

Species Status Assessment

Class: Bivalvia
Family: Unionidae
Scientific Name: *Ligumia recta*
Common Name: Black sandshell

Species synopsis:

Ligumia recta belongs to the subfamily Ambleminae and the tribe Lampsilini, which includes 17 extant and 6 likely extirpated New York species of the genera *Actinonaias*, *Epioblasma*, *Lampsilis*, *Leptodea*, *Ligumia*, *Obovaria*, *Potamilus*, *Ptychobranhus*, *Toxolasma*, *Truncilla*, and *Villosa* (Haag 2012, Graf and Cummings 2011). *L. recta* is grouped in the *Ligumia* genus, which means seed or pod of a legume, referring to its long and pod-shaped structure. The species name *recta*, which means straight, refers to the elongated shape and parallel dorsal margins (Watters et al. 2009).

Since 1970, *L. recta* has been found in ten New York waterbodies. This species has three ranges in New York: the Allegheny basin, the Erie-Ontario basin, and the St. Lawrence-Champlain basin, reflecting its three routes of entry into the state. *L. recta* lives in large creeks, rivers, and large shallow lakes (Strayer and Jirka 1997).

In New York, *L. recta* is ranked as imperiled, although it is apparently secure throughout its range (NatureServe 2013). In North America, approximately $\frac{2}{3}$ to $\frac{3}{4}$ of native mussel species are extinct, listed as endangered or threatened, or are in need of conservation status (Williams *et al.* 1993, Stein *et al.* 2000). While population trends in New York are unknown, based on sparse historical information, it is assumed that they too are declining due to a myriad of environmental stressors.

Status

a. Current and Legal Protected Status

- i. Federal None Candidate? No
- ii. New York None – Species of Greatest Conservation Need

b. Natural Heritage Program Rank

- i. Global G4G5 – Apparently Secure / Secure
- ii. New York S2S3 – Imperiled / Vulnerable Tracked by NYNHP? Yes

Other Rank:

IUCN Red List Category: Near threatened

American Fisheries Society Status: Special Concern (1993)

Status Discussion:

L. recta is widespread in eastern and central U.S. and Canada, occurring from the Great Lakes basin south into Mississippi River drainage, to Louisiana and in some Gulf Coast drainages. There have been some declines throughout its range. Lately it has become increasingly more difficult to find with many occurrences represented by few individuals, often without evidence of recruitment. Declines appear to be localized and the species continues to maintain a wide distribution with many stable populations (NatureServe 2013).

II. Abundance and Distribution Trends

a. North America

i. Abundance

X declining ___ increasing ___ stable ___ unknown

ii. Distribution:

X declining ___ increasing ___ stable ___ unknown

Time frame considered: _____

b. Regional

i. Abundance

___ declining ___ increasing ___ stable X unknown

ii. Distribution:

___ declining ___ increasing ___ stable X unknown

Regional Unit Considered: Northeast

Time Frame Considered: _____

c. Adjacent States and Provinces

CONNECTICUT Not Present X No data _____

MASSACHUSETTS Not Present X No data _____

NEW JERSEY Not Present X No data _____

ONTARIO Not Present _____ No data _____

i. Abundance

___ declining ___ increasing X stable ___ unknown

ii. Distribution:

___ declining ___ increasing X stable ___ unknown

Time frame considered: 2003-2013

Listing Status: S3

PENNSYLVANIA Not Present _____ No data _____

i. Abundance

____ declining ____ increasing ____ stable X unknown

ii. Distribution:

____ declining ____ increasing ____ stable X unknown

Time frame considered: _____

Listing Status: _____ S3S4 _____ SGCN? No

QUEBEC Not Present _____ No data _____

i. Abundance

____ declining ____ increasing X stable ____ unknown

ii. Distribution:

____ declining ____ increasing X stable ____ unknown

Time frame considered: _____

Listing Status: _____ S3 _____

VERMONT Not Present _____ No data _____

i. Abundance

 X declining ____ increasing ____ stable ____ unknown

ii. Distribution:

 X declining ____ increasing ____ stable ____ unknown

Time frame considered: _____

Listing Status: _____ S1 - Endangered _____ SGCN? Yes

d. NEW YORK

No data _____

i. Abundance

 X declining ___increasing ___stable ___unknown

ii. Distribution:

 X declining ___increasing ___stable ___unknown

Time frame considered: _____

Monitoring in New York.

As part of a State Wildlife Grant, NYSDEC Region 8 Fisheries and Wildlife staff is conducting a baseline survey of tributaries in central and western New York for native freshwater mussels 2009 – 2017.

Trends Discussion:

Trends for New York populations are difficult to determine as most historic data comes from opportunistic naturalist collections, as opposed to more comprehensive baseline surveys. For example, mussels were documented for the first time in 50 of the 106 streams surveyed to date by the Southern Lake Ontario mussel inventory project (Mahar and Landry 2013). This is because many of these streams had never before been surveyed for mussels, not because mussel distribution has dramatically increased. In North America, approximately 2/3 to 3/4 of native mussel species are extinct, listed as endangered or threatened, or are in need of conservation status (Williams *et al.* 1993, Stein *et al.* 2000). Based on New York's Natural Heritage S-rank, sparse historical data, and the plight of North America's freshwater mussels, it is assumed that trends are declining due to a myriad of environmental stressors.

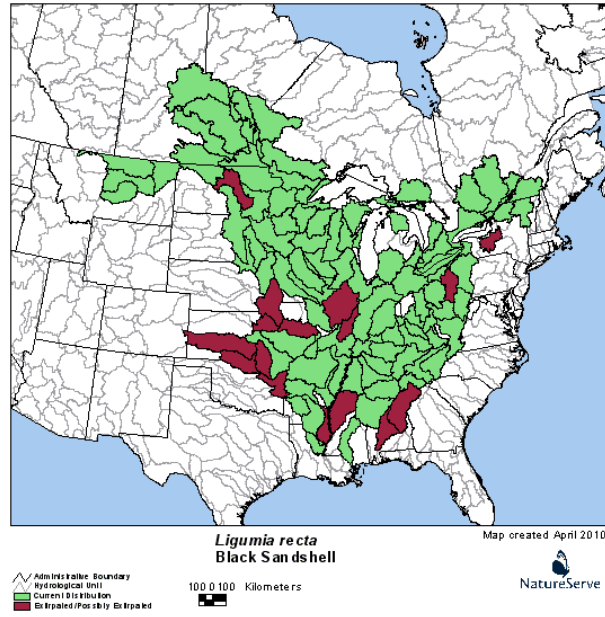


Figure 1. *L. recta* distribution in North America (NatureServe 2013).

