PCBs and Target Analyte List (TAL) list of twenty-three (23) Metals.

The laboratory did not report the following analytes for Volatile analysis: Methyl Tert Butyl Ether, 2-Hexanone and Carbon Disulfide. These analytes have relevant TAGM/GW Standards and therefore, raw data was evaluated for the presence of these compounds since the laboratory has analyzed initial and continuing calibration standards. In cases where the mass spectrum meets the qualitative criteria for identification, the data validator quantitated values have been presented in the summary spreadsheets provided in Appendix E. It is recommended that the laboratory resubmit relevant Form I's for VOCs for the project file.

The laboratory did not report the following analytes for Semivolatile analysis: Aniline, Benzidine, Benzoic Acid and Pyridine. These analytes have relevant TAGM/GW Standards and therefore, raw data was evaluated for the presence of these compounds since the laboratory has analyzed initial and continuing calibration standards. In cases where the mass spectrum meets the qualitative criteria for identification, the data validator quantitated values have been presented in the summary spreadsheets provided in Appendix E. It is recommended that the laboratory resubmit relevant Form I's for SVOCs for the project file.

The data was evaluated in accordance with the National Functional Guidelines for Organic and Inorganic Data Review and in conjunction with the analytical methodologies for which the samples were analyzed, where applicable and relevant.

The data validation report pertains to the following field groundwater/soil and quality control samples:

Sample Identification	Laboratory Identification	Sample Matrix	Collection Date
GW-1	0709206-001	Groundwater	08/07/07
GW-2	0709206-002	Groundwater	08/08/07
GW-3	0709206-003	Groundwater	08/08/07
GW-4	0709206-004	Groundwater	08/08/07
GW-5	0709206-005	Groundwater	08/08/07
GW-6	0709206-006	Groundwater	08/08/07
GW-7	0709206-007	Groundwater	08/08/07
GW-8	0709206-008	Groundwater	08/07/07
Field Blank Water	0709206-009	Aqueous	08/07/07
Trip Blank 8/08/07	0709206-010	Aqueous	08/08/07
Storage Blank 8/09/07	0709206-011	Aqueous	08/09/07
GW-9	0709405-001	Groundwater	08/13/07
GW-10	0709405-002	Groundwater	08/13/07
GW-11	0709405-003	Groundwater	08/13/07
Trip Blank 08/13/07	0709405-004	Aqueous	08/13/07
B-1	0709059-001	Soil	08/06/07
FD-1	0709059-002	Soil	08/06/07
FD-2	0709059-003	Soil	08/06/07
Trip Blank 08/06/07	0709059-004	Aqueous	08/06/07
Storage Blank 08/07/07	0709059-005	Aqueous	08/06/07
B-2 (5-7')	0709204-001	Soil	08/07/07
B-3 (4-4.5')	0709204-002	Soil	08/07/07
B-4 (10-12')	0709204-003	Soil	08/07/07
B-5 (0-6')	0709204-004	Soil	08/07/07
B-6 (2.5-3')	0709204-005	Soil	08/07/07
B-7 (9-10')	0709204-006	Soil	08/07/07
B-8 (6-10')	0709204-007	Soil	08/07/07
B-9 (9-10')	0709204-008	Soil	08/08/07
B-10 (4-6')	0709204-009	Soil	08/08/07
B-11 (9.5-10.5')	0709204-010	Soil	08/08/07
Field Blank Soil	0709204-011	Aqueous	08/08/07
Trip Blank 08/07/07	0709204-012	Aqueous	08/07/07
B-12 (6-8')	0709403-001	Soil	08/13/07
B-13 (7-9')	0709403-002	Soil	08/13/07
B-14 (4-6')	0709403-003	Soil	08/13/07
B-15 (20-26')	0709403-004	Soil	08/13/07

Data Qualifier Definitions:

The following definitions provide brief explanations of the qualifiers assigned to results in the data review process.

U - The analyte was analyzed for, but was not detected above the reported sample quantitation limit.

J - The analyte was positively identified; the associated numerical value is the approximate concentration of the analyte in the sample.

UJ - The analyte was not detected above the reported sample quantitation limit. However, the reported quantitation limit is approximate and may or may not represent the actual limit of quantitation necessary to accurately and precisely measure the analyte in the sample.

R - The sample results are rejected due to deficiencies in the ability to analyze the sample and meet quality control criteria. The presence or absence of the analyte cannot be verified.

N - The analysis indicates the presence of an analyte for which there is presumptive evidence to make a "tentative identification."

NJ - The analysis indicates the presence of an analyte that has been "tentatively identified" and the associated numerical value represents its approximate quantity.

Sample Receipt:

The Chain of Custody documents indicates that the samples were received at H2M Labs, Inc. via courier upon completion of the sampling event. Sample login notes were generated. The temperature was not recorded and therefore data could not be evaluated based on temperature compliance. It is assumed that no loss of Volatiles occurred and data qualifiers were not applied. According to the laboratory correspondence provided in the data package it was noted that one (1) VOA vial each for GW-2, GW-5 and Trip Blank 08/07/07 contained headspace. It is also assumed that the laboratory performed the analysis utilizing the vial that did not contain any air bubbles. No additional problems and/or discrepancies were noted, consequently, the integrity of the samples has been assumed to be good.

The data summary sheets included in Appendix E includes all usable (qualified) and unusable (rejected) results for the samples identified above. These sheets summarize the detailed narrative section of the report. All data validation qualifications have been highlighted in bold for ease of review and verification.

NOTE:
L.A.B. Validation Corp. believes it is appropriate to note that the data validation criteria utilized for data evaluation is different than the method requirements utilized by the laboratory. Qualified data does not necessarily mean that the laboratory was non-compliant in the analysis that was performed.

1.0 Volatile Organics by GC/MS SW846 Method 8260

The following method criteria were reviewed: holding times, SMCs, MS, MSD, LCS, Blanks, Tunes, Calibrations, Internal Standards, Target Component Identification, Quantitation, Reported Quantitation Limits and Overall System Performance. The volatile results were considered to be valid and useable as noted on the data summary sheets in Appendix E and within the following text:

1.1 Holding Time

The amount of an analyte in a sample can change with time due to chemical instability, degradation, volatilization, etc. If the technical holding time is exceeded, the data may not be considered valid. Those analytes detected in the samples whose holding time has been exceeded will be qualified as estimates, "J". The non-detects (sample quantitation limits) are required to be flagged as estimated, "J", or unusable, "R", if the holding times are grossly exceeded.

All samples pertaining to these SDGs were performed within the ASP Holding Time from sample receipt. The SW846 Method 8260 requirement of 14 days for initial and reanalysis was met for all samples. No data validation qualifiers were required based upon holding time.

1.2 System Monitoring Compound (Surrogate) Recovery

All samples are spiked with surrogate compounds prior to sample analysis to evaluate overall laboratory performance and efficiency of the analytical technique. If the measure of surrogate concentrations is outside contact specification, qualifications are required to be applied to associated samples and analytes.

Surrogate recoveries (%R) were found to be within acceptable limits for surrogate compounds pertaining to these SDGs for water analysis. No data validation qualifiers were required based on surrogate recovery data for the groundwater portion of this project.

Surrogate recoveries (%R) fell outside acceptance ranges for the following soil analysis:

B-8 (6-10') – Toluene d8; 148%
B-8 (6-10') MS – Toluene d8; 157%
B-8 (6-10') MSD – Toluene d8; 158%

***no laboratory action is required since a matrix effect is confirmed.**

Additionally, due to matrix interferences, B-3 (4-4.5') and B-6 (2.5-3') also resulted in high surrogate recovery values. With subsequent dilution and lowered matrix interferences acceptable surrogate recovery values were obtained.

All detected concentrations have been qualified, "J" estimated and non-detects as "UJ" as required.

1.3 Matrix Spikes (MS)/ Matrix Spike Duplicates (MSD)/Matrix Spike Blank (MSB)

The MS/MSD data are generated to determine the long-term precision and accuracy of the analytical method in various matrices.

GW-2 was selected for aqueous MS/MSD analysis. As a result of high Trichloroethene concentrations in the original unspiked sample, the recoveries for this analyte fell outside acceptance limits. No qualifications to the data were required. Additionally, Benzene, Toluene and Chlorobenzene recovered low in the MSD, however, acceptable recoveries were observed in the MS. As a result, the RPD values fell outside limits. Based on professional judgment and complicated sample matrices, no qualifications to the data were made. Acceptable recoveries were observed for all spiked components in the MSB.

B-8 (6-10') was selected for soil MS/MSD analysis. As a result of high Trichloroethene concentrations in the original unspiked sample, the recoveries for this analyte fell outside acceptance limits. No qualifications to the data was required. Additionally, 1,-dichloroethene recovered low in both the MS and MSD and benzene recovered low in the MS. Acceptable RPD were obtained. Based on professional judgment and complicated sample matrices, no qualifications to the data were made. Acceptable recoveries were observed for all spiked components in the MSB.

1.4 Laboratory Control Sample/Laboratory Fortified Blank

The LCS data for laboratory control samples (LCS) are generated to provide information on the accuracy of the analytical method and on the laboratory performance.

Acceptable LCS was analyzed for aqueous samples. All recovery values fell within established limits.

Three recoveries fell just below acceptance limits in the soil LFB but were determined to be reasonable per the methodology and therefore no qualifications were applied.

1.5 Blank Contamination

Quality assurance (QA) blanks; i.e. method, trip and field blanks are prepared to identify any contamination which may have been introduced into the samples during sample preparation or field activity. Method blanks measure laboratory contamination. Trip blanks measure cross-contamination of samples during shipment. Field blanks measure cross-contamination of samples during field operations. Storage blanks measure cross-contamination during sample storage of the field samples and are not mandated by SW-846 Method 8260.

The following table was utilized to qualify target analyte results due to contamination. The largest value from all the associated blanks is required to be utilized:

For:	Flag Sample Result with a "U" when:	Report CRQL & Qualify "U" when:	No Qualification is Needed when:
Methylene Chloride, Acetone, Toluene & 2-Butanone	Sample Conc. Is >CRQL, but </=10x blank value	Sample Conc. is <CRQL and </=10x blank value	Sample Conc. is >CRQL and >10x blank value
Other Contaminants	Sample Conc. Is >CRQL, but </=5x blank value	Sample Conc. Is <CRQL and </=5x blank value	Sample Conc. is >CRQL and >5x blank value

Below is a summary of the compounds in the sample and the associated qualifications that have been applied:

A) **Method Blank Contamination:**

Blank concentrations were evaluated compared to associated sample values based on the above criteria.

No target analytes were detected in the method blank applicable to 08/15/07 water analysis. No qualifications to the data were made.

Tetrachloroethene was detected at 3 ug/L in the method blank associated with samples GW-2DL, GW-4DL, GW-3DL, GW-5DL, GW-6DL, GW-7, GW-8DL, GW-1DL, GW-9DL, GW-10, GW-11 and GW-1. Since all sample concentrations with the exception of GW-1 were determined to be greater than 5x the blank level, the presence of Tetrachloroethene in these samples must be considered real and the "B" qualifier has been removed.

The laboratory reported values in GW-1, GW-9, GW-10 and GW-11 were negated due to blank contamination (5.0U ug/L).

Methylene Chloride was detected at acceptable levels in the soil blanks analyzed on 08/13/07 and 08/15/07. Sample results were evaluated based on the above criteria and negated as required. Dilutions were taken into account when applying the criteria.

Tetrachloroethene was detected at 5 ug/kg in the soil method blank analyzed on 08/15/07. Sample results were evaluated based on the above criteria and negated where the sample concentration was </= 5x the blank level.

B) **Field Blank Contamination:**

Carbon Disulfide was detected at acceptable levels in the Field Blank Water. Since the laboratory reported concentrations in GW-1 and GW-5 were less than 5x the blank level, these analytes were negated based on the above criteria.

Tetrachloroethene was detected as acceptable levels in the Field Blank Soil at 1 ug/L. No additional qualifications to the data were required.

C) **Trip Blank Contamination:**
Target analytes were not detected in Trip Blank 08/06/07, Trip Blank 08/07/07 and Trip Blank 08/08/07.

Acetone was detected in the Trip Blank 08/13/07 at acceptable levels. The laboratory reported concentrations in GW-9 and GW-11 were negated due to blank contamination.

D) **Storage Blank Contamination:**
Target analytes were not detected in the Storage Blanks from 08/07/07 and 08/09/07.

1.6 GC/MS Instrument Performance Check

Tuning and performance criteria are established to ensure adequate mass resolution, proper identification of compounds and to some degree, sufficient instrument sensitivity. These criteria are not sample specific. Instrument performance is determined using standard materials. Therefore, these criteria should be met in all circumstances. The Tuning standard for volatile organics is Bromofluorobenzene (BFB).

Instrument performance was generated within acceptable limits and frequency for Bromofluorobenzene (BFB) for all analyses conducted for these SDGs.

1.7 Initial and Continuing Calibrations

Satisfactory instrument calibration is established to ensure that the instrument is capable of producing acceptable quantitative data. An initial calibration demonstrates that the instrument is capable of giving acceptable performance at the beginning of an experimental sequence.

The continuing calibration checks document that the instrument is giving satisfactory daily performance.

A) Response Factor GC/MS:
The response factor measures the instrument's response to specific chemical compounds. The response factor for all compounds must be >/= 0.05 in both initial and continuing calibrations. A value <0.05 indicates a serious detection and quantitation problem (poor sensitivity). Analytes detected in the sample will be qualified as estimated, "J". All non-detects for that compound in the corresponding samples will be rejected, "R".

All the response factors for the target analytes reported were found to be within acceptable limits (>/=0.05), for the initial and continuing calibrations.

B) Percent Relative Standard Deviation (%RSD) and Percent Difference (%D):
Percent RSD is calculated from the initial calibration and is used to indicate the stability of the specific compound response factor over increasing concentrations. Percent D compares the response factor of the continuing calibration check to the mean response factor (RRF) from the initial calibration. Percent D is a measure of the instrument's daily performance. Percent RSD must be <30% and %D must be <25%. A value outside of these limits indicates potential detection and quantitation errors. For these reasons, all positive results are flagged as estimated, "J" and non-detects are flagged "UJ". If %RSD and %D grossly exceed QC criteria, non-detect data may be qualified, "R", unusable. Additionally, in cases where the %RSD is >30% and eliminating either the high or the low point of the curve does not restore the %RSD to less than or equal to 30% then positive results are qualified, "J". In cases where removal of either the low or high point restores the linearity, then only low or high level results will be qualified, "J" in the portion of the curve where non linearity exists.

Initial Calibrations: The initial calibrations provided and the %RSD were within acceptable limits (30%) for all compounds with the following exceptions:

COMPOUND	CALIBRATION DATE/INSTRUMENT	%RSD	ACTION
Acetone	5973-HP 08/11/07	42.7	"UJ" non-detects and 'J" hits in samples B-1 FD-1, FD-2, B-2 (5-7'), B-8 (6-10'), B-5 (0- B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3 B-12 (6-8'), B-13 (7-9'), B-14 (4-6'),B-15 (2 26')
Carbon Disulfide	5973-HP 08/11/07	35.6	"UJ" non-detects and 'J" hits in samples B-1 FD-1, FD-2, B-2 (5-7'), B-8 (6-10'), B-5 (0- B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3 B-12 (6-8'), B-13 (7-9'), B-14 (4-6'),B-15 (2 26')
Isopropylbenzene	5973-HP 08/11/07	35.7	"UJ" non-detects and 'J" hits in samples B-1 FD-1, FD-2, B-2 (5-7'), B-8 (6-10'), B-5 (0- B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3 B-12 (6-8'), B-13 (7-9'), B-14 (4-6'),B-15 (2 26')
Sec-butylbenzene	5973-HP 08/11/07	38.9	"UJ" non-detects and 'J" hits in samples B-1 FD-1, FD-2, B-2 (5-7'), B-8 (6-10'), B-5 (0- B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3

			B-12 (6-8'), B-13 (7-9'), B-14 (4-6'),B-15 (2 26')
4-Isopropyltoluene	5973-HP 08/11/07	32.8	"UJ" non-detects and 'J" hits in samples B-1 FD-1, FD-2, B-2 (5-7'), B-8 (6-10'), B-5 (0- B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3 B-12 (6-8'), B-13 (7-9'), B-14 (4-6'),B-15 (2 26')
p-Diethylbenzene	5973-HP 08/11/07	42.2	"UJ" non-detects and 'J" hits in samples B-1 FD-1, FD-2, B-2 (5-7'), B-8 (6-10'), B-5 (0- B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3 B-12 (6-8'), B-13 (7-9'), B-14 (4-6'),B-15 (2 26')
Napthalene	5973-HP 08/11/07	33.8	"UJ" non-detects and 'J" hits in samples B-1 FD-1, FD-2, B-2 (5-7'), B-8 (6-10'), B-5 (0- B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3 B-12 (6-8'), B-13 (7-9'), B-14 (4-6'),B-15 (2 26')
1,2,4,5-Tetramethylbenzene	5973-HP 08/11/07	46.0	"UJ" non-detects and 'J" hits in samples B-1 FD-1, FD-2, B-2 (5-7'), B-8 (6-10'), B-5 (0- B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3 B-12 (6-8'), B-13 (7-9'), B-14 (4-6'),B-15 (2 26')
Hexachlorobutadiene	5973-HP 08/11/07	36.3	"UJ" non-detects and 'J" hits in samples B-1 FD-1, FD-2, B-2 (5-7'), B-8 (6-10'), B-5 (0- B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3 B-12 (6-8'), B-13 (7-9'), B-14 (4-6'),B-15 (2 26')
1,2,4-Trichlorobenzene	5973-HP 08/11/07	36.2	"UJ" non-detects and 'J" hits in samples B-1 FD-1, FD-2, B-2 (5-7'), B-8 (6-10'), B-5 (0- B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3 B-12 (6-8'), B-13 (7-9'), B-14 (4-6'),B-15 (2 26')

Results for the above initial calibration compounds, non-detects have been qualified, "UJ" and hits qualified "J."

Continuing Calibrations: The continuing calibrations provided and the %D was within acceptable limits (25%) for all compounds with the following exceptions:

COMPOUND	CALIBRATION DATE/INSTRUMENT	%D	AFFECTED SAMPLES
Chlorodifluoromethane	HP5971 08/15/07	28.0	Trip Blank 08/13/07, Field Blank Aqueous, Trip Blank 08/08/07, Storage Blank 08/09/07, GW-2, GW-3, GW-4, GW-5,

			GW-6, GW-8, GW-9, Trip Blank 08/06/07, Storage Blank 08/07/07, Field Blank Soil, Trip Blank 08/07/07
Dichlorodifluoromethane	HP5971 08/15/07	50.4	Trip Blank 08/13/07, Field Blank Aqueous, Trip Blank 08/08/07, Storage Blank 08/09/07, GW-2, GW-3, GW-4, GW-5, GW-6, GW-8, GW-9, Trip Blank 08/06/07, Storage Blank 08/07/07, Field Blank Soil, Trip Blank 08/07/07
Chloromethane	HP5971 08/15/07	42.4	Trip Blank 08/13/07, Field Blank Aqueous, Trip Blank 08/08/07, Storage Blank 08/09/07, GW-2, GW-3, GW-4, GW-5, GW-6, GW-8, GW-9, Trip Blank 08/06/07, Storage Blank 08/07/07, Field Blank Soil, Trip Blank 08/07/07
2-Butanone	HP5971 08/15/07	55.2	Trip Blank 08/13/07, Field Blank Aqueous, Trip Blank 08/08/07, Storage Blank 08/09/07, GW-2, GW-3, GW-4, GW-5, GW-6, GW-8, GW-9, Trip Blank 08/06/07, Storage Blank 08/07/07, Field Blank Soil, Trip Blank 08/07/07
Naphthalene	HP5971 08/15/07	28.1	Trip Blank 08/13/07, Field Blank Aqueous, Trip Blank 08/08/07, Storage Blank 08/09/07, GW-2, GW-3, GW-4, GW-5, GW-6, GW-8, GW-9, Trip Blank 08/06/07, Storage Blank 08/07/07, Field Blank Soil, Trip Blank 08/07/07
1,2,3-Trichlorobenzene	HP5971 08/15/07	27.0	Trip Blank 08/13/07, Field Blank Aqueous, Trip Blank 08/08/07, Storage Blank 08/09/07, GW-2, GW-3, GW-4, GW-5, GW-6, GW-8, GW-9, Trip Blank 08/06/07, Storage Blank 08/07/07, Field Blank Soil, Trip Blank 08/07/07
1,2,4,5-Tetramethylbenzene	HP5971 08/15/07	55.2	Trip Blank 08/13/07, Field Blank Aqueous, Trip Blank 08/08/07, Storage Blank 08/09/07, GW-2, GW-3, GW-4, GW-5, GW-6, GW-8, GW-9, Trip Blank 08/06/07, Storage Blank 08/07/07, Field Blank Soil, Trip Blank 08/07/07
Chlorodifluoromethane	HP5971 08/16/07	27.6	GW-7, GW-11, GW-10, GW-1
Dichlorodifluoromethane	HP5971 08/16/07	54.9	GW-7, GW-11, GW-10, GW-1
Chloromethane	HP5971 08/16/07	31.0	GW-7, GW-11, GW-10, GW-1
2-Butanone	HP5971 08/16/07	42.3	GW-7, GW-11, GW-10, GW-1
1,2,3-Trichloropropane	HP5971 08/16/07	28.3	GW-7, GW-11, GW-10, GW-1
p-Diethylbenzene	HP5971 08/16/07	34.9	GW-7, GW-11, GW-10, GW-1
Naphthalene	HP5971 08/16/07	27.8	GW-7, GW-11, GW-10, GW-1
1,2,3-Trichlorobenzene	HP5971 08/16/07	29.2	GW-7, GW-11, GW-10, GW-1
1,2,4,5-Tetramethylbenzene	HP5971 08/16/07	59.1	GW-7, GW-11, GW-10, GW-1
1,2-Dibromo-3-	HP5971 08/16/07	28.3	GW-7, GW-11, GW-10, GW-1

Chloropropane			
Dichlorodifluoromethane	HP5973-1 08/13/07	25.4	B-1, FD-1, FD-2, B-2 (5-7'), B-8 (6-10')
Trichlorofluoromethane	HP5973-1 08/13/07	29.3	B-1, FD-1, FD-2, B-2 (5-7'), B-8 (6-10')
Acetone	HP5973-1 08/13/07	57.4	B-1, FD-1, FD-2, B-2 (5-7'), B-8 (6-10')
2-Butanone	HP5973-1 08/13/07	50.3	B-1, FD-1, FD-2, B-2 (5-7'), B-8 (6-10')
Tetrachloroethene	HP5973-1 08/13/07	37.9	B-1, FD-1, FD-2, B-2 (5-7'), B-8 (6-10')
p-Diethylbenzene	HP5973-1 08/13/07	43.2	B-1, FD-1, FD-2, B-2 (5-7'), B-8 (6-10')
1,2,4,5-Tetramethylbenzene	HP5973-1 08/13/07	45.0	B-1, FD-1, FD-2, B-2 (5-7'), B-8 (6-10')
Chlorodifluoromethane	HP5973-1 08/15/07	26.9	B-5 (0-6'), B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (9.5-10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3'), B-12 (6-8'), B-13 (7-9'), B-14 (4-6'), B-15 (20-26')
Isopropylbenzene	HP5973-1 08/15/07	36.7	B-5 (0-6'), B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (9.5-10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3'), B-12 (6-8'), B-13 (7-9'), B-14 (4-6'), B-15 (20-26')
n-Propylbenzene	HP5973-1 08/15/07	25.1	B-5 (0-6'), B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (9.5-10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3'), B-12 (6-8'), B-13 (7-9'), B-14 (4-6'), B-15 (20-26')
Tert-Butylbenzene	HP5973-1 08/15/07	39.4	B-5 (0-6'), B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (9.5-10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3'), B-12 (6-8'), B-13 (7-9'), B-14 (4-6'), B-15 (20-26')
Sec-Butylbenzene	HP5973-1 08/15/07	37.0	B-5 (0-6'), B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (9.5-10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3'), B-12 (6-8'), B-13 (7-9'), B-14 (4-6'), B-15 (20-26')
4-Isopropyltoluene	HP5973-1 08/15/07	33.1	B-5 (0-6'), B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (9.5-10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3'), B-12 (6-8'), B-13 (7-9'), B-14 (4-6'), B-15 (20-26')
Naphthalene	HP5973-1 08/15/07	28.5	B-5 (0-6'), B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (9.5-10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3'), B-12 (6-8'), B-13 (7-9'), B-14 (4-6'), B-15 (20-26')
Hexachlorobutadiene	HP5973-1 08/15/07	53.0	B-5 (0-6'), B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (9.5-10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3'), B-12 (6-8'), B-13 (7-9'), B-14 (4-6'), B-15 (20-26')
1,2,4-Trichlorobenzene	HP5973-1 08/15/07	27.7	B-5 (0-6'), B-7 (9-10'), B-9 (9-10'), B-10 (4-6'), B-11 (9.5-10.5'), B-3 (4-4.5'), B-4 (10-12'), B-6 (2.5-3'), B-12 (6-8'), B-13 (7-9'), B-14 (4-6'), B-15 (20-26')
Chlorodifluoromethane	HP5971 08/17/07	28.3	Diluted reanalysis for B-1, B-8 (6-10'), B-6 (2.5-3'), B-13 (7-9'), B-3 (4-4.5'), B-5 (0-6')
Dichlorodifluoromethane	HP5971 08/17/07	49.3	Diluted reanalysis for B-1, B-8 (6-10'), B-6 (2.5-3'), B-13 (7-9'), B-3 (4-4.5'), B-5 (0-6')
Chloromethane	HP5971 08/17/07	53.6	Diluted reanalysis for B-1, B-8 (6-10'), B-6

			(2.5-3'), B-13 (7-9'), B-3 (4-4.5'), B-5 (0-6')
Vinyl Chloride	HP5971 08/17/07	26.4	Diluted reanalysis for B-1, B-8 (6-10'), B-6 (2.5-3'), B-13 (7-9'), B-3 (4-4.5'), B-5 (0-6')
2-Butanone	HP5971 08/17/07	42.6	Diluted reanalysis for B-1, B-8 (6-10'), B-6 (2.5-3'), B-13 (7-9'), B-3 (4-4.5'), B-5 (0-6')
1,2,4,5-Tetramethylbenzene	HP5971 08/17/07	33.4	Diluted reanalysis for B-1, B-8 (6-10'), B-6 (2.5-3'), B-13 (7-9'), B-3 (4-4.5'), B-5 (0-6')
Hexachlorobutadiene	HP5971 08/17/07	41.0	Diluted reanalysis for B-1, B-8 (6-10'), B-6 (2.5-3'), B-13 (7-9'), B-3 (4-4.5'), B-5 (0-6')

Results for the above continuing calibration compounds, non-detects have been qualified, "UJ" (ND) and hits qualified as estimated, "J."

1.8 Internal Standards

Internal Standards (IS) performance criteria ensure that the GC/MS sensitivity and response are stable during every experimental run. The internal standard area count must not vary by more than a factor of 2 (-50% to +100%) from the associated continuing calibration standard. The retention time of the internal standard must not vary more than +/- 30 seconds from the associated continuing calibration standard. If the area count is outside the (-50% to +100%) range of the associated standard, all of the positive results for compounds quantitated using That IS are qualified as estimated, "J", and all non-detects as "UJ", or "R" if there is a severe loss of sensitivity.

If an internal standard retention time varies by more than 30 seconds, professional judgment will be used to determine either partial or total rejection of the data for that sample fraction.

All internal standards met acceptance limits with the exception of 1,4-Dichlorobenzene-d4 in sample B-3 (4-4.5') which resulted in low area response. Detected analytes quantitated from this internal standard must be considered estimated, "J" biased high. With reduced matrix interferences and subsequent dilution, acceptable area responses were observed.

1.9 Target/Non Target Compound List Identification

TCL compounds are identified on the GC/MS by using the analyte's relative retention time (RRT) and by comparison to the ion spectra obtained from known standards. For the results to be a positive hit, the sample peak must be within =/- 0.06RRT units of the standard compound and have an ion spectra which has a ratio of the primary

and secondary m/e intensities within 20% of that in the standard compound.

GC/MS spectra met the qualitative criteria for identification. All retention times were within required specifications.

TICs (Tentatively Identified Compounds) were generated and are presented in Appendix D of this report. The majority of constituents consist of unknown hydrocarbons and substituted aromatic compounds (including naphthalene isomers).

1.10 Compound Quantitation and Reported Detection Limits

GC/MS quantitative analysis is considered to be acceptable. Correct internal standards per SW846, response factors and percent moisture were used to calculate final concentrations.

All samples were initially analyzed undiluted. Based on target analyte concentrations a diluted reanalysis was performed where required. Based on the raw concentrations, the results were hybridized on the spreadsheets in Appendix E to assist the end user which data must be utilized.

It was observed that these samples contained wide range of target concentrations and in some cases, the samples were diluted for Tetrachloroethene and other compounds were subsequently lost in the diluted reanalysis. In these cases, providing the peak was not supersaturated and good peak shape was observed, the value from the initial run was utilized and qualified as estimated, "J."

Results were reported to the CRQL's with "J" values to the MDL. This is common laboratory practice.

1.11 Overall System Performance

Good resolution and chromatographic performance were observed.

Several samples were analyzed at dilutions based upon high target analyte concentrations. In cases where duplicate analyses were submitted by the laboratory, the Form I's has been hybridized to summarize which analytical result should be utilized by the end user.

2.0 Semivolatile Organics by GC/MS SW846 Method 8270

The following method criteria were reviewed: holding times, Surrogates, MS, MSD, LCS, Blanks, Tunes, Calibrations, Internal Standards, Target Component Identification, Quantitation, Reported Quantitation Limits and overall system performance. The semivolatile results were considered to be valid and as noted on the data summary sheets in Appendix E and within the following text:

2.1 Holding Time

The amount of an analyte in a sample can change with time due to chemical instability, degradation, volatilization, etc. If the technical holding time is exceeded, the data may not be considered valid. Those analytes detected in the samples whose holding time has been exceeded will be qualified as estimates, "J". The non-detects (sample quantitation limits) are required to be flagged as estimated, "J", or unusable, "R", if the holding times are grossly exceeded.

All samples were extracted and analyzed within the method required holding times and the technical holding times required for data validation. No qualifications were applied based upon holding time criteria.

2.2 Surrogate Recovery

All samples are spiked with surrogate compounds prior to sample preparation/extraction to evaluate overall laboratory performance and efficiency of the analytical technique. Additionally, the sample itself may produce effects due to such factors as interferences and high concentrations of analytes. Since the effects of the sample matrix are frequently outside the control of the laboratory and may present relatively unique problems, the evaluation of the data is dependent upon reextraction and/or reanalysis to confirm/negate laboratory error or matrix related problems. Discussion of surrogate recoveries that fell outside (above/below) QC guidelines is itemized below:

Nitrobenzene-d5 recovered above acceptance limits in GW-4, GW-5, B-1 and the diluted reanalysis of B-1 due to matrix interferences. No laboratory action is required since the method allows one surrogate per fraction to fall outside ranges providing the recovery value is >10%.

Remaining surrogate recovery values fell within QC limits for these SDGs.

2.3 Matrix Spikes (MS)/Matrix Spike Duplicates (MSD)/Matrix Spike Blank (MSB)

The MS/MSD/MSB data are generated to determine the long-term precision and accuracy of the analytical method in various matrices.

MS/MSD/MSB analysis was not conducted on any samples pertaining to these SDGs. Since acceptable LFB was analyzed no qualifications to the data was made.

2.4 Laboratory Control Sample/Laboratory Fortified Blank

The LCS/LFB data for laboratory control samples (LCS/LFB) are generated to provide information on the accuracy of the analytical method and on the laboratory performance.

Acceptable LCS/LFB was analyzed for these SDGs. All analytes recovered within acceptance limits.

2.5 Method Blanks

Quality assurance (QA) blanks; i.e. method, trip and field blanks are prepared to identify any contamination which may have been introduced into the samples during sample preparation or field activity. Method blanks measure laboratory contamination. Field blanks measure cross-contamination of samples during field operations.

The following table was utilized to qualify target analyte results due to contamination. The largest value from all the associated blanks is required to be utilized:

For:	Flag Sample Result with a "U" when:	Report CRQL & Qualify "U" when:	No Qualification is Needed when:
Phthalates (common laboratory contaminants)	Sample Conc. is >CRQL, but </=10x blank value	Sample Conc. Is <CRQL and </=10x blank value	Sample Conc. is >CRQL and >10x blank value
Other Contaminants	Sample Conc. is >CRQL, but </=5x blank value	Sample Conc. Is <CRQL and </=5x blank value	Sample Conc. is >CRQL and >5x blank value

Below is a summary of the compounds in the sample and the associated qualification that have been applied:

A) **Method Blank Contamination:**

Target analytes were not detected in the methods blanks associated with groundwater or soil analysis.

B) **Field Blank Contamination:**
Field blank analysis was not required for these SDGs for Semivolatile analysis.

2.6 GC/MS Instrument Performance Check

Tuning and performance criteria are established to ensure adequate mass resolution proper identification of compounds and to some degree, sufficient instrument sensitivity. These criteria are not sample specific. Instrument performance is determined using standard materials. Therefore, these criteria should be met in all circumstances. The Tuning standard for semivolatile organics is decafluorotriphenylphosphine (DFTPP).

Instrument performance was generated within acceptable limits and frequency for decafluorotriphenylphosphine (DFTPP) for all analyses.

2.7 Initial and Continuing Calibrations

Satisfactory instrument calibration is established to ensure that the instrument is capable of producing acceptable quantitative data. An initial calibration demonstrates that the instrument is capable of giving acceptable performance at the beginning of an experimental sequence. The continuing calibration checks document that the instrument is giving satisfactory daily performance.

C) Response Factor GC/MS:
The response factor measures the instrument's response to specific chemical compounds. The response factor for all compounds must be >/= 0.05 in both initial and continuing calibrations. A value <0.05 indicates a serious detection and quantitation problem (poor sensitivity). Analytes detected in the sample will be qualified as estimated, "J". All non-detects for that compound in the corresponding samples will be rejected, "R".

All the response factors for the target analytes reported were found to be within acceptable limits (>/=0.05), for the initial (average RRF) and continuing calibrations.

D) Percent Relative Standard Deviation (%RSD) and Percent Difference (%D):
Percent RSD is calculated from the initial calibration and is used to indicate the stability of the specific compound response factor over increasing concentrations. Percent D compares the response factor of the continuing calibration check to the mean response factor (RRF) from the initial calibration. Percent D is a measure of the instrument's daily performance. Percent RSD must be <30% and %D must be <25%. A value outside of these limits indicates potential detection and quantitation errors. For these reasons, all positive results are flagged as estimated, "J" and non-detects are flagged "UJ". If %RSD and %D grossly exceed QC criteria, non-detect data may be qualified, "R", unusable. Additionally, in cases where the %RSD is >30% and eliminating either the high or the low point of the curve does not restore the %RSD to less than or equal to 30% then positive results are qualified, "J". In cases where removal of either the low or high point restores the linearity, then only low or high level results will be qualified, "J" in the portion of the curve where non linearity exists.

Initial Calibrations: The initial calibrations provided and the %RSD were within acceptable limits (30%) for all compounds with the following exceptions:

COMPOUND	CALIBRATION DATE/INSTRUMENT	%RSD	ACTION
2,4-Dinitrophenol	08/14/07 HP5973N	32.3	Non-detects, "UJ" samples GW-1, GW-2, GW-4, GW-5, GW-11, B-1, B-8 (6-10'), B-5 (0-6'), B-12 (6-8'), B-13 (7-9')
Benzo (k) Fluoranthene	08/14/07 HP5973N	33.1	Non-detects, "UJ" samples GW-1, GW-2, GW-4, GW-5, GW-11, B-1, B-8 (6-10'), B-5 (0-6'), B-12 (6-8'), B-13 (7-9')

Continuing Calibrations: The continuing calibrations provided and the %D was within acceptable limits (25%) for all compounds with the following exceptions:

COMPOUND	CALIBRATION DATE/INSTRUMENT	%RSD	ACTION
None			

2.8 Internal Standards

Internal Standards (IS) performance criteria ensure that the GC/MS sensitivity and response are stable during every experimental run. The internal standard area count must not vary by more than a factor of 2 (-50% to +100%) from the associated continuing calibration standard. The retention time of the internal standard must not vary more than +/- 30 seconds from the associated continuing calibration standard. If the area count is outside the (-50% to +100%) range of the associated standard, all of the positive results for compounds quantitated using that IS are qualified as estimated, "J", and all non-detects as "UJ", or "R" if there is a severe loss of sensitivity.

If an internal standard retention time varies by more than 30 seconds, professional judgment will be used to determine either partial or total rejection of the data for that sample fraction.

Data qualifications are summarized below:

Internal Standard Outlier	Sample Identification	Qualified Compounds
Phenanthrene-d10	B-1	Detected compounds "J" (later eluting PAHs)
Chrysene-d12	B-8 (6-10'), B-5 (0-6')	Detected compounds "J" (later eluting PAHs)
Perylene-d12	B-8 (6-10'), B-5 (0-6')	Detected compounds "J" (later eluting PAHs)

2.9 Target/Non Target Compound List Identification

TCL compounds are identified on the GC/MS by using the analyte's relative retention time (RRT) and by comparison to the ion spectra obtained from known standards. For the results to be a positive hit, the sample peak must be within =/- 0.06RRT units of the standard compound and have an ion spectra which has a ratio of the primary and secondary m/e intensities within 20% of that in the standard compound.

GC/MS Spectra meets qualitative criteria for identification for all reported analytes.

TICs are presented in Appendix D and primarily consist of hydrocarbons and substituted naphthalene isomers.

2.10 Compound Quantitation and Reported Detection Limits

GC/MS quantitative analysis is considered to be acceptable. Correct internal standards, response factors and moisture content were used to calculate final concentrations.

Sample GW-1 was analyzed undiluted and Naphthalene and 2-Methylnaphthalene was detected at concentrations exceeding the instruments' linear calibration range. A 1:10 diluted reanalysis was performed. Sample results have been hybridized on the Form I's for ease of the end user as to which sample concentration should be utilized.

Samples GW-2, GW-4 and GW-5 could not be analyzed undiluted initially. A secondary diluted reanalysis was still required and performed in order to bring target concentrations within the instruments linear calibration range. Results were hybridized on the spreadsheet tables provided in Appendix E. Appropriate dilutions were performed and raw data supports the presence of target/non-target compounds.

GW-9 was analyzed at a 1:10 dilution and the extract for GW-11 permitted analysis at full concentration.

Samples B-1, B-5 (0-6') and B-8 (6-10') were initially analyzed undiluted. A reanalysis at a secondary dilution was required to bring target analytes within the upper range of the instrumental linear calibration range. Results were hybridized on the summary spreadsheets provided in Appendix E.

Results were reported to the CRQL's with "J" values to the MDL. This is common laboratory practice.

2.11 Overall System Performance

Acceptable system performance was maintained throughout the analysis.

Phthalates were detected in the soil samples, however, due to the lack of presence in the corresponding method blanks could not be negated during the review process. The end user should proceed with caution when making decisions based on phthalate concentrations.

3.0 Pesticides/PCBs by GC-ECD SW846 Methods 8081/8082

The following method criteria were reviewed: holding times, Surrogates, MS, MSD, LCS, Blanks, Analytical Sequences, Calibrations, Target Component Identification, Quantitation, Reported Quantitation Limits and overall system performance. The Pesticide and PCB results were considered to be valid and usable as noted on the data summary sheets in Appendix E and within the following text:

3.1 Holding Time

The amount of an analyte in a sample can change with time due to chemical instability, degradation, volatilization, etc. If the technical holding time is exceeded, the data may not be considered valid. Those analytes detected in the samples whose holding time has been exceeded will be qualified as estimates, "J". The non-detects (sample quantitation limits) are required to be flagged as estimated, "J", or unusable, "R", if the holding times are grossly exceeded.

All samples were extracted and analyzed within the method required holding times and the technical holding times required for data validation. No qualifications were applied based upon holding time criteria.

3.2 Surrogate Recovery

All samples are spiked with surrogate compounds prior to sample preparation/extraction to evaluate overall laboratory performance and efficiency of the analytical technique. Additionally, the sample itself may produce effects due to such factors as interferences and high concentrations of analytes. Since the effects of the sample matrix are frequently outside the control of the laboratory and may present relatively unique problems, the evaluation of the data is dependent upon reextraction and/or reanalysis to confirm/negate laboratory error or matrix related problems. Discussion of surrogate recoveries that fell outside (above/below) QC guidelines is itemized below:

Surrogate recoveries were acceptable for all analyses conducted for these SDGs with the exception of DCB on the primary column for initial analysis of B-8 (6-10') due to interferences. No qualifications to the data were required since this surrogate recovered well on confirmatory column analysis.

3.3 Matrix Spikes (MS)/Matrix Spike Duplicates (MSD)/Matrix Spike Blank (MSB)

The MS/MSD/MSB data are generated to determine the long-term precision and accuracy of the analytical method in various matrices.

MS/MSD/MSB analysis was not conducted on any samples pertaining to these SDGs. Since acceptable LFB was analyzed no qualifications to the data was made.

3.4 Laboratory Control Sample/Laboratory Fortified Blank

The LCS/LFB data for laboratory control samples (LCS) and laboratory fortified blanks (LFB) are generated to provide information on the accuracy of the analytical method and on the laboratory performance.

Acceptable LCS/LFB was analyzed. No qualifications were applied based upon LCS data. All recoveries fell within specified ranges.

3.5 Blanks

Quality assurance (QA) blanks; i.e. method, instrument, trip and field blanks are prepared to identify any contamination which may have been introduced into the samples during sample preparation or field activity. Method blanks measure laboratory contamination. Instrument blanks measure carryover for cross contamination. Field blanks measure cross-contamination of samples during field operations.

The following table was utilized to qualify target analyte results due to contamination. The largest value from all the associated blanks is required to be utilized:

For:	Flag Sample Result with a "U" when:	Report CRQL & Qualify "U" when:	No Qualification is Needed when:
Any Contaminant	Sample Conc. is >CRQL, but </=5x blank value	Sample Conc. Is <CRQL and </=5x blank value	Sample Conc. is >CRQL and >5x blank value

Extraction and Instrument blanks were performed at the appropriate frequency.

Below is a summary of blank contamination:

A) **Method Blank Contamination:**
No target analytes were detected in the associated method blank and therefore, no qualifications are required.

B) **Field Blank Contamination:**
Field blank analysis was not required for these SDGs.

3.6 Calibration Verification

Initial calibration sequence was performed as required for individual aroclor and pesticide standards. GC resolution is acceptable. No qualifications have been applied.