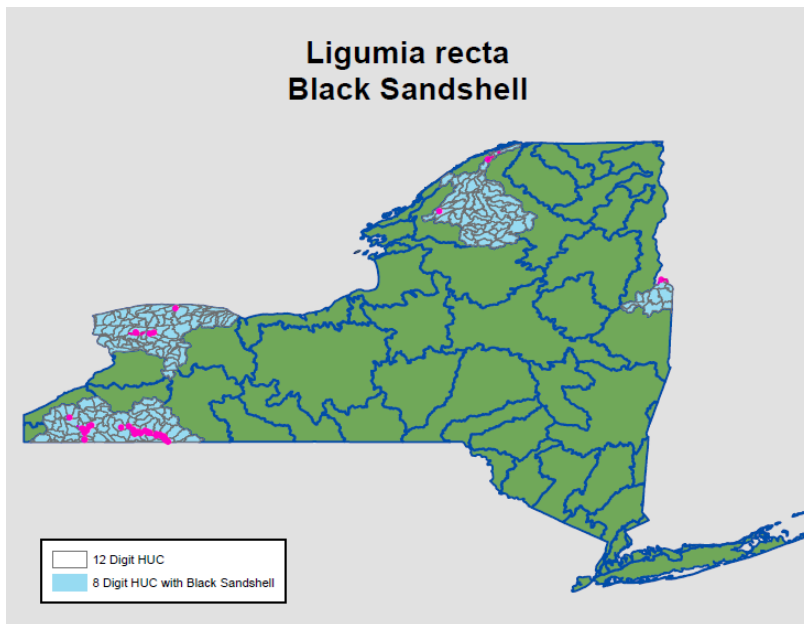


Figure 2. New York distribution of *L. recta*, post-1970 (Mahar and Landry 2013, Harman and Lord 2010, The Nature Conservancy 2009, New York Natural Heritage Program 2013, White et al. 2011).

III. New York Rarity, if known:

Historic	<u># of Animals</u>	<u># of Locations</u>	<u>% of State</u>
prior to 1970	<u>Unknown</u>	<u>At least 8 waterbodies</u>	<u>10 of 56 HUC watersheds</u>
prior to 1980	<u>_____</u>	<u>_____</u>	<u>_____</u>
prior to 1990	<u>_____</u>	<u>_____</u>	<u>_____</u>

Details of historic occurrence:

L. recta has three ranges in New York: the Allegheny basin, the Erie-Ontario basin, and the St. Lawrence-Champlain basin, reflecting its three routes of entry into the state. In the Allegheny basin, it was historically known from the Allegheny River and Cassadaga Creek (Strayer and Jirka 1997). It has been recorded from Lakes Erie and Oneida, and in rivers and large creeks in between these lakes. In the St. Lawrence-Champlain basin, there are scattered records from the St. Lawrence River and its tributary, the Grass River, Lake Champlain, and the Poultney River (Strayer and Jirka 1997), which flows into Lake Champlain.

Current	<u># of Animals</u>	<u># of Locations</u>	<u>% of State</u>
	<u>>127 live</u>	<u>10 waterbodies</u>	<u>6 of 56 HUC 8 watersheds</u>

Details of current occurrence:

L. recta currently exists in eight waterbodies in New York State (not counting waterbodies in which only highly weathered shells “fossil shells” were found) (Figure 2).

A 2009 survey of the Allegheny basin by The Nature Conservancy found 80 live *L. recta*, with populations at most sites in the Allegheny River between Portville and Salamanca, and in the lower sections of Conewango and Cassadaga Creeks, and in Oswayo Creek. Shells were also found in Olean Creek. The greatest catches (up to 3.3 per hr) were in the Allegheny River near and upstream of Olean, NY. *L. recta* were considered viable at 13% (5 of 38 sites) of the sites where they were found (The Nature Conservancy 2009).

In 2010, an old weathered shell of this species was found in lower Oak Orchard Creek, a first record for this species in the Western Lake Ontario basin. In the Erie basin, more than 35 live specimens were confirmed at four locations on Tonawanda Creek between the towns of Pendleton and Alabama (Mahar and Landry 2013, NY Natural Heritage Program 2013). Additional recent occurrences include the Poultney River at the southern end of Lake Champlain, and in the St. Lawrence basin the Grass River and the Oswegatchie River (fossil) (Strayer and Jirka 1997, NY Natural Heritage Program 2013). Although widespread in New York, *L. recta* is usually seen in small numbers. It has not recently been found at historic sites between Rochester and Syracuse (Mahar and Landry 2013).

In 2011 and 2012 Zanatta, Burlakova, Karatayev et al. surveyed 6 locations (9 sites) in Lake Erie, and 54 sites at 33 locations in Lake Ontario (2012 only), and did not found *L. recta* at any of NYS

locations.) Three live and 10 shells of *L. recta* were collected in 2012 at Chittenango Creek, a tributary of Oneida Lake (Bridgeport, Onondaga Co., State Hw 31 crossing), and 9 live *L. recta* were found in Tonawanda Creek in 2011-2013 (Burlakova, Karatayev, Karatayev, unpublished data).