3.7 Target Compound Identification

Qualitative criteria for compound identification have been established to minimize the number of false positives and false negatives. The retention times of all target analytes have been verified in the samples to that of the analyzed reference standards

Positive sample result for Endrin in GW-1 was determined to have %Difference >25% when quantitated on the two columns and has been qualified, "P" as required by the laboratory. Sample chromatograms were reviewed for the presence of interference. The following qualifications were applied where neither column shows interference:

%Difference	Qualifier
0-25%	None
26-70%	"J"
71-100%	"JN"
>100%	"R"
100-200% (interference detected)*	"JN"
>50% (Pesticide value is <CRQL) **	"U"

***When the reported %D is 100-200%, but interference is determined on either column, the results shall be qualified, "J"**

****** When the reported pesticide value is lower than the CRQL and the % D is >50%, raise the value to the CRQL and qualify "U", undetected.**

Endrin was negated in GW-1 based on these criteria and as a result the reported concentration of 0.2 ug/L by the laboratory has been negated due to suspected interference.

Beta-BHC and Endrin was also negated in B-8 (6-10') due to high percent difference.

3.8 Compound Quantification and Reported Detection Limits

TCL compounds are identified on the GC by using the analyte's relative retention time (RRT) and by comparison to the primary column and the secondary confirmation column data.

Results were reported to the CRQL's with "J" values to the MDL. This is common laboratory practice.

Sample B-8 (6-10') was analyzed at several dilutions in order to obtain 4,4-DDT and the breakdown products within the instrumental linear calibration range. Results were hybridized on the summary tables provided in Appendix E.

Pesticides and PCBs were not detected in sample B-13 (7-9'). Results are useable at the concentrations provided in Appendix E.

3.9 Overall System Performance

Acceptable system performance was maintained throughout the analysis of all samples. Good resolution and chromatographic performance were observed.

4.0 Target Analyte List (TAL) Metals by ICP/Cold Vapor SW846 Method 6010/7471

The following method criteria were reviewed: holding times, CRDL standards, calibration, blanks, MS, laboratory duplicates, LCS, interference check sample, ICP serial dilutions and sample results verification. The metals results were considered to be valid and usable with the appropriate qualifiers, as noted on the data summary forms in Appendix E as notated in the following text:

4.1 Holding Times

The amount of an analyte in a sample can change with time due to chemical instability, degradation, volatilization, etc. If the technical holding time is exceeded, the data may not be considered valid. Those analytes detected in the samples whose holding time has been exceeded will be qualified as estimates, "J". The non-detects (sample quantitation limits) are required to be

flagged as estimated, "J", or unusable, "R", if the holding times are grossly exceeded.

All samples were digested and analyzed for Total TAL Metals within the method required holding times and the technical holding times for data validation. No qualifications were applied based upon holding time criteria.

4.2 Calibration (ICV/CCV)

Satisfactory instrument calibration is established to ensure that the instruments are capable of producing acceptable quantitative data. An initial calibration demonstrated that the instruments are capable of giving acceptable performance at the beginning of an experimental sequence. The continuing calibration checks document that the instruments are giving satisfactory sequential performance and that the initial calibration is still valid.

The ICP and Mercury instruments were calibrated utilizing four-point curve in addition to blanks at the beginning of each analytical run. The calibrations had been determined to be acceptable, yielding correlation coefficients of 0.995 or greater.

For ICP analysis, satisfactory instrument performance near the Contract Required Detection Limit (CRDL) was demonstrated by analyzing a CRDL standard at the beginning and end of the analytical run. The instruments were calibrated properly by analyzing the CRDL solution at the correct levels of two times the CRDL's and analyzed at the required frequency at the beginning and end of each analytical run.

All recoveries were within acceptable limits of 90-110 % for initial calibration.

Continuing calibrations were within acceptable limits of 90-110% recovery of the true values for ICP and Mercury.

No qualifications were applied based upon ICV/CCV analysis.

4.3 Blanks

Quality assurance (QA) blanks, i.e. method, field or preparation blanks are prepared to identify any contamination that may have been introduced into the samples during sample preparation or field activity. Preparation blanks measure laboratory contamination. Field blanks measure cross-contamination of samples during field operations.

All blanks were generated within acceptable limits and detected elements were below the CRDL.
The following actions are required:

Blank Type	Blank Result	Sample Result	Action
ICB/CCB	>/= IDL but </= CRDL	Non-Detect	No Action
ICB/CCB/PBW	>/= IDL but </= CRDL	>/= IDL but </= CRDL (<5x Blank Level)	Report CRDL value with a "U"
ICB/CCB	>/= IDL but </= CRDL	>CRDL	Professional Judgment
PREP BLANK	>/= IDL but </= CRDL, or non-detect	</= IDL, or non-detect	No Action

Blank results were evaluated based upon the above criteria.

All blanks were generated within acceptable limits. Sample results >IDL but <5x the blank value have been qualified, "U" as required for Silver and Selenium in GW-1.

Low level concentrations of Beryllium, Cadmium, Silver and Thallium were negated in the soil samples due to ICB/CCB contamination.

Field blank data are not included in this data set for inorganic analyses.

4.4 Spiked Sample Recovery

The spike data are generated to determine the long terms precision and accuracy of the analytical method in various matrices.

The following qualifications are required for spike recovery outliers:

Recovery > 125%	**All detects qualify "J", biased high No action for non-detects.**
Recovery < 75%	**All detects qualify "J" biased low Non detects qualify "UJ"**
Recovery <30%	**All detects qualify "J" biased low Non detects qualify "R" unusable**

Acceptable Spike analysis was conducted on GW-1. Selenium recovered low (58.3%). Non-detects must be considered estimated, biased low, "UJ." A post digestion spike was performed as required and acceptable recoveries were observed. No additional qualifications were required based upon spike data.

B-8 (6-10') was also selected for MS analysis. Antimony recovered low (53.5%). Acceptable post spike analysis was conducted. Results have been qualified as "J/UJ" as required.

No additional qualifications were required based upon spike data.

4.5 Laboratory Duplicates

The laboratory uses duplicate sample determinations to demonstrate acceptable method precision at the time of analysis. Duplicate analyses are also performed to generate data in order to determine the long-term precision of the analytical method on various matrices.

Acceptable duplicate analysis was performed on GW-1. Mercury RPD fell outside acceptance limits. Soil duplicate analysis was conducted on B-8 (6-10'). Copper RPD was outside limits. Results must be considered estimated, "J."

4.6 Laboratory Control Sample

The laboratory Control Sample (LCS) serves as a monitor of the overall performance of each step during the analysis, including the sample preparation. Aqueous and solid Laboratory Control samples shall be analyzed for each analyte utilizing the same sample preparation, analytical methods and QA/QC procedures as employed for the samples.

The LCS was analyzed and reported for all ICP and Mercury analysis. For each batch of sample results, associated LCS recoveries were within the acceptable limits.

4.7 Interference Check Sample

The interference check sample (ICS) verifies the laboratory's interelement and background correction factors. The ICS consists of two solutions A and AB. Solution A consists of interference, and solution AB consists of the analytes mixed with interferents.

SW846 Method 6010 requires solution A and solution AB to be analyzed separately. The recoveries for the ICP interference check sample were all within the acceptable limits of 80-120%.

4.8 ICP Serial Dilution

The serial dilution of samples quantitated by ICP determines whether or not significant physical or chemical interferences exist due to sample matrix. An ICP serial dilution analysis must be performed on a sample for each group of samples with a similar matrix type and concentration, or for each Sample Delivery Group (SDG), whichever is more frequent.

Acceptable ICP serial dilution was performed at a 5-fold dilution as required by the method on GW-1. The serial dilution analysis agrees within a 10% difference of the original determination after correction for dilution for all elements with the exception of Potassium.

Detected concentrations must be considered estimated and have been qualified, "J" as required.

Acceptable serial dilution results were obtained for B-8 (6-10').

4.9 Sample Results Verification

Analyte quantitation was generated in accordance with protocols. The raw data was verified and found within the linear range of each instrument used for quantitation. Raw data supplied corresponds with reported values. Verification of the calculations yielded reported results.

4.10 Overall Assessment of Data

The data generated were of acceptable quality.

Sample results that have been determined to be less than the CRDL but greater than the IDL and qualified "B" by the laboratory have been removed.

Reviewer's Signature Lori A. Beyer **Date** 10/22/07

Appendix A
NYSDEC ASP
Summary Forms

LABValidation@aol.com

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

SAMPLE IDENTIFICATION AND ANALYTICAL REQUIREMENT SUMMARY

SDG: BER043

Analytical Requirements

Customer Sample Code	Laboratory Sample Code	GCSEMI	ME	MSSEMI	MSVOA
GW-1	0709206-001	X	X	X	X
GW-2	0709206-002			X	X
GW-3	0709206-003				X
GW-4	0709206-004			X	X
GW-5	0709206-005			X	X
GW-6	0709206-006				X
GW-7	0709206-007				X
GW-8	0709206-008				X
FIELD BLANK WATER	0709206-009				X
TRIP BLANK 8/8/2007	0709206-010				X
STORAGE BLANK 8/9/2007	0709206-011				X
GW-9	0709405-001			X	X
GW-10	0709405-002				X
GW-11	0709405-003			X	X
TRIP BLANK 8/13/07	0709405-004				X

CLP, (Non-CLP) (Please indicate year of protocol) ASP B 6/2000 CEL 9/4/07

BER043 S3

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

SAMPLE PREPARATION AND ANALYSIS SUMMARY
VOLATILE (VOA)
ANALYSES

SDG: BER043

Laboratory Samp ID	Client Sample ID	Matrix	Analytical Protocol	Date Collected	DateRecd at Lab	Date Extracted	Date Analyzed	Extraction Method	DF	Level	Aux Cleanup
0709206-001A	GW-1	Aqueous	SW8260B	07-Aug-07	09-Aug-07		16-Aug-07		1	LOW	
0709206-001ADL	GW-1DL	Aqueous	SW8260B	07-Aug-07	09-Aug-07		16-Aug-07		5	LOW	
0709206-002A	GW-2	Aqueous	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709206-002ADL	GW-2DL	Aqueous	SW8260B	08-Aug-07	09-Aug-07		16-Aug-07		100	LOW	
0709206-002AMS	GW-2MS	Aqueous	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709206-002AMSD	GW-2MSD	Aqueous	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709206-003A	GW-3	Aqueous	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709206-003ADL	GW-3DL	Aqueous	SW8260B	08-Aug-07	09-Aug-07		16-Aug-07		500	LOW	
0709206-004A	GW-4	Aqueous	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709206-004ADL	GW-4DL	Aqueous	SW8260B	08-Aug-07	09-Aug-07		16-Aug-07		5000	LOW	
0709206-005A	GW-5	Aqueous	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709206-005ADL	GW-5DL	Aqueous	SW8260B	08-Aug-07	09-Aug-07		16-Aug-07		10	LOW	
0709206-006A	GW-6	Aqueous	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709206-006ADL	GW-6DL	Aqueous	SW8260B	08-Aug-07	09-Aug-07		16-Aug-07		200	LOW	
0709206-007A	GW-7	Aqueous	SW8260B	08-Aug-07	09-Aug-07		16-Aug-07		1	LOW	
0709206-008A	GW-8	Aqueous	SW8260B	07-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709206-008ADL	GW-8DL	Aqueous	SW8260B	07-Aug-07	09-Aug-07		16-Aug-07		5	LOW	
0709206-009A	FIELD BLANK	Aqueous	SW8260B	07-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709206-010A	TRIP BLANK 8/8/2007	Aqueous	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709206-011A	STORAGE BLANK 8/9/2	Aqueous	SW8260B	09-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709405-001A	GW-9	Aqueous	SW8260B	13-Aug-07	14-Aug-07		15-Aug-07		1	LOW	
0709405-001ADL	GW-9DL	Aqueous	SW8260B	13-Aug-07	14-Aug-07		16-Aug-07		500	LOW	
0709405-002A	GW-10	Aqueous	SW8260B	13-Aug-07	14-Aug-07		16-Aug-07		1	LOW	
0709405-003A	GW-11	Aqueous	SW8260B	13-Aug-07	14-Aug-07		16-Aug-07		1	LOW	
0709405-004A	TRIP BLANK 8/13/07	Aqueous	SW8260B	13-Aug-07	14-Aug-07		15-Aug-07		1	LOW	

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

SAMPLE PREPARATION AND ANALYSIS SUMMARY
SEMIVOLATILE (BNA)
ANALYSES

SDG: BER043

Laboratory Samp ID	Client Sample ID	Matrix	Analytical Protocol	Date Collected	DateRecd at Lab	Date Extracted	Date Analyzed	Extraction Method	DF	Level	Aux Cleanup
0709206-001B	GW-1	Aqueous	ASPB 8270	07-Aug-07	09-Aug-07	13-Aug-07	15-Aug-07	SEPF	1	LOW	
0709206-001BDL	GW-1DL	Aqueous	ASPB 8270	07-Aug-07	09-Aug-07	13-Aug-07	20-Aug-07	SEPF	10	LOW	
0709206-002B	GW-2	Aqueous	ASPB 8270	08-Aug-07	09-Aug-07	13-Aug-07	20-Aug-07	SEPF	100	LOW	
0709206-004B	GW-4	Aqueous	ASPB 8270	08-Aug-07	09-Aug-07	13-Aug-07	20-Aug-07	SEPF	10	LOW	
0709206-004BDL	GW-4DL	Aqueous	ASPB 8270	08-Aug-07	09-Aug-07	13-Aug-07	20-Aug-07	SEPF	100	LOW	
0709206-005B	GW-5	Aqueous	ASPB 8270	08-Aug-07	09-Aug-07	13-Aug-07	20-Aug-07	SEPF	10	LOW	
0709206-005BDL	GW-5DL	Aqueous	ASPB 8270	08-Aug-07	09-Aug-07	13-Aug-07	20-Aug-07	SEPF	100	LOW	
0709405-001B	GW-9	Aqueous	ASPB 8270	13-Aug-07	14-Aug-07	16-Aug-07	20-Aug-07	SEPF	10	LOW	
0709405-003B	GW-11	Aqueous	ASPB 8270	13-Aug-07	14-Aug-07	16-Aug-07	20-Aug-07	SEPF	1	LOW	

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

SAMPLE PREPARATION AND ANALYSIS SUMMARY
PESTICIDE/PCB/HERB
ANALYSES

SDG: BER043

Laboratory Samp ID	Client Sample ID	Matrix	Analytical Protocol	Date Collected	DateRecd at Lab	Date Extracted	Date Analyzed	Extraction Method	DF	Level	Aux Cleanup
0709206-001B	GW-1	Aqueous	SW8081/8082	07-Aug-07	09-Aug-07	13-Aug-07	14-Aug-07	CONT	1	LOW	

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

SAMPLE PREPARATION AND ANALYSIS SUMMARY
INORGANIC ANALYSIS

SDG : BER043

Laboratory Samp ID	Client Sample ID	Matrix	Metals Requested	DateRecd at Lab	Date Analyzed
0709206-001	GW-1	WATER	AG,AL,AS,BA,BE,CA,CD,CO,CR,CU,FE,HG,K,MG,MN,NA,NI,PB,SB,SE,TL,V,ZN,	09-Aug-07	08/07
0709206-001DUP	GW-1D	WATER	AG,AL,AS,BA,BE,CA,CD,CO,CR,CU,FE,HG,K,MG,MN,NA,NI,PB,SB,SE,TL,V,ZN,	09-Aug-07	08/07
0709206-001MS	GW-1S	WATER	AG,AL,AS,BA,BE,CA,CD,CO,CR,CU,FE,HG,K,MG,MN,NA,NI,PB,SB,SE,TL,V,ZN,	09-Aug-07	08/07

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

SAMPLE IDENTIFICATION AND ANALYTICAL REQUIREMENT SUMMARY

SDG: BER042

Analytical Requirements

Customer Sample Code	Laboratory Sample Code	GCSEMI	ME	MSSEMI	MSVOA
B-1	0709059-001		X	X	X
FD-1	0709059-002				X
FD-2	0709059-003				X
TRIP BLANK 8/6/07	0709059-004				X
STORAGE BLANK 8/7/07	0709059-005				X
B-2 (5-7')	0709204-001				X
B-3 (4-4 1/2')	0709204-002				X
B-4 (10-12')	0709204-003				X
B-5 (0-6')	0709204-004			X	X
B-6 (2 1/2-3')	0709204-005				X
B-7 (9-10')	0709204-006				X
B-8 (6-10')	0709204-007	X	X	X	X
B-9 (9-10')	0709204-008				X
B-10 (4-6')	0709204-009				X
B-11 (9 1/2-10 1/2)	0709204-010				X
FIELD BLANK (SOIL)	0709204-011				X
TRIP BLANK 8/7/2007	0709204-012				X
B-12 6'-8'	0709403-001			X	X
B-13 7'-9'	0709403-002	X	X	X	X
B-14 4'-6'	0709403-003				X
B-15 20'-26'	0709403-004				X

CLP (Non-CLP) (Please indicate year of protocol) ASP B 6/2000 CEL 9/4/07

BER042 S3

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

SAMPLE PREPARATION AND ANALYSIS SUMMARY
VOLATILE (VOA)
ANALYSES

SDG: BER042

Laboratory Samp ID	Client Sample ID	Matrix	Analytical Protocol	Date Collected	DateRecd at Lab	Date Extracted	Date Analyzed	Extraction Method	DF	Level	Aux Cleanup
0709059-001A	B-1DL	Soil	SW8260B	06-Aug-07	07-Aug-07		17-Aug-07		1	MED	
0709059-001A	B-1	Soil	SW8260B	06-Aug-07	07-Aug-07		13-Aug-07		5	LOW	
0709059-002A	FD-1	Soil	SW8260B	06-Aug-07	07-Aug-07		13-Aug-07		1	LOW	
0709059-003A	FD-2	Soil	SW8260B	06-Aug-07	07-Aug-07		13-Aug-07		1	LOW	
0709059-004A	TRIP BLANK 8/6/07	Aqueous	SW8260B	06-Aug-07	07-Aug-07		15-Aug-07		1	LOW	
0709059-005A	STORAGE BLANK 8/7/0	Aqueous	SW8260B	07-Aug-07	07-Aug-07		15-Aug-07		1	LOW	
0709204-001A	B-2 (5-7')	Soil	SW8260B	07-Aug-07	09-Aug-07		13-Aug-07		1	LOW	
0709204-002A	B-3 (4-4 1/2')DL	Soil	SW8260B	07-Aug-07	09-Aug-07		17-Aug-07		1	MED	
0709204-002A	B-3 (4-4 1/2')	Soil	SW8260B	07-Aug-07	09-Aug-07		15-Aug-07		10	LOW	
0709204-003A	B-4 (10-12')	Soil	SW8260B	07-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709204-004A	B-5 (0-6')	Soil	SW8260B	07-Aug-07	09-Aug-07		15-Aug-07		10	LOW	
0709204-004A	B-5 (0-6')DL	Soil	SW8260B	07-Aug-07	09-Aug-07		17-Aug-07		1	MED	
0709204-005A	B-6 (2 1/2-3')DL	Soil	SW8260B	07-Aug-07	09-Aug-07		17-Aug-07		500	MED	
0709204-005A	B-6 (2 1/2-3')	Soil	SW8260B	07-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709204-006A	B-7 (9-10')	Soil	SW8260B	07-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709204-007A	B-8 (6-10')DL	Soil	SW8260B	07-Aug-07	09-Aug-07		17-Aug-07		1000	MED	
0709204-007A	B-8 (6-10')	Soil	SW8260B	07-Aug-07	09-Aug-07		14-Aug-07		1	LOW	
0709204-007AMS	B-8 (6-10')MS	Soil	SW8260B	07-Aug-07	09-Aug-07		14-Aug-07		1	LOW	
0709204-007AMSD	B-8 (6-10')MSD	Soil	SW8260B	07-Aug-07	09-Aug-07		14-Aug-07		1	LOW	
0709204-008A	B-9 (9-10')	Soil	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709204-009A	B-10 (4-6')	Soil	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709204-010A	B-11 (9 1/2-10 1/2)	Soil	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709204-011A	FIELD BLANK (SOIL)	Aqueous	SW8260B	08-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709204-012A	TRIP BLANK 8/7/2007	Aqueous	SW8260B	07-Aug-07	09-Aug-07		15-Aug-07		1	LOW	
0709403-001A	B-12 6'-8'	Soil	SW8260B	13-Aug-07	14-Aug-07		15-Aug-07		1	LOW	
0709403-002A	B-13 7'-9'	Soil	SW8260B	13-Aug-07	14-Aug-07		15-Aug-07		1	LOW	
0709403-002A	B-13 7'-9'DL	Soil	SW8260B	13-Aug-07	14-Aug-07		17-Aug-07		1	MED	
0709403-003A	B-14 4'-6'	Soil	SW8260B	13-Aug-07	14-Aug-07		15-Aug-07		1	LOW	

Laboratory Samp ID	Client Sample ID	Matrix	Analytical Protocol	Date Collected	DateRecd at Lab	Date Extracted	Date Analyzed	Extraction Method	DF	Level	Aux Cleanup
0709403-004A	B-15 20'-26'	Soil	SW8260B	13-Aug-07	14-Aug-07		15-Aug-07		1	LOW	

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

SAMPLE PREPARATION AND ANALYSIS SUMMARY
SEMIVOLATILE (BNA)
ANALYSES

SDG: BER042

Laboratory Samp ID	Client Sample ID	Matrix	Analytical Protocol	Date Collected	DateRecd at Lab	Date Extracted	Date Analyzed	Extraction Method	DF	Level	Aux Cleanup
0709059-001D	B-1	Soil	ASPB 8270	06-Aug-07	07-Aug-07	13-Aug-07	15-Aug-07	PFEX	1	LOW	
0709059-001DDL	B-1DL	Soil	ASPB 8270	06-Aug-07	07-Aug-07	13-Aug-07	15-Aug-07	PFEX	5	LOW	
0709204-004B	B-5 (0-6')	Soil	ASPB 8270	07-Aug-07	09-Aug-07	13-Aug-07	15-Aug-07	PFEX	1	LOW	
0709204-004BDL	B-5 (0-6')DL	Soil	ASPB 8270	07-Aug-07	09-Aug-07	13-Aug-07	15-Aug-07	PFEX	5	LOW	
0709204-007B	B-8 (6-10')	Soil	ASPB 8270	07-Aug-07	09-Aug-07	13-Aug-07	15-Aug-07	PFEX	1	LOW	
0709204-007BDL	B-8 (6-10')DL	Soil	ASPB 8270	07-Aug-07	09-Aug-07	13-Aug-07	15-Aug-07	PFEX	5	LOW	
0709403-001B	B-12 6'-8'	Soil	ASPB 8270	13-Aug-07	14-Aug-07	15-Aug-07	20-Aug-07	PFEX	1	LOW	
0709403-002B	B-13 7'-9'	Soil	ASPB 8270	13-Aug-07	14-Aug-07	15-Aug-07	20-Aug-07	PFEX	1	LOW	

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

SAMPLE PREPARATION AND ANALYSIS SUMMARY
PESTICIDE/PCB/HERB
ANALYSES

SDG: BER042

Laboratory Samp ID	Client Sample ID	Matrix	Analytical Protocol	Date Collected	DateRecd at Lab	Date Extracted	Date Analyzed	Extraction Method	DF	Level	Aux Cleanup
0709204-007B	B-8 (6-10')	Soil	SW8081/8082	07-Aug-07	09-Aug-07	14-Aug-07	19-Aug-07	PFEX	1	LOW	GPC
0709204-007BDL	B-8 (6-10')DL	Soil	SW8081/8082	07-Aug-07	09-Aug-07	14-Aug-07	21-Aug-07	PFEX	10	LOW	GPC
0709204-007BDL2	B-8 (6-10')DL	Soil	SW8081/8082	07-Aug-07	09-Aug-07	14-Aug-07	21-Aug-07	PFEX	25	LOW	GPC
0709204-007BDL3	B-8 (6-10')DL	Soil	SW8081/8082	07-Aug-07	09-Aug-07	14-Aug-07	21-Aug-07	PFEX	50	LOW	GPC
0709403-002B	B-13 7'-9'	Soil	SW8081/8082	13-Aug-07	14-Aug-07	14-Aug-07	19-Aug-07	PFEX	1	LOW	GPC

NEW YORK STATE DEPARTMENT OF ENVIRONMENTAL CONSERVATION

SAMPLE PREPARATION AND ANALYSIS SUMMARY
INORGANIC ANALYSIS

SDG : BER042

Laboratory Samp ID	Client Sample ID	Matrix	Metals Requested	DateRecd at Lab	Date Analyzed
0709059-001	B-1	SOIL	AG,AL,AS,BA,BE,CA,CD,CO,CR,CU,FE,HG,K,MG,MN,NA,NI,PB,SB,SE,TL,V,ZN,	07-Aug-07	08/07
0709204-007	B-8 (6-10')	SOIL	AG,AL,AS,BA,BE,CA,CD,CO,CR,CU,FE,HG,K,MG,MN,NA,NI,PB,SB,SE,TL,V,ZN,	09-Aug-07	08/07
0709204-007DUP	B-8 (6-10')D	SOIL	AG,AL,AS,BA,BE,CA,CD,CO,CR,CU,FE,HG,K,MG,MN,NA,NI,PB,SB,SE,TL,V,ZN,	09-Aug-07	08/07
0709204-007MS	B-8 (6-10')S	SOIL	AG,AL,AS,BA,BE,CA,CD,CO,CR,CU,FE,HG,K,MG,MN,NA,NI,PB,SB,SE,TL,V,ZN,	09-Aug-07	08/07
0709403-002	B-13 7'-9'	SOIL	AG,AL,AS,BA,BE,CA,CD,CO,CR,CU,FE,HG,K,MG,MN,NA,NI,PB,SB,SE,TL,V,ZN,	14-Aug-07	08/07

Appendix B
Chain of Custody Documents

H2M LABS, INC.

575 Broad Hollow Rd, Melville, NY 11747-5076
Tel: (631) 694-3040 Fax: (631) 420-8436

2588

EXTERNAL CHAIN OF CUSTODY

CLIENT:

H2M SDG NO: BER 043

PROJECT NAME/NUMBER: Schmuckler's Cleaners

SAMPLERS: (signature)/Client: Berninger Environmental/ INC.

DELIVERABLES: NYSDEC ASP-B

TURNAROUND TIME:

NOTES: (*) MS/MSD on VOCs only.

Project Contact: B. Mills / Walter B.

Phone Number: 631 589 6541

PIS/Quote #

P.O. # 7082112

DATE	TIME	MATRIX	FIELD I.D.	Total No. of Containers	ANALYSIS REQUESTED: ORGANIC VOA	ORGANIC BNA	ORGANIC Pest/PCB	INORG. Metal	INORG. CN	LAB I.D. NO.	REMARKS:
8/8/07		L	Trip Blank 8/8/07 (*)	2	X					0709201-010	
8/7/07		S	B-8 6-10' (MS/MSD)	8	X	X	X	X			
8/7/07		S	B-7 9-10'	1	X						
8/7/07	1:05	S	B-3 4-4½	2	X						
8/7/07		S	B-2 5-7'	1	X						
8/7/07		S	B-1 2½-3'	1	X						
8/7/07		S	B-4 10-12'	1	X						
8/7/07		L	Trip Blank 8/7/07	2	X						
8/7/07		L	GW-1	10	X	X	X	X		0709201-001	
8/7/07		L	GW-8	2	X					0709201-008	

Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
Peter Damish	8/9/07	12:40		8/9/07	12:40
Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time

LABORATORY USE ONLY

Discrepancies Between Sample Labels and COC Record? Y or N
Explain:

Samples were:
1. Shipped___ or Hand Delivered___ Airbill# _____
2. Ambient or chilled, Temp_____
3. Received in good condition: Y or N
4. Properly preserved: Y or N

COC Tape was:
1. Present on outer package: Y or N
2. Unbroken on outer package: Y or N
3. COC record present & complete upon sample receipt: Y or N

WHITE COPY - ORIGINAL (BER043.SSV)

YELLOW COPY - CLIENT

PINK COPY - LABORATORY

H2M LABS, INC.

575 Broad Hollow Rd, Melville, NY 11747-5076
Tel: (631) 694-3040 Fax: (631) 420-8436

2580

EXTERNAL CHAIN OF CUSTODY

CLIENT:

H2M SDG NO: B0704?

PROJECT NAME/NUMBER
Schmuckler's Cleaners
358-362 North Ave

SAMPLERS: (signature)/Client
Berninger Environmental Inc.

DELIVERABLES:
NJ/SDEC ASP-B

TURNAROUND TIME: STANDARD

NOTES:
*MS/MSD
VOCs only

Project Contact:
Bills/Walter S

Phone Number:
631 589 6521

PIS/Quote #

P.O.# 7082112

DATE	TIME	MATRIX	FIELD I.D.	Total No. of Containers	ANALYSIS REQUESTED: ORGANIC VOA (Sample Container Description: VOCs 8260 w/ TICs)	ORGANIC BNA (Sample Container Description: SVOCs 8270)	ORGANIC Pest/PCB	INORG. Metal	INORG. CN	LAB I.D. NO.	REMARKS:
8/8/07	3:21	L	GW-7	2	X	X				07092084-007	
8/8/07		L	GW-6	2	X					-006	
8/8/07	2:37	S	B-10 4-4½'	1	X						
8/7/07		S	B-5 (0-6")	2	X	X					
8/8/07	11:10	L	GW-4	3	X	X				07092084-004	
8/8/07	3:15	S	B-11 9½-10½	1	X						
8/8/07		L	GW-3	2	X					07092084-003	
8/8/07		L	GW-2 (MS/MSD)*	5	X	X				-002	
8/8/07		L	GW-5	3	X	X				-005	
8/8/07		S	B-9 (9-10')	1	X						

Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
Peter Danish	8/9/07	12:40		8/9/07	12:40

LABORATORY USE ONLY

Discrepancies Between Sample Labels and COC Record? Y or N
Explain:

Samples were:
1. Shipped___ or Hand Delivered___ Airbill# _____
2. Ambient or chilled, Temp_____
3. Received in good condition: Y or N
4. Properly preserved: Y or N

COC Tape was:
1. Present on outer package: Y or N
2. Unbroken on outer package: Y or N
3. COC record present & complete upon sample receipt: Y or N

WHITE COPY - ORIGINAL (B070492) YELLOW COPY - CLIENT PINK COPY - LABORATORY

H2M LABS, INC.

575 Broad Hollow Rd, Melville, NY 11747-5076
Tel: (631) 694-3040 Fax: (631) 420-8436

258

EXTERNAL CHAIN OF CUSTODY

CLIENT:

H2M SDG NO: BER2049

PROJECT NAME/NUMBER: Schmuckler's Cleaners

SAMPLERS: (signature)/Client: Berdinger Environmental Inc

DELIVERABLES:

TURNAROUND TIME:

NOTES:

Project Contact: Bill S Walter B.

Phone Number: 631 582 6551

PIS/Quote #

P.O# 7082112

DATE	TIME	MATRIX	FIELD I.D.	Total No. of Containers	ANALYSIS REQUESTED: ORGANIC VOA	ORGANIC BNA	ORGANIC Pest/PCB	INORG. Metal	INORG. CN	LAB I.D. NO.	REMARKS:
8/8/07	8/7/07	L	Field Blank water	X	X					BER2049-001	
8/8/07		L	Field Blank Soil	X	X					002	

Sample Container Description: VOCs & HAL plus TICs

Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
Peter Daigh	8/9/07	12:40		8/9/07	1240
Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time

LABORATORY USE ONLY

Discrepancies Between Sample Labels and COC Record? Y or N

Explain:

Samples were:
1. Shipped___ or Hand Delivered___ Airbill# _____
2. Ambient or chilled, Temp_____
3. Received in good condition: Y or N
4. Properly preserved: Y or N

COC Tape was:
1. Present on outer package: Y or N
2. Unbroken on outer package: Y or N
3. COC record present & complete upon sample receipt: Y or N

WHITE COPY - ORIGINAL YELLOW COPY - CLIENT PINK COPY - LABORATORY

BER0431511

H2M LABS, INC.

575 Broad Hollow Rd, Melville, NY 11747-5076
Tel: (631) 694-3040 Fax: (631) 420-8436

25884

EXTERNAL CHAIN OF CUSTODY

BER042

CLIENT: | **H2M SDG NO:** BER043

PROJECT NAME/NUMBER: Schmuckler's

SAMPLERS: (signature)/Client: Bernwager Environmental Inc.

DELIVERABLES: NYSDEC ASP-B

TURNAROUND TIME:

NOTES:

Project Contact: B. M. S. Walter B

Phone Number: 917 715 0752 / 631 589 6521

PIS/Quote #

DATE	TIME	MATRIX	FIELD I.D.	Total No. of Containers	Sample Container Description: VOCs 8260 plus TICs	SVOCs 8270	Pest/PCBs	TAL Metals	LAB I.D. NO.	REMARKS:
8/13/07	8:35	S	B-12 6-8'	2	X	X			0709403-001	
8/13/07	9:45	S	B-13 7-9'	3	X	X	X	X	-002	
8/13/07	10:40	S	B-14 4-6'	1	X				-003	
8/13/07	1:05	S	B-15 20-26"	1	X				-004	
8/13/07	2:51	L	GW-9	3	X	X			0709405-001	
8/13/07	2:45	L	GW-10	2	X				-002	
8/13/07	3:25	L	GW-11	3	X	X			-003	
8/13/07	N/A	L	Trip Blank 8/13/07	2	X				-004	

ANALYSIS REQUESTED — ORGANIC: VOA, BNA, Pest/PCB; INORG.: Metal, CN

Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
Butch Meyer	8/14/07	3:15		8/14/07	15:15
	8/14/07	15:20		8/14/07	15:20

LABORATORY USE ONLY

Discrepancies Between Sample Labels and COC Record? Y or N

Explain:

Samples were:
1. Shipped___ or Hand Delivered___ Airbill# _____
2. Ambient or chilled, Temp_____
3. Received in good condition: Y or N
4. Properly preserved: Y or N

COC Tape was:
1. Present on outer package: Y or N
2. Unbroken on outer package: Y or N
3. COC record present & complete upon sample receipt: Y or N

WHITE COPY - ORIGINAL — YELLOW COPY - CLIENT — PINK COPY - LABORATORY

BER043 S19

H2M LABS, INC.

575 Broad Hollow Rd, Melville, NY 11747-5076
Tel: (631) 694-3040 Fax: (631) 420-8436

Page 2 of 2 2582 PO # 1082407

EXTERNAL CHAIN OF CUSTODY

CLIENT: | H2M SDG NO: BER042

PROJECT NAME/NUMBER
Schmukler's Cleaner
358-364 North Ave
New Rochelle, N.Y.

SAMPLERS: (signature)/Client
BER / William Schlageter

DELIVERABLES: NYSDEC ASP B

TURNAROUND TIME:

NOTES:
NYSDEC
ASP B Deliverables

Project Contact: Brad Walter B
Phone Number: 631 589 6521
PIS/Quote #

BERNINGER ENVIRONMENTAL, INC
90-B KNICKERBOCKER AVE
BOHEMIA, NEW YORK 11716

DATE	TIME	MATRIX	FIELD I.D.	Total No. of Containers	ANALYSIS REQUESTED: ORGANIC: VOA (8260 VOCs plus TICs)	ORGANIC: BNA (8270 SVOCs)	ORGANIC: Pest/PCB	ORGANIC: TAL Metals	INORG.: Metal	INORG.: CN	LAB I.D. NO.	REMARKS:
8/6/07	5:05	S	B-1	4	X	X	X	X			0709059-001	
8/6/07	~~5:05~~	L	Trip Blank 8/6/07								0709059-002	
8/6/07	6:35	S	FD-1	1	X						0709059-002	
8/6/07	5:51	S	FD-2	1	X						0709059-003	

Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
William Schlageter	8/7/07	10:30	[illegible]	8/7/07	10:30
Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time
Relinquished by: (Signature)	Date	Time	Received by: (Signature)	Date	Time

LABORATORY USE ONLY

Discrepancies Between Sample Labels and COC Record? Y or N
Explain:

Samples were:
1. Shipped___ or Hand Delivered___ Airbill# _____
2. Ambient or chilled, Temp._____
3. Received in good condition: Y or N
4. Properly preserved: Y or N

COC Tape was:
1. Present on outer package: Y or N
2. Unbroken on outer package: Y or N
3. COC record present & complete upon sample receipt: Y or N

WHITE COPY - ORIGINAL YELLOW COPY - CLIENT PINK COPY - LABORATORY

Appendix C
SDG Narratives

H2M LABS, INC.

SDG NARRATIVE FOR VOLATILE ORGANICS
SAMPLES RECEIVED: 8/9/07, & 8/14/07
SDG #: BER043

For Samples:

GW-1	GW-6	STORAGE BLANK 8/9/2007
GW-2 MSS/MSD	GW-7	GW-9
GW-3	GW-8	GW-10
GW-4	FIELD BLANK WATER	GW-11
GW-5	TRIP BLANK 8/8/2007	TRIP BLANK 8/13/07

The above water samples and blanks were analyzed for volatile organics by EPA method 8260B in accordance with the NYSDEC ASP, Rev. 6/2000.

All QC data and calibrations met the requirements of the method, unless discussed below. The following should be noted:

- Sample GW-2 was analyzed as the matrix spike/matrix spike duplicate. Due to the level of trichloroethene present in the sample, the spiking level was not a multiple of the sample concentration, and QC limits therefore do not apply for this analyte.
- Dilutions were required for seven samples to keep the concentrations within the calibration range. Results for both sets of data are reported.

I certify that this data package is in compliance with the terms and conditions of the contract, both technically and for completeness, for other than the conditions detailed above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or his designee, as verified by the following signature.

Date Reported: August 31, 2007

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* *
****

Ursula Middel
Technical Manager

BER043 S26

H2M LABS, INC.

SDG NARRATIVE FOR SEMIVOLATILES ANALYSES
SAMPLES RECEIVED: 8/9/07 - 8/14/07
SDG #: BER043

For Samples:

GW-1	GW-5
GW-2	GW-9
GW-4	GW-11

The above water samples were analyzed for a select list of semivolatile organic analytes by EPA method 8270C in accordance with the NYSDEC ASP, Rev. 6/2000.

All QC data and calibrations met the requirements of the method, unless listed below. The following should be noted:

- No matrix spike/matrix spike duplicate was submitted. Two lab-fortified blanks were analyzed. In both, all recoveries met acceptance criteria.
- Due to insufficient volume available, only 500 mL were extracted for sample GW-11, however, the same concentration factor was achieved by concentrating the final extract volume to 0.5 mL.
- Due to matrix, the final extract volume for sample GW-2 was 2 mL.
- With the exception of sample GW-11, all samples required dilutions. The undiluted analyses for sample GW-2, GW-4, and GW-5 are not submitted. Severe matrix interferences were observed with the internal standards that made results unusable.
- Recovery for the surrogate nitrobenzene-d5 was above the QC limit in samples GW-4 and GW-5.
- No surrogate recoveries are reportable for GW-4DL, GW-5DL, and GW-2, because the surrogates were diluted out.
- In the initial calibration, the RSD for benzo(k)fluoranthene was above 20.5% but below the limit of 40%, and the response factor (RF) for bis(2chloroethoxy)methane was below 0.3.
- The response factor for bis(2chloroethoxy)methane was also low in the continuous calibration checks (CCV) for 8/15/07 and 8/20/07.

I certify that this data package is in compliance with the terms and conditions of the contract, both technically and for completeness, for other than the conditions detailed above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or his designee, as verified by the following signature.

Date Reported: August 29, 2007

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* *
* *
****

Ursula Middel
Technical Manager

BER043 S27

H2M LABS, INC.

SDG NARRATIVE FOR PESTICIDES AND PCB ANALYSES
SAMPLES RECEIVED: 8/9/07
SDG #: BER043

For Sample:

GW-1

The above water sample was prepared and analyzed for pesticides and PCBs by EPA methods 3510B and 8081A/ 8082 in accordance with the NYSDEC ASP, Rev. 6/2000.

All QC data and calibrations met the requirements of the method, unless discussed below. The following should be noted:

- No matrix spike / matrix spike duplicate was requested. Two lab-fortified blanks were analyzed, and results indicate good method efficiency.
- Positives are reported to the practical detection limit of ½ of the reporting limits (PQL).

I certify that this data package is in compliance with the terms and conditions of the contract, both technically and for completeness, for other than the conditions detailed above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or his designee, as verified by the following signature.

Date Reported: August 29, 2007

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* *
* *
****

Ursula Middel
Technical Manager

BER043 S28

H2M LABS, INC.

SDG NARRATIVE FOR METALS ANALYSIS
SAMPLES RECEIVED: 8/9/07
SDG#: BER043

For Samples:

GW-1

One water sample was received by H2M Labs, Inc on 8/9/07 for TAL metals analysis.

Samples were prepared and analyzed using EPA methods 6010B with a TJA 61E trace ICP instrument and 7470A with a Leeman Hydra mercury analyzer.

Sample GW-1 was utilized for QC analysis and reporting.

Spike analysis did not recover within 75-125% for aluminum, copper, iron, manganese and mercury. Since the sample values were greater than four times the spike concentrations, post spikes and data qualifiers were not required.

Selenium spike analysis did not recover within acceptance ranges. The sample was post spiked, reanalyzed and recovered at 76.7%. Selenium data was reported flagged "N" on Forms 1 and 5a.

Duplicate analysis did not reproduce within acceptance ranges for mercury. Mercury data was reported flagged "*" on Forms 1 and 6.

ICP serial dilution analysis did not reproduce within acceptance ranges for potassium. Potassium data was reported flagged "E" on Forms 1 and 9.

Samples were diluted and reanalyzed as required to keep instrument readings within linear ranges.

No other issues were noted during the analysis of this sample group.

I certify that this data package is in compliance with the terms and conditions of the contract, both technically and for completeness, for other than the conditions detailed above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or his designee, as verified by the following signature.

Date Reported: August 30, 2007

Vincent Stancampiano
Vice President

BER043 S29

H2M LABS, INC.

SDG NARRATIVE FOR VOLATILE ORGANICS
SAMPLES RECEIVED: 8/7/07, 8/9/07, & 8/14/07
SDG #: BER042

For Samples:

B-1	B-4 (10-12')	B-11 (9 1/2-10 1/2)
FD-1	B-5 (0-6')	FIELD BLANK (SOIL)
FD-2	B-6 (2 1/2-3')	TRIP BLANK 8/7/2007
TRIP BLANK 8/6/07	B-7 (9-10')	B-12 6'-8'
STORAGE BLANK 8/7/07	B-8 (6-10') MS/MSD	B-13 7'-9'
B-2 (5-7')	B-9 (9-10')	B-14 4'-6'
B-3 (4-4 1/2')	B-10 (4-6')	B-15 20'-26'

The soil samples and blanks were analyzed for volatile organics by EPA method 8260B in accordance with the NYSDEC ASP, Rev. 6/2000.