New York’s Contribution to Species North American Range:

% of NA Range in New York	Classification of New York Range
<input type="checkbox"/> 100 (endemic)	<input type="checkbox"/> Core
<input type="checkbox"/> 76-99	<input checked="" type="checkbox"/> Peripheral
<input type="checkbox"/> 51-75	<input type="checkbox"/> Disjunct
<input type="checkbox"/> 26-50	Distance to core population:
<input checked="" type="checkbox"/> 1-25	<u>700 km</u>

IV. Primary Habitat or Community Type:

1. Medium River; Low Gradient; Assume Moderately Buffered (Size 3+ rivers); Warm
2. Medium River; Low Gradient; Assume Moderately Buffered (Size 3+ rivers); Transitional Cool
3. Medium River; Low-Moderate Gradient; Assume Moderately Buffered (Size 3+ rivers); Warm
4. Summer-stratified Monomictic Lake

Habitat or Community Type Trend in New York:

Declining Stable Increasing Unknown

Time frame of decline/increase: _____

Habitat Specialist? Yes No

Indicator Species? Yes No

Habitat Discussion:

L. recta is typically found in medium to large rivers (Cummings and Mayer 1992, McMurry et al. 2012, Metcalfe-Smith et al. 2005, Watters et al. 2009). It can also be found in large creeks, and some large, shallow lakes (Strayer and Jirka 1997). It is commonly cited that this species is associated

with gravel or firm sand substrate (Parmalee 1967, Watters et al. 2009, McMurry et al. 2012, Metcalfe-Smith et al. 2005), but can occasionally be found in mud, silt, or cobbles (Parmalee and Bogan 1998, Metcalfe-Smith et al. 2005, NatureServe 2013). It is typically found in locations with strong current (riffles or raceways) in water depths from several inches to six feet or more (Metcalfe-Smith et al. 2005, Parmalee and Bogan 1998).

V. New York Species Demographics and Life History

- Breeder in New York**
 - Summer Resident**
 - Winter Resident**
 - Anadromous**
- Non-breeder in New York**
 - Summer Resident**
 - Winter Resident**
 - Catadromous**
- Migratory only**
- Unknown**

Species Demographics and Life History Discussion:

Upstream males release sperm into the water. Females downstream take up the sperm with incoming water. Fertilization success may be related to population density, with a threshold density required for any reproductive success to occur. Eggs are fertilized within the female. Like nearly all North American mussels, this species must parasitize an often specific vertebrate host to complete its life cycle. It is suspected that some mussel populations are not recruiting because their hosts no longer occur with them. Once released by the female, glochidia must acquire a suitable host or die, usually within 24-48 hours. After attaching to a suitable host, glochidia encyst, usually at the fish's gills or fins and receive food and dispersal. Once the glochidia metamorphose into juveniles, they drop from the host. If they land in suitable habitat, they will burrow into the substrate, where they may remain for several years (Watters et al. 2009).

In the adult form, freshwater mussels are basically sessile; movement is limited to a few meters of the lake or river bottom. The only time that significant dispersal can take place is during the

parasitic phase. Infected host fishes can transport the larval unionids into new habitats, and can replenish depleted populations with new individuals. Dispersal is particularly important for genetic exchange between populations. Dispersal is likely to be a slow process for mussels which use resident fishes with limited home ranges as their hosts (COSEWIC as cited in NatureServe 2013).

It has a periodic life history strategy, characterized by moderate to high growth rate, low to intermediate life span, age at maturity, and fecundity, but generally smaller body size than opportunistic species. Most species are long-term brooders. This life history strategy is considered an adaptation to allow species to persist in unproductive habitats or habitats that are subject to large-scale, cylindrical environmental variation or stress (Haag 2012).

Individuals of this species may live to be at least 30 years old, with female shell characteristics becoming apparent in the third year. *L. recta* is thought to be bradytictic, with eggs found in August, glochidia developing by September, and overwintering in the female until the following July (Watters et al. 2009).

L. recta glochidia have been shown to transform on rock bass (*Ambloplites rupestris*), central stoneroller (*Campostoma anomalum*), banded killifish (*Fundulus diaphanus*), redbreast sunfish (*Lepomis auritus*), green sunfish (*Lepomis cyanellus*), pumpkinseed (*Lepomis gibbosus*), orangespotted sunfish (*Lepomis humilis*), bluegill (*Lepomis macrochirus*), longear sunfish (*Lepomis megalotis*), largemouth bass (*Micropterus salmoides*), white perch (*Morone americana*), rosyface shiner (*Notropis rubellus*), yellow perch (*Perca flavescens*), white crappie (*Pomoxis annularis*), black crappie (*Pomoxis nigromaculatus*), sauger (*Stizostedion canadense*), and walleye (*Stizostedion vitreum*) (Watters et al. 2009). Khym and Layzer, (2000), found 10 times more juveniles metamorphosing on sauger than any other fish tested, including largemouth bass, bluegill, white crappie, black crappie. Other potential hosts include American eel (*Anguilla rostrata*) and common carp (*Cyprinus carpio*) (Watters et al. 2009).

VI. Threats:

Agricultural Runoff

The watershed upstream of Olean in the Allegheny River where some of New York's largest populations of *L. recta* has been found is primarily a forested landscape (New York State Landcover 2010). However, cultivated cropland is present adjacent to the Allegheny River in portions of this area, as well as in important downstream habitat between Olean and Salamanca, and in streams with *L. recta* populations in the Conewango basin, Grass River, and Tonawanda Creek. Recently harvested forest land was also prevalent upstream of and adjacent to *L. recta* habitat on the Grass River (New York State Landcover 2010). Aquatic habitats lacking vegetated buffers of adequate width are threatened by runoff from urban areas, roads, lawns, and agricultural land (Gillis 2012). If best management practices are not closely adhered to, mussel habitat adjacent to wood harvest or agricultural land is subjected to pesticide, fertilizer, and silt/sediment runoff. During recent mussel surveys in western and central New York, it has been documented that sufficient vegetated riparian

buffers are often lacking along known mussel streams (Mahar and Landry 2013), indicating that runoff is a major threat to resident mussel populations.