All QC data and calibrations met the requirements of the method, unless discussed below. The following should be noted:

- Sample B-8 (6-10') was analyzed as the matrix spike/matrix spike duplicate. Due to the level of trichloroethene present in the sample, the spiking level was not a multiple of the sample concentration, and QC limits therefore do not apply for this analyte.

- Dilutions were required for five samples to keep the concentrations within the calibration range. These five samples were analyzed with low and medium level procedure, and results for both methods are reported. Three of the low level analyses were performed with lower sample weight "at a dilution".

- In the lab fortified blank (LFB) for the low level method recoveries for acetone, p-diethylbenzene and 1,2,4,5-tetramethylbenzene were below the QC limit. The recoveries for p-diethylbenzene and 1,2,4,5-tetramethylbenzene were 71% and only slightly below the limit of 72%. All recoveries for the medium level LFB met the acceptance criteria.

- %D in the continuous calibration checks (CCV) exceeded 25% (but met the limit of 40%) for the following analytes: Tetrachloroethene on 8/13/07 and vinylchloride on 8/17/07.

I certify that this data package is in compliance with the terms and conditions of the contract, both technically and for completeness, for other than the conditions detailed above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or his designee, as verified by the following signature.

Date Reported: August 31, 2007

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Ursula Middel
Technical Manager

BER042 A10

H2M LABS, INC.

SDG NARRATIVE FOR SEMIVOLATILES ANALYSES
SAMPLES RECEIVED: 8/7/07 - 8/14/07
SDG #: BER042

For Samples:

B-1	B-12 6'-8'
B-5 (0-6')	B-13 7'-9'
B-8 (6-10')	

The above samples were analyzed for a select list of semivolatile organic analytes by EPA method 8270C in accordance with the NYSDEC ASP, Rev. 6/2000.

All QC data and calibrations met the requirements of the method, unless listed below. The following should be noted:

- No matrix spike/matrix spike duplicate was submitted. Two lab-fortified blanks were analyzed. In both, all recoveries met acceptance criteria.
- Concentrations in three samples, B-1, B-8(6-10'), and B-5(0-6'), exceeded the calibration range, and the samples were reanalyzed at a dilution. Both sets of data are reported.
- In the undiluted analyses of the three samples, one or more internal standard areas were low, but they were within the acceptance limits in the dilutions.
- Recovery for the surrogate nitrobenzene-d5 was above the QC limit in sample B-1 and the dilution B-1DL.
- In the initial calibration, the RSD for benzo(k)fluoranthene was above 20.5% but below the limit of 40%, and the response factor (RF) for bis(2chloroethoxy)methane was below 0.3.
- The response factor for bis(2chloroethoxy)methane was also low in the continuous calibration checks (CCV) for the three days of analysis.
- Some TIC compounds were flagged with the qualifier "Y" to indicate that the value reported is biased low due to unresolved matrix interference with the internal standard that was used to quantify the TIC compounds. The TIC results for the dilution should be used instead.

I certify that this data package is in compliance with the terms and conditions of the contract, both technically and for completeness, for other than the conditions detailed above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or his designee, as verified by the following signature.

Date Reported: August 28, 2007

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Ursula Middel
Technical Manager

BER042 A11

H2M LABS, INC.

SDG NARRATIVE FOR PESTICIDES AND PCB ANALYSES
SAMPLES RECEIVED: 8/9/07 & 8/14/07
SDG #: BER042

For Samples:

B-8 (6-10')
B-13 7'-9'

The above soil samples were prepared and analyzed for pesticides and PCBs by EPA methods 3545 and 8081A/ 8082 in accordance with the NYSDEC ASP, Rev. 6/2000..

All QC data and calibrations met the requirements of the method, unless discussed below. The following should be noted:

- No matrix spike / matrix spike duplicate was requested. Two lab-fortified blanks were analyzed, and results indicate good method efficiency.
- The concentrations of analytes in sample B-8 (6-10') exceeded the highest calibration level. Several dilutions were reanalyzed to keep the concentrations for all analytes within the calibration range. Four sets of data are reported.
- No surrogate recoveries are reported for the dilutions, because the surrogates are diluted out.
- Recovery for the surrogate DCB was above the QC limit on one column sample B-8 (6-10').
- Positives are reported to the practical detection limit of ½ of the reporting limits (PQL) for pesticides. Due to interference level, no aroclors are reportable below the PQL.

I certify that this data package is in compliance with the terms and conditions of the contract, both technically and for completeness, for other than the conditions detailed above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or his designee, as verified by the following signature.

Date Reported: August 29, 2007

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Ursula Middel
Technical Manager

BER042 A12

H2M LABS, INC.

SDG NARRATIVE FOR METALS ANALYSIS
SAMPLES RECEIVED: 8/7/07, 8/9/07 & 8/14/07
SDG#: BER042

For Samples:

B-1
B-8 (6-10')
B-13 7'-9'

Three soil samples were received by H2M Labs, Inc on 8/7/07, 8/9/07 and 8/14/07 for TAL metal analysis.

Samples were prepared and analyzed using EPA methods 6010B with a TJA 61E trace ICP instrument and 7471A with a Leeman Hydra mercury analyzer.

Sample B-8 (6-10') was utilized for QC analysis and reporting.

Spike analysis did not recover within 75-125% for iron. Since the sample value was greater than four times the spike concentration, post spikes and data qualifiers were not required.

Antimony spike analysis did not recover within acceptance ranges. The sample was post spiked, reanalyzed and recovered at 101.7%. Antimony data was reported flagged "N" on Forms 1 and 5a.

Duplicate analysis did not reproduce within acceptance ranges for copper. Copper data was reported flagged "*" on Forms 1 and 6.

No other issues were noted during the analysis of this sample group.

I certify that this data package is in compliance with the terms and conditions of the contract, both technically and for completeness, for other than the conditions detailed above. Release of the data contained in this hardcopy data package has been authorized by the Laboratory Manager or his designee, as verified by the following signature.

Date Reported: August 30, 2007

Vincent Stancampiano
Vice President

BER042 A13

L.A.B. Validation Corp. 14 West Point Drive, East Northport, N.Y. 11731 (516) 523-7891

Appendix D
Tentatively Identified Compounds ((TICs) for VOA/SVOA

LABValidation@aol.com

1F

EPA SAMPLE NO.

GW-1

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-001A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55481.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/16/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 6 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST. CONC.	Q
1. 000108-87-2	Cyclohexane, methyl-	8.43	26	JN
2.	c4 substituted benzene (14.75)	14.75	33	J
3.	c4 substituted benzene (15.11)	15.11	14	J
4.	c4 substituted benzene (15.53)	15.53	20	J
5.	c4 substituted benzene (16.44)	16.44	16	J
6.	c4 substituted benzene (16.72)	16.72	24	J

BER043 S36

1G

EPA SAMPLE NO.

GW-1

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: ____

Lab Code: 10478 Case No.: BER SAS No.: ____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-001B

Sample wt/vol: 1000 (g/mL) ML Lab File ID: 7\N21160.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: Decanted:(Y/N) N Date Extracted: 08/13/07

Concentrated Extract Volume: 1000 (µl) Date Analyzed: 08/15/07

Injection Volume: 2 (µl) Dilution Factor: 1.00

GPC Cleanup: (Y/N) N pH: ____ Extraction: (Type) SEPF

CONCENTRATION UNITS:

Number TICs found: 18 (µg/L or µg/Kg) UG/L

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.		ethyl naphthalene isomer	8.82	160	J
2.		dimethyl naphthalene isomer (8.98)	8.98	370	J
3.		dimethyl naphthalene isomer (9.12)	9.12	360	J
4.		dimethyl naphthalene isomer (9.17)	9.17	230	J
5.		dimethyl naphthalene isomer (9.31)	9.31	91	J
6.		(DEL) Alkane: Branched (9.38)	9.38	93	J
7.		dimethyl naphthalene isomer (9.47)	9.47	92	J
8.		unknown (9.56)	9.56	95	J
9.		methyl, ethyl ~~trimethyl~~ naphthalene isomer (9.98)	9.98	160	J
10.		methyl, ethyl ~~trimethyl~~ naphthalene isomer (10.05)	10.05	55	J
11.		trimethyl naphthalene isomer (10.17)	10.17	86	J
12.		trimethyl naphthalene isomer (10.25)	10.25	110	J
13.		trimethyl naphthalene isomer (10.38)	10.38	120	J
14.		trimethyl naphthalene isomer (10.44)	10.44	100	J
15.		trimethyl naphthalene isomer (10.53)	10.53	64	J
16.	027193-28-8	Phenol, (1,1,3,3-tetramethylbutyl)	10.89	260	JN
17.		unknown (11.39)	11.39	64	J
18.		(DEL) Alkane: Branched (11.78)	11.78	180	J
19.		unknown (11.95)	11.95	84	J
20.		substituted PNA	13.34	70	J

8/29/07

BER043 S42

1F

EPA SAMPLE NO.

GW-2

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-002A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55448.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 7 (µg/L or µg/Kg) UG/L

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.		c3 substituted benzene	13.56	18	J
2.		c4 substituted benzene (15.53)	15.53	30	J
3.		c4 substituted benzene (15.94)	15.94	18	J
4.		c4 substituted benzene (16.44)	16.44	21	J
5.		c4 substituted benzene (16.71)	16.71	38	J
6.	000119-64-2	Naphthalene, 1,2,3,4-tetrahydro-	17.04	18	JN
7.		c5 substituted benzene (17.45)	17.45	20	J

BER043 S50

1G

EPA SAMPLE NO.

GW-2

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-002B

Sample wt/vol: 1000 (g/mL) ML Lab File ID: 7\N21260.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: Decanted:(Y/N) N Date Extracted: 08/13/07

Concentrated Extract Volume: 2000 (µl) Date Analyzed: 08/20/07

Injection Volume: 2 (µl) Dilution Factor: 100.00

GPC Cleanup: (Y/N) N pH: ___ Extraction: (Type) SEPF

CONCENTRATION UNITS:

Number TICs found: 9 (µg/L or µg/Kg) UG/L

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.		(DEL) Alkane: Straight-Chain (4.95)	4.95	5000	J
2.		(DEL) Alkane: Branched (7.69)	7.69	4800	J
3.		(DEL) Alkane: Straight-Chain (7.97)	7.97	11000	J
4.		dimethyl naphthalene isomer (8.1)	8.10	7400	J
5.		dimethyl naphthalene isomer (8.25)	8.25	8300	J
6.		dimethyl naphthalene isomer (8.3)	8.30	5200	J
7.		(DEL) Alkane: ~~Straight~~ Chain (8.57) Branched	8.57	6800	J
8.		(DEL) Alkane: Straight-Chain (9)	9.00	12000	J
9.		trimethyl naphthalene isomer	9.68	5900	J
10.		(DEL) Alkane: Straight-Chain (9.99)	9.99	10000	J
11.		unknown (10.02)	10.02	6400	J
12.		(DEL) Alkane: Branched (10.41)	10.41	11000	J
13.		(DEL) Alkane: Straight-Chain (10.93)	10.93	34000	J
14.		unknown (11.05)	11.05	6800	J
15.	000605-39-0	2,2'-Dimethylbiphenyl	11.28	5700	JN
16.		unknown (11.48)	11.48	6500	J
17.		(DEL) Alkane: Straight-Chain (12.6)	12.60	14000	J
18.		substituted PNA	12.82	5000	J
19.		(DEL) Alkane: Straight-Chain (13.15)	13.15	10000	J
20.		(DEL) Alkane: Straight-Chain (13.59)	13.59	7800	J

8/29/07

BER043 S56

1F

EPA SAMPLE NO.

GW-3

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-003A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55451.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 7 (µg/L or µg/Kg) UG/L

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.		c4 substituted benzene (15.98)	15.98	8	J
2.		c4 substituted benzene (16.71)	16.71	8	J
3.		c4 substituted benzene (17.16)	17.16	10	J
4.		c5 substituted benzene (17.46)	17.46	7	J
5.		c4 substituted benzene (18.42)	18.42	7	J
6.		c4 substituted benzene (18.81)	18.81	11	J
7.	000090-12-0	Naphthalene, 1-methyl	20.62	9	JN

BER043 S59

1F

EPA SAMPLE NO.

GW-4

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-004A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55452.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 8 (µg/L or µg/Kg) UG/L

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.		c3 substituted benzene	13.25	50	J
2.		(DEL) Alkane: Branched	13.48	48	J
3.		(DEL) Alkane: Cyclic	14.08	98	J
4.		c4 substituted benzene (15.16)	15.16	57	J
5.		unknown	15.77	66	J
6.		c4 substituted benzene (15.95)	15.95	45	J
7.		c4 substituted benzene (16.44)	16.44	45	J
8.		c4 substituted benzene (16.72)	16.72	46	J

BER043 S62

1G

EPA SAMPLE NO.

GW-4

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-004B

Sample wt/vol: 1000 (g/mL) ML Lab File ID: 7\N21255.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: Decanted:(Y/N) N Date Extracted: 08/13/07

Concentrated Extract Volume: 1000 (µl) Date Analyzed: 08/20/07

Injection Volume: 2 (µl) Dilution Factor: 10.00

GPC Cleanup: (Y/N) N pH: ____ Extraction: (Type) SEPF

CONCENTRATION UNITS:

Number TICs found: 9 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	C3 substituted benzene	3.94	420	J
2.	(DEL) Alkane: Straight-Chain (4.18)	4.18	930	J
3.	C4 substituted benzene	4.63	420	J
4.	(DEL) Alkane: Straight-Chain (4.97)	4.97	720	J
5.	(DEL) Alkane: Branched (7.73)	7.73	410	J
6.	(DEL) Alkane: Straight-Chain (8.03)	8.03	880	J
7.	dimethyl naphthalene isomer (8.16)	8.16	860	J
8.	dimethyl naphthalene isomer (8.31)	8.31	770	J
9.	dimethyl naphthalene isomer (8.36)	8.36	520	J
10.	dimethyl naphthalene isomer (8.52)	8.52	530	J
11.	(DEL) Alkane: ~~Straight~~-Chain (8.62) Branched	8.62	420	J
12.	(DEL) Alkane: Straight-Chain (9.06)	9.06	790	J
13.	trimethyl naphthalene isomer (9.15)	9.15	460	J
14.	trimethyl naphthalene isomer (9.57)	9.57	420	J
15.	trimethyl naphthalene isomer (9.72)	9.72	530	J
16.	(DEL) Alkane: Straight-Chain (10.05)	10.05	1300	J
17.	(DEL) Alkane: Branched (11.03)	11.03	1400	J
18.	(DEL) Alkane: Straight-Chain (12.64)	12.64	600	J
19.	(DEL) Alkane: Straight-Chain (13.18)	13.18	410	J

8/29/07

BER043 S68

1F

EPA SAMPLE NO.

GW-5

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-005A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55453.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

Number TICs found: 8 CONCENTRATION UNITS: (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1. 000108-87-2	Cyclohexane, methyl-	8.42	31	JN
2.	c3 substituted benzene	13.24	25	J
3.	c4 substituted benzene (14.74)	14.74	50	J
4.	c4 substituted benzene (15.15)	15.15	21	J
5.	c4 substituted benzene (15.52)	15.52	24	J
6.	c4 substituted benzene (16.44)	16.44	21	J
7.	c4 substituted benzene (16.71)	16.71	29	J
8. 000119-64-2	Naphthalene, 1,2,3,4-tetrahydro-	17.03	18	JN

BER043 S74

1G

EPA SAMPLE NO.

GW-5

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: ____

Lab Code: 10478 Case No.: BER SAS No.: ____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-005B

Sample wt/vol: 1000 (g/mL) ML Lab File ID: 7\N21256.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: Decanted:(Y/N) N Date Extracted: 08/13/07

Concentrated Extract Volume: 1000 (µl) Date Analyzed: 08/20/07

Injection Volume: 2 (µl) Dilution Factor: 10.00

GPC Cleanup: (Y/N) N pH: ____ Extraction: (Type) SEPF

CONCENTRATION UNITS:

Number TICs found: 15 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	C3 substituted benzene	3.94	300	J
2.	(DEL) Alkane: Branched (4.59)	4.59	360	J
3.	C4 substituted benzene (4.62)	4.62	340	J
4.	C4 substituted benzene (4.71)	4.71	340	J
5.	C4 substituted benzene (4.81)	4.81	270	J
6.	(DEL) Alkane: Cyclic	7.45	280	J
7.	(DEL) Alkane: Branched (7.75)	7.75	300	J
8.	ethyl naphthalene isomer	8.03	390	J
9.	dimethyl naphthalene isomer (8.17)	8.17	800	J
10.	dimethyl naphthalene isomer (8.33)	8.33	790	J
11.	dimethyl naphthalene isomer (8.38)	8.38	530	J
12.	dimethyl naphthalene isomer (8.53)	8.53	510	J
13.	(DEL) Alkane: ~~Straight~~-Chain Branched	8.65	640	J
14.	trimethyl naphthalene isomer (9.18)	9.18	380	J
15.	trimethyl naphthalene isomer (9.37)	9.37	250	J
16.	trimethyl naphthalene isomer (9.45)	9.45	290	J
17.	trimethyl naphthalene isomer (9.58)	9.58	330	J
18.	unknown	10.08	390	J
19.	(DEL) Alkane: Branched (11.04)	11.04	740	J
20.	substituted PNA	13.41	290	J

8/29/07

BER043 S80

1F

EPA SAMPLE NO.

GW-6

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-006A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55454.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 9 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	c3 substituted benzene (13.16)	13.16	17	J
2.	c3 substituted benzene (13.56)	13.56	23	J
3.	c3 substituted benzene (14.62)	14.62	14	J
4.	c4 substituted benzene (14.75)	14.75	22	J
5.	c4 substituted benzene (15.26)	15.26	13	J
6.	c4 substituted benzene (15.53)	15.53	22	J
7.	c4 substituted benzene (15.94)	15.94	14	J
8.	c4 substituted benzene (16.43)	16.43	16	J
9.	c4 substituted benzene (16.71)	16.71	24	J

BER043 S86

1F

EPA SAMPLE NO.

GW-7

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-007A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55475.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/16/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 9 (µg/L or µg/Kg) UG/L

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.		c3 substituted benzene	13.57	25	J
2.		c4 substituted benzene (14.76)	14.76	44	J
3.		c4 substituted benzene (15.26)	15.26	22	J
4.		c4 substituted benzene (15.53)	15.53	40	J
5.		c4 substituted benzene (15.95)	15.95	25	J
6.		c4 substituted benzene (16.44)	16.44	30	J
7.		c4 substituted benzene (16.72)	16.72	44	J
8.	000119-64-2	Naphthalene, 1,2,3,4-tetrahydro-	17.04	22	JN
9.		c5 substituted benzene (17.18)	17.18	21	J

BER043 S92

1F

EPA SAMPLE NO.

GW-8

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-008A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55456.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 7 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1. 000108-87-2	Cyclohexane, methyl-	8.42	22	JN
2.	c3 substituted benzene	13.24	18	J
3.	c4 substituted benzene (14.74)	14.74	53	J
4.	c4 substituted benzene (15.52)	15.52	32	J
5.	c4 substituted benzene (16.44)	16.44	20	J
6.	c4 substituted benzene (16.71)	16.71	31	J
7. 000119-64-2	Naphthalene, 1,2,3,4-tetrahydro-	17.03	32	JN

BER043 S95

1F

EPA SAMPLE NO.

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VOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-009A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55445.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 0 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

BER043 S101

1F

EPA SAMPLE NO.

TRIP BLANK 8/8/200

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-010A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55446.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 0 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

BER043 S104

1F

EPA SAMPLE NO.

STORAGE BLANK 8/9/2

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709206-011A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55447.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 0 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

BER043 S107

1F

EPA SAMPLE NO.

GW-9

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709405-001A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55457.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 8 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1. 000108-87-2	Cyclohexane, methyl-	8.42	16	JN
2.	c3 substituted benzene (13.24)	13.24	30	J
3.	c3 substituted benzene (13.56)	13.56	24	J
4.	c4 substituted benzene (14.74)	14.74	26	J
5.	c4 substituted benzene (15.52)	15.52	21	J
6.	c4 substituted benzene (16.43)	16.43	19	J
7.	c4 substituted benzene (16.71)	16.71	26	J
8. 000090-12-0	Naphthalene, 1-methyl	20.62	17	JN

BER043 S110

1G

EPA SAMPLE NO.

GW-9

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709405-001B

Sample wt/vol: 1000 (g/mL) ML Lab File ID: 7\N21252.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: Decanted:(Y/N) N Date Extracted: 08/16/07

Concentrated Extract Volume: 1000 (µl) Date Analyzed: 08/20/07

Injection Volume: 2 (µl) Dilution Factor: 10.00

GPC Cleanup: (Y/N) N pH: ____ Extraction: (Type) SEPF

Number TICs found: 11 CONCENTRATION UNITS: (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	C3 substituted benzene (3.94)	3.94	73	J
2.	C3 substituted benzene (4.36)	4.36	49	J
3.	C4 substituted benzene (4.61)	4.61	46	J
4.	C4 substituted benzene (5.45)	5.45	66	J
5.	(DEL) Alkane: Straight-Chain (6.89)	6.89	92	J
6. 000090-12-0	Naphthalene, 1-methyl-	7.11	160	JN
7.	(DEL) Alkane: Straight-Chain (7.93)	7.93	140	J
8.	dimethyl naphthalene isomer (8.08)	8.08	140	J
9.	dimethyl naphthalene isomer (8.22)	8.22	150	J
10.	dimethyl naphthalene isomer (8.26)	8.26	100	J
11.	dimethyl naphthalene isomer (8.42)	8.42	56	J
12.	(DEL) Alkane: Straight-Chain (8.96)	8.96	90	J
13.	trimethyl naphthalene isomer	9.07	48	J
14.	(DEL) Alkane: Straight-Chain (9.95)	9.95	86	J
15.	unknown	9.99	59	J
16.	(DEL) Alkane: Straight-Chain (10.89)	10.89	110	J
17.	(DEL) Alkane: Straight-Chain (11.78)	11.78	82	J
18.	(DEL) Alkane: Straight-Chain (12.57)	12.57	68	J
19.	(DEL) Alkane: Straight-Chain (13.14)	13.14	49	J

BER043 S116

1F

EPA SAMPLE NO.