The presence of pesticides and fertilizers in our rural watersheds is nearly ubiquitous (Haag 2012). And because pesticides and their associated surfactants adsorb onto sediment particles, sedimentation may act as a vector for their transport into the aquatic system (Haag 2012). Mussels are more sensitive to pesticides than many other animals (Watters et al. 2009). Although effects of pesticides are species-specific, sub-lethal levels of PCBs, DDT, malathion, and other compounds inhibit respiratory efficiency and accumulate in the tissues. Atrazine and permethrin at sublethal concentrations reduced juvenile growth (Bringolf et al. 2007a, 2007b) and environmental levels of atrazine altered mussel movement and aggregation behavior (Flynn and Spellman 2009). Pesticides can affect mussels in many ways, but the full range of long-term effects remains unknown (Haag 2012).

Fertilizer runoff is also a concern. High inputs of nitrogen from fertilizers can cause increases in ammonia in the water and the substrate, leading to direct toxicity for a wide range of mussel species. Mussels, especially in their early life stages, are more sensitive to un-ionized ammonia than other organisms, and high sensitivity is seen across a range of species and life histories (Haag 2012). In addition, ammonia adsorbs to sediment particles, resulting in higher nitrogen concentrations in the substrate than in the overlying water. The nitrogen present in the interstitial spaces in the substrate is thought to result in juvenile mortality and to prevent recruitment by some mussel species (Strayer and Malcom 2012). Studies have suggested decreasing sediment loads entering aquatic systems as the best way to decrease the impact of numerous stressors for mussels in general (Roley et al. 2012).

Runoff from Developed Land

All eight New York waterbodies that host *L. recta* populations are intermittently bordered by interstate highways, state routes, and/or local roads. *L. recta* habitat receives stormwater runoff from the municipalities of Olean, Portville, Salamanca and Allegany on the Allegheny River, Jamestown through a tributary to Conewango Creek, and Madrid on the Grass River. Residential development along Tonawanda Creek also contributes to the non-point source pollution received into *L. recta* habitat. These waterbodies are likely threatened by stormwater runoff containing metals and road salts (Gillis 2012). Mussels are particularly sensitive to heavy metals, more so than many other animals used in toxicological tests (Keller and Zam 1991). Low levels of metals may interfere with the ability of glochidia to attach to the host (Huebner and Pynnonen 1992), suggesting that U.S. EPA ambient water quality criteria may not adequately protect mussels from toxic metals (Wang et al. 2011). In addition, increases in salinity from the runoff of salt used for clearing roads in winter may be lethal to glochidia and juvenile mussels (Keller and Zam 1991, Liquori and Insler 1985, Pandolfo et al. 2012). Based on these studies, the U.S. EPA's ambient water quality criterion for acute chloride exposures may not be protective of all freshwater mussels (Pandolfo et al. 2012).

Treated Waste Water

L. recta habitat treated waste water from the municipalities of Olean and Portville on the Allegheny River, Jamestown through a tributary to Conewango Creek, and Madrid on the Grass River (SPDES 2007). Recent studies show that mussel richness and abundance decrease with increased proximity to sewage effluent (Wildenberg 2012). The input of biomaterial from waste water treatment plants depletes dissolved oxygen levels, negatively impacting mussels. Ammonia from wastewater treatment plants has been found to be toxic to glochidia (Goudraeu et al. 1993) and at sub-lethal exposure, adult mussels exhibit decreased respiratory efficiency (Anderson et al. 1978). Endocrine disruptors from pharmaceuticals are also present in municipal sewage effluents and are increasingly common in rivers and lakes (Haag 2012). In mussels, chronic exposure to estrogenic compounds in effluents caused feminization of male mussels, but these individuals did not produce eggs, suggesting major disruption of reproductive function (Gagne et al. 2011). The long term effects of these compounds on mussels are unknown (Haag 2012). It should be noted that in the Susquehanna Basin, Harman and Lord (2010) found no evidence that waste water treatment plants were responsible for reductions in mussel species of greatest conservation need.

Flood Control Projects

In the Upper Allegheny basin, large stretches of *L. recta* habitat has been found within or adjacent to stream reaches shaped by levee and/or floodwall flood control projects in Olean and Salamanca on the Allegheny River, and in Portville on the Allegheny River and Oswayo Creek (“New York State Flood Protection” 2013). These structures confine larger rivers, preventing the river from inundating its natural floodplains and wetlands to minimize flood damage. Additionally, many smaller streams have been channelized and bermed by landowners and highway departments to protect farm fields and other structures. Channelization and dredging associated with flood control projects are catastrophic to mussels and have been implicated in the decline of some populations (Watters et al. 2009). The result of these projects is altered seasonality of flow and temperature regimes, increased stream velocities, unstable substrates, changed patterns of sediment scour and deposition, including streambank erosion, altered transport of particulate organic matter (the food base for mussels), and a general degradation of stream habitat (Benke 1999, Yeager 1993, Nedeau 2008).

Other Habitat Modifications

In addition to channelization and regular channel dredging for maintenance of flood control structures, other ecosystem modifications such as instream work associated with bridge replacement, gravel mining, and vegetation removal kill mussels and destroy their habitat. For example, dredging for vegetation removal has been shown to remove up to 23% of mussels in spoils (Aldridge 2000). Further evidence for disruption was provided by mussel surveys adjacent to approximately 20 river miles of Conewango Creek that had been channelized and straightened in the first half of the 20th century. The resulting “dredge” had no riffle or run habitat and sites just below and above this channelized section contained few or no mussels (The Nature Conservancy 2009). Although limited in geographic scope these habitat modification activities have long term impacts on mussels and their distribution (Aldridge 2000).

Invasive Species

Invasive mussels are a potential threat to *L. recta* populations in the Conewango basin and in the Poultney River. Zebra mussels (*Dreissena polymorpha*) are present in *L. recta* habitat in the lower reaches of Cassadaga and Conewango Creeks (The Nature Conservancy 2009). They have also been detected in the Champlain Canal, approximately 2 miles downstream of *L. recta* habitat in the Poultney River (iMapInvasives 2013). Zebra mussels have been repeatedly cited as a threat to native mussel populations and have effectively eliminated native mussels from the western basin of Lake Erie (Strayer and Jirka 1997, Watters et al. 2009). En masse, Dreissenids outcompete native mussels by efficiently filtering food and oxygen from the water. They reduce reproductive success by filtering native mussel male gametes from the water column and they can foul the shells of the native mussels to the point that their valves can no longer open. In heavily invested areas, they may transform a habitat by hardening the substrate, such that dislodged mussels are not able to rebury (USFWS 1994).