GW-10

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709405-002A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55479.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: not dec. Date Analyzed: 08/16/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

Number TICs found: 0 CONCENTRATION UNITS: (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

BER043 S119

1F

EPA SAMPLE NO.

GW-11

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709405-003A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55480.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: not dec. Date Analyzed: 08/16/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 1 (µg/L or µg/Kg) UG/L

	CAS NUMBER	COMPOUND NAME	RT	EST. CONC.	Q
1.		unknown	3.56	9	J

BER043 S122

1G

EPA SAMPLE NO.

GW-11

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS. INC. Contract: ____

Lab Code: 10478 Case No.: BER SAS No.: ____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709405-003B

Sample wt/vol: 500 (g/mL) ML Lab File ID: 7\N21254.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: Decanted:(Y/N) N Date Extracted: 08/16/07

Concentrated Extract Volume: 500 (µl) Date Analyzed: 08/20/07

Injection Volume: 2 (µl) Dilution Factor: 1.00

GPC Cleanup: (Y/N) N pH: ___ Extraction: (Type) SEPF

CONCENTRATION UNITS:

Number TICs found: 3 (µg/L or µg/Kg) UG/L

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	000149-57-5	Hexanoic acid, 2-ethyl-	5.06	2	JN
2.		dichlorobenzoic acid isomer	9.20	15	J
3.	000057-10-3	n-Hexadecanoic acid	12.95	3	JN

BER043 S125

1F

EPA SAMPLE NO.

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VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER043

Matrix: (soil/water) WATER Lab Sample ID: 0709405-004A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55443.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 0 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

BER043 S128

1F

EPA SAMPLE NO.

B-2 (5-7')

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-001A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30655.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. 10.2 Date Analyzed: 08/13/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

Number TICs found: 10 CONCENTRATION UNITS: (µg/L or µg/Kg) UG/KG

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.		straight-chain alkane (10.14)	10.14	18	J
2.		straight-chain alkane (11.23)	11.23	46	J
3.		branched alkane (11.38)	11.38	21	J
4.		branched alkane (12.01)	12.01	21	J
5.		Straight branched alkane (12.33)	12.33	35	J
6.		Naphthalene,tetrahydro iso (12.59)	12.59	19	J
7.		Naphthalene,tetrahydro iso (12.96)	12.96	15	J
8.		Naphthalene,tetrahydro iso (13.16)	13.16	16	J
9.		straight-chain alkane (13.39)	13.39	19	J
10.		Naphthalene,tetrahydro iso (13.84)	13.84	17	J

8/31/07

BER042 S40

1F

EPA SAMPLE NO.

B-3 (4-4 1/2')

VOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-002A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30679.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. 10.4 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 10.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 10 (µg/L or µg/Kg) UG/KG

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.		branched alkane (7.69)	7.69	2300	J
2.		branched alkane (8.16)	8.16	1200	J
3.		cyclic alkane (8.26)	8.26	1200	J
4.		straight-chain alkane	9.02	2100	J
5.		c3-subs_benzene (9.15)	9.15	800	J
6.		cyclic alkane (9.57)	9.57	1100	J
7.		c3-subs_benzene (9.74)	9.74	860	J
8.		branched alkane (10.21)	10.21	2800	J
9.		c4-subs_benzene (10.41)	10.41	910	J
10.		c4-subs_benzene (11.25)	11.25	1100	J

BER042 S43

1F

EPA SAMPLE NO.

B-4 (10-12')

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-003A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30680.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. 10.8 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

Number TICs found: 8 CONCENTRATION UNITS: (µg/L or µg/Kg) UG/KG

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.		straight-chain alkane (10.15)	10.15	15	J
2.		straight-chain alkane (11.23)	11.23	32	J
3.		branched alkane (12.33)	12.33	28	J
4.		c5-subs_benzene	12.57	16	J
5.		branched alkane (13.4)	13.40	18	J
6.		Naphthalene,dimethyl- isomer (14.15)	14.15	30	J
7.		Naphthalene,dimethyl- isomer (14.3)	14.30	24	J
8.		Naphthalene, dimethyl- isomer	14.33	15	J

BER042 S49

1F

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

EPA SAMPLE NO.

B-5 (0-6')

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-004A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30674.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. 19.7 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 10.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 9 (µg/L or µg/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	c4-subs_benzene (10.38)	10.38	1000	J
2. 1000152-47-3	trans-Decalin, 2-methyl-	10.63	920	JN
3.	c4-subs_benzene (11.08)	11.08	780	J
4.	c4-subs_benzene (11.24)	11.24	2000	J
5.	branched alkane	11.39	1900	BJ
6.	c4-subs_benzene (11.62)	11.62	770	J
7.	c5-subs_benzene (12.11)	12.11	960	J
8.	c5-subs_benzene (12.35)	12.35	1300	J
9.	c5-subs_benzene (12.58)	12.58	1100	J

BER042 S52

1G

EPA SAMPLE NO.

B-5 (0-6')

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-004B

Sample wt/vol: 15 (g/mL) G Lab File ID: 7\N21149.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: 19.7 Decanted:(Y/N) N Date Extracted: 08/13/07

Concentrated Extract Volume: 1000 (µl) Date Analyzed: 08/15/07

Injection Volume: 2 (µl) Dilution Factor: 1.00

GPC Cleanup: (Y/N) N pH: 8.0 Extraction: (Type) PFEX

CONCENTRATION UNITS:

Number TICs found: 16 (µg/L or µg/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1. 000123-42-2	2-Pentanone, 4-hydroxy-4-methyl-	3.68	12000	JNA
2.	C3 substituted benzene	4.23	5900	J
3.	C4 substituted benzene (5.25)	5.25	2300	J
4.	C4 substituted benzene (5.31)	5.31	2900	J
5.	unknown (5.39)	5.39	1900	J
6.	(DEL) Alkane: Straight-Chain	5.65	1900	J
7.	unknown (6.35)	6.35	2000	J
8.	(DEL) Alkane: Branched (6.79)	6.79	1900	J
9.	dimethyldihydroindene isomer	7.28	2200	J
10.	(DEL) Alkane: Branched (7.43)	7.43	3500	J
11.	unknown (7.74)	7.74	2100	J
12. 000090-12-0	Naphthalene, 1-methyl-	8.03	4000	JN
13.	dimethyl naphthalene isomer (9.04)	9.04	2900	JY
14.	dimethyl naphthalene isomer (9.18)	9.18	3000	JY
15.	unknown (11.46)	11.46	2300	J
16. 000529-05-5	Azulene, 7-ethyl-1,4-dimethyl-	11.65	2700	JN
17.	(DEL) Alkane: Branched (11.88)	11.88	12000	J
18.	unknown (12.03)	12.03	2900	J
19.	unknown (12.91)	12.91	2000	J
20.	substituted PNA	13.41	2400	J

BER042 S58

1F

EPA SAMPLE NO.

B-6 (2 1/2-3')

VOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-005A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30681.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. 11.2 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 10 (µg/L or µg/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1. 000108-87-2	Cyclohexane, methyl-	5.13	56	JN
2.	cyclic alkane (6.96)	6.96	70	J
3.	branched alkane (7.22)	7.22	89	J
4.	unknown	8.20	120	J
5.	cyclic alkane (8.3)	8.30	140	J
6.	branched alkane (8.6)	8.60	90	J
7.	branched alkane (8.71)	8.71	95	J
8.	c3-subs_benzene (9.01)	9.01	92	J
9.	c3-subs_benzene (9.19)	9.19	67	J
10.	c3-subs_benzene (9.77)	9.77	62	J

BER042 S64

1F

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

EPA SAMPLE NO.

B-7 (9-10')

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-006A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30675.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. 8.4 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

Number TICs found: 6 CONCENTRATION UNITS: (µg/L or µg/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	Naphthalene,ethyl- isomer	14.05	6	J
2.	unknown	14.08	7	J
3.	Naphthalene,dimethyl- isomer	14.15	16	J
4.	Naphthalene, dimethyl- isomer (14.3)	14.30	15	J
5.	Naphthalene, dimethyl-isomer	14.33	10	J
6.	Naphthalene, dimethyl- isomer (14.53)	14.53	6	J

BER042 S70

1F

VOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

EPA SAMPLE NO.
B-8 (6-10')

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-007A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30661.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. 9.7 Date Analyzed: 08/14/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 5 (µg/L or µg/Kg) UG/KG

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.		unknown	8.11	42	J
2.		branched alkane (9.77)	9.77	35	J
3.		branched alkane (9.8)	9.80	48	J
4.		branched alkane (10.45)	10.45	40	J
5.		branched alkane (10.95)	10.95	110	J

BER042 S73

1G

EPA SAMPLE NO.

B-8 (6-10')

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-007B

Sample wt/vol: 15 (g/mL) G Lab File ID: 7\N21148.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: 9.7 Decanted:(Y/N) N Date Extracted: 08/13/07

Concentrated Extract Volume: 1000 (µl) Date Analyzed: 08/15/07

Injection Volume: 2 (µl) Dilution Factor: 1.00

GPC Cleanup: (Y/N) N pH: 7.0 Extraction: (Type) PFEX

CONCENTRATION UNITS:

Number TICs found: 4 (µg/L or µg/Kg) UG/KG

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	000123-42-2	2-Pentanone, 4-hydroxy-4-methyl-	3.69	5700	JNAY
2.		C3 substituted benzene	4.24	2400	JY
3.		(DEL) Alkane: Branched (5.25)	5.25	1800	JY
4.		(DEL) Alkane: Branched (5.33)	5.33	1400	JY
5.		(DEL) Alkane: Branched (5.45)	5.45	1700	JY
6.		(DEL) Alkane: Branched (5.87)	5.87	3000	J
7.		(DEL) Alkane: Branched (6.27)	6.27	5500	J
8.		(DEL) Alkane: Branched (6.32)	6.32	5000	J
9.		(DEL) Alkane: Branched (6.78)	6.78	1600	J
10.		(DEL) Alkane: Branched (7.24)	7.24	2500	J
11.		(DEL) Alkane: Branched (7.36)	7.36	1600	J
12.		(DEL) Alkane: Straight-Chain (7.71)	7.71	1900	J
13.		(DEL) Alkane: Straight-Chain (8.78)	8.78	2000	J
14.		(DEL) Alkane: Branched (9.41)	9.41	1600	J
15.		(DEL) Alkane: Straight-Chain (9.84)	9.84	2300	J
16.		(DEL) Alkane: Straight-Chain (10.86)	10.86	3100	J
17.		(DEL) Alkane: Straight-Chain (11.83)	11.83	3000	JY
18.		unknown	13.90	1400	JY
19.	007683-64-9	Squalene	16.15	1500	JN

BER042 S79

1F

EPA SAMPLE NO.

B-9 (9-10')

VOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-008A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30676.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. 8.1 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 10 (µg/L or µg/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	branched alkane (11.38)	11.38	15	J
2.	branched alkane (12.01)	12.01	18	J
3.	branched alkane (13.2)	13.20	21	J
4.	unknown (13.58)	13.58	16	J
5.	Naphthalene, 1~~,2,3,4~~-tetrahydro iso	13.84	15	J
6.	branched alkane (13.92)	13.92	25	J
7.	unknown (14.08)	14.08	27	J
8.	Naphthalene, dimethyl- isomer	14.16	15	J
9.	unknown (14.36)	14.36	15	J
10.	unknown (14.42)	14.42	15	J

8/31/07

BER042 S90

1F

EPA SAMPLE NO.

B-10 (4-6')

VOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-009A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30677.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. 17.3 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

Number TICs found: 0 CONCENTRATION UNITS: (µg/L or µg/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

BER042 S93

1F

EPA SAMPLE NO.

B-11 (9 1/2-10 1/2

VOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709204-010A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30678.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. 9.2 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

Number TICs found: 1 CONCENTRATION UNITS: (µg/L or µg/Kg) UG/KG

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	000110-54-3	Hexane	3.05	6	JN

BER042 S96

1F

EPA SAMPLE NO.

FIELD BLANK (SOIL)

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) WATER Lab Sample ID: 0709204-011A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55441.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 0 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

BER042 S99

1F

EPA SAMPLE NO.

TRIP BLANK 8/7/200

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) WATER Lab Sample ID: 0709204-012A

Sample wt/vol: 5 (g/mL) ML Lab File ID: A\A55442.D

Level: (low/med) LOW Date Received: 08/09/07

% Moisture: not dec. Date Analyzed: 08/15/07

GC Column: ZB-624 ID: .18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 0 (µg/L or µg/Kg) UG/L

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

1F

EPA SAMPLE NO.

B-12 6'-8'

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709403-001A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30685.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: not dec. 8.4 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 0 (µg/L or µg/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

BER042 S105

1G

EPA SAMPLE NO.

B-12 6'-8'

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709403-001B

Sample wt/vol: 15 (g/mL) G Lab File ID: 7\N21248.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: 8.4 Decanted:(Y/N) N Date Extracted: 08/15/07

Concentrated Extract Volume: 1000 (µl) Date Analyzed: 08/20/07

Injection Volume: 2 (µl) Dilution Factor: 1.00

GPC Cleanup: (Y/N) N pH: 7.0 Extraction: (Type) PFEX

Number TICs found: 7 CONCENTRATION UNITS: (µg/L or µg/Kg) UG/KG

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	000141-79-7	3-Penten-2-one, 4-methyl-	2.87	1300	JNA
2.	004161-60-8	2-Pentanone, 4-hydroxy-	3.04	680	JNA
3.	000123-42-2	2-Pentanone, 4-hydroxy-4-methyl-	3.26	17000	JNA
4.		C3 substituted benzene	3.72	5300	J
5.		(DEL) Alkane: Branched (4.57)	4.57	250	J
6.		(DEL) Alkane: Branched (4.65)	4.65	310	J
7.		(DEL) Alkane: Branched (4.75)	4.75	280	J
8.		unknown (5.34)	5.34	180	J
9.	000112-84-5	Erucylamide	15.54	340	JN
10.		unknown (15.7)	15.70	150	J

BER042 S108

1F

EPA SAMPLE NO.

B-13 7'-9'

VOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709403-002A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30686.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: not dec. 8.5 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 9 (µg/L or µg/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	unknown	6.84	11	J
2.	branched alkane (9.41)	9.41	22	J
3.	branched alkane (9.52)	9.52	50	J
4.	branched alkane (9.69)	9.69	21	J
5.	branched alkane (9.76)	9.76	33	J
6.	branched alkane (9.79)	9.79	61	J
7.	branched alkane (9.94)	9.94	15	J
8.	branched alkane (10.14)	10.14	17	J
9.	branched alkane (10.21)	10.21	20	J

BER042 S111

1G

EPA SAMPLE NO.

B-13 7'-9'

SEMIVOLATILE ORGANICS ANALYSIS DATA SHEET
TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS. INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709403-002B

Sample wt/vol: 15 (g/mL) G Lab File ID: 7\N21249.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: 8.5 Decanted:(Y/N) N Date Extracted: 08/15/07

Concentrated Extract Volume: 1000 (µl) Date Analyzed: 08/20/07

Injection Volume: 2 (µl) Dilution Factor: 1.00

GPC Cleanup: (Y/N) N pH: 7.0 Extraction: (Type) PFEX

Number TICs found: 6 CONCENTRATION UNITS: (µg/L or µg/Kg) UG/KG

	CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q
1.	000141-79-7	3-Penten-2-one, 4-methyl-	2.87	1300	JNA
2.	004161-60-8	2-Pentanone, 4-hydroxy-	3.04	720	JNA
3.	000123-42-2	2-Pentanone, 4-hydroxy-4-methyl-	3.26	17000	JNA
4.		C3 substituted benzene	3.72	5400	J
5.		(DEL) Alkane: Branched (4.57)	4.57	240	J
6.		(DEL) Alkane: Branched (4.65)	4.65	310	J
7.		(DEL) Alkane: Branched (4.75)	4.75	260	J
8.		unknown	5.35	180	J
9.	000112-84-5	Erucylamide	15.54	340	JN

BER042 S117

1F

EPA SAMPLE NO.

B-14 4'-6'

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709403-003A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30687.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: not dec. 17.3 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 0 (µg/L or µg/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

BER042 S122

1F

EPA SAMPLE NO.

B-15 20'-26'

VOLATILE ORGANICS ANALYSIS DATA SHEET

TENTATIVELY IDENTIFIED COMPOUNDS

Lab Name: H2M LABS, INC. Contract: _____

Lab Code: 10478 Case No.: BER SAS No.: _____ SDG No.: BER042

Matrix: (soil/water) SOIL Lab Sample ID: 0709403-004A

Sample wt/vol: 5 (g/mL) G Lab File ID: V\F30688.D

Level: (low/med) LOW Date Received: 08/14/07

% Moisture: not dec. 20 Date Analyzed: 08/15/07

GC Column: DB-624 ID: 0.18 (mm) Dilution Factor: 1.00

Soil Extract Volume: (µl) Soil Aliquot Volume: 0 (µL)

CONCENTRATION UNITS:

Number TICs found: 0 (µg/L or µg/Kg) UG/KG

CAS NUMBER	COMPOUND NAME	RT	EST.CONC.	Q

Appendix E
Data Summary Tables
With Qualifications

Laboratory Method: EPA Method 8260

PARAMETER	CAS No.	TAGM 4046 Rec. Soil Cleanup Obj. ug/kg	NYS DEC Class GA Ambient Water Qual. Standards Guidance Values ug/L	GW-1 Groundwater ug/L		GW-2 Groundwater ug/L		GW-3 Groundwater ug/L		GW-4 Groundwater ug/L		GW-5 Groundwater ug/L	
				Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
1,1,1,2-Tetrachloroethane	630-20-6	*	5	5	U	5	U	5	U	5	U	5	U
1,1,1-Trichloroethane	71-55-6	800	5	5	U	5	U	5	U	5	U	5	U
1,1,2,2-Tetrachloroethane	79-34-5	600	5	5	U	2	J	5	U	700	J	5	U
1,1,2-Trichloroethane	79-00-5	*	5	5	U	5	U	5	U	170		5	U
1,1-Dichloroethane	75-34-3	200	5	5	U	5	U	5	U	7		5	U
1,1-Dichloroethene	75-35-4	400	5	5	U	1	J	5	U	27		5	U
1,1-Dichloropropene	563-58-6	*	5	5	U	5	U	5	U	5	U	5	U
1,2,3-Trichlorobenzene	87-61-6	*	5	5	UJ	5	UJ	5	UJ	5	UJ	5	UJ
1,2,3-Trichloropropane	96-18-4	400	5	5	UJ	5	U	5	U	5	U	5	U
1,2,4,5-Tetramethylbenzene	95-93-2	*	*	41	J	38	J	5	J	220	J	64	J
1,2,4-Trichlorobenzene	120-82-1	3400	5	5	U	5	U	5	U	5	U	5	U
1,2,4-Trimethylbenzene	95-63-6	*	5	200		110		7		840	J	410	
1,2-Dibromo-3-chloropropane	96-12-8	*	0.04	5	UJ	5	U	5	U	5	U	5	U
1,2-Dibromoethane	106-93-4	*	*	5	U	5	U	5	U	5	U	5	U
1,2-Dichlorobenzene	95-50-1	7900	4.7	5	U	1	J	5	U	58		5	U
1,2-Dichloroethane	107-06-2	100	5	5	U	5	U	5	U	5	U	5	U
1,2-Dichloropropane	78-87-5	*	1	5	U	5	U	5	U	5	U	5	U
1,3,5-Trimethylbenzene	108-67-8	*	5	50		81		5		690	J	350	
1,3-Dichlorobenzene	541-73-1	1600	5	5	U	5	U	5	U	5	U	5	U
1,3-Dichloropropane	142-28-9	300	5	5	U	5	U	5	U	5	U	5	U
1,4-Dichlorobenzene	106-46-7	8500	5	5	U	5	U	5	U	5	U	5	U
2,2-Dichloropropane	594-20-7	*	5	5	U	5	U	5	U	5	U	5	U
2-Butanone	78-93-3	300	50	1	J	5	UJ	5	UJ	5	UJ	30	J
2-Chlorotoluene	95-49-8	*	5	5	U	5	U	5	U	5	U	5	U
2-Hexanone	591-78-6	*	50	5	U	5	U	5	U	5	U	5	U
4-Chlorotoluene	106-43-4	*	5	5	U	5	U	5	U	5	U	5	U
4-Isopropyltoluene	99-87-6	*	5	12		13		5	U	59		18	
4-Methyl-2-Pentanone	108-10-1	1000	50	5	U	5	U	5	U	5	U	5	U
Acetone	67-64-1	200	50	4	J	12		4	J	30		24	
Benzene	71-43-2	60	0.7	1	J	5	U	5	U	34		62	
Bromobenzene	108-86-1	*	5	5	U	5	U	5	U	5	U	5	U
Bromochloromethane	74-97-5	*	5	5	U	5	U	5	U	5	U	5	U
Bromodichloromethane	75-27-4	*	50	5	U	5	U	5	U	5	U	5	U
Bromoform	75-25-2	*	50	5	U	5	U	5	U	5	U	5	U
Bromomethane	74-83-9	*	5	5	U	5	U	5	U	5	U	5	U
Carbon Disulfide	75-15-0	2700	50	5	U	5	U	5	U	5	U	5	U
Carbon Tetrachloride	56-23-5	600	5	5	U	5	U	5	U	5	U	5	U
Chlorobenzene	108-90-7	1700	5	5	U	5	U	5	U	5	U	5	U
Chlorodifluoromethane	75-45-6	*	*	5	UJ	5	UJ	5	UJ	5	UJ	5	UJ
Chloroethane	75-00-3	1900	50	5	U	5	U	5	U	5	U	5	U
Chloroform	67-66-3	300	7	5	U	5	U	5	U	9		5	U
Chloromethane	74-87-3	*	*	5	UJ	5	UJ	5	UJ	5	UJ	5	UJ
cis-1,2-Dichloroethene	156-59-2	*	5	5	U	2100		16		1000	J	3	J
cis-1,3-Dichloropropene	10061-01-5	*	5	5	U	5	U	5	U	5	U	5	U
Dibromochloromethane	124-48-1	*	*	5	U	5	U	5	U	5	U	5	U
Dibromomethane	74-95-3	*	5	5	U	5	U	5	U	5	U	5	U
Dichlorodifluoromethane	75-71-8	*	5	5	UJ	5	UJ	5	UJ	5	UJ	5	UJ
Ethylbenzene	100-41-4	5500	5	78		6		5	U	150		150	
Freon-113	76-14-2	*	*	5	U	5	U	5	U	5	U	5	U
Hexachlorobutadiene	87-68-3	*	0.5	5	U	5	U	5	U	2	J	5	U
Isopropylbenzene	98-82-8	*	5	22		7		5	U	47		35	
m+p Xylene	108-38-3/106-42-3	1200	5	53		8		1	J	520	J	320	
Methyl tert-butyl ether	1634-04-4	*	5	5	U	5	U	5	U	5	U	5	U
Methylene Chloride	75-09-2	100	5	1	J	5	U	5	U	5	U	1	J
Naphthalene	91-20-3	*	10	640	J	150	J	7	J	700	J	670	J
n-Butylbenzene	104-51-8	*	5	5	U	5	U	5	U	5	U	5	U
n-Propylbenzene	103-65-1	*	5	33		9		5	U	94		55	
o-Xylene	95-47-6	1200	5	5	U	23		1	J	290	J	22	
p-Diethylbenzene	105-05-5	*	*	18	J	61		9		390	J	110	
sec-Butylbenzene	135-98-8	*	5	8		6		5	U	44		14	
Styrene	100-42-5	*	5	5	U	5	U	5	U	5	U	5	U
tert-Butylbenzene	98-06-6	*	5	5	U	5	U	5	U	5	U	45	
Tetrachloroethene	127-18-4	1400	5	5	U	4500		5	U	830000		150	
Toluene	108-88-3	1500	5	5	U	5	U	5	U	130		2	J
trans-1,2-Dichloroethene	156-60-5	300	5	1	J	56		5	U	14		5	U
trans-1,3-Dichloropropene	10061-02-6	*	*	5	U	5	U	5	U	5	U	5	U
Trichloroethene	79-01-6	700	5	5	U	11000		26000	B	58000		29	
Trichlorofluoromethane	75-69-4	*	5	5	U	5	U	5	U	5	U	5	U
Vinyl Chloride	75-01-4	200	2	5	U	5	U	5	U	28		5	U