In addition, the regular use of lampricide in the Poultney River to reduce sea lamprey populations was identified by Natural Heritage Program as a possible threat to *L. recta* populations in this waterbody (NY Natural Heritage Program 2013).

Climate Change

In a recent assessment of the vulnerability of at-risk species to climate change in New York, Schesinger et al. (2011) ranked this species as “extremely vulnerable.” This indicates that abundance and/or range extent within New York is extremely likely to substantially decrease or disappear by 2050.

Impoundments – Range wide

Range wide, impoundments likely contributed to the reduced distribution that we see today. Vaughn and Taylor (1999) observed a mussel extinction gradient with a gradual, linear increase in mussel species richness and abundance with increasing distance downstream from impoundments. Species and their hosts that require shallow, oxygenated, fast-flowing water quickly are eliminated. Continuously cold water from both increased water depth upstream of the dam and dam discharges downstream of the dam may prevent reproduction. Impoundment increases silt load and eutrophication, resulting in changes in the fish fauna, and therefore the availability of hosts. Dams represent distributional barriers to fish hosts, and therefore to the mussels themselves. The zoogeographic patterns of several species suggest a dam-limited range. Dams also act as sediment traps, often having many feet of silt and debris caught on their upstream side. These areas generally are without mussels. Below the dam, the tailwaters often have dense mussel beds, as these reaches are the only areas left that still have oxygenated, fast moving water. This is exemplified by the distribution of beds in the lower Muskingum River, Ohio (Stansbery and King 1983, ESI 1993c).

In addition, improperly sized and poorly installed or poorly maintained culverts have impacts similar to dams in that they fragment habitat, preventing the movement by host fish, and effectively

isolating mussel populations. And because culverts are located at nearly every road-stream intersection, there is the potential for landscape level fragmentation of mussel habitat.

Are there regulatory mechanisms that protect the species or its habitat in New York?

No **Unknown**

Yes

Mussel habitats receive some generic protection under several New York State regulations (NYCRR) promulgated under the authority of the New York Environmental Conservation Law (ECL), specifically Part 608 of the NYCRR: Use and Protection of Waters, and Part 617 of the NYCRR: State Environmental Quality Review (SEQR). Part 608 provides protection of some mussel habitats by regulating and requiring environmental review of the modification or disturbance of any “protected stream”, its bed or bank, and removal of sand, gravel or other material from its bed or banks (608.2 Disturbance of Protected Streams). This does not provide adequate protection of mussels and their habitats as it only protects streams or particular portions of a streams for which there has been adopted by NYSDEC or any of its predecessors any of the following classifications or standards: AA, AA(t), A, A(t), B, B(t) C(t), or Streams designated (t)(trout) also include those more specifically designated (ts)(trout spawning). Mussel habitats may also receive some additional protections as the construction, repair, breach or removals of dams, and the excavation and placement of fill in navigable waters are subject to regulation and environmental review under Part 608, 608.3 and 608.5 respectively. Under part 608, projects requiring a permit can be conditioned by NYSDEC to include best management practices, such as sediment and erosion protections. Through the review process, these projects can also be modified to reduce impacts in order to meet permit issuance standards.

Under Part 608, protection of unlisted species of mussels is general and relatively limited. More importantly, Class C and D waters with mussels do not receive protection under these regulations. A significant portion of the New York’s mussel resources occur within Class C and D waters. An additional but not insignificant gap in protection occurs because agricultural activities consisting of the crossing and re-crossing of a protected stream by livestock or wheeled farming equipment normally used for traditional agricultural purposes or of withdrawing irrigation water in a manner which does not otherwise alter the stream, are exempt from these regulations and environmental review.

Water quality certifications required by Section 401 of the Federal Water Pollution Control Act, Title 33 United States Code 1341(see subdivision (c) of this Section) may provide protection for freshwater mussels and their habitats from some activities that would potentially have adverse impacts by regulating construction or operation of facilities that may result in any discharge into navigable waters. Water quality certifications set water quality-related effluent limitations, water quality standards, thermal discharge criteria, effluent prohibitions and pretreatment standards for projects on navigable waters.

The State Environmental Quality Review (SEQR, Part 617 NYCRR) may also protect mussels and their habitats by requiring the consideration of environmental factors into the existing planning, review and decision-making processes of state, regional and local government agencies for activities that require discretionary approval. SEQR requires the preparation of an Environmental Impact Statement, including an alternatives analysis, for those activities that may result in a substantial adverse change in ground or surface water quality; a substantial increase in potential for erosion, flooding, leaching or drainage problems; the removal or destruction of large quantities of vegetation or fauna; substantial interference with the movement of any resident or migratory fish or wildlife species; impacts on a significant habitat area; substantial adverse impacts on a threatened or endangered species of animal or plant, or the habitat of such a species; other significant adverse impacts to natural resources; or, a substantial change in the use, or intensity of use, of land including agricultural, open space or recreational resources, or in its capacity to support existing uses.

New York State has numerous laws and regulations that both directly or indirectly protect waters of the state (mussel habitats) including regulations governing direct discharges to surface and groundwater, storm water, agricultural activities, pesticides, flood control, and dams. Without these regulations, mussels would certainly be in worse shape; however, most of these generic protections are not adequate in scope or specific enough to mussel threats to protect the mussel resources of New York State.