* Not Applicable.

GW-6 Groundwater ug/L		GW-7 Groundwater ug/L		GW-8 Groundwater ug/L		Field Blank Water Aqueous ug/L		Trip Blank 08/06/07 Aqueous ug/L		Storage Blank 08/09/07 Groundwater ug/L	
Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
18		5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	UJ	5	UJ	5	UJ	5	UJ	5	UJ	5	U
5	U	5	UJ	5	U	5	U	5	U	5	U
27	J	25	J	40	J	5	UJ	5	UJ	5	U
5	U	5	U	5	U	5	U	5	U	5	U
140		66		180		5	U	5	U	5	U
5	U	5	UJ	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
1	J	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
84		25		73		5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	UJ	5	UJ	5	J	5	UJ	5	UJ	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
3	J	6		10		5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5		13		5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	4	J	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	UJ	5	UJ	5	UJ	5	UJ	5	UJ	5	U
5	U	11		2	J	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	UJ	5	UJ	5	UJ	5	UJ	5	UJ	5	U
8700		44		2	J	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	UJ	5	UJ	5	UJ	5	UJ	5	UJ	5	U
21		16		39		5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
13		9		19		5	U	5	U	5	U
20		9		18		5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
1	J	5	U	5	U	5	U	5	U	5	U
83	J	29	J	350	J	5	UJ	5	UJ	5	U
5	U	5	U	5	U	5	U	5	U	5	U
23		13		29		5	U	5	U	5	U
10		12		7		5	U	5	U	5	U
54		4	J	48		5	U	5	U	5	U
6		8		13		5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
310	J	25		31		5	U	5	U	5	U
5	U	2	J	5	U	5	U	5	U	5	U
44		5	U	5	U	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
170		5		4	J	5	U	5	U	5	U
5	U	5	U	5	U	5	U	5	U	5	U
2	J	3	J	5	U	5	U	5	U	5	U

GW-9 Groundwater ug/L		GW-10 Groundwater ug/L		GW-11 Groundwater ug/L		Trip Blank 08/13/07 Aqueous ug/L		B-1 Soil ug/kg (dry weight)		FD-1 Soil ug/kg (dry weight)	
Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	UJ	5	UJ	5	UJ	5	UJ	65	U	11	U
5	U	5	UJ	5	UJ	5	U	65	U	11	U
37	J	5	UJ	5	UJ	5	UJ	830	J	11	UJ
5	U	5	U	5	U	5	U	65	UJ	11	UJ
280	J	5	U	5	U	5	U	8200		11	U
5	U	5	UJ	5	UJ	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
2	J	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
280		5	U	5	U	5	U	410		11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	UJ	2	J	5	UJ	5	UJ	65	UJ	11	UJ
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
8		5	U	5	U	5	U	410	J	11	UJ
5	U	5	U	5	U	5	U	65	U	11	U
7	U	5	U	5	U	8		44	J	4	J
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	UJ	11	UJ
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	UJ	5	UJ	5	UJ	5	UJ	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	UJ	5	UJ	5	UJ	5	UJ	65	U	11	U
690	J	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	UJ	5	UJ	5	UJ	5	UJ	65	UJ	11	UJ
20		5	U	5	U	5	U	160		11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	UJ	11	UJ
15		5	U	5	U	5	U	200	J	11	UJ
69		5	U	5	U	5	U	130		11	U
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
340	J	5	UJ	5	UJ	5	UJ	1000	J	5	J
5	U	5	U	5	U	5	U	520		11	U
28		5	U	5	U	5	U	420		11	U
25		5	U	5	U	5	U	65	U	11	U
70		5	UJ	5	UJ	5	U	440	J	11	UJ
9		5	U	5	U	5	U	340	J	11	UJ
5	U	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
25000		140		34		5	U	17	J	14	J
3	J	5	U	5	U	5	U	65	U	11	U
3	J	5	U	5	U	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	U	11	U
250	J	3	J	2	J	5	U	65	U	11	U
5	U	5	U	5	U	5	U	65	UJ	11	UJ
5	U	5	U	5	U	5	U	65	U	11	U

FD-2 Soil ug/kg (dry weight)		Trip Blank 08/06/07 Aqueous ug/L		Storage Blank 08/07/07 Aqueous ug/L		B-2 (5-7') Soil ug/kg (dry weight)		B-3 (4-4.5') Soil ug/kg (dry weight)		B-4 (10-12') Soil ug/kg (dry weight)	
Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	UJ	5	UJ	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
170	J	5	U	5	U	1	J	10000	J	4	J
11	UJ	5	U	5	U	11	UJ	110	UJ	11	UJ
110		5	U	5	U	11	U	20000	J	2	J
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
280		5	U	5	U	11	U	12000	J	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	UJ	5	UJ	5	UJ	11	UJ	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
150	J	5	U	5	U	11	UJ	1000	J	11	UJ
11	U	5	U	5	U	11	U	110	UJ	11	U
11	UJ	5	U	5	U	4	J	60	J	14	J
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	UJ	5	U	5	U	11	UJ	110	UJ	11	UJ
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	UJ	5	UJ	11	UJ	110	UJ	11	UJ
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	UJ	5	UJ	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	3000	J	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	UJ	5	UJ	5	UJ	11	UJ	110	UJ	11	U
11	U	5	U	5	U	11	U	880	J	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	UJ	5	U	5	U	11	UJ	110	UJ	11	UJ
29	J	5	U	5	U	11	UJ	2200	J	11	UJ
11	U	5	U	5	U	11	U	870	J	11	U
11	U	5	U	5	U	11	U	110	UJ	11	UJ
11	UJ	5	U	1	J	11	UJ	110	UJ	11	UJ
19	J	5	UJ	5	UJ	11	UJ	6300	J	8	J
150		5	U	5	U	11	U	5300	J	11	U
57		5	U	5	U	11	UJ	3400	J	11	UJ
11	U	5	U	5	U	11	U	240	J	11	U
67	J	5	U	5	U	11	UJ	1200	J	6	J
120	J	5	U	5	U	11	UJ	1700	J	11	UJ
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	UJ	110	UJ	11	UJ
30	J	5	U	5	U	5	J	4100	J	17	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U
11	U	5	U	5	U	11	U	1300	J	11	U
11	UJ	5	U	5	U	11	UJ	110	UJ	11	U
11	U	5	U	5	U	11	U	110	UJ	11	U

B-5 (0-5') Soil ug/kg (dry weight)		B-5 (2.5-3') Soil ug/kg (dry weight)		B-7 (9-10') Soil ug/kg (dry weight)		B-8 (6-10') Soil ug/kg (dry weight)		B-10 (4-5') Soil ug/kg (dry weight)		B-11 (9.5-10.5) Soil ug/kg (dry weight)	
Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
120	U	11	UJ	11	U	310	J	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	330	J	12	U	11	U
120	U	11	UJ	11	U	16		12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	2	J	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
2000	J	41000	J	11	UJ	3	J	12	UJ	11	UJ
120	UJ	11	UJ	11	UJ	3	J	12	UJ	11	UJ
8500		98000	J	11	U	13		12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	300	J	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
2100		63000	J	11	U	12		12	U	11	U
120	U	11	UJ	11	U	10	J	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	110		12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	UJ	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
480	J	11000	J	11	UJ	11	U	12	UJ	11	UJ
120	U	11	UJ	11	U	11	U	12	U	11	U
54	J	11	UJ	6	J	11	UJ	8	J	12	J
120	U	37	J	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	UJ	11	UJ	11	UJ	11	UJ	12	UJ	11	UJ
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	UJ	11	UJ	11	UJ	11	U	12	UJ	11	UJ
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	6	J	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
220		6600	J	11	U	4	J	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	UJ	11	UJ	11	UJ	19	U	12	UJ	11	UJ
310	J	5200	J	11	UJ	2	J	12	UJ	11	UJ
330		15000	J	11	U	11	U	12	U	11	U
120	UJ	11	UJ	11	UJ	11	U	12	U	11	U
120	UJ	11	UJ	11	UJ	11	UJ	12	UJ	11	UJ
1000	J	85000	J	3	J	18		12	UJ	11	UJ
850		18000	J	11	U	11	U	12	U	11	U
560	J	16000	J	11	UJ	10	J	12	UJ	11	UJ
180		84	J	11	U	5	J	12	U	11	U
310	J	65000	J	11	UJ	11	UJ	12	UJ	11	UJ
440	J	9800	J	11	UJ	3	J	12	UJ	11	UJ
120	U	11	UJ	11	U	11	U	12	U	11	U
120	UJ	11	UJ	11	UJ	11	U	12	UJ	11	UJ
230		89	J	11	U	210000	J	12	U	11	U
120	U	4	J	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U
120	U	11	UJ	11	U	280	J	12	U	11	U
120	U	11	UJ	11	U	11	UJ	12	U	11	U
120	U	11	UJ	11	U	11	U	12	U	11	U

Field Blank Soil Aqueous ug/L		Trip Blank 05/07/07 Aqueous ug/L		B-12 (5-8') Soil ug/kg (dry weight)		B-13 (7-9') Soil ug/kg (dry weight)		B-14 (4-6') Soil ug/Kg (dry weight)		B-15 (20-26') Soil ug/Kg (dry weight)	
Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	UJ	5	UJ	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	2	J	12	UJ	12	UJ
5	U	5	U	11	UJ	11	UJ	12	UJ	12	UJ
5	U	5	U	11	U	7	J	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	6	J	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	UJ	5	UJ	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	UJ	11	UJ	12	UJ	12	UJ
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	4	J	14	J	7	J	7	J
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	UJ	12	UJ	12	UJ
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	UJ	5	UJ	11	UJ	11	UJ	12	UJ	12	UJ
5	U	5	U	11	U	11	U	12	U	12	U
5	U	5	U	11	U	11	U	12	U	12	U
5	UJ	5	UJ	11	U	11	U	12	U	12	U
5	U	5	U

Laboratory Method: EPA Method 8270

PARAMETER	CAS No.	TAGM 4046 Rec. Soil Cleanup Obj. ug/kg	NYS DEC Class GA Ambient Water Qual. Standards ug/L	GW-1 Groundwater ug/L		GW-2 Groundwater ug/L		GW-4 Groundwater ug/L		GW-5 Groundwater ug/L		GW-9 Groundwater ug/L	
				Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
1,2,4-Trichlorobenzene	120-82-1	3400	5	10	U	2000	U	100	U	100	U	100	U
1,2-Dichlorobenzene	95-50-1	7900	4.7	10	U	2000	U	53	J	100	U	100	U
1,3-Dichlorobenzene	541-73-1	1600	5	10	U	2000	U	100	U	100	U	100	U
1,4-Dichlorobenzene	106-46-7	8500	5	10	U	2000	U	19	J	100	U	100	U
2,4,5-Trichlorophenol	95-95-4	*	*	25	U	5000	U	250	U	250	U	250	U
2,4,6-Trichlorophenol	88-06-2	1000	*	10	U	2000	U	100	U	100	U	100	U
2,4-Dichlorophenol	102-83-2	400	1	10	U	2000	U	100	U	100	U	100	U
2,4-Dimethylphenol	105-67-9	*	*	10	U	2000	U	100	U	100	U	100	U
2,4-Dinitrophenol	51-28-5	200 or MDL	*	25	UJ	5000	UJ	250	UJ	250	UJ	250	UJ
2,4-Dinitrotoluene	121-14-2	1000	*	10	U	2000	U	100	U	100	U	100	U
2,6-Dinitrotoluene	606-20-2	1000	5	10	U	2000	U	100	U	100	U	100	U
2-Chloronaphthalene	91-58-7	*	**10	10	U	2000	U	100	U	100	U	100	U
2-Chlorophenol	95-57-8	800	*	10	U	2000	U	100	U	100	U	100	U
2-Methylnaphthalene	91-57-6	36400	*	320		7700		3400		3700		300	
2-Methylphenol	95-48-7	100 or MDL	*	10	U	2000	U	100	U	100	U	100	U
2-Nitroaniline	88-74-4	430 or MDL	*	25	U	5000	U	250	U	250	U	250	U
2-Nitrophenol	88-75-5	330 or MDL	*	10	U	2000	U	100	U	100	U	100	U
3,3'-Dichlorobenzidine	91-94-1	n/a	*	10	U	2000	U	100	U	100	U	100	U
3+4-Methylphenol	108-39-4 / 106-44-5	*	*	10	U	2000	U	100	U	100	U	100	U
3-Nitroaniline	99-09-2	0.5 or MDL	*	25	U	5000	U	250	U	250	U	250	U
4,6-Dinitro-2-methylphenol	534-52-1	*	*	25	U	5000	U	250	U	250	U	250	U
4-Bromophenyl phenyl ether	101-55-3	*	*	10	U	2000	U	100	U	100	U	100	U
4-Chloro-3-methylphenol	59-50-7	240 or MDL	*	10	U	2000	U	100	U	100	U	100	U
4-Chloroaniline	106-47-8	220 or MDL	*	10	U	2000	U	100	U	100	U	100	U
4-Chlorophenyl phenyl ether	7005-72-3	*	*	10	U	2000	U	100	U	100	U	100	U
4-Nitroaniline	100-01-6	*	*	25	U	5000	U	250	U	250	U	250	U
4-Nitrophenol	100-02-7	100 or MDL	*	25	U	5000	U	250	U	250	U	250	U
Acenaphthene	83-32-9	50000	**20	13		710	J	140		150		10	J
Acenaphthylene	208-96-8	41000	*	10	U	2000	U	100	U	100	U	100	U
Aniline	62-53-3	1000	5	10	U	2000	U	100	U	100	U	100	U
Anthracene	120-12-7	50000	**50	5	J	280	J	31	J	38	J	100	U
Benzidine	92-87-5	*	5	10	U	2000	U	100	U	100	U	100	U
Benzo(a)anthracene	56-55-3	224 or MDL	**0.002	10	U	2000	U	100	U	100	U	100	U
Benzo(a)pyrene	50-32-8	61 or MDL	*	10	U	2000	U	100	U	100	U	100	U
Benzo(b)fluoranthene	205-99-2	1100	**0.002	10	U	2000	U	100	U	100	U	100	U
Benzo(g,h,i)perylene	191-24-2	50000	*	10	U	2000	U	100	U	100	U	100	U
Benzo(k)fluoranthene	207-08-9	1100	**0.002	10	UJ	2000	UJ	100	UJ	100	UJ	100	UJ
Benzoic Acid	65-85-0	2700	*	10	U	2000	U	100	U	100	U	100	U
Bis(2-chloroethoxy)methane	111-91-1	*	*	10	U	2000	U	100	U	100	U	100	U
Bis(2-chloroethyl)ether	111-44-4	*	1	10	U	2000	U	100	U	100	U	100	U
Bis(2-ethylhexyl)Phthalate	117-81-7	50000	50	15		2000	U	70	J	100	U	100	U
Butyl benzyl Phthalate	85-68-7	50000	**50	10	U	2000	U	100	U	100	U	100	U
Carbazole	86-74-8	*	*	10	U	2000	U	28	J	38	J	10	U
Chrysene	218-01-9	400	**0.002	10	U	2000	U	100	U	100	U	100	U
Dibenzo(a,h)anthracene	53-70-3	14 or MDL	*	10	U	2000	U	100	U	100	U	100	U
Dibenzofuran	132-64-9	6200	*	12		410	J	91	J	98	J	100	U
Diethyl Phthalate	84-66-2	7100	**50	10	U	2000	U	120		100	U	100	U
Dimethyl Phthalate	131-11-3	2000	*50	10	U	2000	U	100	U	100	U	100	U
Di-n-butyl Phthalate	84-74-2	8100	50	10	U	2000	U	15	J	100	U	100	U
Di-n-octyl Phthalate	117-84-0	50000	**50	10	U	2000	U	100	U	100	U	100	U
Fluoranthene	206-44-0	50000	**50	1	J	2000	U	12	J	13	J	100	U
Fluorene	86-73-7	50000	**50	20		1000	J	190		220		12	J
Hexachlorobenzene	118-74-1	410	0.35	10	U	2000	U	100	U	100	U	100	U
Hexachlorobutadiene	87-68-3	*	5	10	U	2000	U	100	U	100	U	100	U
Hexachlorocyclopentadiene	77-47-4	*	5	10	U	2000	U	100	U	100	U	100	U
Hexachloroethane	67-72-1	*	*	10	U	2000	U	100	U	100	U	100	U
Indeno(1,2,3-c,d)pyrene	193-39-5	3200	**0.002	10	U	2000	U	100	U	100	U	100	U
Isophorone	78-59-1	4400	50*	10	U	2000	U	100	U	100	U	100	U
Naphthalene	91-20-3	13000	**10	250		1300	J	1200		1300		150	
Nitrobenzene	98-95-3	200 or MDL	5	10	U	2000	U	100	U	100	U	100	U
N-Nitrosodi-n-propylamine	621-64-7	*	*	10	U	2000	U	100	U	100	U	100	U
N-Nitrosodiphenylamine	86-30-6	*	**50	10	U	2000	U	100	U	100	U	100	U
Pentachlorophenol	87-86-5	1,000 or MDL	1	25	U	5000	U	250	U	250	U	250	U
Phenanthrene	85-01-8	50000	**50	41		2400		450		510		23	J
Phenol	108-95-1	30 or MDL	1	10	U	2000	U	100	U	100	U	100	U
Pyrene	129-00-0	50000	**50	3	J	2000	U	22	J	24	J	100	U
Pyridine	110-86-1	*	**50	10	U	2000	U	100	U	100	U	100	U

* Not Applicable.
** No Standard Exists. Value is a Guidance Value.