Describe knowledge of management/conservation actions that are needed for recovery/conservation, or to eliminate, minimize, or compensate for the identified threats:

- Priority conservation efforts for this species should focus on, but not be limited to, Tonawanda Creek and the Allegheny River near and upstream of Olean.
- Modify marine mussel regulations or the definition of protected wildlife in NYCRR to clarify that freshwater mussels are protected under ECL. Current regulations could be interpreted that freshwater mussels may only be protected as shellfish without a season within the Marine District.
- Through landowner incentive programs or regulation, riparian buffers, particularly those that also provide shade, should be added/maintained/widened, along agricultural fields, subdivisions, and along major roads to decrease the levels of nitrogen, pesticides, sediment, heavy metals, and salts from entering these aquatic systems, as well as to moderate water temperature. Studies have suggested decreasing sediment loads entering aquatic systems as the best way to decrease the impact of numerous stressors for mussels in general (Roley and Tank 2012).
- Require all state agencies to maintain appropriate vegetative buffers along streams, rivers and lakes on state-owned or state managed properties.

- Coordinate with local wastewater treatment facilities to improve ammonia removal of treated discharge. This has been documented as a threat to Unionids at multiple life stages, and therefore needs to be addressed (Gillis 2012).
- Establish a protocol where as DEC staff work closely with flood control management to reduce or impacts to native mussels during maintenance flood control projects.
- Establish a protocol whereas DEC staff work closely with state and local highway departments to reduce impacts to native mussels during maintenance and construction projects.
- Replace culverts that disrupt aquatic habitat connectivity to allow for passage of small fish species.
- Develop and implement a comprehensive monitoring strategy that identifies protocols, including locations and specific intervals, for regular monitoring of known mussel populations to detect assess trends and detect dangerous declines.

- Within the Great Lakes and Champlain watersheds, lamprey control efforts should consider specific, potentially adverse, impacts to native freshwater mussels when determining methods, including selection of lampricide formulations and concentrations.

Lampricide treatment managers should use caution when using the combination of TFM and niclosamide in streams with known mussel populations and every effort should be made to maintain lampricide concentrations at or near the MLC for sea lamprey to minimize the risk to this important faunal group (Boogaard 2006).

- NYSDEC should consider sensitivity of freshwater mussels to specific pollutants in the establishment and setting of water quality standards and TMDLs for waters containing freshwater mussels. A Total Maximum Daily Load (TMDL) specifies the maximum amount of a pollutant that a waterbody can receive and still meet water quality standards. TMDLs account for all contributing sources (e.g. point and nonpoint sources, and natural background levels), seasonal variations in the pollutant load, and incorporate a margin of safety that accounts for unknown or unexpected sources of the pollutant. In essence, a TMDL defines the capacity of the waterbody to absorb a pollutant and still meet water quality standards. The Clean Water Act requires states to identify waterbodies that do not meet water quality standards after application of technology-based effluent limitations. For these "impaired waters," states must consider the development of alternative strategies, including TMDLs, for reducing the pollutants responsible for the failure to meet water quality standards.
- Mussel sensitivity to particular pollutants should be considered or addressed in the regulation of wastewater and stormwater discharges to groundwater and surface waters, State Pollutant Discharge Elimination Systems (SPDES). This should be reflected in effluent limitations for discharges, including discharges from P/C/I facilities

(Private/Commercial/Industrial), CAFO facilities (Concentrated Animal Feeding Operations), High Volume Hydraulic Fracturing Discharges, and Wastewater treatment plants, etc. Discharges whose receiving waters have mussels, particularly those with known populations of mussels listed as Endangered, Threatened, Special concern or SGCN, should be carefully reviewed for potential impacts to mussels. For example, deleterious levels of ammonia (a component of many types of discharges) and molluscicides (a commonly used water treatment chemical in discharged water) should not be permitted.

The Comprehensive Wildlife Conservation Strategy (NYSDEC 2006) includes recommendations for the following actions for freshwater mussels:

Habitat management:

- Manage areas of important mussel populations by controlling degradation factors (e.g. Controlling livestock access, point source or non-point source pollution, flow alteration, etc.)
- Develop methods to improve and restore freshwater bivalve habitat.

Habitat research:

- Conduct research to determine habitat parameters necessary for good populations of each species of species-at-risk listed mussels.
- Research flow requirements of freshwater bivalves and model the effects of flow changes both in volume and timing.
- Research all parameters of mussel habitat requirements including temperature, substrate, fish, flow, food, etc.

Habitat restoration:

- Restore degraded habitat areas to allow for recolonization or reintroduction of listed mussels.

Invasive species control:

- Develop a monitoring/control plan that includes measures to detect invasive species problematic to freshwater bivalves in all New York watersheds and actions that will be taken to control them before they become threats.
- Conduct research on control of exotic bivalve species that compete with native mussels and exotic crustaceans or fish which may prey on them.

Life history research:

- Research effects of pesticides and other chemicals, including ammonia, on all life stages of freshwater bivalves: sperm/egg, glochidia, larva, adults.
- Research potential interbreeding between *Alasmidonta varicosa* and *Alasmidonta marginata* and, if occurring, evaluate the potential threat to *A. varicosa* population integrity.
- Determine fish hosts for species where this is not known for populations living in New York.
- Research population dynamics of listed mussel species including connectivity of populations or subpopulations and genetic distinctness of populations or subpopulations.

- Determine or confirm breeding phenology and habitat conditions necessary for successful breeding for listed mussels (e.g., mussel density, pop. level of fish host, temp, flow).

Modify regulation:

- Modify marine mussel regulations to be clearer that freshwater mussels are protected under ECL.

New regulation:

- Ban the importation of fish that feed on freshwater mollusks (e.g., black carp).
- Require inclusion of all stages of freshwater mussels in testing for approval of new pesticides in New York.

Other action:

- Develop an outreach program to private landowners through the Landowner Incentive Program to educate the public about freshwater mussel protection and initiate projects to prevent or repair impacts from land use on mussels.
- Increase regional permit control of development and highway projects that may impact native mussels.
- Develop standard monitoring/survey protocols for development projects in all watersheds in New York.
- Evaluate threats to mussels in each New York watershed and prioritize areas for actions to address the threats.
- Research the best survey methods both for detection of rare species and evaluation of population status and trends.
- Begin evaluation of members of the family Sphaeridae (fingernail clams) for inclusion into the species at risk list.