GW-11 Groundwater ug/L		B-1 Soil ug/kg (dry weight)		B-5 (0-6') Soil ug/kg (dry weight)		B-8 (6-10') Soil ug/kg (dry weight)		B-12 (6-8') Soil ug/kg (dry weight)		B-13 (7-9') Soil ug/kg (dry weight)	
Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	1700		360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	520		360	U	360	U
25	U	1100	U	1000	U	920	U	910	U	910	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
25	**UJ**	**1100**	**UJ**	**1000**	**UJ**	**920**	**UJ**	**910**	**UJ**	**910**	**UJ**
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	**24000**	**J**	**19000**		370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
25	U	1100	U	1000	U	920	U	910	U	910	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
25	U	1100	U	1000	U	920	U	910	U	910	U
25	U	1100	U	1000	U	920	U	910	U	910	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	**1500**	**J**	**590**	**J**	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	**UJ**	**430**	**UJ**	**410**	**UJ**	**370**	**UJ**	**360**	**UJ**	**360**	**UJ**
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	170	J	1100		**14000**		220	J	220	J
10	U	430	U	410	U	400		360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	2100		740		370	U	360	U	360	U
10	U	880		290	J	410		620		620	
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	2000		160	J	160	J
10	U	430	U	410	U	910		360	U	360	U
10	U	**660**	**J**	**97**	**J**	370	U	360	U	360	U
10	U	**4900**	**J**	**1200**	**J**	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	390		360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	6700		4300		170	J	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
25	U	1100	U	1000	U	920	U	910	U	910	U
10	U	**15000**	**J**	**3600**	**J**	370	U	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U
10	U	**490**	**J**	**430**	**J**	**79**	**J**	360	U	360	U
10	U	430	U	410	U	370	U	360	U	360	U

Laboratory Method: EPA Method 8081

PARAMETER	CAS No.	TAGM 4046 Rec. Soil Cleanup Obj. mg/kg	Ambient Water Qual. Standards ug/L	GW-1 Groundwater ug/L		B-8 (6-10') Soil ug/kg (dry weight)		B-13 (7-9') Soil ug/kg (dry weight)	
				Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
4,4'-DDD	72-54-8	2.9	0.01	0.1	U	**750**		3.6	U
4,4'-DDE	72-55-9	2.1	0.01	0.1	U	33		3.6	U
4,4'-DDT	50-29-3	2.1	0.01	0.1	U	**1700**		3.6	U
Aldrin	309-00-2	0.041	0.01	0.05	U	1.9	U	1.9	U
Alpha-BHC	319-84-6	0.11	0.05	0.05	U	**1.1**	**J**	1.9	U
Beta-BHC	319-85-7	0.2	0.05	0.05	U	**1.9**	**U**	1.9	U
Chlordane (alpha/gamma)	5103-71-9/5103-74-2	0.54	0.1	0.046	J	**26**	**J**	1.9	U
Delta-BHC	319-86-8	0.3	0.05	0.05	U	1.9	U	1.9	U
Dieldrin	60-57-1	0.044	0.1	0.1	U	**3.4**	**J**	3.6	U
Endosulfan I	959-98-8	0.9	0.1	0.05	U	**2.1**	**J**	1.9	U
Endosulfan II	33212-65-9	0.9	0.1	0.1	U	3.7	U	3.6	U
Endosulfan Sulfate	1031-07-8	1	0.1	0.1	U	3.7	U	3.6	U
Endrin	72-20-8	0.1	0.01	**0.2**	**U**	**4.1**	**U**	3.6	U
Endrin Aldehyde	7421-93-4	*	*	0.1	U	**8.6**	**J**	3.6	U
Endrin Ketone	53494-70-5	*	NA	0.1	U	6.3		3.6	U
Gamma-BHC (Lindane)	58-89-9	0.06	0.05	0.05	U	**4.1**	**J**	1.9	U
Heptachlor	76-44-8	0.1	0.01	0.05	U	4.3		1.9	U
Heptachlor Epoxide	1024-57-3	0.02	0.01	0.05	U	**2.9**	**J**	1.9	U
Methoxychlor	72-43-5	<10 total	35	0.5	U	**370**		19	U
Toxaphene	8001-35-2	*	*	5.0	U	190	U	190	U

* Not Applicable

Laboratory Method: EPA Method 8082

PARAMETER	CAS No.	TAGM 4046 Rec. Soil Cleanup Obj. mg/kg	Ambient Water Qual. Standards ug/L	GW-1 Groundwater ug/L		B-8 (6-10') Soil ug/kg (dry weight)		B-13 (7-9') Soil ug/kg (dry weight)	
				Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
Aroclor-1016	12674-11-2	*	0.1	1.0	U	37	U	36	U
Aroclor-1221	1104-28-2	*	0.1	2.0	U	74	U	73	U
Aroclor-1232	11141-16-5	*	0.1	1.0	U	37	U	36	U
Aroclor-1242	53469-21-9	*	0.1	1.0	U	37	U	36	U
Aroclor-1248	12672-29-6	*	0.1	1.0	U	37	U	36	U
Aroclor-1254	11097-69-1	*	0.1	1.0	U	37	U	36	U
Aroclor-1260	11096-82-5	*	0.1	1.0	U	37	U	36	U

* 1 surface / 10 subsurface

Laboratory Method: EPA Method 6010/7470

PARAMETER	CAS No.	NYS Background Levels mg/kg	TAGM 4046 Rec. Soil Cleanup Obj. mg/kg	Class GA Ambient Water Qual. Standards mg/L	GW-1 Groundwater ug/L		B-1 Soil mg/kg (dry weight)		B-13 (7-9') Soil mg/kg (dry weight)		B-8 (6-10') Soil mg/kg (dry weight)	
					Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit	Concentration	Detection Limit
Aluminum, Al	7429-90-5	33,000	SB	*	28900		3750		8900		8970	
Antimony, Sb	7440-36-0	*	SB	**.003	11.1		1.1	J	1.0	J	1.0	J
Arsenic, As	7440-38-2	3 - 12	7.5 or SB	0.025	2.0	U	0.55		0.22	U	0.22	U
Barium, Ba	7440-39-3	15 - 600	300 or SB	1	1160		34.7		97.9		84.4	
Beryllium, Be	7440-41-7	0 - 1.75	0.16 or SB	**0.003	3.4		0.18	U	0.27	U	0.29	U
Cadmium, Cd	7440-43-9	0.1 - 1	1 or SB	0.01	27.3		0.46	U	0.16	U	0.12	U
Calcium, Ca	7440-70-2	130 - 35,000	SB	*	269000		3090		1580		1670	
Chromium, Cr	7440-47-3	1.5 - 40	10 or SB	0.05	75.7		11.3		24.2		22.1	
Cobalt, Co	7440-48-4	2.5 - 60	30 or SB	*	116		4.9		10.4		9.6	
Copper, Cu	7440-50-8	1 - 50	25 or SB	0.2	1710		54.1	J	18.6	J	17.6	J
Iron, Fe	7439-89-6	2,000 - 550,000	2,000 or SB	0.3	225000		11800		16900		16200	
Lead, Pb	7439-92-1	*	SB	0.025	652		50.2		2.7		3.5	
Magnesium, Mg	7439-95-4	100 - 5,000	SB	**35	41000		1510		4120		3900	
Manganese, Mn	7439-96-5	50 - 5,000	SB	0.3	7970		100		223		320	
Mercury, Hg	7439-97-6	0.001 - 0.2	0.1	0.002	11.2	J	0.23		0.018	U	0.030	
Nickel, Ni	7440-02-0	0.5 - 25	13 or SB	*	275		12.5		19.5		16.4	
Potassium, K	9/7/7440	8,500 - 43,000	SB	*	36500	J	997		4620		3130	
Selenium, Se	7782-49-2	0.1 - 3.9	2 or SB	0.01	6.5	UJ	0.43	U	0.33	U	0.34	U
Silver, Ag	7440-22-4	*	SB	0.05	5.0	J	0.31	U	0.18	U	0.17	U
Sodium, Na	7440-23-5	6,000 - 8,000	SB	20	553000		312		178		145	
Thallium, Tl	7440-28-0	*	SB	**.004	2.2	U	0.55		0.24	U	0.24	U
Vanadium, V	7440-62-2	1 - 300	150 or SB	*	111		10.4		28.8		25.3	
Zinc, Zn	7440-66-6	9 - 50	20 or SB	0.3	12300		159		37.7		33.1	

SB Site Background.
* Not Applicable
** No Standard Exists. Value is a Guidance Value.

Appendix-A
Professional Resumes

Nicholas A. Andrianas, P.E.
Principal

Mr. Andrianas is a professional engineer with over 40 years of experience, solving complex environmental regulatory issues, conducting environmental site investigations, and designing and constructing treatment systems for sanitary and industrial wastewater, contaminated groundwater, soil and industrial waste.

Mr. Andrianas was founder and president of **NAC CONSULTANTS, INC.** for over 25 years. Prior to founding **NAC CONSULTANTS, INC.** he was the principal in charge of the environmental and remediation section of a New York based, 200 person, international environmental consulting firm. Mr. Andrianas directs the environmental compliance programs, industrial air and waste treatment system projects, soil and groundwater investigations, feasibility studies, decommissioning and remediation of commercial, industrial, institutional, and municipal facilities. He has completed hundreds of investigation and cleanup projects under the supervision of the United States Environmental Protection Agency and various State and local environmental regulatory agencies. He is a recognized expert in wastewater treatment system design, environmental investigations, remediation and environmental compliance. He served on the technical and regulatory committees of the National Association of Chemical Recyclers for over five years, developing draft environmental statutes, tracking and interpreting environmental regulations, and developing remediation programs. He has taught programs in environmental compliance, treatment system design and operation, and chemical management for major transportation and manufacturing companies. Mr. Andrianas has prepared professional engineer's expert reports and has been accepted by the United States Tax Court as an expert witness in environmental engineering and compliance. Mr. Andrianas is a member of the National Society of Professional Engineers is a licensed professional engineer in New York and Michigan.

- **Education**

  Bachelor of Science in Environmental Engineering Rensselaer Polytechnic Institute (1979)
  Master of Science studies in Environmental Engineering, Rensselaer Polytechnic Institute

- **Employment Experience**

  2020 to Present
  Owner/Principal, Nicholas A. Andrianas, P.E.

  1995 to 2020
  President, NAC CONSULTANTS, INC.

  1983 to 1995
  Principal, Eder Associates

Project Experience

- *Photofinishing Company*

  Directed the environmental compliance program for a national film and paper photofinishing company. Tracked wastewater discharge system operation and monitoring for 23 facilities and compiled corporate permit records. Negotiated permit compliance programs, developed process monitoring programs for environmental and QC purposes, evaluated wastewater treatment equipment and developed programs to maintain and upgrade equipment to meet metals, nitrogen, and COD effluent limits. Prepared Best Management Practices and Spill Prevention Control and Countermeasures Plans for US facilities. Reviewed wastewater discharge permits and negotiated permit limits and wastewater discharge surcharge programs.

- *Airline Fixed Base Operator (FBO) International Company*

  Reviewed the onsite sanitary septic wastewater treatment system condition and obtained variance for existing system. Performed the sampling and analyses for the closure of the existing septic system and leaching system. Designed the new sanitary wastewater sewer system for the facility charter airline terminal to meet NYSDOT and local sewer authority requirements. Provided oversight of the sewer system construction and connection to the public sewer. Designed the oil-water separator pretreatment systems and double walled washwater collection systems for the facility aircraft hangars.

- *State Psychiatric Center*

  Directed the environmental assessment of waste disposal and sanitary wastewater surface impoundment discharge areas at the Pilgrim State Psychiatric center, where hazardous materials were placed in landfills and surface impoundments and contamination was discharged to the soil and groundwater. The investigation included the installation of shallow and deep groundwater monitoring wells, test pit excavation and sampling of waste materials, and sampling of soil and groundwater. Developed the remedial alternatives and prepared the cost estimates to close the landfill and waste disposal areas, and to prepare the waste management areas for redevelopment as a warehouse distribution center.

- *Fasteners manufacturer*

  Developed the remediation plan to address contamination emanating from surface impoundments and an on-site landfill where hazardous wastes were discharged. Directed the operation and monitoring of the groundwater remediation system installed to remove solvents from groundwater prior to discharging the treated water to surface water. Prepared reports required by New York State Department of Environmental Conservation to document soil and groundwater conditions at the site.

- *Armament Manufacturer*

  Operated the wastewater pretreatment system at the facility for 15 months. Modified the treatment system chemistry to improve effluent quality and meet City of Glen Cove discharge permit limits. Negotiated nitrogen and heavy metals permit limits and prepared monthly reports to the City. Obtained permits for bulk chemical, petroleum, and metal finishing operations. Prepared and implemented spill prevention program and waste reduction program. Prepared and obtained air emission permits for scrubbers and process exhausts.

- *Precision Screw Machine Manufacturing*

  Prepared wastewater discharge monitoring reports. Prepared waste minimization reports to reduce wastewater discharge quantities and treatment system costs. Operated and maintained the plant's wastewater pretreatment system and directed the upgrades to the system to reduce metals, cyanide, nitrogen, fluoride and COD. Prepared and obtained permits for hazardous materials storage. Prepared facility emergency response and Spill Prevention Control Plans for hazardous materials. Performed compliance audits and trained personnel in treatment system operation and hazardous materials management.

- *Brookhaven National Laboratories*

  Directed the installation of soil vapor extraction wells and air sparging wells designed to remediate the groundwater plume contaminated with petroleum products and solvents. Directed the preparation of the summary report including lithologic information, well construction details, development data, and well permits.

- *Abandoned waste disposal site*

  Directed the investigation and remediation of a 4 acre parcel of vacant land that had been used as an illegal dump for construction and demolition (C&D) debris, plus petroleum contaminated soil. Prepared work plans for review by the State and conducted a soil boring investigation to map the extent of contaminated soil. Developed a remediation plan approved by the State. The remedial plan consisted of excavating and separating C&D debris from soil contaminated with semi-volatile and volatile petroleum hydrocarbons and recycling the soil as aggregate in an asphalt sub-base material. Provided field oversight of the soil excavation, separation of C&D debris and processing of the contaminated soil into asphalt sub-base pursuant to the State beneficial use permit.

- *Brookhaven National Laboratories*

  Directed the installation of the recovery wells designed to capture the groundwater plume contaminated with tritium. Directed the preparation of the reports on the lithologic information, well construction details, development data, and well permits to document the installation. Provided oversight of the soil vapor extraction system to remediate contaminated soil.

- *Electronics manufacturer*

  Designed and managed the construction of the plant's wastewater pretreatment system. Negotiated permit limits for nitrogen and heavy metals with the sewer agency based on effluent dilution calculations for the sewer district's outfall. Developed monitoring programs for the process operations and treatment system. Designed the hazardous materials storage areas.

- *USEPA Superfund Site, New Jersey*

  Directed the preparation of the feasibility study for groundwater extraction, treatment, and recharge remedy for this site. Directed the field pilot scale treatability and recharge tests, and the design and construction of the groundwater remediation system using extraction wells and two granular activated carbon (GAC) treatment units.

- *Wire and cable manufacturer*

  Directed the site investigation to evaluate soil and groundwater conditions at this closed industrial facility. Reviewed the results of the sampling programs and directed the preparation of the data review and development of the site-specific cleanup levels using a risk assessment for the property. Reviewed the remediation work completed at the site and prepared the petition to delete the property from the New York State Registry of Inactive Waste Disposal Site.

- *Martin Marrietta Astronautics Group*

  Directed the feasibility study prepared to develop, evaluate and recommend soil and groundwater remediation alternatives for over twenty waste disposal areas at this 5200 acre Federal Superfund site. The study included bench scale and pilot tests of technologies to immobilize and treat inorganic and organic contamination in groundwater. Groundwater extraction methods including shallow and deep wells and interceptor trenches were designed and evaluated for recovery effectiveness and implementability.

- *Pump and motor rebuild*

  Directed the investigation, feasibility study and the design, construction and operation of the remediation system for soil and groundwater contaminated with solvents. The remediation system consisted of an air sparging system to strip solvents from the groundwater and a soil vapor extraction system to recover solvents from the soil column. The remediation system was installed in lieu of a conventional pump and treat system that would have generated a substantial volume of contaminated treatment system residue.

  Negotiated the permit for the plant's wastewater discharge for heavy metals, nitrogen, and VOCs from the rebuild operations, designed the pretreatment system, and obtained permits for hazardous materials storage and parts cleaning processes.

  Designed and operated the remediation system to capture soil vapors contaminated with VOC solvents. Designed the sub-slab depressurization system to prevent VOC vapor migration to indoor air.

- *International Delivery/Carrier Company*

  Reviewed the onsite sanitary wastewater treatment system condition and prepared report on upgrade requirements for facility expansion. Performed the sampling and analyses for the closure of the existing septic system and leaching system. Designed the new sanitary wastewater treatment system and provided oversight of the new system construction. Designed the hazardous materials storage systems and petroleum product storage tanks and dispensing systems.

- *Former Military Aircraft Manufacturing Facility*

  Designed and provided construction oversight of sub-slab depressurization systems (SSDS) to prevent solvent VOC soil vapor migration to the redeveloped former aircraft manufacturing facility with over 1 million square feet. Provide operation and maintenance services of the SSDS.

- *Semiconductor Manufacturer*

  Prepared the design of the hazardous materials storage areas and tank systems. Designed the process ventilation systems for over 50 new facility operations. Prepared the industrial wastewater engineering reports for onsite wastewater treatment and discharge. Reviewed the onsite sanitary wastewater treatment system condition and prepared report on upgrade requirements for facility expansion. Performed the sampling and analyses for the closure of the existing septic system and leaching system. Designed the wastewater treatment system to meet groundwater standards for metals and nitrogen, and designed the new sewer connection to the public sewer system. Designed the industrial wastewater pretreatment system to meet sewer discharge standards and designed the sewer connection system.

- *Former Machine Shop*

  Designed the sub-slab depressurization system (SSDS) to control VOC vapor migration at the redeveloped facility. Performed system start-up testing and prepared annual performance reports for submittal to NYSDEC.

- *Subscription-Based Streaming Service Studio Facility*

  Designed the sub slab chemical resistant vapor barrier for the building constructed on geopiers and designed the SSDS to control VOC vapor migration at the 5 studio facility constructed on a former printing facility.

- *Military Subcontractor Composites Manufacturing Facility*

  Designed the sanitary sewage pump station and force main for the manufacturing facility. Obtained permits to connect to public sewer works and provided oversight of the system construction.

- *Former Institutional Facility*

  Provide the routine stormwater pollution prevention engineering services for the redevelopment of this 100 acre site. Oversee sampling, analyses and remediation of the site wide stormwater drainage structures.

- *Starlight Park, Bronx, New York NYC Design and Development Corp.*

  Prepared the Field Sampling Plan to characterize the soil as part of the park redevelopment for the "BRONX RIVER GREENWAY: STARLIGHT PARK" PHASE II, directed the soil characterization sampling and analyses program at the Bronx River Park remediation site, reviewed the sampling results for non-hazardous and hazardous soil, determined soil disposal options and provided oversight of the hazardous soil excavation removal and off-site disposal.

Justin Halpin

631-461-5371

Education: **B.A. Major in Earth Science**

B.S **Minor in Chemistry -** ***Dowling*** **College** Oakdale, New York.

Work Experience:

January 2008- August 2014- Berninger Environmental Inc.

WRS/Champion Environmental Services August 2014- November 2024)

Owner/Principal of BEI Consulting LLC January 2025 - current

Environmental Cunsultant/Scientist

- Experience working with and identifying underground utility structures with private locating companies and New York 811 to interpret distribution maps and read blueprints
- Project Manager/Scientist with over fifteen (15) years' experience directing and conducting Environmental Site Assessments (ESA's)
- Preparation of monthly/quarterly monitoring reports for NYS DEC spill cases
- Experience with managing a diverse staff of Union Workers, Scientists, Geologists and Engineers and overseeing Geoprobing field work with regard to sub-surface investigation and monitoring well installations
- Underground Storage Tank (UST) Work Plans and Closure Reports
- Experience working in conjunction with various government agencies including NYS DEC, NYS DOH, SCDOH, NCDOH, NYC DEP managing client compliance needs with regard to the specific jurisdiction governing oversight
- Soil boring logs (AGI), monitoring well logs, air sampling logs
- Phase 2 and Phase 3 Environmental Site Assessment/Remediation reporting including: Investigation Work Plans (IWP), Interim Remedial Measures (IRM), Remedial Action Plans (RAP), Operation Monitoring and Maintenance Plans (OM M) Final Engineering Reports (FER), Site Management Plans (SMP) etc.
- Experience with Phase 1 Environmental Assessment Reports and Environmental Impact Statements (EIS)
- Slug Test Analysis (STA) Geoprobe Computer Software
- Portable Gas Chromatograph (GC) testing in-field
- Performance of commercial environmental due diligence for various industries (Real Estate, Gas retail, Dry-Cleaning industry, etc.)
- Scaled site map construction, remedial design, As-built drawings

- Laboratory analysis; i.e.: spreadsheets and graphing of results with regard to NYS DEC drinking water standards
- In-field site surveying for construction of groundwater flow/gradient maps and scaled site plans
- TEA analysis (field and lab); nutrient analysis (Sulfates, Nitrates, Iron etc.)
- Groundwater, soil and air sampling collection and handling of Chain of Custody's
- Experience with various NYS DEC Technical Guidance's (DER-10, DER-13) and EPA regulations
- Achieved numerous closures for NYS DEC Spill Response Program sites
- Managing of transportation and disposal projects and performing of waste characterization for facility approvals
- Designing of pump and treat systems – dewatering operations and groundwater treatment systems including (Carbon treatment, ion-exchange, particulate and reverse osmosis) in order to treat a wide array of chemical constituents
- Water treatment system troubleshooting for submersible pump systems, vapor extraction systems, sub-slab depressurization systems and in-situ chemical injection remediation projects
- Experience with site restoration, wetland restoration and surveying (flora, fauna analysis)