Population monitoring:

- Conduct population estimates of species-at-risk listed mussel species in NY
- Conduct surveys to determine distribution of species-at-risk listed mussel species in NY.

Regional management plan:

- Incorporate freshwater mussel goals and objectives into regional water quality and fish management plans and policies.

Relocation/reintroduction:

- Where appropriate, reintroduce listed mussels into appropriate habitat within their historic range.

Statewide management plan:

Incorporate freshwater mussel goals and objectives into statewide water quality and fish management plans and policies.

VII. References

- Aldridge, D. C. (2000). The impacts of dredging and weed cutting on a population of freshwater mussels (Bivalvia: Unionidae). *Biological Conservation*, 95(3), 247-257.
- Anderson, K. B., Sparks, R. E., and Paparo, A. A. (1978). Rapid assessment of water quality, using the fingernail clam, *Musculium transversum*: Final Report. University of Illinois, Urbana. 130p.
- Benke, A.C. (1990). A perspective on America's vanishing streams. *Journal of the N. American Benthological Society*: 9: 77-88
- Boogaard, Michael A., *Acute Toxicity of the Lampricides TFM and Niclosamide to Three Species of Unionid Mussels*, USGS Open-File Report 2006-1106, April 2006.
- Bringolf, R. B., Cope, W. G., Eads, C. B., Lazaro, P. R., Barnhart, M. C., and Shea, D. (2007). Acute and chronic toxicity of technical-grade pesticides to glochidia and juveniles of freshwater mussels (unionidae). *Environmental Toxicology and Chemistry*, 26(10), 2086-2093.
- Bringolf, R. B., Cope, W. G., Barnhart, M. C., Mosher, S., Lazaro, P. R., and Shea, D. (2007). Acute and chronic toxicity of pesticide formulations (atrazine, chlorpyrifos, and permethrin) to glochidia and juveniles of *Lampsilis siliquoidea*. *Environmental Toxicology and Chemistry*, 26(10), 2101-2107.
- COSEWIC. 2003. COSEWIC assessment and status report on the kidneyshell *Ptychobranhus fasciolaris* in Canada. Committee on the Status of Endangered Wildlife in Canada. Ottawa, Canada. 32 pp.
- Cummings, K. S., and Mayer, C. A. 1992. *Field guide to freshwater mussels of the Midwest* (p. 194). Champaign, Illinois: Illinois Natural History Survey.
- Flynn, K., and Spellman, T. (2009). Environmental levels of atrazine decrease spatial aggregation in the freshwater mussel, *Elliptio complanata*. *Ecotoxicology and Environmental Safety*, 72(4), 1228-1233.
- Gagné, F., Bouchard, B., André, C., Farcy, E., and Fournier, M. (2011). Evidence of feminization in wild *Elliptio complanata* mussels in the receiving waters downstream of a municipal effluent outfall. *Comparative Biochemistry and Physiology Part C: Toxicology and Pharmacology*, 153(1), 99-106.
- Gillis, P. L. (2012). Cumulative impacts of urban runoff and municipal wastewater effluents on wild freshwater mussels (*Lasmigona costata*). *Science of the Total Environment*, 431, 348-356.
- Goudraeu, S. E., Neves, R. J., and Sheehan, R. J. (1993). Effects of wastewater treatment plant effluents on freshwater mollusks in the upper Clinch River, Virginia, USA. *Hydrobiologia*, 252(3), 211-230.

- Graf, D. and K. Cummings. (2011). MUSSELp Evolution: North American Freshwater Mussels. The MUSSEL Project. The University of Wisconsin. http://mussel-project.uwsp.edu/evol/intro/north_america.html
- Haag, W. R. (2012). *North American freshwater mussels: natural history, ecology, and conservation*. Cambridge University Press.
- Harman, W.N. and P.H. Lord (2010). Susquehanna Freshwater Mussel Surveys, 2008-2010. Final report submitted to New York State Department of Environmental Conservation. SUNY Oneonta Biological Field Station. Oneonta, NY. 24 pp + plus appendix.
- Huebner, J. D., and Pynnönen, K. S. (1992). Viability of glochidia of two species of Anodonta exposed to low pH and selected metals. *Canadian Journal of Zoology*, 70(12), 2348-2355.
- iMapInvasives: an online mapping tool for invasive species locations. 2013. The Nature Conservancy. Available at: iMapInvasives.org. [Date accessed: 03,06,2013].
- Keller, A. E., and Zam, S. G. (1991). The acute toxicity of selected metals to the freshwater mussel, *Anodonta imbecilis*. *Environmental Toxicology and Chemistry*, 10(4), 539-546.
- Khym, J.R. and J.B. Layzer. 2000. Host fish suitability for glochidia of *Ligumia recta*. *American Midland Naturalist* 143: 178-184.
- Liquori, V. M., and Insler, G. D. (1985). Gill parasites of the white perch: Phenologies in the lower Hudson River. *New York Fish and Game Journal*, 32(1), 71-76.
- Mahar, A.M. and J.A. Landry. (2013). State Wildlife Grants Final Report: Inventory of Freshwater Mussels in New York's Southeast and Southwest Lake Ontario Basins, 2008-2013. New York State Department of Environmental Conservation. Avon, NY. *In progress*.
- Metcalf-Smith, J., A. MacKenzie, I. Carmichael, and D. McGoldrick. (2005). Photo Field Guide to the Freshwater Mussels of Ontario. St. Thomas Field Naturalist Club. St. Thomas, ON, 60pp.
- McMurray, S.E., Faiman, J.S., Roberts, A., Simmons, B., and Barnhart, C.M. 2012. *A guide to Missouri's freshwater mussels*. Missouri Department of Conservation, Jefferson City, Missouri.
- Natural Heritage Program Element Occurrences [ARC/INFO coverages] (2013). New York Natural Heritage Program, Albany, NY. Available: NYS Department of Environmental Conservation Master Habitat Data Bank's Data Selector.
- NatureServe. 2013. NatureServe Explorer: An online encyclopedia of life [web application]. Version 7.1. NatureServe, Arlington, Virginia. Available <http://www.natureserve.org/explorer>. (Accessed: February 12, 2013).

- Nedeau, E.J. 2008. *Freshwater Mussels and the Connecticut River Watershed*. Connecticut River Watershed Council, Greenfield, Massachusetts. xviii+ 132 pp.
- New York State Department of Environmental Conservation. (2006). *New York State Comprehensive Wildlife Conservation Strategy*. Albany, NY: New York State Department of Environmental Conservation.
- New York State Flood Control Project Details and Maps. (2013). Retrieved from *NYS Dept. of Environmental Conservation* website: <http://www.dec.ny.gov/lands/59934.html>. N.p., n.d. Web. 03. June 2013.
- New York State Landcover, Version 1. [SDE raster digital data] (2010). National Gap Analysis Program. Moscow, Idaho. Available: NYS Department of Environmental Conservation Master Habitat Data Bank's Data Selector.
- Northeastern Aquatic Habitat Classification System (NAHCS) GIS map for streams and rivers, [vector digital data] (2010). US Environmental Protection Agency, the US Geological Survey, and The Nature Conservancy Eastern Conservation Science. Boston, MA. Available: NYS Department of Environmental Conservation Master Habitat Data Bank's Data Selector.
- Pandolfo, T. J., Cope, W. G., Young, G. B., Jones, J. W., Hua, D., and Lingenfelter, S. F. (2012). Acute effects of road salts and associated cyanide compounds on the early life stages of the unionid mussel *Villosa iris*. *Environmental Toxicology and Chemistry*, 31(8), 1801-1806.
- Parmalee, P.W. and A.E. Bogan. 1998. *The Freshwater Mussels of Tennessee*. University of Tennessee Press: Knoxville, Tennessee. 328 pp.
- Parmalee, P. W. (1967). The fresh-water mussels of Illinois. *Popular Science Series*, 8.
- Roley, S.S. 2012. The influence of floodplain restoration on stream ecosystem function in an agricultural landscape. (unpublished doctoral dissertation). University of Notre Dame, Notre Dame, Indiana. Submitted for publishing with Tank, J.L.
- Roley, S. S., J. Tank, and M. A. Williams (2012), Hydrologic connectivity increases denitrification in the hyporheic zone and restored floodplains of an agricultural stream, *J. Geophys. Res.*
- Schlesinger, M.D., J.D. Corser, K.A. Perkins, and E.L. White. 2011. Vulnerability of at-risk species to climate change in New York. New York Natural Heritage Program, Albany, NY.
- Stansbery, D. H., and King, C. C. (1983). Management of Muskingum River mussel (unionid mollusk) populations. Final Report to the U.S. Department of Commerce, and the Ohio Department of Natural Resources. *Ohio State University Museum of Zoology Reports*. 79 p.

- State Pollutant Discharge Elimination System (SPDES)- New York State [vector digital data]. (2007). Albany, New York: New York State Department of Environmental Conservation. Available: <http://gis.ny.gov/gisdata/inventories/details.cfm?dsid=1010&nysgis=>
- Stein, B. A., Kutner, L. S., Hammerson, G. A., Master, L. L., and Morse, L. E. 2000. State of the states: geographic patterns of diversity, rarity, and endemism. *Precious heritage: the status of biodiversity in the United States*. Oxford University Press, New York, 119-158.
- Strayer, D.L. and K.J. Jirka. 1997. The Pearly Mussels of New York State. New York State Museum Memoir (26): 113 pp., 27 pls.
- Strayer, D.L. and Malcom, H.M. 2012. Causes of recruitment failure in freshwater mussel populations in southeastern New York. *Ecological Applications* 22:1780–1790.
- The Nature Conservancy (2009). *Freshwater Mussel (Unionidae) Distributions, Catches, and Measures of their Viability across the Catches, and Measures of their Viability across the Allegheny River Basin in New York*. Report submitted New York State Department of Environmental Conservation. The Nature Conservancy, Central and Western NY Chapter. Rochester, NY. 63 pp.
- U.S. Fish and Wildlife Service. 1994. Clubshell (*Pleurobema clava*) and Northern Riffleshell (*Epioblasma tondosa rangiana*) Recovery Plan. Hadley, Massachusetts. 68 pp.
- Vaughn, C. C. and Taylor, C. M. (1999), Impoundments and the Decline of Freshwater Mussels: a Case Study of an Extinction Gradient. *Conservation Biology*, 13: 912–920
- Wang, N., Mebane, C. A., Kunz, J. L., Ingersoll, C. G., Brumbaugh, W. G., Santore, R. C., ... and Arnold, W. (2011). Influence of dissolved organic carbon on toxicity of copper to a unionid mussel (*Villosa iris*) and a cladoceran (*Ceriodaphnia dubia*) in acute and chronic water exposures. *Environmental Toxicology and Chemistry*, 30(9), 2115-2125.
- Watters, G. T., Hoggarth, M. A., and Stansbery, D. H. 2009. *The freshwater mussels of Ohio*. Columbus: Ohio State University Press.
- Wildenberg, A. (2012, August). Mussel Community Response to Wastewater Effluent in a Midwestern River. In *AFS 142nd Annual Meeting*. AFS
- White, E.L., J.J. Schmid, T.G. Howard, M.D. Schlesinger, and A.L. Feldmann. 2011. New York State freshwater conservation blueprint project, phases I and II: Freshwater systems, species, and viability metrics. New York Natural Heritage Program, The Nature Conservancy. Albany, NY. 85 pp. plus appendix.
- Williams, J.D., M.L. Warren, K.S. Cummings, J.L. Harris and R.J. Neves. 1993. Conservation status of freshwater mussels of the United States and Canada. *Fisheries* 18(9):6-22 .

Yeager, B. (1993). Dams. Pages 57-92 in C.F. Bryan and D. A Rutherford, editors. Impacts on warm water streams: guidelines for evaluation. *American Fisheries Society*, Little Rock, Arkansas.

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